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Nutrición Hospitalaria, vol. 22, núm. 2, marzo-abril, 2007, pp. 244-251

Grupo Aula Médica
Madrid, España

Available in: http://www.redalyc.org/articulo.oa?id=309226715015
Recovery from experimental malnutrition with soymilk: immunological and genetic aspects

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Abstract

Experimental malnutrition models have been useful to study the effects of malnutrition at early ages. Substantial evidence exists that malnutrition in critical stages of development could result in chromosomal damages. The effect of nutritional rehabilitation with soymilk as a complement of a restricted diet, on plasma and muscle proteins, chromosomal integrity, and unspecific and mucosa immune responses, was studied.

Adult male and female Wistar rats (5 weeks old) were assigned to different nutritional conditions: a) 14 days on protein restricted diet (corn flour and water), followed by 14 days in which water was replaced by soymilk, as nutritional rehabilitation; b) the same conditions above but periods of 28 days of a protein restricted diet, and 28 days of nutritional rehabilitation and c) age-matched malnourished (protein restricted diet without nutritional rehabilitation) and normally nourished controls.

After both nutritional rehabilitation periods, the weights reached were significantly higher ($p < 0.001$) than the malnourished control values, but lower than the normal control ones. Plasma protein concentrations were similar in all groups. Muscle proteins that were diminished during the restricted diet, reached normal control values after both rehabilitation periods. The protein restricted diet, produced numeric and structural chromosomal abnormalities. Nutritional rehabilitation was only partially able to revert these abnormalities. The phagocytic activity and gut mucosa IgA-secreting cells were significantly reduced ($p < 0.001$) during the restricted diet; both nutritional rehabilitation periods induced a significant increase of both, phagocytic activity and IgA secreting cells. These values were similar to controls.

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Aceptado: 19-V-2006.
Our results show that the supplementation of a protein-restricted diet with soymilk improved tissue protein content, as well as unspecific and gut mucosa immune responses, even though it was not able to reinstate fully normal body weight and a normal chromosome karyotype.

Key words: Malnourish. Soymilk. Chromosome abnormalities. Mucosa immunity. IgA.

Background

Protein-energy malnutrition in children has been recognized as the most widespread nutritional problem around the world. This disorder produces biochemical alterations leading to poor growth. The growth retardation varies in accordance with the severity and duration of the nutritional deficiency.

Experimental malnutrition models have been useful to study the effects of malnutrition at early ages. Substantial evidence exists that malnutrition in critical stages of development could result in chromosomal damages. Effects of starvation and marginal malnutrition on rat lymphocytes have been evaluated by chromosomal analysis before and after rehabilitation; reported results indicated that young animals exposed to conditions like starvation or chronic malnutrition are prone to permanent damage of the genetic system. Cortés et al. reported that malnutrition during lactation in rearing rats is associated with significantly increased DNA damage in spleen, peripheral blood, and bone marrow cells, as well as in isolated lymphocytes or lymphoid cells from the same tissues. This damage could be due to the deficiency of several essential nutrients required for protein synthesis that are associated with DNA integrity, impaired DNA repair mechanisms, and/or to the unavailability of molecules necessary to protect cells against DNA oxidative damage.

On the other hand, the increased level of DNA damage in peripheral blood lymphocytes and leukocytes could be related to negative effects, such as a deficient immune response, resulting in severe consequences, like relapse of previous infectious diseases or emergence of new ones. In fact, malnutrition results in reduced a number and functions of T-cells, phagocytic cells, and secretory immunoglobulin A antibody response. In addition, levels of many complement components are reduced.

Protein malnutrition often coexists with deficiencies of specific micronutrients, such as zinc, iron, vitamin A, folate, and others, suggesting that the immune-depressing effects of protein deficiency may be the result of these associated micronutrient deficiencies.

An ideal food for the prevention and management of malnutrition should be of high nutritive value. Taking into account its nutritional profile and health effects, soy could be considered as an ideal food for the prevention and management of malnutrition. In this line, we performed this study in order to determine the effect of the nutritional rehabilitation with soymilk, as a complement of a restricted diet, for as long as the malnutrition had lasted, on plasma and muscle protein levels, chromosomal abnormalities, and the unspecific and mucosa immune responses.

Material and methods

Animals

Male and female Wistar rats, supplied by the School of Medicine Central Animal Facilities (Universidad Nacional de Tucumán, Argentina), were used. Animals aged 5 weeks and weighing 85 ± 5 g each, were housed in plastic cages, in a temperature-controlled room (21 ± 1°C) with a controlled 12-hour light-darkness cycle (light on at 7:00 AM). Liquids and food of the different diets were available ad libitum. All experimental procedures were done in accordance with the relevant directives of the European Union (86/609/EEC) and the rules and recommendations of the FESSCAL (Federacion de Sociedades Sudamericanas de la Ciencia de Animales de Laboratorio).

Experimental Design

Animals were subject to a protein-restricted diet (corn flour and water), for two different periods of time, after which water was replaced by soymilk (AdeS, Unilever, Argentina) for nutritional rehabilitation. The first group (d28) received a 14 day period of nutritional rehabilitation after a similar time (14 days) of a protein-restricted diet. For the second group (d56) the periods of malnutrition and nutritional rehabilitation lasted 28 days each. Normally nourished and malnourished (without nutritional rehabilitation) age matched animals were used as controls (fig. 1).

On the 14th, 28th and 56th day, 6 rats from each control and experimental group were weighed and then anesthetized with ethyllic ether and a sample of peripheral blood was taken. After that, the animals were killed by overexposure to ether. Then, the other samples such as peritoneal cells, quadriiceps muscles and small intestine were taken (fig. 1).
The commercial soymilk used in this study (AdesS, Unilever, Argentina) is elaborated from a non-transgenic variety of soy. Its components are: protein: 2.6 g%; carbohydrates: 4 g%; fat: 1.5 g%; fiber: 0.5 g%; minerals: Ca, 11 mg%; Fe, 0.5 mg%; P, 40 mg%; Mg, 16 mg%; Na, 80 mg%; K, 150 mg%; vitamins: A, 60 mg%; B6, 0.2 mg%; B9, 20 mg%; B12, 0.1 mg%; D, 0.5 mg%; Isoflavones, 6.4 mg% (daidzein: 2.4 mg% ; genistein: 4 mg%).

Normally nourished rats received a commercial laboratory food (Cargill, Buenos Aires, Argentina) as a standard. Its components are: protein: 24.6% of proteins from animal and vegetal origin.

Plasma and Muscle Protein concentration

Plasma protein concentration was determined by the Biuret method (Proti-2, Wiener Lab, Rosario, Argentina) using a control serum (Proti-2 Suero Patrón, Wiener Lab, Rosario, Argentina) as a standard.

In order to determine muscle protein, right hind limbs were removed and skinned. Quadriceps muscles were rapidly dissected, rinsed with saline solution to eliminate blood remnants, dried with tissue paper, and weighed. Tissues were homogenized in phosphate buffer pH 7 (w/v = 1/5). Then, 0.1 ml of Triton X-100 was added and mixed for 5 minutes. Finally, the homogenate was centrifuged at 20,000 rpm for 30 minutes. Proteins were determined in the supernatants by the Biuret method (Proti-2, Wiener Lab, Rosario, Argentina).

Results for both plasma and muscle protein concentrations are expressed in g%.

Cytogenetic analysis

In order to perform cytogenetic analysis, peripheral blood samples to which 1% Sodium Heparin, 5,000 U/ml (PL. Rivero y cia. SA, Industria Argentina) were obtained from the jugular vein and was added at the periods described above. Thirty drops of heparinized whole blood were added to each of two 15-ml polypropylene centrifuge tubes (Cellstar™, Greiner bioone, Germany) with 5 ml of RPMI 1640 culture medium (Gibco BRL, Grand Island, NY, USA) each, supplemented with 10% fetal calf serum (Gibco BRL, Grand Island, NY, USA), 2% glutamine (Gibco BRL, Grand Island, NY, USA), 1% phytohemagglutinin M-form (PHA-M) (Gibco BRL, Grand Island, NY, USA) and 0.1% gentamycin (Shering-Plough SA, Key Pharma S.A. Argentina) using a 21-gauge needle (Terumo, Japan). Cultures were incubated at 37 °C for 72 h. At harvest time, to each culture 2 drops of Colcemid (Gibco BRL, Grand Island, NY, USA), were added, using a 25-gauge needle (Terumo, Japan), incubated for 30 more minutes at 37 °C, and centrifuged for 8 minutes at 1,000 rpm. Then, the supernatant was removed, 4 ml of 0.075 M KCl were added to each pellet while kept in a 37°C water bath, and resuspension performed. Immediately, cell suspensions were centrifuged for 8 min at 1,000 rpm, supernatants were removed, pellets were resuspended into 4 ml of fresh 0.075 M, KCl each, 5 drops of 3:1 methanol-acetic acid mix added in order to begin fixing the cells, centrifuged immediately for 8 min at 1,000 rpm, and resuspended into fresh 3:1 methanol-acetic acid mix. Pelleting and resuspension were repeated two more times. For making slides, 3–6 drops of cell suspension were poured along the upper long edge of a cold, wet slide, held in parallel to the lab counter with its surface at a 45º angle; the back and the bottom of the slide were dried immediately with tissue paper. Phase contrast microscopy (CH Olympus, Japan) was used to make sure that chromosomes were well spread and metaphases devoid of cytoplasm; if not, drying time were adjusted accordingly. Slides were placed in the oven at 95 ºC for 20 min for aging. Aged slides were immersed in 10% trypsin solution in 0.9% NaCl (Gibco BRL, Grand Island, NY, USA), for 50 seconds, and then in two baths of 0.9% NaCl solution; immediately, each slide was stained for 2 minutes with a fresh mix of pHydrion buffer (Micro Essential Laboratory Inc. U.S.A.) (3 ml) and 0.25% Wright’s stain (1 ml), (Sigma Chemical Co., St. Louis, MO, USA).

In order to determine the mitotic index, the total number of metaphases was counted in 100 nuclei and expressed as percentage.

The chromosomal abnormalities were analyzed in 50 G-banded metaphases per animal. Then, they were photographed, and their chromosomes counted on black and white prints. 3-5 of these metaphases were selected and karyotyped. Karyotypes were carried out according to nomenclature rules for rat G-banded chromosomes.

Phagocytosis Test

The non-specific immune response was performed by an ex vivo phagocytosis test using peritoneal macrophages according to Perdigón et al., 1986. Peritoneal cells from rats of different groups and feeding periods were recovered. Macrophages were suspended at a concentration of 10⁶ cells/ml in 5 ml of RPMI-1640 medium (Gibco BRL, Grand Island, NY, USA) supplemented with 0.3% (v/v) sodium heparin (PL. Rivero y cia. S.A., Industria Argentina). Then, 0.2 ml of Candida albicans suspension at a concentration of 5 x 10⁶
10^6 microorganisms/ml opsonized with rat autologous serum was added to 0.2 ml of each macrophage suspension and incubated for 15 min at 37 °C. The reaction was stopped by icing. Immediately, smears were made, air dried, and stained with Giemsa (Biopack, Sistemas Analíticos SA, Buenos Aires, Argentina). The phagocytosis rate was measured by counting 100 cells from each of three smears per animal. The results were expressed as the mean ± SD of the percentage of macrophages that had one or more yeast cells phagocytized.

**Counting of IgA Secreting Cells by Immunofluorescence**

As indicated above, samples from the small intestine were removed. The intestinal fluids were washed out with 0.01 M phosphate buffered saline solution, pH = 7.2. Tissues were placed in cold ethanol (96%) for processes with the Saint-Marie’s technique. Once fixed, dehydrated and embedded in paraffin at 56 °C, they were cut in 4 mm serial sections, and used to perform the immunofluorescence test in order to determine the number of IgA producing cells.

Histological samples from the small intestine were incubated with 0.2 ml of goat anti-rat IgA (a chain specific, Bethyl Inc., Montgomery, Texas, USA) at 1/20 dilution for 30 min, in a wet chamber at room temperature. Then, they were washed three times with 0.01 M phosphate buffered saline, pH = 7.2. A 1/100 dilution of the second antibody, fluorescein-conjugated rabbit F(AB’)2 fragment to goat IgG, whole molecule (Cappel, ICN Pharmaceuticals Inc., Costa Mesa, California, USA), was added and incubated for 30 min at room temperature. Afterward, the slides were rinsed again three times with the same buffer, air-dried, and mounted with 1:9 glycerol-buffer, pH 9.

Three histological sections from each animal and for each period were analyzed. The number of IgA-positive cells within villi lamina propria was determined in 30 fields (magnification 40x). The results were expressed as the mean ± SD of positive cells in the lamina propria per 10 villi.

**Statistical Analysis**

All statistical calculation was carried out with Sigma Stat software package (Jandel Scientific, San Raphael, California, USA). Differences between diet groups were analyzed by one way analysis of variance (ANOVA), and Dunnet’s test was used as a post-hoc test. Differences were considered significant if p < 0.05.

**Results**

**Effect of Soymilk rehabilitation on body weight records**

Figure 2 shows weight evolution during the experiment periods across different diet groups. At the beginning of the study, the mean body weight was 85 ± 5 g. A normally nourished animal regularly increased in body weight while malnourished does not. Weights reached, after both periods of nutritional rehabilitation were significantly superior (p < 0.001) to the malnourished control values, but significantly diminished when compared to age-matched normal control ones (d28: p < 0.01, d56: p < 0.001).

**Effect of Soymilk rehabilitation on Plasma and Muscle Protein**

Plasma Protein concentration did not show significant differences between experimental and control groups at any data point (table I). However, muscle proteins tested were significantly decreased during the restricted diet (p < 0.01) in relation to the normal group. After both rehabilitation periods, muscle protein levels were similar to those showed by normal control groups. These results are detailed in figure 3.

**Effect of Soymilk rehabilitation on the mitotic index and on induced chromosomes abnormalities**

During the restricted diet, the mitotic index was significantly lower in malnourished rats when compared with age-matched normal controls (p < 0.005). Nutritional rehabilitation with soymilk was able to restore normal values when malnutrition had lasted 14 days.

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Table I

<table>
<thead>
<tr>
<th>Days</th>
<th>Normal control</th>
<th>Malnourished control</th>
<th>D28</th>
<th>D56</th>
</tr>
</thead>
<tbody>
<tr>
<td>14</td>
<td>4.9 ± 0.8</td>
<td>4.5 ± 0.6</td>
<td>4.5 ± 0.6</td>
<td></td>
</tr>
<tr>
<td>28</td>
<td>5.2 ± 0.6</td>
<td>4.0 ± 0.6</td>
<td>5.2 ± 0.4</td>
<td>4.0 ± 0.6</td>
</tr>
<tr>
<td>56</td>
<td>5.0 ± 0.4</td>
<td>4.4 ± 0.5</td>
<td>5.1 ± 0.7</td>
<td></td>
</tr>
</tbody>
</table>

Malnutrition did not significantly affect the plasma protein.
(d28), but not when it lasted 28 days (d56); in this case, the mitotic index of nutritionally rehabilitated animals was only 80-85% of that showed by normal control (table II).

After 14 days of the restricted diet, numeric chromosomes abnormalities consisting of hypodiploid metaphases with 38-39 chromosomes, nullisomies (chromosome pair 20 lost) and monosomies (e.g.: one chromosome lost from pair 17, 14, and 11). Other numeric abnormalities like trisomies (e.g. pair 17) were also observed (fig. 4).

After 28 days of protein malnutrition, besides the numerical abnormalities, we observed structural anomalies like robertsonian translocations [T(8;8)]. Unidentified marker chromosomes were also observed in 80% of the metaphases analyzed (fig. 5). A robertsonian translocation is originated by a centric fusion of the long arms, of the acrocentric chromosomes, usually with simultaneous loss of both short arms15. A marker chromosome (mar) is a structurally abnormal chromosome in which no part can be identified by routine banding. In the description of karyotypes the presence of a mar must be preceded by a plus sign (+). When several different markers are clonally present, they may be indicated by an Arabic number after the symbol mar, e.g. mar 1, mar 2, etc. It must be stressed that this does not mean derivation of the

<table>
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<th>Normal control</th>
<th>Malnourished control</th>
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<th>D56</th>
</tr>
</thead>
<tbody>
<tr>
<td>14</td>
<td>7.3 ± 2</td>
<td>3.0 ± 1*</td>
<td>3.0 ± 1</td>
<td></td>
</tr>
<tr>
<td>28</td>
<td>7.0 ± 2</td>
<td>1.5 ± 2*</td>
<td>6.8 ± 1</td>
<td>1.5 ± 2</td>
</tr>
<tr>
<td>56</td>
<td>7.2 ± 1</td>
<td>1.5 ± 1*</td>
<td>5.8 ± 1</td>
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</tr>
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</table>

Malnutrition significantly decreased the mitotic index. Nutritional rehabilitation with soymilk fully reverted this effect at D28 but only partially at D56. *=p<0.01

Effect of Soymilk on the Phagocytic Activity

Figure 7 shows the phagocytic activity of rat peritoneal macrophages in the different experimental
groups. It can be seen that phagocytosis was significantly reduced ($p < 0.001$) in the restricted diet groups compared with the normal control. After both periods of soy milk rehabilitation, phagocytosis percentages incremented to values similar to those displayed by age-matched normal controls.

**Effect of a Soymilk Complement on the Number of IgA-secreting Cells**

This assay was performed in order to determine the effect of the rehabilitation with soymilk as a complement of the restricted diet on the gut mucosa immunity.

In this case, results were similar to the phagocytic assays, i.e. numbers of IgA-positive cells were significantly reduced ($p < 0.001$) among malnourished animals when compared to normal control ones. The soymilk administrated during both rehabilitation periods produced a remarkable increment in the number of IgA-producing cells in the lamina propria of villi of intestinal mucosa, reaching values similar to the normal ones (fig. 8).

**Discussion**

Malnutrition is multifactorial; however, the immediate cause of this condition is inadequate intake and poor utilization of nutrients due to inadequate food supply or poor quality foods$^{18}$. There is considerable evidence that malnutrition has effects on physical growth, morbidity, mortality, cognitive development, reproduction, physical work capacity and risks for several adult onset chronic diseases$^{19,20}$.

Soymilk, as well as a combination of soybeans with maize, has been found to be valuable in the management of malnutrition$^{21}$.

In our study we used a rat model of chronic malnutrition, in which corn flour was the only solid food intake. This diet initially produced some weight loss, but its most remarkable effect in this regard was the absolute failure to gain weight (fig. 2). This result is similar to that reported by Qureshi et al$^{22}$. Rehabilitation of malnourished animals by complementing the restricted diet with soymilk, allowed a significant but incomplete weight recovery (fig. 2).

Moreover, malnutrition produced a significant decrease of muscle but not plasma protein (fig 3 and table I). Similar findings have been reported in other studies that showed that there was a marked reduction in the rate of muscle proteins synthesis in rats fed a low protein diet$^{23}$, and of proteins in muscle and other organs and tissues in rats fed a protein-free diet$^{24}$. It was postulated that chronic protein deficiency leads to an overall reduction of protein synthesis that does not include certain plasma proteins. In others words, the protein metabolic response to the nutritional stress could be a redistribution of amino acids from the peripheral tissues to the liver for the synthesis of proteins that have a rapid turn over and are critical for survival$^{25}$.

The nutritional quality of a protein food depends on content, digestion, absorption, and utilization of amino acids. Availability of amino acids varies with protein source, food processing treatment, and interaction...
with other components of the diet. Thus in the case of the diets used in our experiments it is important to note that corn proteins are of poor quality because of their low tryptophan and lysine content29. On the other hand, there are data showing that well-processed soy-protein isolates and soy-protein concentrates can serve as the major, or even sole, source of protein intake and that their protein value is essentially equivalent to that of food proteins of animal origin27. The soy milk we used (ADES) had all the essential amino acids. In this way it could be explained how nutritional rehabilitation with soy milk as a complement of corn flour, induced a tissue protein recovery to almost normal values (fig. 3).

Some authors3,28 have reported that malnutrition provokes chromosome anomalies, while others postulate the opposite3,29. In the present work, we detected abnormalities of the cell chromosome number after two weeks of malnutrition (fig. 4). Our data show that the longer period of the protein restricted diet, the more affected is the integrity of the chromosomes (fig. 5). Once chromosomal damage has occurred, nutritional rehabilitation with soymilk consistently has a limited effect in reverting it (fig. 6). This limitation is more noticeable when the length of the malnutrition period increases in spite of the fact that the rehabilitation period also increased. In this regard, our results agree with Alu and Murthy3. The lack of chromosomal abnormalities reported by others29,30 may be due to differences in the restricted diet and the techniques used for detecting them.

We have also observed that the restricted diet provokes a significant decrease in the lymphocyte mitotic index, which was reverted to a considerable degree when we complemented the restricted diet with soy-milk (table II). This agrees with papers using other cell systems in experimental animals31,32. We think that this effect could be due mainly to the deficiency of essential amino acids and micronutrients that are necessary for the protein synthesis of mitotic spindles.

It is a well known fact that protein malnutrition induces a decrease in phagocytic activity of peritoneal macrophages33. Our results corroborate that the phagocytic function of macrophages is susceptible to protein malnutrition (fig. 7). It has been suggested that functional alterations of macrophages may be associated with micronutrient deficiencies, since protein restriction often coexists with them34. IgA-secreting cells play an important role in the local immunological defense against pathogens and damage resulting from absorption of foreign antigens35. Also, dietary protein, minerals and vitamins are important in the mucosal IgA response of the small intestine36. Results reported here confirm this, since a significative decrease of IgA producing cells in the gut lamina propria was observed during the experimental malnutrition (fig. 8).

Early nutritional rehabilitation is needed to improve survival of malnourished individuals and to support immune responses37. Our results show that the rehabilitation with soymilk-complemented corn flour increased notably not only the peritoneal macrophage activity, but the mucosal immune function as well by stimulating the IgA-secreting cells (fig. 7 and 8). These data would suggest that high quality soy protein38 and/or micronutrients present in the soymilk administered like A, B2, B12, B6, vitamins and iron, may enhance or preserve immune function. The latter is supported by reports that the use of micronutrient supplements, singly or in combination, stimulates immune response and thus influences the susceptibility of a host to infectious diseases39.

The mechanisms by which the soymilk complement may exert immunomodulator effects have to be investigated. We hypothesize that the nutritive components of the soymilk could induce, within the lamina propria, T cells to produce Th1 IgA stimulating cytokines, which could modulate IgA production by B cells.

On the other hand, soy oligosaccharides (raffinose and stachyose) could have a prebiotic effect. Prebiotics are indigestible food ingredients that can beneficially affect the host by selectively stimulating the growth and/or activity of certain endogenous microbial populations such as bifidobacteria and lactobacilli40. It has been reported that bifidobacteria enhances immune responses by increasing antibody production and proliferation of B cells in Peyer’s patches41.

In our experimental model, the supplementation of a restricted diet with soymilk, for a period of time identical to that of the previous malnutrition, normalized tissue protein concentration, and appears to do the same with unspecific and gut mucosa immune responses; however the diet does not permit the recovery of fully normal body weight and the elimination of chromosomal abnormalities. We consider that a longer rehabilitation time would be necessary for complete recovery.

Acknowledgements

This work was carried out with the facilities of the School of Medicine at the National University of Tucumán, Argentina.

The authors thank UNILEVER (Argentina) for free supplying the AdeS soymilk used in this work. We also grateful thank Patricia Black-Decima, PhD for their helpful discussions during the preparation of the manuscript.

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