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Balanced cytokine-producing pattern in mice immunized with an avirulent Trypanosoma cruzi

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ABSTRACT

We have previously demonstrated that inoculation of BALB/c mice with trypomastigotes of CL-14, an avirulent Trypanosoma cruzi clone, prevents the development of parasitemia and mortality after challenge with virulent CL strain. In this report, we investigated the cytokine and antibody profiles induced by inoculation with CL-14 clone. Groups of mice were inoculated with trypomastigotes of CL-14 clone and challenged with infective CL strain. Challenged CL-14-inoculated mice had lower levels of IFN-γ and higher production of IgG1 antibodies as compared to CL strain-infected mice. Previous inoculation with CL-14 clone partially prevented the suppression of IL-2 production caused by CL strain infection. No significant differences were found regarding IL-4 production by splenocytes from CL-14-inoculated or control groups after challenge with CL-strain. Our results show that protection against acute T. cruzi infection induced by CL-14 inoculation correlates with a balanced T1/T2 cytokine production, a profile likely to be beneficial for the host.

Key words: Trypanosoma cruzi, protection, cytokines, humoral response.

INTRODUCTION

Cytokines are soluble mediators which regulate and act as effector mechanisms of immunity. They are critical factors in driving the establishment of protective or exacerbating responses against a variety of pathogens. In an attempt to identify the profile of a protective response against Trypanosoma cruzi, the etiologic agent of Chagas’ disease, several studies have been carried on the production of cytokines by different mouse strains upon infection (DosReis 1997). While the presence of IFN-γ and IL-4 in the supernatants from lymphoid cell cultures was associated with resistance and susceptibility, respectively (Hoft et al. 1993), in situ analysis showed no such correlation (Zhang and Tarleton 1996). Moreover, both susceptible and resistant mice produced similar levels of IFN-γ (Zhang and Tarleton 1996), suggesting the involvement of other factors in the control of infection.

Despite the lack of association between T1/T2 phenotypes and resistance of some particular mouse strains against T. cruzi infection, IFN-γ is known to participate in the control of the acute phase parasitemia. Administration of exogenous IFN-γ increases resistance (Reed 1988), while treatment with anti-IFN-γ antibodies renders the mice more sensitive to infection (Torrico et al. 1991). In fact, IFN-γ activates macrophages to produce nitric ox-
ide and destroy T. cruzi in vitro (Gazzinelli et al. 1992), whereas the lack of either IFN-γ receptor or iNOS expression renders mice highly susceptible to T. cruzi infection (Hölscher et al. 1998). The polarization towards a Th1 profile has also been described in mice protected against T. cruzi by immunization with T. cruzi proteins (Miller et al. 1996, Rodrigues et al. 1999, Schnapp et al. 2002). In order to understand the mechanisms of protective immunity against T. cruzi, we studied a model of inoculation with trypomastigotes of the CL-14 avirulent clone, derived from the CL strain of T. cruzi (Lima et al. 1991). Mice inoculated with live trypomastigotes of the CL-14 clone show complete absence of parasitemia or pathology, and develop an efficient immunity against a lethal challenge with virulent parasites (Lima et al. 1991, 1995). Here we studied the cytokine profile and the production of anti-T. cruzi antibodies that help to keep CL-14-inoculated mice protected against virulent T. cruzi.

**MATERIALS AND METHODS**

BALB/c mice (6-8 weeks of age) were used in all experiments. CL-14 is a T. cruzi clone isolated from the CL strain. CL-14 trypomastigotes were obtained by in vitro metacyclogenesis and purified by DEAE-chromatography, as described elsewhere (Lima et al. 1991). Inoculation with clone CL-14 was performed by injecting 10^7 live purified trypomastigotes i.p. in 0.2 ml of phosphate-buffered saline (PBS). The original CL strain was maintained by serial blood passages through BALB/c mice. Infections were performed by intraperitoneal (i.p.) inoculation of contaminated blood containing 10^4 trypomastigotes.

Splenocytes from mice sacrificed at different time points (see figure legends) were plated in 24 well plates (5 x 10^5/well) in 1 ml of RPMI supplemented with 0.5% normal mouse serum, 2 mM L-glutamine, and 5 x 10^-5 M 2-mercaptoethanol (Life Technologies, GIBCO-BRL, Gaithersburg, MD). Cultures were stimulated with 1 μg of Concanavalin A (Con A; Sigma, St. Louis, MO) for 24 or 48 hours at 37°C and 5% CO2. Cell-free supernatants were then collected and stored at –20°C for cytokine detection by ELISA technique, using capture and detection antibodies purchased from PharMingen (San Diego, CA), according to the manufacturer’s instructions. Samples of plasma were tested for IFN-γ production, whereas supernatants were tested for IL-2, IL-4 and IFN-γ production. After incubation with streptavidin-peroxidase conjugate (Sigma), the assay was developed with OPD substrate (o-phenylenediamine-dihydrochloride, Sigma) and stopped with HCL 3 N. The plate was then read at 570 nm in a Benchmark microplate reader Bio-Rad (Richmond, CA) Benchmark. Curve regression was performed with the help of Microplate Manager software (Bio Rad).

Anti-T.cruzi isotype production was evaluated by ELISA assays. Briefly, ELISA plates were coated with T. cruzi antigen (prepared by freeze-and-thawing CL strain epimastigote suspensions) overnight at 4°C. After washing plates, 50μl of diluted plasma samples (1/20-1/12500, duplicate) were incubated for 8 h at 4°C. Plates were then washed with PBS-0.05% Tween 20 and incubated with isotype-specific anti-mouse IgG1 and IgG2a conjugated to alkaline phosphatase (Southern Biotechnology Associates, Birmingham, AL) to reveal bound antibodies. After washing, assays were developed with p-NPP (Zymed, San Francisco, CA) and read under a 410 nm filter in a microplate reader.

All experimental procedures were conducted in accordance with guidelines for care and use of laboratory animals (CAUAP) of the Institute of Biophysics Carlos Chagas Filho, which conform to the National Institutes of Health (Bethesda, MD, USA) guidelines.

**RESULTS AND DISCUSSION**

To evaluate the production of cytokines by the protective response induced by CL-14 trypomastigotes, groups of mice were inoculated with the CL-14 clone and challenged with trypomastigotes of the CL strain six weeks later. Serum levels of IFN-γ and cytokine production in response to stimulation of splenocytes in vitro with Con A were evaluated two weeks after the infective challenge.
At this timepoint, no IFN-γ could be detected in sera from CL-14-inoculated controls (Figure 1A). Challenged-CL-14 inoculated mice had detectable, but lower serum levels of IFN-γ than non-immunized mice infected with the CL strain. Production of IFN-γ after stimulation of splenocytes in vitro with Con A was similar to that described in vivo (Figure 1B). Despite the lower levels of IFN-γ in CL-14-inoculated mice after challenge with CL strain, we cannot rule out a role of this cytokine in protection induced by CL-14 clone. T. cruzi infection induces the production of high levels of IFN-γ during the acute phase of the disease (Hoft et al. 1993, Zhang and Tarleton 1996, our own data), possibly as a result of intense macrophage stimulation by the parasite to produce IL-12 and IL-18, two potent IFN-γ inducing factors (Meyer zum Büschenfelde et al. 1997, Camargo et al. 1997). Since parasitemia is absent after the challenge of CL-14-inoculated mice, it is possible that serum IFN-γ levels are relatively low upon challenge as a result of parasitemia control. The production of IL-4 by stimulated splenocytes varied among experiments, but in general they were similar in the groups challenged-immunized and CL-infected (Figure 1D).

Immunosuppression is a hallmark of acute T. cruzi infection (DosReis 1997). Inoculation of clone CL-14 induces a moderate and transient decrease in the production of IL-2 stimulated by mitogens, while infection with the CL strain induces a longer and more severe decrease in this response (data not shown). Prior inoculation with clone CL-14 partially prevented the suppression of IL-2 production induced by infection with the CL strain (Figure 1C). These results parallel the production of the polyclonal lymphocyte activation induced by T. cruzi in CL-14-immunized mice (Paiva et al. 1999). It is possible that the presence of circulating antibodies in CL-14-immunized mice contribute to quickly control the antigenic burden, thereby preventing the induction of such immune abnormalities.

IFN-γ induces IgG2a secretion by activated lymphocytes, whereas IL-4 induces a switch to IgG1 production in various models (Finkelman et al. 1989). High levels of IFN-γ are produced during acute infection with T. cruzi. Thus, we compared the serum levels of anti-T. cruzi IgG1 and IgG2a between normal or CL-14-inoculated mice two weeks after infective challenge with the CL strain. An increase in the serum titers of IgG1 antibodies to T. cruzi was observed after challenge of CL-14 inoculated mice with the CL strain (Figure 2A). These titers were far higher in mice previously inoculated with CL-14 clone than in CL-infected control mice. In contrast, the titers of T. cruzi-specific IgG2a antibodies were not significantly altered in CL-14 inoculated mice two weeks after challenge with CL strain (Figure 2B). The increased IgG1 production by CL-14-inoculated mice compared to CL infected mice shown here and in a previous report (Pyrrho et al. 1998) may be the result of a more balanced IFN-γ/IL-4 production elicited by CL-14 inoculation.

The role of different IgG subclasses in protection against T. cruzi infection has varied in studies using different models of immunization. In a model of protection induced by DNA immunization, the predominant isotype induced was IgG2a (Planelles et al. 2001), whereas, in another study, the predominant isotype found was IgG1 (Costa et al. 1998). In our model, IgG1 is also the prevalent isotype stimulated by CL-14 inoculation (Pyrrho et al., 1998) and increases considerably after challenge with CL strain. In spite of that, we cannot rule out that both IgG subclasses participate in the protection induced by this avirulent clone as various studies have shown protective roles of IgG1 and IgG2 antibodies against T. cruzi. (Brodsky et al. 1989, Cerban et al. 1992).

This study shows that the efficient immunity induced by the T. cruzi clone CL-14 follows a T1/T2 and immunoglobulin isotype pattern different from other models of immunization described in the literature (Miller et al. 1996, Rodrigues et al. 1999). The finding of polarized T1 responses in other models of immunization against T. cruzi suggested an important role for IFN-γ in protection. Despite its protective role in the control of the acute phase parasitemia, IFN-γ-producing T-cell responses upon T. cruzi infection have also been associated with sever-
Fig. 1 – Cytokine production in mice inoculated with clone CL-14 and challenged with CL-strain trypomastigotes. Groups of BALB/c mice (non-inoculated controls or mice inoculated 6 weeks before with $10^7$ purified CL-14 trypomastigotes) were challenged with $10^4$ CL-strain trypomastigotes. Three mice from each group (CL-14 clone or CL strain) were sacrificed two weeks after challenge. Pools of plasma were assayed by ELISA to determine the IFN-$\gamma$ levels (A). Splenocytes were cultured in 24 well plates and stimulated with Con A ($1 \mu$g/ml), as described in Material and Methods. Cell-free supernatants were collected after 24hs of incubation and assayed to determine the IFN-$\gamma$ (B), IL-2 (C) and IL-4 (D) levels by ELISA. The data represent the ratio between cytokine levels of splenocyte cultures from *T. cruzi* inoculated mice ($10^7$ CL-14 clone: dashed bars; CL strain: open bars; CL strain after inoculation with CL-14 clone: black bars) and the levels of splenocyte cultures from normal mice, upon stimulation with Con A. Figure shows the results of one representative experiment of three performed.

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**RESUMO**

Inoculação de camundongos BALB/c com triatomina- 
gostas do clone avirulento de *Trypanosoma cruzi* CL-14 pro-
tega contra o desenvolvimento de parasitemia e mortali-

...
Fig. 2 – Production of IgG1 and IgG2a by mice inoculated with CL-14 clone and challenged with CL strain trypomastigotes. BALB/c mice were inoculated with 10^7 purified CL-14 trypomastigotes. Groups of normal littermates and CL-14-inoculated mice were challenged with 10^4 CL strain trypomastigotes 6 weeks later. Three mice from each group (CL or CL-14+CL) and from a group that received only the inoculation of 10^7 trypomastigotes of CL-14 clone (CL-14) were bled two weeks after challenge. Pools of plasma were assayed by ELISA to determine the T. cruzi-specific IgG1 (A) and IgG2a (B) titers, as described in Materials and Methods. Similar results were obtained in two independent experiments.

**Cytokine Profile Induced by an Avirulent T. cruzi**

After challenge with the virulent CL strain, we investigated the cytokine profile induced by inoculation with the CL-14 clone. BALB/c mice were inoculated with 10^7 CL-14 trypomastigotes and challenged with 10^4 CL strain trypomastigotes 6 weeks later. Three mice from each group (CL or CL-14+CL) and from a group that received only the inoculation of 10^7 trypomastigotes of CL-14 clone (CL-14) were bled two weeks after challenge. Pools of plasma were assayed by ELISA to determine the T. cruzi-specific IgG1 and IgG2a titers, as described in Materials and Methods. Similar results were obtained in two independent experiments.

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