

Revista Facultad Nacional de Agronomía Medellín

ISSN: 0304-2847 ISSN: 2248-7026

Facultad de Ciencias Agrarias - Universidad Nacional de

Colombia

Criollo-Escobar, Hernando; Muñoz-Belalcázar, Johanna Gamma ray irradiation (Co 60) of lulo with and without thorns calluses and seedlings (*Solanum quitoense* Lam.) produced *in vitro* Revista Facultad Nacional de Agronomía Medellín, vol. 73, no. 3, 2020, September-December, pp. 9263-9271 Facultad de Ciencias Agrarias - Universidad Nacional de Colombia

DOI: https://doi.org/10.15446/rfnam.v73n3.82362

Available in: https://www.redalyc.org/articulo.oa?id=179964927004



Complete issue

More information about this article

Journal's webpage in redalyc.org



Scientific Information System Redalyc

Network of Scientific Journals from Latin America and the Caribbean, Spain and Portugal

Project academic non-profit, developed under the open access initiative

Revista Facultad Nacional de**Agronomía**

Gamma ray irradiation (Co⁶⁰) of lulo with and without thorns calluses and seedlings (*Solanum quitoense* Lam.) produced *in vitro*



Irradiación con rayos gamma (Co⁶⁰) de callos y plántulas de lulo con y sin espinas (*Solanum quitoense* Lam.) producidas *in vitro*

doi: 10.15446/rfnam.v73n3.82362

Hernando Criollo-Escobar^{1*} and Johanna Muñoz-Belalcázar¹

ABSTRACT

Keywords:

Genetic variation Mutagenesis Naranjilla RAM Survival The cultivation of lulo de Castilla (*Solanum quitoense* Lam.) in Colombia is subject to a series of sanitary problems, which has forced many producers to abandon the crop as a result of the total loss of plantations or to transfer the crop to new areas. It is necessary to implement breeding programs in order to produce varieties that are tolerant to the limiting problems. Since these programs require broad genetic variability in the progenitors, the present study aimed to evaluate the possibility of inducing *in vitro* variability in explants subjected to different doses of gamma radiation using a Co⁶⁰ source. The evaluated radiation doses were 0 Gy, 15 Gy, 30 Gy, 45 Gy, and 60 Gy in calluses induced with cotyledonary leaves and in seedlings from *in vitro* cultures of lulo with and without thorns. The survival and regeneration potential were also evaluated. The calluses were the explants that showed the highest survival, and the lulo seedlings without thorns were the most radiosensitive with a mortality of 100% at a dose of 30 Gy. The lulo seedlings with thorns had 100% mortality at a dose of 45 Gy. The irradiated lulo seedlings with thorns had a greater regeneration capacity than the lulo without thorns, with 1.52 seedling per explant and 1.12 seedling per explant, respectively, and the RAM markers showed genetic variability in all the irradiation treatments.

RESUMEN

Palabras clave:

Variación genética Mutagénesis Naranjilla RAM Supervivencia El cultivo de lulo de Castilla (Solanum quitoense Lam.) en Colombia está sometido a una serie de problemas principalmente de índole sanitario, que han obligado a muchos productores al abandono del cultivo por la pérdida total de las plantaciones o al traslado del cultivo a zonas nuevas, por lo cual se hace necesario la implementación de programas de mejoramiento para la búsqueda de variedades tolerantes a los problemas limitantes. Teniendo en cuenta que estos programas requieren una amplia variabilidad genética de sus progenitores, el presente estudio se orientó a evaluar la posibilidad de inducir variabilidad in vitro, de explantes sometidos a diferentes dosis de radiaciones gamma utilizando una fuente de Co⁶⁰. Se evaluaron dosis de radiación con 0 Gy, 15 Gy, 30 Gy, 45 Gy y 60 Gy en callos inducidos a partir de hojas cotiledonares y en plántulas provenientes de cultivos in vitro de lulo con y sin espinas. La supervivencia y el potencial de regeneración también fueron evaluados. Los callos fueron los explantes que presentaron mayor supervivencia y las plántulas de lulo sin espinas las más radiosensitivas con una mortalidad del 100% a una dosis de 30 Gy; las plántulas de lulo con espinas tuvieron una mortalidad del 100% con una dosis de 45 Gy. Las plántulas irradiadas de lulo con espinas tuvieron mayor capacidad de regeneración que las de lulo sin espinas con 1,52 plántulas por explante y 1,12 plántulas por explante, respectivamente; además, los marcadores RAM mostraron variabilidad genética con todos los tratamientos de irradiación estudiados.



¹ Facultad de Ciencias Agrícolas. Universidad de Nariño. Calle 18 No. 50-02. CP 520002, Pasto, Nariño, Colombia.

^{*} Corresponding author: <hcriolloescobar@gmail.com>

he cultivation of lulo in Colombia is of great importance, and the international potential of this fruit is promising. The increase in the harvested area went from 7,039 to 8,655 ha between 2010 and 2016. In production, it rose from 59,091 to 78,610 t in the same period, demonstrating the interest of producers and the economic advantages that this fruit offers (Agronet, 2018). In addition, the recent free trade agreements (FTAs) signed with several countries around the world opened new markets for Colombian agriculture, which requires strong technological boosts in crops to make Colombia highly competitive, as seen in the case of the lulo.

Lulo producers in Colombia face the susceptibility of cultivars to pests and diseases, farm heterogeneity, and reduction of vegetative and reproductive development. Agronomic problems that limit productivity and, in some cases, cause total crop loss (Lobo, 2006; Cruz *et al.*, 2007; Muñoz, 2013). Despite the losses caused by these problems, the cultivation area and production increased by 7.58% and 3.34%, respectively, in Nariño (Colombia) between 2010 and 2016 (Agronet, 2018). By searching suitable culturable areas, the low technological level of lulo farming systems has caused deforestation, making it necessary to rethink current cultivation techniques in order to avoid environmental problems (Muñoz, 2013).

These drawbacks mean that breeding programs aimed at increasing productivity are needed. It pursues lulo varieties tolerant to the major diseases and greater efficiency in transforming light into chemical energy, along with better distribution of carbohydrates (Angulo, 2006).

In Colombia, minimal efforts in lulo breeding have been geared toward obtaining cultivars resistant or tolerant to pathogens and pest, such as *Fusarium* sp., *Phytophthora infestans*, *Neolucinodes elegantalis*, and *Meloidogyne* sp. It has been necessary to resort to related species and obtain hybrids with resistance, but with lower quality, since all lulo varieties show susceptibility (Betancourth *et al.*, 2005; Muñoz, 2013). Despite its low domestication, the lulo has stability in specific niches and restrict general adaptability (Lobo, 2006).

Genetic variability is essential in plant breeding programs, it can be a definitive solution in crop breeding, especially in crops with limited variability (Subramanian, 2011; Rimieri,

2017). The induction of mutations is a widely used method to produce genetic variability, but it is necessary to evaluate the effectiveness and efficiency of each technique for the frequency of desirable mutants before considering one a potent tool for the improvement of plants (Prina *et al.*, 2010).

Artificial mutagenic agents can be physical or chemical. Treatments with ionizing radiation produce high degrees of chromosomal damage with breaks in the DNA chain and physiological effects that cause cell death and sterility in first-generation plants. Currently, the most commonly used radiation for plant breeding purposes includes x-rays and gamma rays (Sonone et al., 2010; Prina et al., 2010; Velmurugan et al., 2010; Silvera, 2017). However, the mutation rate can also be increased by in vitro cultures (somaclonal variation). It is possible to select plants by obtaining agronomically useful mutants and combine mutation techniques with in vitro propagation (Patade et al., 2008).

The improvement of the tolerance to water stress conditions of many crops has been achieved with in vitro selection of mutants obtained with radiation or chemical methods. These procedures have been tested on multiple crops, such as potatoes (Velmurugan et al., 2010), peanuts (Sonone et al., 2010), soybeans (Satpute and Fultambkar, 2012; Kavithamani et al., 2010; Ortiz et al., 2008), rice (Baloch et al., 2004), sorghum (Javaramachandran et al., 2010), cherries (Roman et al., 2009), sunflower (Kumar and Ratnam, 2010), sugar cane (Singh, 1993; Desai, 2006). among others. The improvement of crops with genetic mutations has also been successful in obtaining new colors and shapes in ornamental plants, and many mutants are induced by gamma radiation and other mutagens that are currently marketed (Datta et al., 2005; Matsumura et al., 2010; Sahariya et al., 2017)

Mutagenic induction with chemicals and radiation has been successfully used to obtain desirable agronomic characteristics. The 89% of mutant varieties have been obtained with irradiation methods, from which 64% were obtained using gamma radiation (Patade, 2008).

According to Predieri and Zimmerman (2001) and Mostafa *et al.* (2015), the effectiveness and efficiency of irradiation depend on the dose used, the genotype, and the type of irradiated explant. Therefore, the present study aimed to

carry out an *in vitro* evaluation of the effect of Co⁶⁰ gamma ray irradiation on the somaclonal variation of Iulo (*Solanum quitoense* Lam) with and without thorns.

MATERIALS AND METHODS

This study was carried out in the Tissue Laboratory of the Biology program of the Faculty of Sciences at the Universidad Nacional de Colombia (Medellín Headquarters); the gamma radiation was carried out in the nuclear reactor of the Instituto Colombiano de Geología y Minería (INGEOMINAS), using a Co⁶⁰ source.

The plants of lulo with and without thorns were obtained with three subcultures in an MS medium and calluses produced with foliar segments of lulo with thorns in an MS medium supplemented with ANA (6.0 mg L⁻¹) and sucrose at 9%, which were incubated in darkness for 45 days, and then were irradiated.

The plants and calluses were cultivated in Petri dishes $(\emptyset=9 \text{ cm})$ in an MS medium before being irradiated in a pilot irradiation plant, with an activity of 10000 curies and a cobalt60 source; the irradiation doses

were 0 Gy (Control), 15 Gy, 30 Gy, 45 Gy, and 60 Gy. Immediately after the treatment, the plants and calluses were transplanted to containers with a fresh MS culture medium. A completely randomized design, with a bifactorial arrangement, was used to analyze the irradiated plants' prolificacy and survival. The factors corresponded to the two varieties of lulo (Factor A) and the five doses of irradiation (Factor B). Each irradiated treatment (plants and callus) consisted of eight repetitions (Petri dishes), with four explants each.

To analyze the effect of the irradiation on the genetic fidelity, foliar samples were taken from the live explants after six months for DNA extraction following the methodology of Dellaporta *et al.* (1983) at a concentration of 10 ng μ L⁻¹, based on dilutions of phage lambda DNA (Sigma, St. Louis, MO, USA).

In the PCR, eight primers were tested, and seven were analyzed, considering that the CGA primer showed no amplification. The primers used in the RAM technique to determine the effect of the gamma irradiation on the lulo plants and calluses are presented in Table 1.

Table 1. Primers and sequences used in the RAM technique to determine the genetic stability of the lulo seedlings and calluses irradiated with gamma rays.

Primer	Sequence 5' - 3'		
СТ	DBDCTCTCTCTCTCTC		
CGA	DHBCGACGACGACGA		
CA	DBDACACACACACACACA		
AG	HBHAGAGAGAGAGAGAG		
TG	HVHTGTGTGTGTGTGTGT		
CCA	DDBCCACCACCA		
GT	VHV GTG TGT GTG TG		
ACA	BDB ACA ACA ACA ACA		

Designations used for degenerated sites: H (A or T or C); B (G or T or C); V (G or A or C) and D (G or A or T).

The hybridization and amplification conditions were established according to the selected primer (Table 2). The products were separated and visualized in 7% polyacrylamide gels (37:1 acrylamide-bisacrylamide) ran at 160 V for 1 h and 10 min, and stained with ethidium bromide and silver salts as described by Sambrook *et al.* (1989).

The observation of the gels provided a binary matrix for presence (1) and absence (0). Based on this matrix, descriptive analyses were carried out, and the genetic distances and similarities were established. The unbiased minimum distance criterion (Nei, 1978) was used for the classification analysis following the UPGMA method.

Table 2. Hybridization and amplification conditions for the RAM technique with the selected primers.

Cycle	Temperature (°C)				Time		
	Primer					Stage	
	AGCAACA	TGCTCCA	GTCGA	AGCAACA	TGCTCCA	GTCGA	-
1		95			5 min		Initial denaturation
2		95			30 s		Denaturation
3	50	55	58		45 s		Hybridization
4		72			2 min		Amplification
5	37 cycles from step two						
6		72			7 min		Final amplification
7		16			5 min		End

These analyses were carried out using TFPGA (Tools for Population Genetic Analysis), as described by Miller (1997).

RESULTS AND DISCUSSION Survival

The analysis of variance for the percentage of live explants two months after the irradiation showed highly significant differences between the lulo varieties, between the irradiation doses and for the variety×dose

interaction (Table 3). At the variety level, the lulo with thorns presented greater survival at the higher radiation doses, requiring an approximate dose of 28 Gy to cause 50% mortality. The lulo variety without thorns proved to be more susceptible to irradiation, with the same level of mortality at a radiation dose of 19 Gy (Figure 1). In general, the plants submitted to high doses presented foliar chlorosis, necrosis of the basal regions of the petioles, leaf fall, growth arrest, and death.

Table 3. Analysis of variance for the survival and regeneration capacity variables of lulo seedlings subjected to different doses of radiation with gamma rays.

Source	Sum of Squares					
	Su	ırvival	Regeneration			
	DF	SS	DF	SS		
Variety	1	3781.25 **	1	1.95 **		
Dose	4	121140.62 **	2	14.92 **		
Variety×Dose	4	5828.12 **	2	0.84 *		
Error	70		42			

^{**}Significant differences at 99% probability

The reaction among the lulo varieties has been observed in other species, such as soybeans (Satpute and Fultambkar, 2012), mung bean (Singh, 2007), rice (Baloch, *et al.*, 2004), beans (Rocha *et al.*, 2010) and pears (Predieri and Zimmerman, 2001). These differences can be attributed to the differential sensitivity of plants to radiation, because their adaptation to light (WT) and understory (WOT) conditions, may affect

physiological properties at the cellular and nuclear level (Baloch *et al.*, 2004; Rocha *et al.*, 2010); an examination of the leaf histology indicates that the two varieties present very similar characteristics (Medina *et al.*, 2008)

Multiplication capacity

The regeneration capacity of the plants from the initial explants after irradiation presented statistical differences

^{*} Significant differences at 95% probability

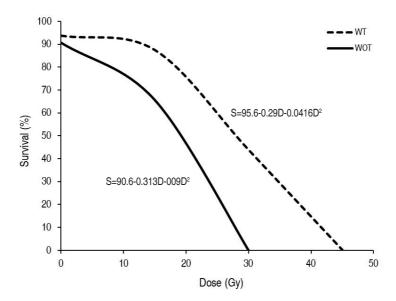


Figure 1. Survival percentage of lulo seedlings (*Solanum quitoense* Lam.) with thorns (WT) and without thorns (WOT) subjected to different doses of gamma radiation with Co⁶⁰.

between the varieties, irradiation dose and in the varietyxdose interaction (Table 3). At the variety level, the lulo with thorns had a greater multiplication capacity (1.52 seedlings per explant) than the thornless variety (1.12 seedlings per explant), confirming that the latter is more radiosensitive since, at the control level (0 Gy), they had a similar behavior (2.33 seedlings per explant and 2.37 seedlings per explant, respectively). The ability

of the initial explants to regenerate plants in the first 30 days after irradiation was correlated with the intensity; as the irradiation dose increased, the prolificacy of the explants was markedly reduced (Figure 2). The radiation-induced necrosis caused this result at the meristematic level, which caused general growth arrest that prevented or reduced the formation of appropriate phytomers for multiplication.

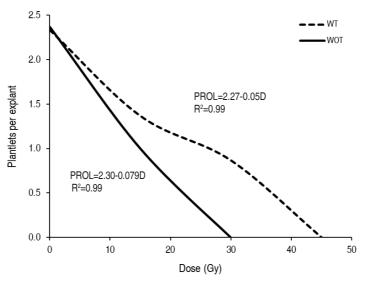


Figure 2. Effect of the of Co⁶⁰ gamma radiation dose on the regeneration capacity (plantlets per explant) of the plants in explants of two lulo varieties (*Solanum quitoense* Lam) with thorns (WT) and without thorns (WOT).

Genetic Variability

The band count for the seven primers showed 115 bands for the irradiated seedlings and 114 for the

irradiated calluses; the number of bands was between 12 for the TG primer and 21 for the ACA primer (Figure 3).

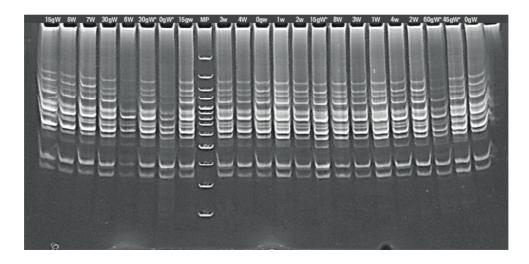


Figure 3. Banding patterns generated with RAM primer in Iulo (*Solanum quitoense* Lam.) calluses with thorns (*) and in Iulo seedlings with (W) and without (w) thorns, irradiated with gamma rays. g: indicate the irradiation doses; MP: marker.

The lulo calluses irradiated with different doses of gamma rays showed average unbiased heterozygosity of 0.22 and polymorphism of 50%. The analysis of the genetic distances provided values between 0.40 (between 0 Gy and 30 Gy) and 0.17 (between 0 Gy and 60 Gy). Likewise, the control treatment (without irradiation) presented similar values of 0.66 and 0.84 when compared with the irradiation doses of 30 Gy and

60 Gy, respectively (Table 4). The observed variability confirmed the effectiveness of gamma radiation in the induction of genetic variants at the level of the treated lulo explants (callus) and the sensitivity of the species to gamma radiation. This sensitivity depends on factors such as the physiological properties of the cells and nuclei, DNA content, chromosome size and ploidy level (Baloch *et al.*, 2004, Subramanian *et al.*, 2011).

Table 4. Matrix of genetic distance (above the diagonal) of Nei (1978) and similarity (below the diagonal) obtained in lulo with thorns calluses irradiated with Co⁶⁰ gamma rays.

Treatment	1(0 Gy)	2(15Gy)	3(30Gy)	4(45Gy)	5(60Gy)
1	-	0.34	0.40	0.34	0.17
2	0.71	-	0.31	0.25	0.27
3	0.66	0.72	-	0.16	0.37
4	0.71	0.77	0.85	-	0.27
5	0.84	0.76	0.68	0.76	-

The cluster analysis using the UPGMA criterion with Nei's distances (1972) formed four nodes. Node 1, with a distance of 0.16, contained the doses 30 Gy and 45 Gy, which, along with the 15 Gy dose, formed the second

node, while the control dose (0 Gy) and the maximum dose of 60 Gy formed a third group with a distance of 0.17. All treatments were included in the fourth node (Figure 4).

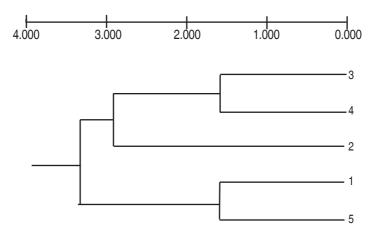


Figure 4. Dendogram of the genetic variability of calluses induced with lulo cotyledonary leaves and irradiated with Co^{60} gamma ray doses (Based on Nei's distances, combined data of seven RAM primers and UPGMA classification criteria; 1 = 0 Gy, 2 = 15 Gy, 3 = 30 Gy, 4 = 45 Gy, 5 = 60 Gy).

The analysis of the irradiation effect on lulo seedlings with and without thorns revealed unbiased heterozygosity of 0.13 and polymorphism of 31.03%; heterozygosity estimates the probability that two randomly drawn alleles from a set of genes in a population are different. The greatest genetic distance (0.22) was observed between the lulo with and without thorns controls, with a similarity of 0.80. The distance between the lulo with thorns control and the doses 15 Gy and 30 Gy was 0.15 and 0.18, with a similarity of 85% and 83%, respectively. There was a genetic distance of 0.15 and a similarity of 85% between the lulo without thorns control and the dose 15 Gy. The heterogeneity of responses to radiation between species and between genotypes of the same species

was reported by Arena *et al.* (2017), attributed mainly to differences between DNA contents (Subramanian *et al.*, 2011); however, the IAEA (1977) stated that radiosensitivity differences between genotypes of the same species are much lower than between other species.

The cluster analysis formed four groups. The treatments with 15 Gy on lulo without thorns and with 30 Gy on lulo with thorns formed group one, located at a distance of 0.13, which, along with the control treatment without thorns, formed group two at a distance of 0.17. Group three contained the treatments 0 Gy and 15 Gy with thorns at a distance of 0.14, and group four included all treatments at a distance of 0.18 (Figure 5).

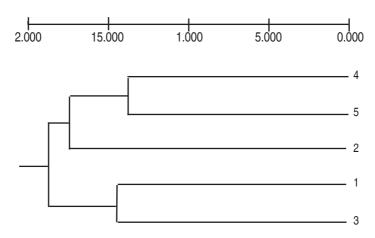


Figure 5. Dendogram of the genetic variability of lulo seedlings with (CE) and without thorns (SE) irradiated with different Co^{60} gamma ray doses (Based on Nei's distances, combined data of seven RAM primers and UPGMA classification criteria; 1 = 0Gy CE, 2 = 0Gy SE, 3 = 10Gy CE, 4 = 15Gy SE, 5 = 30Gy SE).

Rev. Fac. Nac. Agron. Medellín 73(3): 9263-9271. 2020

CONCLUSIONS

The Gamma ray irradiation of calluses induced from cotyledonary leaves of lulo and seedlings regenerated from lulo with and without thorns, generates genetic variability. The survival of the explants at the different doses of gamma radiation showed that the calluses were less radiosensitive (28 Gy, 50% mortality) and the lulo seedlings without thorns, were the most sensitive to radiation (19 Gy, 50% mortality). The ability of the irradiated explants (seedlings) to regenerate plants was inversely proportional to the dose of gamma rays received; the effects of the radiation were genotype-dependent.

ACKNOWLEDGMENTS

The authors express their gratitude to the Dirección de Asuntos Nucleares del Ministerio de Minas y Energía, the Universidad Nacional de Colombia, Bogotá and Palmira, Dr. Margarita Perea, Dr. Jaime Eduardo Muñoz, and the Grupo de Investigación en Producción de Frutales Andinos of the Universidad de Nariño.

REFERENCES

Agronet. 2018. Area, producción y rendimiento nacional por cultivo. http://www.agronet.gov.co/estadistica/Paginas/default.aspxConsulta: noviembre, 2018.

Angulo R. 2006. Lulo: el cultivo. Fundación Universidad de Bogotá Jorge Tadeo Lozano, Bogotá. 100 p.

Arena C, Turano M, Hay Mele B, Cataletto P, Furia M, Pugliese M and De Micco V. 2017. Anatomy, photochemical activity, and DNA polymorphism in leaves of dwarf tomato irradiated with X-rays. Biologia Plantarum 61(2):305-314. doi: 10.1007/s10535-016-0668-5

Baloch AW, Soomro AM, Bughio HR, Bughio MS, Mohammed T and Mastoi NN. 2004. Gamma irradiation induced chlorophyll mutations in rice (*Oryza sativa* L.). SAARC Journal of Agriculture 2: 257-261.

Betancourth C, Zambrano M y Narvaez C. 2005. Reacción de diferentes genotipos de lulo (*Solanum quitoense*) al ataque de *Fusarium oxysporum*. Revista de Ciencias Agrícolas 22(1-2):79-90.

Cruz P, Acosta K, Cure JR y Rodríguez D. 2007. Desarrollo y fenología del lulo *Solanum quitoense* var. *septentrionale* bajo polisombra desde siembra hasta primera fructificación. Agronomía Colombiana 25(2): 288-298.

Datta SK, Misra P and Mandal AKA. 2005. *In vitro* mutagenesis a quick method for establishment of solid mutant in chrysanthemum. Current Science 88: 155-158.

Dellaporta SL, Wood J and Hicks JB. 1983. A rapid method for DNA extraction from plant tissue. Plant Molecular Biology Reporter 1: 19-21.

Desai N, Joseph D, Suprasanna P and Bapat V. 2006. Study of elemental variations during somatic embryogenesis in sugarcane using photon induced X-ray probe. Nuclear Instruments and Methods in Physics Research 252(2): 299-302. doi: 10.1016/j.nimb.2006.08.009

IAEA - International Atomic Energy Agency. 1977. Manual on

mutation breeding. Second edition. Technical reports series No. 119, Vienna, Austria. 288 p.

Jayaramachandran M, Kumaravadivel N, Eapen S and Kandasamy G. 2010. Gene Action for yield attributing characters in segregating generation (M2) of Sorghum (*Sorghum bicolor* L.). Electronic Journal of Plant Breeding 1(4): 802-805.

Kavithamani D, Kalamani A, Vanniarajan C and Uma D. 2010. Development of new vegetable soybean (*Glycine max* L. Merill) mutants with high protein and less fibre content. Electronic Journal of Plant Breeding 1(4): 1060-1065.

Kumar P and Ratnam S. 2010. Mutagenic effectiveness and efficiency in varieties of sunflower (*Helianthus annuus* L.) by separate and combined treatment with gamma-rays and sodium azide. African Journal of Biotechnology 9(39): 6517-6521.

Lobo M. 2006. Recursos genéticos y mejoramiento de frutales andinos: una visión conceptual. Ciencia y Tecnología Agropecuaria 7(2): 40-54. doi: 10.21930/rcta.vol7_num2_art:68

Matsumura A, Nomizu T, Furutani F, Hayashi K, Minamiyama Y and Hase Y. 2010. Ray florets color and shape mutants induced by ¹²C⁵⁺ ion beam irradiation in chrysanthemum. Scientia Horticulturae 123: 558-561. doi: 10.1016/j.scienta.2009.11.004

Medina C, Sanchez D, Camayo G, Lobo M, y Martínez E. 2008. Anatomía foliar comparative de materiales de Iulo (*Solanum quitoense* Lam.) con y sin espinas. Ciencia y Tecnología Agropecuaria 9(1): 5-13.

Miller MP. 1997. Tools for population genetic analysis (TFPGA) 1.3: a Windows program for the analysis of allozyme and molecular population genetic data. Computer software distributed by the author.

Mostafa HHA, Wuan H, Shen D, Qiu Y, and Li X. 2015. Sprout differentiation and mutation induction of garlic (*Allium sativum* L.) callus exposed to gamma radiation. Plant Growth Regulation 75: 465-471. doi: 10.1007/s10725-014-0009-7

Muñoz JA, Rodríguez LF y Bermúdez LT. 2013. Análisis de la competitividad del sistema de producción de lulo (*Solanum quitoense* Lam.) en tres Municipios de Nariño. Revista Colombiana de Ciencias Hortícolas 7(2): 173-185. doi: 10.17584/rcch.2013v7i2.2233

Nei M. 1978. Estimation of average heterozygosity and genetic distance from a small number of individuals. Genetics 89(3): 583-590.

Ortiz R, De la Fé C y Ponce M. 2008. Incasoy-36: variedad de soya obtenida en cuba a partir de la inducción de mutaciones con los rayos gamma de ⁶⁰Co. Cultivos Tropicales 29(3):73.

Patade VY, Suprasanna P and Bapat VA. 2008. Gamma Irradiation of Embryogenic Callus Cultures and *In vitro* Selection for Salt Tolerance in Sugarcane (*Saccharum officinarum* L.). Agricultural Sciences in China 7(9): 1147-1152. doi: 10.1016/S1671-2927(08)60158-3

Predieri S and Zimmerman RH. 2001. Pear mutagenesis: *In vitro* treatment with gamma-rays and field selection for productivity and fruit traits. Euphytica 117(3): 217–227. doi: 10.1023/A:1026594103277

Prina A, Landau A, Pacheco M, y Hopp E. 2010. Mutagénesis, TILLING y EcoTILLING. pp. 218-228. En: Levitus G, Echenique V, Rubinstein C, Hopp E y Mroginski L (eds.). Biotecnología y Mejoramiento Vegetal II. ArgenBio, Ediciones Instituto Nacional de Tecnología Agropecuaria, Argentina. 648p.

Rocha F, Coan M, Coimbra J, Bertoldo J, Guidolin A and Kopp M. 2010. Root distribution in common bean populations used in breeding programs. Crop Breeding and Applied Biotechnology 10: 40-47.: doi: 10.12702/1984-7033.v10n01a06

Roman G, Harsan E and Wagner S. 2009. New cherry cultivars

obtained by mutagenesis at the fruit research station Cluj, Romania. Bulletin UASVM Horticulture 66(1): 215-217.

Rimieri P. 2017. La diversidad genética y la variabilidad genética: dos conceptos diferentes asociados al germoplasma y al mejoramiento genético vegetal. BAG. Journal of basic and applied genetics 28(2): 7-13.

Sahariya K, Kaushik RA, Khan R and Sarolia D. 2017. Influence of Gamma Irradiation on Flowering of Gladiolus (*Gladiolus hybrida* L.). International Journal of Current Microbiology and Applied Sciences 6(11): 1362-1368. doi: 10.20546/ijcmas.2017.611.162

Satpute RA and Fultambkar RV. 2012. Mutagenic effectiveness and efficiency of gamma rays and EMS in soybean (*Glycine max* (L.) Merrill). Current Botany 3(2): 18-20

Sambrook J, Fritsch EF and Maniatis T. 1989. Molecular Cloning: A Laboratory Manual. Second Edition. Cold Spring Harbor Laboratory Press, New York. 1546 p.

Silvera G. 2017. Mutaciones mediante radiaciones gamma en

arroz. Revista Observatorio de la Economía Latinoamericana 235:1-16.

Singh S and Singh S. 1993. Effect of gamma rays on callus growth and plant regeneration in sugarcane CV Co 687. Indian Sugar 43(3): 181-182.

Singh A. 2007. Mutagenic effectiveness and efficiency of gamma rays and ethyl methane sulphonate in mungbean. Madras Agricultural Journal 94(1-6): 7-13.

Sonone NG, Deshmukh SN, Lanjewar SB and Bhakre RL. 2010. Induced variability in quantitative characters of groundnut (*Arachis hypogaea* L.). Electronic Journal of Plant Breeding 1(6): 1409-1414.

Subramanian A, Nirmalakumari A. and Veerabadhiran P. 2011. Mutagenic efficiency and effectiveness in kodomillet (*Paspalum scrobiculatum* L.). Madras Agricultural Journal 98 (1-3): 22-25.

Velmurugan M, Rajamani K, Paramaguru P, Gnanam R, Bapu K, Harisudan C and Hemalatha P. 2010. *In vitro* mutation in horticultural crops. Agricultural Reviews 31(1): 63-67.