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YIELD AND QUALITY OF STRAWBERRY FRUITS FERTILIZED WITH BOVINE BIOFERTILIZER¹

ELISANGELA MARIA DOS SANTOS², THALES VINÍCIUS DE ARAÚJO VIANA², GEOCLEBER GOMES DE SOUSA^{3*}, BENITO MOREIRA DE AZEVEDO², JOÃO GUTEMBERG LEITE MORAES³

ABSTRACT - The objective of this study was to evaluate the effect of different doses of bovine biofertilizer on yield and post harvest of the strawberry crop in different growing environments. The experiment was carried out in two cycles of production, in Fortaleza, Ceará. The experimental design was in randomized blocks, arranged in subdivided plots, where the plots were of three cultivated environments (A1 = protected environment with screens, cold-water spraying, and white floor; A2 = environment with full sun, cold-water spraying, and white floor; A3 = environment with full sun, no cold-water spraying, and grey floor). The subplots consisted of five doses of biofertilizer (D1 = 0,0, D2 = 500, D3 = 750, D4 = 1000 and D5 = 1250 mL plant⁻¹ week⁻¹), with five replications. In the two-year period, the dose was higher in the protected environment of the weed type, with nebulization and on the white floor, at the dose of 500 mL week⁻¹ plant⁻¹ in the first cycle and 325 mL week⁻¹ plant⁻¹ in the second cycle. The dose of bovine biofertilizer of 1250 mL week⁻¹ plant⁻¹ promotes a longer duration and duration of the first cycle. During the second cycle, as the bovine biofertilizer doses of 585 and 620 provide higher and lower output than the protected environment. The environment without nebulization and on the floor without painting, the best soluble practices (° Brix) in relation to the protected environment of the type screened and a full sun with nebulization.

Keywords: *Fragaria x ananassa* duch. Organic Input. Postharvest.

PRODUTIVIDADE E QUALIDADE DE FRUTOS DE MORANGUEIRO ADUBADOS COM BIOFERTILIZANTE BOVINO

RESUMO - Objetivou-se avaliar o efeito de diferentes doses de biofertilizante bovino na produtividade e na pós-colheita da cultura do morango em diferentes ambientes de cultivo. O experimento foi desenvolvido, em dois ciclos de produção, em Fortaleza, Ceará. O delineamento experimental foi em blocos ao acaso, arranjados em parcelas subdivididas, onde as parcelas foram três ambientes de cultivo (A1= ambiente protegido do tipo telado, com nebulização com água gelada (temperatura variando entre 18 e 20 °C) e sobre piso branco, A2 = a pleno sol, com nebulização com água gelada e sobre piso branco, A3 = a pleno sol sem nebulização e sobre piso concretado sem pintura) e as subparcelas, cinco doses de biofertilizante (D1=0,0; D2=500; D3= 750; D4=1000; e D5= 1250 mL planta⁻¹. semana⁻¹), com cinco repetições. Nos dois anos, a produtividade foi maior no ambiente protegido do tipo telado, com nebulização e sobre piso branco, na dose de 500 mL semana⁻¹ planta⁻¹ no primeiro ciclo e 325 mL semana⁻¹ planta⁻¹ no segundo ciclo. A dose de biofertilizante bovino de 1250 mL semana⁻¹ planta⁻¹ promovem melhor diâmetro e comprimento do fruto durante o primeiro ciclo. Durante o segundo ciclo, as doses de biofertilizante bovino de 585 e 620 proporcionam maior comprimento e diâmetro do fruto, respectivamente, no ambiente protegido do tipo telado, com nebulização e sobre piso branco. O ambiente a pleno sol sem nebulização e sobre piso sem pintura, apresentou melhores sólidos solúveis (°Brix) em relação ao ambiente protegido do tipo telado e a pleno sol com nebulização.

Palavras-Chave: *Fragaria x ananassa* duch. Insumo orgânico. Pós-colheita.

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INTRODUCTION

Strawberry (*Fragaria x ananassa* Duch.) belongs to the Rosacea family; it is produced and appreciated in most regions of the world and is one of the most economic important small fruit species (OLIVEIRA et al., 2006). According to Mazaro et al. (2013), strawberry crops have socioeconomic importance in the South and Southeast regions of Brazil; it is an alternative of income for small farmers. Strawberry is used for fresh consumption and in the food processing industry. The average fruit yield of strawberry crops in Brazil is 30 Mg ha⁻¹, but it can reach more than 60 Mg ha⁻¹ in high-technological areas (REISSER JÚNIOR, 2015).

Technologies for strawberry production has been contributing to meet the quantitative and qualitative demand of farmers and consumers (HENZ, 2010). The fruit yield of strawberry varies significantly depending on the edaphoclimatic conditions of the growing region (thermal amplitude), and physiological (less stomatal resistance) and genetic (cultivar) factors of the plant (CAMARGO et al., 2010). The great variation of climatic conditions and the increasingly demanding consumer market have caused many producers to search for new production technologies. Protected environments are more advantageous to grow strawberries when compared to field conditions; they protect the crop from wind, hail, rain, frost, and low temperatures, minimize occurrence of diseases and attack from pests, and provide better conditions for the plant growth, increasing their fruiting development, and production for commercial purposes (COSTA et al., 2011; DIAS et al., 2015).

According to Reis et al. (2012), protected environments for crops are used intending to increase fruit yields and improving the quality of agricultural products by mitigating seasonal environmental variations in the production area, and reducing the adverse effects of excessive rainfall, incidence of solar radiation, and air temperatures.

The use of organic sources for soil fertilization combined with protected environments is a sustainable alternative for increasing productivity of agricultural crops. Cattle manure-based biofertilizer have been used satisfactorily for this purpose in recent years as an economic, environmental alternative to chemical fertilizers because it favors the natural cycling of nutrients and is a carbon source for agricultural crops (PENTEADO, 2007). These biofertilizer are liquid compounds produced in aerobic or anaerobic conditions with mixed organic material (e.g. fresh manure) and water (VIANA et al., 2014), they provide essential nutrients to the soil (VIANA et al., 2013) and improve the productivity of crops (DIAS et al., 2015).

In this context, the objective of this work was to evaluate the effect of different rates of a cattle manure-based biofertilizer and different environmental conditions on fruit yield and post-harvest quality of strawberry plants.

MATERIAL AND METHODS

The experiment was conducted at the experimental area of the Department of Agricultural Engineering of the Federal University of Ceará, in Fortaleza, CE, Brazil (03°45'S, 38°33'W, and altitude of 19.6 m), in two crop seasons (2013 and 2014). The first crop season was in the dry season (September to December 2013), and the second crop season was in the end of the rainy season (May to August 2014). The climate of the region is Aw', tropical rainy, according to the Köppen classification, with high temperatures, and rainy season mostly in the autumn. The climatic conditions of the experimental area were monitored by automatic weather stations installed in the environments evaluated. The mean and the relative measures of measurement are shown in Table 1.

Table 1. Monthly average air temperature and relative humidity.

Year	Air temperature (°C)			Relative air humidity (%)		
	E1	E2	E3	E1	E2	E3
2013 [#]	28.22	29.86	30.09	69.66	68.65	66.27
2014 ^{##}	27.76	28.04	29.43	71.39	70.83	69.9

[#] = means from September to December; ^{##} = means from May to August. E1 = protected environment with screens, cold-water spraying, and white floor; E2 = environment with full sun, cold-water spraying, and white floor; E3 = environment with full sun, no cold-water spraying, and grey floor.

Strawberry seedlings of the cultivar Oso Grande were transplanted into 11-liter pots containing substrate at the ratio 4:4:2 (sandy humic soil, coarse sand, and organic compound), spaced 1m × 1m. The chemical characteristics of the substrate

presented 19.16 g kg⁻¹ of organic matter, 0.16 mmol_c dm⁻³ of nitrogen, 26.3 mmol_c dm⁻³ of calcium, 0.11 mmol_c dm⁻³ of potassium, 45 mmol_c dm⁻³ of sodium, 0.34 mg dm³ of phosphorous, and pH of 6.9.

The experiment was conducted in a

randomized block design in split-plot arrangement with three plants per plot, and five replications, using three environments—protected environment of 10 × 5 m, with aluminum-based 50% reflective thermal screen (Aluminet®), cold-water (18 to 20 °C) spraying, and white floor (E1); environment with full sun, cold-water spraying, and white floor (E2); environment with full sun, no cold-water spraying, and grey floor (E3)—in the plots, and five rates (R1 = 0.0, R2 = 500, R3 = 750, R4 = 1,000, and R5 = 1,250 mL plant⁻¹ week⁻¹) of a cattle manure-based biofertilizer in the subplots.

The cold-water spraying was performed using a spray system with non-saline water, activated by a centrifugal pump, and controlled by a timer using water pulses; it was used for E1 and E2 every hour, from 09:30h to 16:30h, for 3 minutes, totaling 8 sprays per day.

The cattle manure-based biofertilizer was prepared with fresh cattle manure and water at 1:1 ratio, based on volume. This mixture was placed in a 500-liter box to be aerobically fermented for 20 days (VIANA et al., 2013). The chemical analysis of the biofertilizer presented N (0.82 g L⁻¹), P (1.4 g L⁻¹), K (1.0 g L⁻¹), Ca (2.5 g L⁻¹), Mg (0.75 g L⁻¹) and Na (0.28 g L⁻¹). The biofertilizer was applied weekly to all treatments (environments) from 15 days after transplanting (DAT) of the seedlings. A total of 15 applications were performed to meet the nutritional requirements of the crop, considering the maximum fertilization recommended for strawberry plants (SANTOS; MEDEIROS, 2003)—180 kg ha⁻¹ of N, 300 kg ha⁻¹ of P₂O₅, and 100 kg ha⁻¹ of K₂O. Thus, the maximum recommended rate per plant⁻¹ in the crop cycle was 14.4 g of N, 24 g of P₂O₅ and 8 g of K₂O.

The nutrient content in the substrate was estimate by multiplying the soil density (1.3 kg dm⁻³) by the volume of soil placed into each pot (10 L), resulting in 13 kg of soil per pot. Thus, the substrate provided 0.16 g kg⁻¹ of N, 0.11 g kg⁻¹ of P, and 0.34 g kg⁻¹ of K, i.e., the total N, P, and K available to the plants before the application of the biofertilizer was 2.08; 1.43, and 4.42 g kg⁻¹, respectively; therefore, the nutritional needs of the plants were 12.32 g plant⁻¹ of N, 22.57 g plant⁻¹ of P, and 3.42 g plant⁻¹ of N.

A drip irrigation system with one dripper per plant and average flow of 1.6 L h⁻¹ per emitter was used to apply the biofertilizer; the fertigation was controlled by valves at the beginning of each row.

The fertigation inside the protected environment was quantified using Equation 1:

$$Ti = \frac{LLi * AV * Faj}{Ei * q_{vi}} \times 60$$

wherein *Ti* is the fertigation duration (min), *LLi* is the liquid depth applied (mm); *Av* is the pot area (m²); *Faj* is the adjustment factor 0.8—internal divided by external water evaporation of USWB Class A tanks; *Ei* is the irrigation efficiency (0.90); and *q_v* is the flow per pot (L h⁻¹). The fertigation duration in the environments with full sun was quantified considering an adjustment factor (*Faj*) of 1.0. The fertigation was performed daily and the water evaporation was measured in USWB Class A tanks installed at 30 meters from the experimental area.

Fruits that presented 75% ripening or red-intense surface were harvested at 60 DAT (first crop season) and at 64 DAT (second crop season), following to the methodology proposed by Camargo et al. (2009). These fruits were counted and weighted using a precision scale to evaluate the number of fruits per plant (NFP), fruit weight (FW), and fruit yield (FY). NFP was quantified by the sum of the number of fruits divided by the number of plants of the plot; FY was represented by the average yield of fruits per area (kg ha⁻¹) in each crop seasons; and FW was represented by the mean weight of fruits per plot of each crop season.

The post-harvest variables evaluated in the two crop seasons were: soluble solids (SS) content (°Brix), determined by the analyze of the juice manually extracted from the samples in a PAL-1 Digital Refractometer (ATAGO); and fruit diameter (FD), and fruit length (FL) measured using a digital caliper.

The data were subjected to analysis of variance by the F test, the means of the plots (environments) were compared by the Tukey's test (*P*<0.05), and the means of the subplots (biofertilizer rates) were subjected to regression equations, using the ASSISTAT. 7.6 Beta program.

RESULTS AND DISCUSSION

According to the analysis of variance, the interaction between the environments and biofertilizer rates was significant in the two crop seasons for fruit yield (FY). It was significant for fruit weight (FW) only in the first crop season; and for number of fruits per plant (NFP) only in the second crop season (Table 2).

Table 2. Analysis of variance and significance levels for number of fruits per plant (NFP), the average fruit mass (FM) and fruit yield (FY) of strawberry plants grown in different environments and fertilized with different rates of a cattle manure-based biofertilizer (BR) in two crop seasons (2013 and 2014).

Source of variation	DF	Mean square					
		2013			2014		
		NFP	FW	FY	NFP	FW	FY
Environments (A)	2	75.57 **	3.561*	58.74 **	2.16 ^{ns}	1.98 ^{ns}	5.20 **
Residue (a)	12	2.25	0.92	1.41	1.95	0.83	1.33
Plot	14						
BR (B)	4	20.61 ^{ns}	3.52 **	3.77 *	1.29 ^{ns}	1.60 ^{ns}	2.66 ^{ns}
A x B interaction	8	9.52 ^{ns}	2.08 *	2.43 *	2.34 *	0.61 ^{ns}	8.99 **
Residue (b)	48	9.31	0.9	5.1	1.8	1.74	1.14
Total	74						
CV A (%)		28.76	26.89	20.25	27.87	15.99	25.6
CV B (%)		29.44	26.11	29.17	29.01	24.25	22.46

DF = degrees of freedom; CV = coefficient of variation; ^{ns} = not significant, * = significant by the F test at 5% probability, ** = significant by the F test at 1% probability.

The environment had independent effect on NFP (Figure 1). Plants in Environment 1 (E1) had higher NFP (5.7) than those in Environment 2 (E2) (3.56), and 3 (E3) (2.28). This result may be due to the better control of leaf temperature by the plants,

which optimized the photosynthetic activity, and turgidity, resulting in a better flowering, fruiting, and consequently, a high NFP (CHAVARRIA et al., 2012).

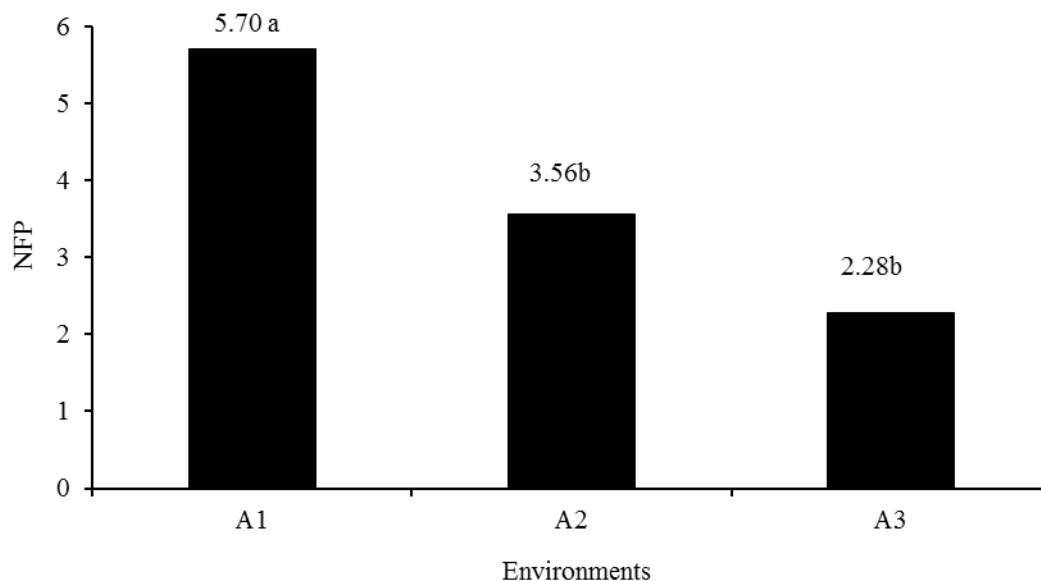


Figure 1. Number of fruits per strawberry plant influenced by cultivated environments, A1 = protected environment with screens, cold-water spraying, and white floor; A2 = environment with full sun, cold-water spraying, and white floor; A3 = environment with full sun, no cold-water spraying, and grey floor.

The lower fruit production found in E2 and E3 were similar to that reported by Andriolo et al. (2010), who explained this result by the abortion of strawberry flowers due to high temperatures. Dias et al. (2015) evaluated strawberry crops in a mountain region in the state of Ceará, Brazil, which has average temperatures of 27.1 °C, and found a higher NFP (10.96).

The interaction between environments and

biofertilizer rates was significant in the second crop season for NFP. The NFP of plants grown in E3 fitted to a linear model, and plants in soils with no biofertilizer had a maximum NFP of 2.5. The NFP of plants grown in E1 and E2 fitted to a quadratic polynomial model; the biofertilizer rate of 583.33 mL plant⁻¹ week⁻¹ resulted in a maximum NFP of 5.37 (E1), and the rate of 650 mL plant⁻¹ week⁻¹ resulted in a maximum NFP of 3.68 (E2) (Figure 2).

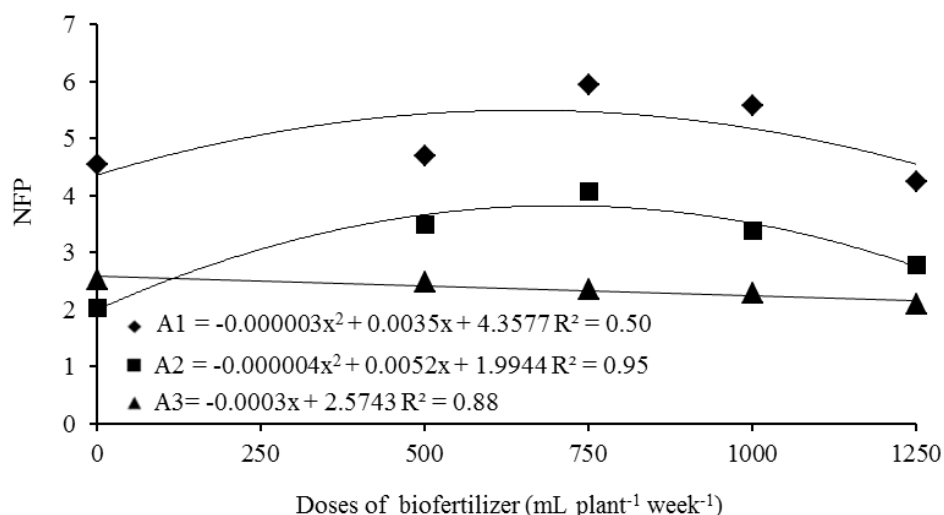


Figure 2. Number of fruits per plant of strawberry as a function of the doses of bovine biofertilizer cultivated in protected environment protected environment of the screened type, protected environment with screens, cold-water spraying, and white floor - A1 (♦), environment with full sun, cold-water spraying, and white floor - A2 (■) and environment with full sun, no cold-water spraying, and grey floor - A3 (▲).

Higher NFP were reported by Vignolo et al. (2011), who evaluated production of strawberry plants of the cultivar Camarosa under full sun conditions and found 43.6 fruits plant⁻¹; and by Pires et al. (2007) who evaluated NFP of strawberry plants under protect environment and found 85.7 fruits plant⁻¹. However, these expressively higher NFP were found in experiments conducted in regions with more favorable climatic conditions to this crop—average temperatures of 22 °C, and air relative humidity of 76%.

The low fruit production found in the coast of the state of Ceará, Brazil, was probably due to the high temperatures (average of 28.2 °C) of this

region, which generated a stressful environment, causing floral abscission and, consequently, reduction of number of fruits (LEDESMA et al., 2008). Sousa et al. (2014) performed an experiment under the same climatic conditions of the present study and found higher NFP for strawberry plants of the cultivars Oso Grande (9.5 fruits plant⁻¹) and Verão (6.45 fruit plant⁻¹) grown under full sun (average temperature of 26.2 °C) conditions.

The interaction between the environments and biofertilizer rates was significant for fruit weight (FW) only in the first season (Figure 3); the FW data in E1 and E2 fitted to a positive linear model, and those of E3 fitted to a negative linear model.

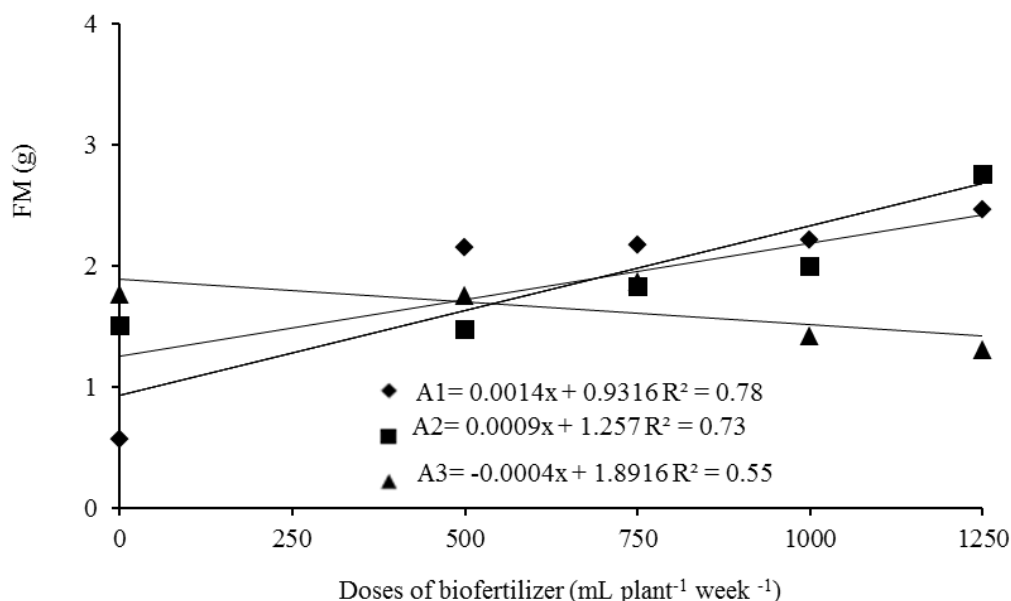


Figure 3. Average mass of fruits per plan of strawberry as a function of the doses of bovine biofertilizer, in protected environment protected environment of the screened type, protected environment with screens, cold-water spraying, and white floor - A1 (♦), environment with full sun, cold-water spraying, and white floor - A2 (■) and environment with full sun, no cold-water spraying, and grey floor - A3 (▲).

FW increased in E1 and E2, and decreased in E3, probably due to the temperature of the environments, i.e., the lower the temperature the lower the plants' metabolic expense and, consequently, the greater the amount of photoassimilates to the fruits. Moreover, high temperatures directly affect stomatal closure, reducing absorption of nutrients and fruit weight.

A high nutrient intake allows high photosynthetic rates and, consequently, large amount of carbohydrates allocated to reserve organs of plants, resulting in heavier fruits (SOUSA et al., 2013). Contrastingly, Dias et al. (2015) found no significant effects for strawberry FW when evaluating different biofertilizer rates in full sun, and screened protected environments. However, according to Sousa et al. (2014), the FW of strawberry plants of the Oso Grande cultivar in a similar environment to E1 of the present study were similar to that found in the environment with full sun.

In Figure 4A and 4B the regression analyses for the data of total fruit yield (FY) as a function of the interaction between the environments and

biofertilizer rates showed a quadratic polynomial model for the first season, with the highest yields of 10.15 Mg ha⁻¹ with a biofertilizer rate of 378.57 mL plant⁻¹ week⁻¹ (E1); 4.62 Mg ha⁻¹ with a biofertilizer rate of 576.16 mL plant⁻¹ week⁻¹ (E2); and 2.95 Mg ha⁻¹ with a biofertilizer rate of 750 mL plant⁻¹ week⁻¹ (E3). The data of FY in E1 and E2 fitted to a quadratic model in the second season, with maximum yields of 6.49 Mg ha⁻¹ with a biofertilizer rate of 500 mL plant⁻¹ week⁻¹ (E1), and 4.65 Mg ha⁻¹ with a biofertilizer rate of 325 mL plant⁻¹ week⁻¹ (E2); and the data of FY in E3 in the second season fitted to a negative linear model, with a maximum yield of 2.52 Mg ha⁻¹ with no biofertilizer.

The results showed an increase in FY from the first to the second crop season; this can be due to the occurrence of lower temperatures in the second season, which improved the plant development and, consequently, fruit yield. The air temperature of the experimental area affects the potential flowering of strawberry plants; since it affects the speed of biochemical reactions, and internal processes of sap transport (RESENDE et al., 2010).

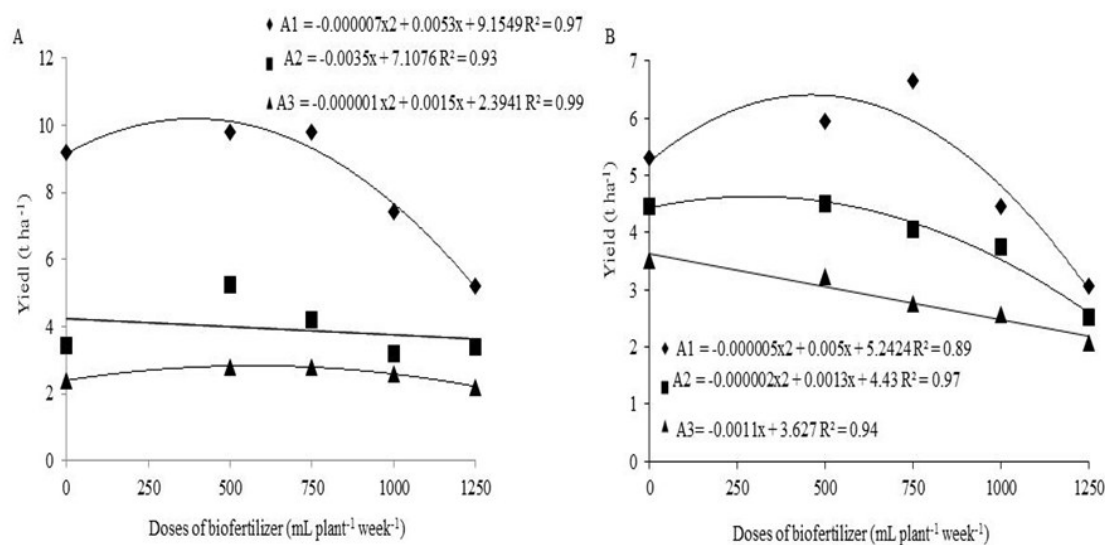


Figure 4. Yield per plan of strawberry as a function of the doses of bovine biofertilizer, in protected environment protected environment of the screened type, protected environment with screens, cold-water spraying, and white floor - A1 (♦), environment with full sun, cold-water spraying, and white floor - A2 (■) and environment with full sun, no cold-water spraying, and grey floor - A3 (▲).

Studies conducted under similar climatic conditions with the same strawberry cultivar (Oso Grande) showed variable results of FY; Sousa et al. (2014) found FY of 3.2 Mg ha⁻¹ when this cultivar were grown under full sun and Lima (2014) reported a FY of 6.7 Mg ha⁻¹ for plants under protected environment. Dias et al. (2015) studied this same cultivar under the climatic conditions of Redenção, CE, Brazil, and found 10.7 Mg ha⁻¹; however, these values are well below those found in some experiments performed in the South and Southeast

regions of Brazil. According to Filgueira (2012), the strawberry FY in Brazil can reach 80 Mg ha⁻¹ when crops are well managed, in areas with favorable climatic conditions, and using adapted cultivars.

According to the analysis of variance (Table 3), no significant interaction between environments and biofertilizer rates were found for fruit length (FL), fruit diameter (FD), and soluble solids (SS) content in the first crop season. However, in the second crop season, this interaction was significant for FL and FD at 5% level of significance.

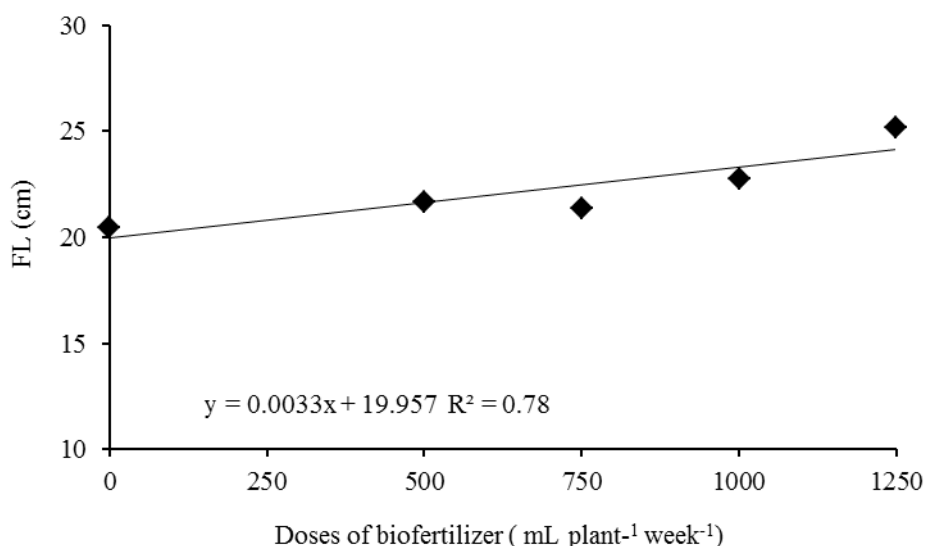
Table 3. Analysis of variance and levels of significance for fruit length (FL), fruit diameter (FD), and soluble solids (SS) content of strawberry plants grown in different environments and fertilized with different rates of cattle manure-based biofertilizer (BR) in two crop seasons (2013 and 2014).

Sources of variation	DF	Mean square					
		2013			2014		
		FL	FD	SS	FL	FD	SS
Environments (A)	2	156.87 ^{ns}	204.82 ^{ns}	37.83 ^{**}	38.38 ^{**}	22.77 [*]	3.16 ^{ns}
Residue (a)	12	7.84	3.52	1.76	4.83	4.87	0.96
Plots	14						
BR (B)	4	49.40 ^{**}	25.77 ^{**}	4.01 ^{ns}	4.37 ^{ns}	2.12 ^{ns}	1.67 ^{ns}
A x B interaction	8	14.55 ^{ns}	6.85 ^{ns}	3.47 ^{ns}	5.41 [*]	6.40 [*]	1.30 ^{ns}
Residue (b)	48	11.41	4.34	3.11	2.38	2.39	1.06
Total	74						
CV A (%)		12.56	10.36	16.19	8.27	11.33	11.83
CV B (%)		15.15	11.5	23.51	5.8	7.94	12.44

DF = degrees of freedom; CV = coefficient of variation; ^{ns} = not significant, * = significant by the F test at 5% probability, ** = significant by the F test at 1% probability.

In Figure 5 one can see the biofertilizer rate had independent effect on FL in the first crop season, with data fitting to a positive linear model, and the highest FL (25 mm) found with a biofertilizer rate of 1,250 mL plant⁻¹ week⁻¹. These results may be due to

the greater supplying of nutrients by the higher biofertilizer rate. A good nutritional condition allows a better crop development and fruit yield (SOUSA et al., 2013; VIANA et al., 2013).

**Figure 5.** Fruit length due to the different doses of bovine biofertilizer.

Contrastingly, Dias et al. (2015) found a decreased in FL with increasing rates of a cattle manure-based biofertilizer for plants of the Oso Grande cultivar grown in the climatic conditions of Redenção, Ceará.

In Figure 6 it is observed that the interaction between environments and biofertilizer rates was

significant for FL in the second crop season. The FL data of the tree environments evaluated fitted to a quadratic polynomial model; however, the highest FL found were 29.22 mm in E1 with a biofertilizer rate of 585 mL plant⁻¹ week⁻¹; 25.04 mm in E2 with a rate of 370 mL plant⁻¹ week⁻¹; and 27.22 mm in E3 with a rate of 466.67 mL plant⁻¹ week⁻¹.

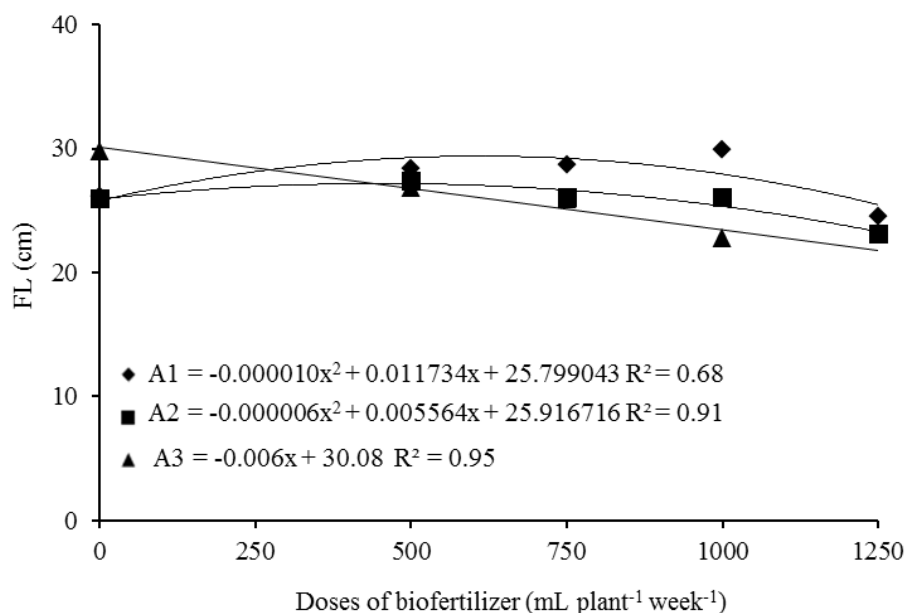


Figure 6. Fruit length as a function of the doses of bovine biofertilizer, in protected environment protected environment of the screened type, protected environment with screens, cold-water spraying, and white floor - A1 (♦), environment with full sun, cold-water spraying, and white floor - A2 (■) and environment with full sun, no cold-water spraying, and grey floor - A3 (▲).

Dias et al. (2015) evaluated strawberry crops in greenhouse and in field conditions subjected to increasing cattle manure-based biofertilizer rates and found higher FL in plants grown in field conditions. Yuri et al. (2012) evaluated the strawberry cultivar Oso Grande grown with different types of mulching in the state of Minas Gerais, Brazil, and found FL of 29.1 mm—higher than that found in the present work.

The model that best fitted to a positive linear model as a function of the biofertilizer rates in the first crop season (Figure 7). According to Pinto et al. (2008), the application of organic products to agricultural soils is important because of their diverse mineral nutrients and positive action as enzymatic activator of plant metabolism, which can contribute to a larger fruit diameter.

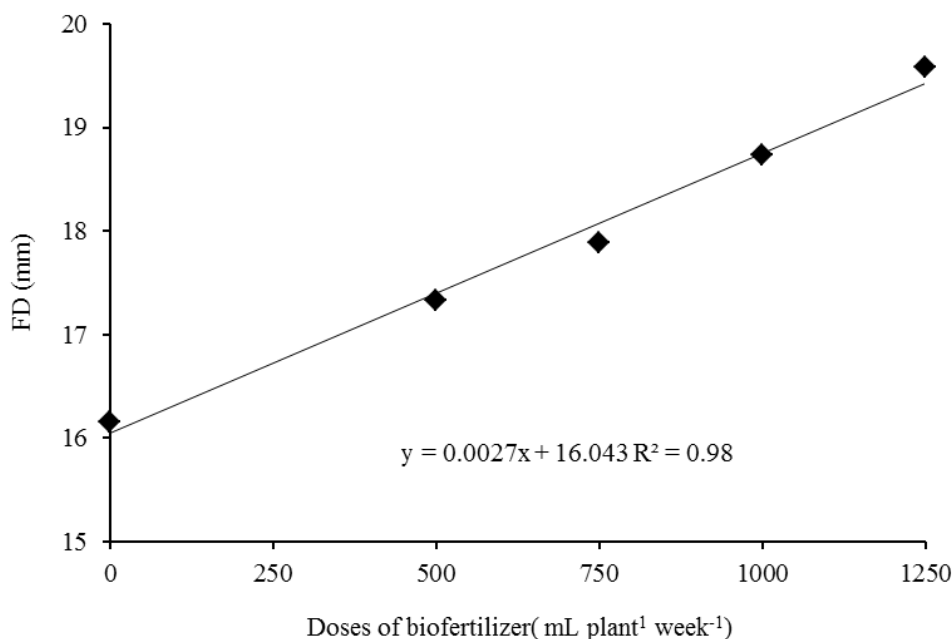


Figure 7. Fruit diameter due to the different doses of bovine biofertilizer.

The result for FD found by Lima (2014) (21.0 mm) was similar to that found in the present work; they evaluated strawberry plants grown with a cattle manure-based biofertilizer under the same climatic conditions. However, Dias et al. (2015) found no significant changes in FD of strawberry fruits with increasing rates of this same biofertilizer.

In Figure 8, the biofertilizer rates and its

interaction with the environments affected the FD in the second cycle. The mathematical models showed largest FD (21.46 mm) in plants in E1 with the rate of 620 mL plant⁻¹ week⁻¹; fruits in E2 presented similar FD with increasing biofertilizer rates; and fruits in E3 had the largest FD (20.05 mm) with a biofertilizer rate of 330 mL plant⁻¹ week⁻¹.

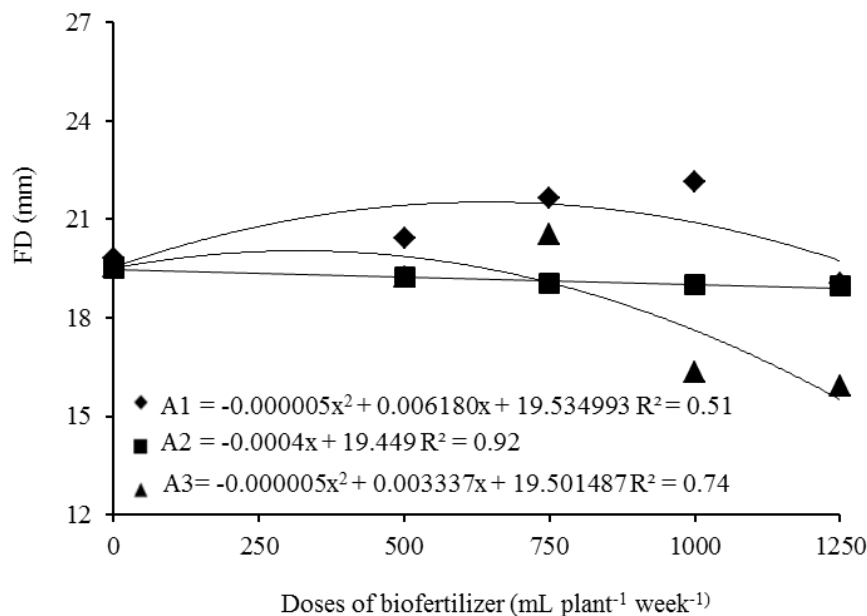


Figure 8. Fruit diameter as a function of the doses of bovine biofertilizer, in protected environment protected environment of the screened type, protected environment with screens, cold-water spraying, and white floor - A1 (♦), environment with full sun, cold-water spraying, and white floor - A2 (■) and environment with full sun, no cold-water spraying, and grey floor - A3 (▲).

Similarly, Rodrigues et al. (2009) reported a positive increase of longitudinal diameter in passion fruit (*Passiflora edulis f. flavicarpa* Dg.) fruits using a Brazilian biofertilizer formula known as Supermagro. Moreover, Silva et al. (2016) found positive FD response of fig (*Ficus carica* L.) plants grown under protected environment, and full sun conditions to a cattle manure-based biofertilizer.

The soluble solids (SS) contents of fruits in E2 (8.54 °Brix) and E3 (9.2 °Brix) were statistically similar by the Tukey's test in the first crop season, but they were higher than that of fruits in E1 (6.82 °Brix) (Figure 9). The SS of fruits of all environments

were similar in the second crop season.

The plants grown under full sun had higher °Brix than those grown under protected environment, differing from the results of Resende et al. (2010) who found 6.9, 6.2, and 5.6 °Brix for strawberries grown in high tunnel greenhouse, low tunnel greenhouse, and full sun conditions, respectively. The °Brix found by Dias et al. (2015) (6.77) was similar to that found in the present work in the protected environment for the same cultivar; however, they found lower °Brix (7.8) in fruits grown under full sun conditions.

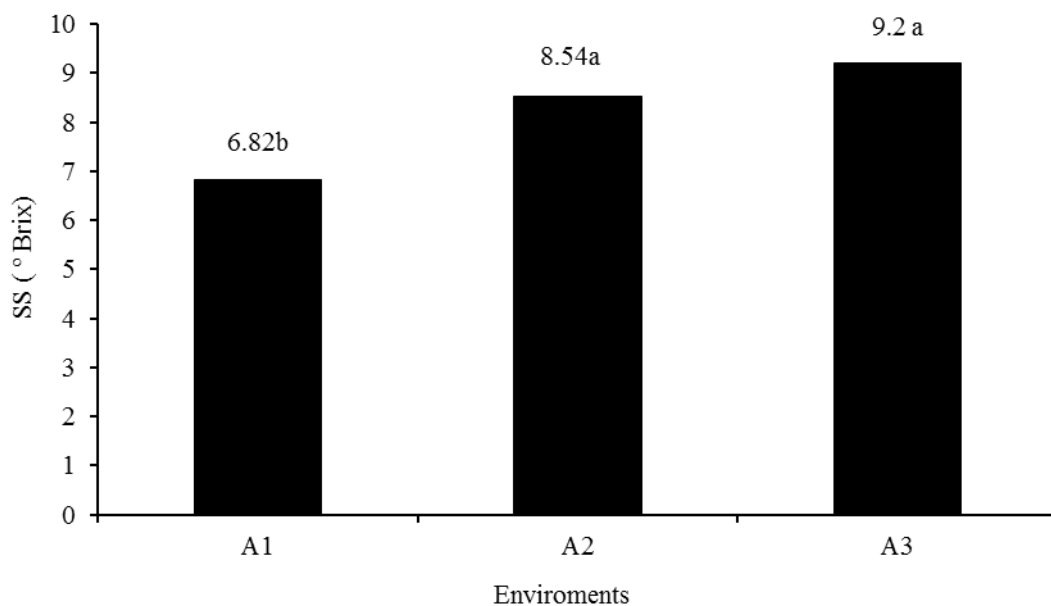


Figura 9. Soluble solids in plant plants influenced by growing environments, A1 = protected environment with screens, cold-water spraying, and white floor; A2 = environment with full sun, cold-water spraying, and white floor; A3 = environment with full sun, no cold-water spraying, and grey floor.

CONCLUSIONS

Plants grown under screened protected environment, with cold-water spraying, and white floor presented the greatest number of strawberry fruits per plant in both first (2013) and second (2014) crop seasons.

The application of the cattle manure-based biofertilizer at rate of 1,250 mL plant⁻¹ week⁻¹ resulted in higher fruit weights when plants were grown in environment with full sun, cold-water spraying, and white floor (second crop season).

The strawberry fruit yield was higher when the plants were grown under protected environment with a biofertilizer rate of 500 mL plant⁻¹ week⁻¹ (first crop season) and 325 mL plant⁻¹ week⁻¹ (second crop season).

The highest fruit diameter and length in the first crop season was found using a biofertilizer rate of 1,250 mL plant⁻¹ week⁻¹.

In the second crop season, the highest fruit length was found with a biofertilizer rate of 585 plant⁻¹ week⁻¹, and the highest fruit diameter was found with a rate of 620 mL plant⁻¹ week⁻¹, in plants grown under protected environment.

Plants grown in the environment with full sun, no cold-water spraying, and gray floor presented fruits with better soluble solid contents (°Brix) than those in the protected environment, and the environment with full sun and cold-water spraying.

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