



Ingeniería, investigación y tecnología

ISSN: 1405-7743

Universidad Nacional Autónoma de México, Facultad de Ingeniería

Loa-Ramírez, Ixchel Adriana; Rodríguez-Montesinos, Yoloxochitl Elizabeth; Arvizu-Higuera, Dora Luz; Hernández-Carmona, Gustavo

Adaptation of the alginate production process for the brown alga *Sargassum horridum* (Fucales: Sargassaceae)

Ingeniería, investigación y tecnología, vol. XXIII, no. 2, e1777, 2022, April-June
Universidad Nacional Autónoma de México, Facultad de Ingeniería

DOI: <https://doi.org/10.22201/fi.25940732e.2022.23.2.010>

Available in: <https://www.redalyc.org/articulo.oa?id=40475446002>

- How to cite
- Complete issue
- More information about this article
- Journal's webpage in redalyc.org

UNAM  redalyc.org

Scientific Information System Redalyc

Network of Scientific Journals from Latin America and the Caribbean, Spain and Portugal

Project academic non-profit, developed under the open access initiative



Adaptation of the alginate production process for the brown alga *Sargassum horridum* (Fucales: Sargassaceae)

Adaptación del proceso de producción de alginato para el alga café *Sargassum horridum* (Fucales: Sargassaceae)

Loa-Ramírez Ixchel Adriana

Universidad del Mar, Campus Puerto Ángel, Oaxaca

Correo: ixchel_loa@hotmail.com

<https://orcid.org/0000-0003-3370-2618>

Rodríguez-Montesinos Yoloxochitl Elizabeth

Instituto Politécnico Nacional

Centro Interdisciplinario de Ciencias Marinas (CICIMAR)

Correo: yrodriguez@ipn.mx

<https://orcid.org/0000-0002-5841-7701>

Arvizu-Higuera Dora Luz

Instituto Politécnico Nacional

Centro Interdisciplinario de Ciencias Marinas (CICIMAR)

Correo: darvizu@ipn.mx

<https://orcid.org/0000-0002-5219-7392>

Hernández-Carmona Gustavo (Corresponding author)

Instituto Politécnico Nacional

Centro Interdisciplinario de Ciencias Marinas (CICIMAR)

Correo: gcarmona@ipn.mx

<https://orcid.org/0000-0002-8931-7148>

Abstract

The genus *Sargassum* is a brown alga which is abundant in México, in the Gulf of California and the Caribbean Sea. The large biomass could be used to production of alginate, a polysaccharide with properties which allow it to form a thick solution or gels for different industries, such as for pharmaceutical and food products. The alginate production process may have some modifications, depending on the type of alga that is used as raw material, reducing water consumption and increasing the alginate yield and quality. In this research, the amount of water and reagents was optimized, reducing the volumes in the production process, based on the volumes used to process the giant kelp *Macrocystis pyrifera*. Hydration was reduced from 1:10 to 1:4 (part of alga:volume of chemical), acid pre-extraction from 1:10 to 1:5, extraction from 1:16 to 1:10, and dilution from 1:55 to 1:15; bleaching was improved from very dark brown to pale brown. The main reason that reduction is possible is that *Sargassum* has a more rigid cell wall than *Macrocystis*, absorbing less water and producing less viscosity. Although the alginate viscosity is relatively low, some applications require low viscosity solutions. However, the gel strength reached up to 1906 g cm^{-2} , which may compete with other alginates to produce gelling products. To avoid the use of hydrochloric acid and decrease the contamination of wastewater in the process, the acid pre-extraction stage was changed for a neutral pre-cooking stage, resulting in an increase of the alginate yield from 8.3 % to 13.5 %; the viscosity was normally kept low, but the gel strength increased up to 2400 g cm^{-2} .

Keywords: Process, polysaccharide, alginate, yield, viscosity, gel strength.

Resumen

El género *Sargassum* es un alga parda que abunda en México, tanto en el Golfo de California como en el Mar Caribe. La gran biomasa podría usarse para la producción de alginato, un polisacárido con propiedades para formar soluciones viscosas o geles para diferentes industrias, como las farmacéuticas y de alimentos. El proceso de producción de alginato puede tener algunas modificaciones, dependiendo del tipo de alga que se utiliza como materia prima, reduciendo el consumo de agua e incrementando el rendimiento y la calidad del alginato. En esta investigación se optimizó la cantidad de agua y de reactivos, reduciendo los volúmenes en el proceso de producción, que se basaron en los volúmenes utilizados para procesar el alga gigante *Macrocystis pyrifera*. La hidratación se redujo de 1:10 a 1:4 (parte de alga: volumen de producto químico); pre-extracción ácida de 1:10 a 1:5, extracción de 1:16 a 1:10, dilución de 1:55 a 1:15 y blanqueamiento de marrón muy oscuro a marrón pálido. La razón principal que hace posible esta reducción es que *Sargassum* tiene una pared celular más rígida que *Macrocystis*, adsorbe menos agua y produce alginato con menor viscosidad. Aunque los valores de viscosidad son relativamente bajos, algunas aplicaciones requieren soluciones de baja viscosidad. Sin embargo, la fuerza de gel alcanzó hasta 1906 g cm^{-2} , la cual puede competir con otros alginatos para producir productos con propiedades gelificantes. Para evitar el uso de ácido clorhídrico y disminuir la contaminación de las aguas residuales en el proceso, se experimentó la eliminación de la etapa de pre-extracción ácida y en su lugar se realizó una etapa de precocción neutra, aumentando el rendimiento de 8.3 a 13.5 %, la viscosidad se mantuvo normalmente baja, pero la fuerza de gel aumentó hasta 2400 g cm^{-2} .

Descriptores: Proceso, polisacárido, alginato, rendimiento, viscosidad, fuerza de gel.

INTRODUCTION

The brown alga *Sargassum* has become a very important marine resource in recent times, since global changes in the sea have generated massive arrivals on different coasts of the world, including México (Casas *et al.*, 1990; Rodríguez *et al.*, 2016). Brown algae are a source of alginates that have properties which allow the production of viscous solutions or the formation of gels, which can be used in different industries such as for textiles, paper, pharmaceuticals, bandages, capsules, dental impression material, immobilization of organisms and plant growth promoters, among many (Di Filippo *et al.*, 2018).

Most of these studies about the polysaccharides from brown seaweeds in México are focused on the giant kelp *Macrocystis pyrifera*, because it is the most abundant and provides the highest alginate yield. Hernández *et al.* (2012) published a detailed process for the extraction of alginate from *M. pyrifera*, which was scaled up to pilot plant level. That method was considered as the control process to carry out this research. The chemical composition and biological activity of *Sargassum horridum* has previously been described for beds in Bahía de La Paz, Baja California Sur, the months with maximum values being found as follows: alginate yield 8.6 % in August, viscosity 73 mPa s (measured in 1 % alginate solution at 22°C with a viscometer (NDJ-1) at 60 rpm using the appropriate spindle) in September and gel strength 1906 g cm⁻² in May, maintained for the rest of the months at around 800 g cm⁻² (Di Filippo *et al.*, 2018).

The hypothesis of this research is that alginate production from *S. horridum* can be adapted from the process used for *M. pyrifera*, reducing the consumption of water and reagents, and therefore reducing the production cost and increasing the alginate yield and quality.

MATERIALS AND METHODS

Samples of *S. horridum* were collected from San Juan de la Costa, in Bahía de La Paz, Baja California Sur, México, in June 2018. The algae were cut near the holdfast with a knife, at 1–2 m depth (n = 30 of 1 m²) and transported in mesh bags to the laboratory. They were washed with tap water and sun-dried. Subsequently, they were ground in a mill and sieved to homogenize the particle size to 30 mesh (0.5 mm²).

SODIUM ALGINATE PRODUCTION PROCESS

All the experiments were carried out in triplicate. The volumes described for the control are those used for *M. pyrifera* in the process described by Hernández *et al.* (2012), which includes the following steps: hydration,

acid pre-extraction, alkaline extraction, dilution, filtration, precipitation, conversion of calcium alginate to alginic acid, conversion of alginic acid to sodium alginate, and drying.

HYDRATION

Ten grams of dried and ground algae were hydrated for 12 h with different volumes of 0.1 % formaldehyde solution at a ratio of 1:9 (alga: formaldehyde) coded as H control, 1:8 (H8), 1:6 (H6), 1:5 (H5) and 1:4 (H4). After this time, the algae were filtered, and the residual liquid was measured. Since the volume of liquid incorporated into the tissues was constant, hydration was carried out with the minimum ratio of 1:4 for the next experiments.

ACID PRE-EXTRACTION

The algae previously hydrated at 1:4 was placed in different volumes of distilled water: 1:20 control (Pre-control), 1:15 (Pre15), 1:10 (Pre10) and 1:5 (Pre5). The pH was adjusted to 4 by adding 1 N hydrochloric acid solution. The algae were maintained under constant stirring with a magnetic stirrer, for 15 min, filtered and transferred to the next stage.

ALKALINE EXTRACTION

The algae, previously hydrated and treated with acid as described above, were placed in water at different ratios (alga:water): 1:20 (Ex control), 1:15 (Ex15) and 1:10 (Ex10). Then, 10 % sodium carbonate solution (w/v) was added until the pH was adjusted to 10, while maintaining the beaker in a water bath at 80 °C, with an external motor stirrer (CAFRAMO®) with a three-blade propeller, at a speed of 800 rpm for 2 h.

DILUTION AND FILTRATION

The alginate solution produced was diluted with distilled water to 1:55 (Control), 1:20 (Di20), 1:15 (Di15) and 1:10 (Di10), based on the initial weight of the algae, and mixed with 10 g of diatomaceous earth (Celite®) as a filtration aid. The mixture was then filtered with a layer of diatomaceous earth over a filter paper, contained in a Buchner funnel under vacuum.

PRECIPITATION AND DRYING (LABORATORY LEVEL METHOD)

The clarified alginate solution was precipitated with alcohol in a proportion of 1:2 to assure that all the alginate in solution was precipitated. Finally, the alginate

fibres produced were oven-dried at 55 °C for 12 h and placed in a desiccator to avoid moisture. The percentage of alginate was calculated based on the initial weight of the alga: $R = (\text{dry weight of alginate} / \text{dry weight of alga}) \times 100$. At industrial level, the alginate is not precipitated with alcohol, because of the high cost, so it is precipitated as calcium alginate and then converted to alginic acid and finally to sodium alginate. The next experiments followed those industrial steps.

QUALITY CONTROL OF THE ALGINATE OBTAINED

VISCOSITY

The viscosity of the samples was obtained by preparing a 1 % alginate solution and measuring the viscosity with a viscometer NDJ-1, measuring at 60 rpm and 22 °C, with the appropriate needle (1–4), before and after adding sodium hexametaphosphate which sequesters any residual calcium (Hernández *et al.*, 1999).

GEL STRENGTH

The gel strength of alginate was measured by preparing a 1 % alginate solution in distilled water and filling a dialysis membrane tube (5 cm in diameter $\times$ 2.5 cm in height); the tubes were immersed in a 10 % calcium chloride solution for 24 h to form the gels. After that time, the gel strength was measured with a texture analyser (TA.XT Plus), programmed to carry out a penetration of 2 cm at a speed rate of 5 s (breaking point) (Camacho & Hernández, 2012).

PRECIPITATION TO CALCIUM ALGINATE, ALGINATE BLEACHING, AND CONVERSION TO SODIUM ALGINATE

A new batch of 10 g of alga was processed until the filtration stage. The clarified alginate solution, instead of being precipitated with alcohol, was precipitated with 37.5 mL of 10 % calcium chloride for every 10 g of dry alga used. The resulting alginate fibres were immersed in 20 mL of water, adding 5 % sodium hypochlorite (commercial chlorine) at different volumes: 0.0 (control, T1), 0.5 (T2), 1.0 (T3), 2.0 (T4) and 3.0 mL (T5). The bleached alginate fibres were subjected to three acid washes for conversion to alginic acid as follows: the fibres were suspended in 20 parts of water for one part of algae; the pH was adjusted to 2 with 1 N hydrochloric acid solution and stirred for 15 min. The sample was drained off and the acid washing was repeated twice, adjusting the pH to 1.8 each time. Sodium alginate was obtained by adding 75 mL of water and 75 mL of alcohol and neu-

tralizing with sodium carbonate until adjusting to pH 10, with constant agitation (Arvizu *et al.*, 2002).

ALGINATE COLOR

The color of a 1 % alginate solution was measured using a UV Visible Spectrophotometer (Thermo Scientific™ GENESYS 10S) at a wavelength of 510 nm (as a percentage of absorbance). Color was compared with commercial sodium alginate (71238 Sigma™ and Algimar™) and with the Munsell charts (2009), to assign a name to the colors.

ALGINATE PURITY

One gram of calcium chloride was dissolved in 100 mL of methanol-water solution (40-60 %). The resulting solution was added to 100 mL of 0.5 % alginate solution, while gently stirring. The precipitate was removed using a fine filter and washed with methanol-water solution (20-80 %). A second washing was carried out with methanol water solution (40-60 %). The precipitated alginate was dried in an oven at 105 °C for 2 h. The alginate was maintained in a desiccator for one hour and weighed. The alginate content (purity) was computed as: (weight of the precipitated / weight of the initial alginate on dry basis $\times$ 100) (Reyes, 2005).

MODIFICATION OF THE OPTIMIZED CONTROL PROCESS WITH MINIMUM WATER CONSUMPTION (TCONT), CHANGING THE PRE-EXTRACTION ACID TREATMENT FOR A NEUTRAL PRE-COOKING STAGE (PCS)

To avoid the use of hydrochloric acid in the process and reduce the contamination from wastewater, experiments were performed with substitution of the acid pre-extraction by a neutral pre-cooking stage. The algae were treated in a water bath at 75 °C for 2 h, with constant agitation with an external motor (CAFRAMO®) and three-bladed propellers at 500 rpm. The algae were subjected to the following stages: hydration: H4 (1:4 alga:water); neutral pre-cooking: 1:7; extraction: Ext10 (1:10); dilution: Di10 (1:10); and filtration.

STATISTICAL ANALYSIS

Analysis of variance (ANOVA with $\alpha = 0.05$) was used to test the best process conditions, based on the alginate yield, viscosity (1 % solution) and gel strength, obtained in each experiment after modifying the consumption of water in each of the stage (Zar, 1984), included in the program STATISTICA version 7.0 (2004).

RESULTS AND DISCUSSION

HYDRATION, ACID PRE-EXTRACTION, ALKALINE EXTRACTION, DILUTION AND FILTRATION TREATMENTS

The statistical analysis showed no significant difference ($p < 0.05$) among any of the treatments for the first four steps. When varying the hydration volume, down to the proportion 1:4 (alga:0.1 % formaldehyde), the alginate yield (average 8.3 %), viscosity (average 89.3 mPa s) and gel strength (average 1611 g cm⁻²) were maintained without significant changes (Figure 1); therefore, the minimum amount for algae rehydration is one part of alga with four parts of formaldehyde solution. With this change, the formaldehyde solution, was reduced by 55 %.

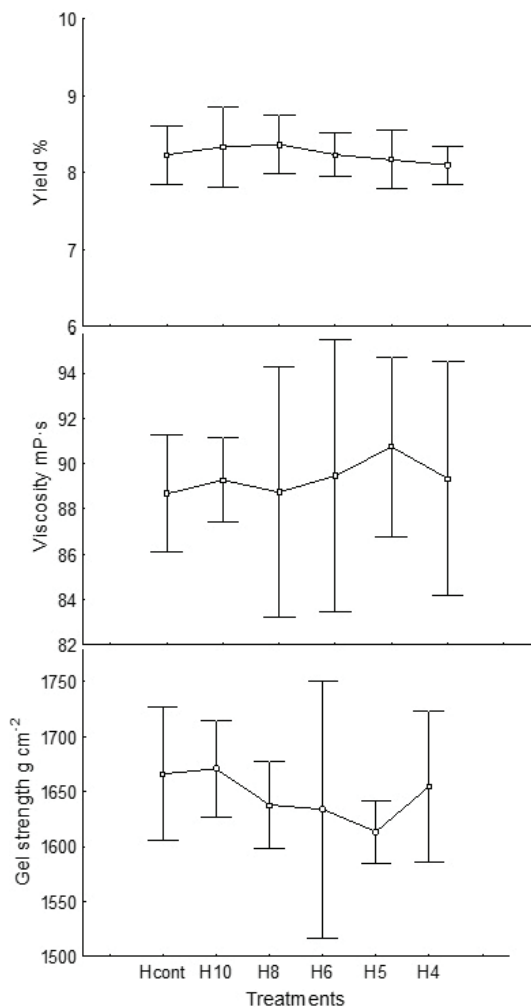


Figure 1. Effect of hydration volume with 0.1 % formaldehyde on the yield (%), viscosity (mPa s) (measuring to 60 rpm) and gel strength (g cm⁻²). Hcont: control (1:9 alga:water), H10: 1:10, H8: 1:8, H6: 1:6, H5: 1:5, H4: 1:4

No significant effect of reducing the volume of water down to the proportion 1:4 (alga:water) was found during the acid treatment, with no significant difference ($p < 0.05$) found for the yield 8–8.6 %, viscosity 89.6 mPa s or gel strength 1641–1668 g cm⁻² (Figure 2). Hernández *et al.* (2012) proposed that for *M. pyrifera*, one part of dry algae requires up to nine parts of water (1:9); however, for *S. horridum*, one part of algae only requires four parts of water (1:4). This can be explained because the cell wall does not allow excessive water entry; for this reason, even if more than four parts of water were used, *Sargassum* does not absorb any more. Haug (1964) suggested that formaldehyde is necessary to fix phenolic compounds that are in the cell wall, otherwise a dark brown alginate is produced. This reduction also reduces water contamination since less formalin is used at this stage. With the proposed rehydration volume, a saving of 55 % of water was achieved (Table 3).

ACID PRE-EXTRACTION

In this stage, ion exchange of Ca²⁺, Mg²⁺ and K¹⁺ occurs, and the phenolic compounds are eliminated. The reaction with the acid allows the alginate mixture to transform into alginic acid and prepares the algae for the alkaline extraction to facilitate the removal of soluble sodium alginate. Using one part of algae to five parts of water, a saving of 50 % is obtained. Also, 44 % of hydrochloric acid was saved and no significant differences in alginate yield, viscosity or gel strength were found.

ALKALINE EXTRACTION

In the alkaline extraction step, it was possible to reduce the water used from 1:15 to 1:10 without significant differences ($p < 0.05$), maintaining the average yield as 8.1–8.5 %, viscosity as 83.2 mPa s and gel strength as 1638–1664 g cm⁻² (Figure 3). Also, during this step, the amount of sodium carbonate was reduced by 22 %.

Before this stage, the algae do not yet show any type of change in color or shape, only the cell wall has been hydrated and is swelled. For the extraction step, the volume can be reduced to 10 parts of water for one part of dried seaweed since this species does not produce high viscosity and it is still possible to stir the solution at that viscosity. The water reduction represents a saving of 37 %. Reduction of the water volume allows a saving of 22 % of sodium carbonate. The extraction parameters proposed by Hernández *et al.* (2012) were maintained, which consisted of increasing the pH to 10, temperature of 80 °C and a time of 120 min. No significant differences in viscosity or gel strength of alginate were obtained by reducing the volume of water used. At the end of

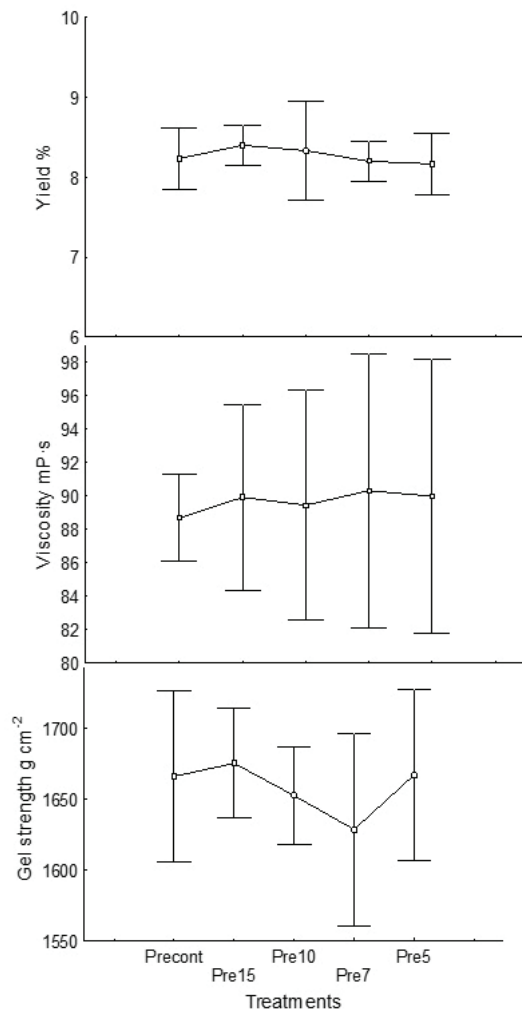


Figure 2. Effect of volume of water in the acid pre-extraction step on yield (%), viscosity (mPa s) (measuring to 60 rpm) and gel strength (g cm⁻²). Precont (1:20 alga:water), Pre15: 1:15, Pre10: 1:10, Pre7: 1:7, Pre5: 1:5

this treatment, the viscosity of the paste was 300–800 mPa s.

During dilution, the water was reduced from 1:20 to 1:10, maintaining ($p < 0.05$) the yield (8–8.6 %), viscosity (90.5 mPa s) and gel strength (1665–1678 g cm⁻²) (Figure 4); this volume saves 72 % of water in this step. The experimental dilutions did not present any significant difference in the parameters evaluated. Total water saving at this stage was 72 %. This is the step in which more water is saved since the viscosity that occurs after extraction is not as high as when *M. pyrifera* is processed. With *S. horridum*, the cell wall contains less alginate (8.6 %) (Di Filippo *et al.*, 2018) and the alginate produced is of lower viscosity. Therefore, to carry out the filtration, a smaller volume of water is needed for filtration (1:10). The total

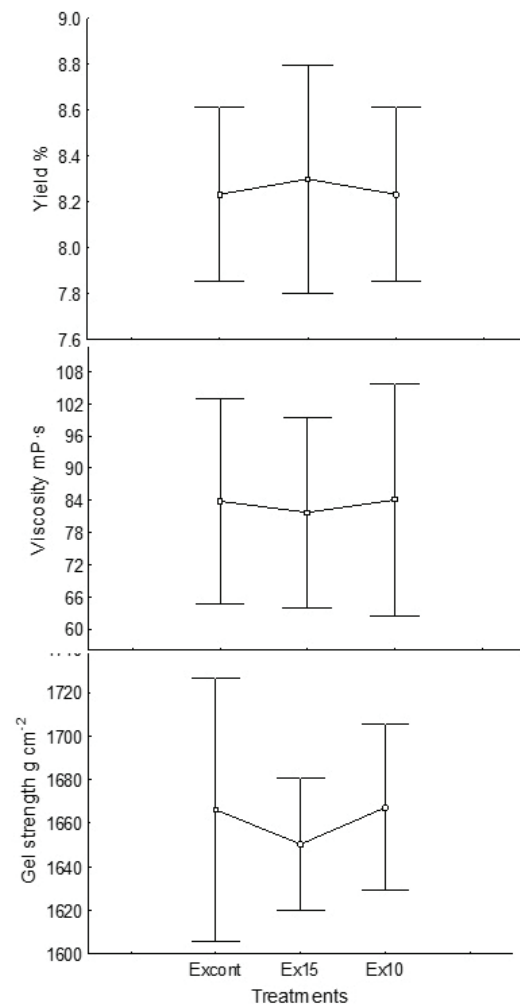


Figure 3. Effect of volume of water in the alkaline extraction step on yield (%), viscosity (mPa s) (measured at 60 rpm) and gel strength (g cm⁻²). Excont: control (1:20 alga:water), Ex15: (1:15), Ex10: (1:10)

water saving in the whole process with the modifications described above was 62.22 % (Table 1).

COMPARISON OF THE CONTROL (T_{CONT}) VS THE MODIFIED PROCESS USING LESS WATER AND CHANGING THE ACID PRE-EXTRACTION FOR NEUTRAL PRE-COOKING (T_{NEW})

The average alginate yield was significantly higher ($p > 0.05$) in the new process (T_{new}, 13.5 %) than in the control treatment (T_{cont}, 8.3 %) (Figure 5).

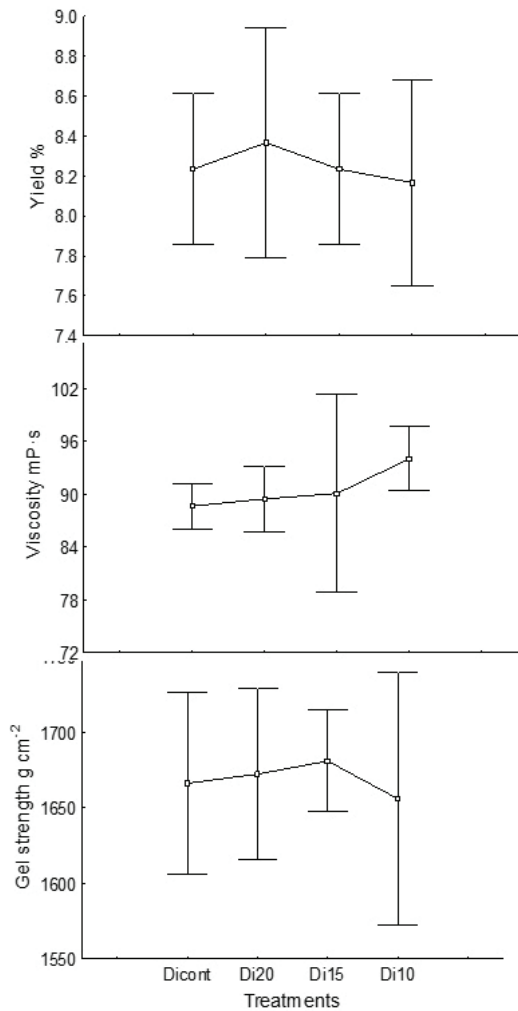


Figure 4. Effect of volume of water in the dilution step on yield (%), viscosity (mPa s) (measuring to 60 rpm) and gel strength (g cm^{-2}). Di:cont: control (1:55 alga:water), Di20: (1:20), Di5: (1:15), Di10: (1:10)

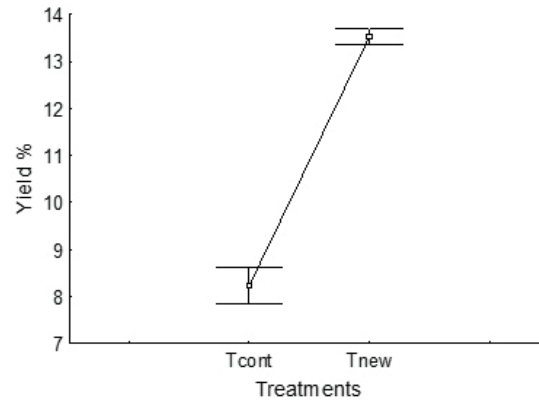


Figure 5. Comparison of yield in the control process (Tcont) and in the process using less water, changing the acid pre-extraction for neutral pre-cooking (Tnew)

For viscosity, there was no significant difference between the two processes; for Tnew on average it was 86.87 mPa s while with Tcont it was on average 88.67 mPa s (Figure 6).

The gel strength was significantly ($p > 0.05$) higher in the new process (Tnew 2442 g cm^{-2}) than the control (Tcont 1666 g cm^{-2}) because the optimized extraction conditions generate less depolymeration of the alginates, that is, the bonds are not broken and the polymer chains remain long, thus producing more rigid gels (Figure 7).

COLOR

Bleaching of the alginate with sodium hypochlorite (NaClO) resulted in a significant change in coloration,

Table 1. Comparison of the water used in the control process of Hernández *et al.* (2012), with *Macrocystis pyrifera* and the process using less water, calculated to process 1 kg of dried seaweed *Sargassum horridum*

Step	Control process		New process	
	Proportion	Volume of water (L)	Proportion	Volume of water (L)
Hydration	1:10	9	1:4	4
Acid pre-extraction	1:10	10	1:5	5
Alkaline extraction	1:16	16	1:10	10
Dilution	1:55	55	1:15	15
Total water used		90		34

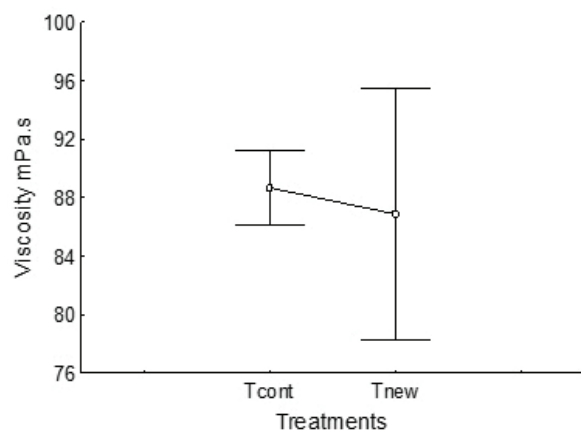


Figure 6. Viscosity in the control process (Tcont) and in the process using less water, changing the acid pre-extraction for neutral pre-cooking (Tnew)

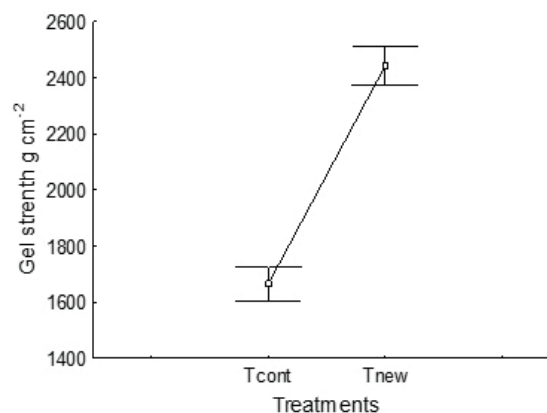


Figure 7. Comparison of gel strength in the control process (Tcont) and in the process using less water, changing the acid pre-extraction for neutral pre-cooking (Tnew)

based on the Munsell charts (2009), for different concentrations of NaClO (Table 2). Sodium alginate Sigma® was used as a control, since it has the lowest absorbance value (0.03) at a wavelength of 510 nm and is a low viscosity product. The treatments with sodium hypochlorite did not affect the alginate viscosity which was in the range 80–90 mPa s (Figure 8); the gel strength was not affected either, being maintained at high values of 2400 g cm⁻² (Figure 9).

CONTRASTING THE CONTROL PROCESS (TCONT) WITH THE MODIFIED PROCESS USING LESS WATER AND CHANGING THE ACID PRE-EXTRACTION FOR NEUTRAL PRE-COOKING (TNEW)

After verifying that reducing the volume of water in all the alginate extraction stages did not affect the rheological properties, experiments were conducted to change the acid pre-extraction stage for a neutral pre-cooking stage in which the algae were kept in a water bath at 75 °C, under constant agitation with an external motor at 500 rpm for 2 h.

The change in this stage had beneficial results. Neutral washing or neutral pre-cooking eliminates more phenolic compounds; this was seen in the wastewater,

Table 2. Color characteristics of alginates by absorbance and comparison with Munsell charts

Alginate solution		Absorbance	Munsell charts	
(1 %)			Hue	Color
Treatment with 5 % NaClO (mL)				
T1	0.0	2.609	7.5 YR (2.5/3)	Very dark brown
T2	0.5	1.499	5 YR (4/4)	Reddish brown
T3	1.0	0.849	5 YR (5/8)	Yellowish red
T4	2.0	0.232	10 YR (6/6)	Yellowish brown
T5	3.0	0.196	2.5 Y (7/4)	Pale brown
AlgCICIMAR®		0.068	10 YR (8/4)	Very pale brown
Sigma®		0.03	2.5 Y (8/1)	Transparent white

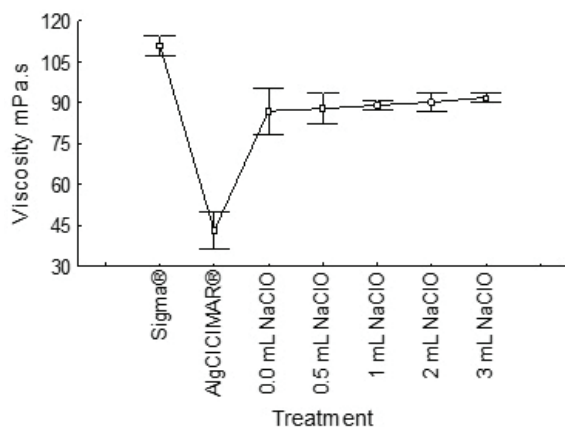


Figure 8. Comparison of the viscosity of alginates for different treatments with sodium hypochlorite in the bleaching stage, the measurements were made with a viscometer NDJ-1 at 60 rpm, at a constant deformation gradient

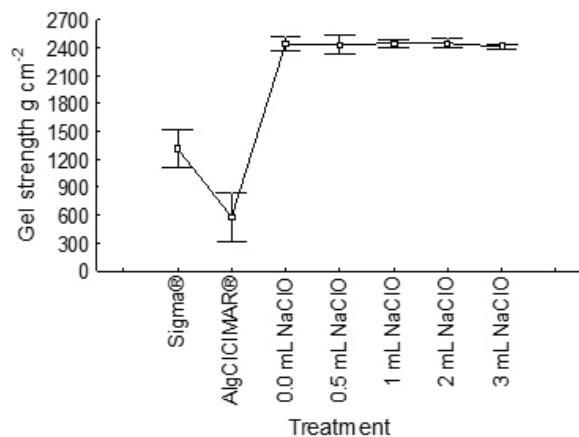


Figure 9. Comparison of the gel strength of alginates for different treatments with sodium hypochlorite in the bleaching stage

since it had a darker brown color and higher viscosity, compared to the process with acid. This washing step was carried out with seven parts of water for one part of seaweed.

YIELD

The results were beneficial using this new process, increasing the yield from 8.3 % to 13.5 % (62 %). This yield was higher than that reported by Di Filippo *et al.* (2018) and the maximum yield of 13.7 % reported by Rodríguez *et al.* (2008) for *S. sinicola*.

The yield in the present study with the pre-cooking modification had a maximum of 13.5 %; however, it was lower than for other species such as *S. ilicifolium* (23 %), *S. carpophyllum* (32 %), *S. siliquosum* (41 %) (Ragaza & Hurtado, 1999), *S. tenerrimum* (18.32 %) (Redekar & Rajee, 2000), *S. oligocystum* (20.5 %), *Sargassum* sp.

(20.9 %) and *S. filipendula* (17 %) (Camacho & Hernández, 2012). The yields obtained in this study with the modification of neutral pre-cooking are higher than those reported for *Dictyota caribaea* (7.4 %), *Padina perindusiata* (5.4 %) (García *et al.*, 2012) and those for *S. horridum* that were carried out in the same season of the year by Di Filippo *et al.* (2018).

Neutral pre-cooking helps to soften the cell wall, giving the alginate a better chance of being successfully extracted; it also improves the color since at this stage many phenolic compounds are removed, and the final alginate is lighter than that produced with acid pre-extraction.

VISCOSITY

For the process with less water and neutral pre-cooking, a normal low viscosity was maintained, ranging between 82 and 89 mPa s. This result is similar to those reported for *Sargassum* by Rodríguez *et al.* (2008) who obtained viscosity of 41–191 mPa s, and higher than the values reported by Di Filippo *et al.* (2018) of 38 to 73 mPa s, and by Camacho and Hernández (2012) of 12 to 21 mPa s. Although the yields were different, the viscosity were not significant different among treatments. This suggest that, under the extraction parameters, the alginate chain remain with approximated the same length and therefore the pre-cooking has no significant effect on the viscosity.

The viscosity of alginates used in the food and pharmaceutical industries is variable, and there is some demand for alginates with low viscosity, for example, as a stabilizer for bottled beverages, for gelatinizing, thickening, deflocculating and in cream ice creams, among other products. In the pharmaceutical industry, alginates are used to stabilize cosmetics such as face masks, and products that remove heavy metals from the human body. Therefore, use could be found for *S. horridum* alginates with varying viscosity, according to the industry needs.

GEL STRENGTH

The process using less water and neutral pre-cooking showed a significant positive effect on gel strength, with high values (2416 to 2472 g cm⁻²) exceeding all previous reports, including that of Di Filippo *et al.* (2018) who reported a minimum in June (815 g cm⁻²) and maximum in May (1906 g cm⁻²). Lower values (709 to 866 g cm⁻²) were reported by Camacho and & Hernández (2012). Draget *et al.* (2002) suggested that the alginate from *S. sinicola* has better stability than *M. pyrifera* alginate, because of the higher quantity of the guluro-

nic acid units and biaxial bonds (GG) of the polymer chain. The increased gel strength, compared to the control process, may be due that cooking process preserved the length of the guluronic acid chains, which are related to the production of stronger gels (Murillo & Hernández, 2007).

Alginates with a high gel strength (above 750 g cm^{-2}) have a wide range of industrial uses. Murillo & Hernández (2007) showed that GG blocks predominate in the alginate from *Sargassum*; for this reason, the gels produced have greater strength. An example of use is in microcapsules, used for the slow release of products, which need a high gel strength to maintain the stability of beneficial bacteria, and thus improving the growth of land plants (Yabur *et al.*, 2007).

COLOR

The color that best suits the industry is that of alginates produced from treatment with 3 mL of sodium hypochlorite. According to Munsell (2009), the color obtained is classified as pale brown; although it is not as white as the alginate from *M. pyrifera*, it is very close.

CONCLUSIONS

The optimized method to process the alga *Sargassum* to obtain sodium alginate includes the following steps and suggested proportions of alga:water: hydration: 1:4; neutral pre-cooking: 1:7 (75°C , 2 h); extraction: 1:10 (pH to 10, 80°C , 120 min); dilution: 1:10; and vacuum filtration. Alginate precipitation uses 37.5 mL of 10 % calcium chloride for every 10 g of dry alga. Calcium alginate fibres are bleached with 3 mL sodium hypochlorite per 10 g of alga. Conversion to alginic acid uses three washes of 10 % hydrochloric acid at a ratio of 1:10 until pH 2; the second and third washes use a proportion of 1:5, adjusting the pH to 1.8 with constant stirring for 15 min. Neutralization to sodium alginate is carried out in 1:1 alcohol:water and the pH is adjusted to 10 with sodium carbonate. Finally, alginate fibres are pressed, dried, and milled to dust.

Reducing the volume of water in all the alginate extraction steps does not affect the rheological properties. This new process increases the yield from 8.3 % to 13.5 % (62 %). Neutral pre-cooking helps to soften the cell wall, and the alginate is better extracted; it also improves the colour, since at this stage some phenolic compounds are removed, and the final alginate is lighter than when using the acid pre-extraction.

ACKNOWLEDGEMENTS

Hernández-Carmona G., Rodríguez-Montesinos Y. R., & Arvizu-Higuera D. L. thanks to the Instituto Politécnico Nacional for the financially supporting to carry out this research, the Beca Exclusividad (COFAA) and Estímulo al Desempeño de los Investigadores (EDI). Loa-Ramírez I. A., thanks to Dr. Cervantes-Hernández Pedro and Universidad del Mar, Puerto Ángel Oaxaca for the advice provided.

REFERENCES

- Armisen, R., & Galatas, M. (1987). Producción, properties and uses of agar. In McHugh D. J. *Production and utilization of products from commercial seaweeds* (p. 194). Roma: FAO Fisheries Technical Paper.
- Arvizu, D. L., Hernández, G., & Rodríguez, Y. E. (2002). Parámetros que afectan la conversión de ácido alginico a alginato de sodio. *Ciencias Marinas*, 28(1), 27-36.
- Camacho, O., & Hernández, G. (2012). Phenology and alginates of two *Sargassum* species from the Caribbean coast of Colombian. *Ciencias Marinas*, 38(2), 381-393. <https://doi.org/10.7773/cm.v38i2.1963>
- Casas, M. M., Fajardo, C., Sánchez, I., Hernández, G., & Rodríguez, Y. E. (1990). Evaluación de *Sargassum* spp. en la Bahía de La Paz, B.C.S., México. *Investigaciones Marinas CICIMAR*, 5(1), 11-18. <http://www.repositoriodigital.ipn.mx/handle/123456789/14499>
- Di Filippo, D. A., Hernández, G., Muñoz, M., Arvizu, D. L., & Rodríguez, Y. E. (2018). Monthly variation in the chemical composition and biological activity of *Sargassum horridum*. *Botanica Marina*, 61(1), 91-102. <https://doi.org/10.1515/bot-2017-0031>
- Dragnet, K. I., Smidsrød, O., & Skjåk-Bræk, G. (2002). Alginates from algae. In A. Steinbüchel, S. De Dantes, & Vandame E. J. *Biopolymers*, (6) 636. Wiley-VCH, Weinheim.
- García, V., Ríos, E., Robledo, D., & Freile, Y. (2012). Polysaccharides composition from tropical brown seaweeds. *Phycological Research*, 60, 305-315. <https://doi.org/10.1111/j.1440-1835.2012.00661.x>
- Haug, A. (1964). Composition and properties of alginates. Report No. 30. Norwegian Institute of Seaweed Research.
- Hernández, G., McHugh, D. J., Arvizu, D. L., & Rodríguez, Y. E. (1999). Pilot plant scale extraction of alginate from *Macrocystis pyrifera*. Part 1. The effect of pre-extraction treatments on the yield and quality of alginate. *Journal of Applied Phycology*, 10(6), 507-513.
- Hernández, G., Rodríguez, Y. E., Arvizu, D. L., Murillo, J. I., Muñoz, M., & Reyes, R. (2012). Avances tecnológicos en la producción de alginatos en México. *Ingeniería Investigación y Tecnología*, XIII(2), 155-168. <https://doi.org/10.22201/fi.25940732e.2012.13n2.015>
- Munsell. (2009). *Munsell soil colour charts*. NY: Gretag Macbeth.
- Murillo, J. I., & Hernández, G. (2007). Monomer composition and sequence of sodium alginate extracted at pilot plant scales

- from three commercially important seaweeds from México. *Journal of Applied Phycology*, 19(5), 545-548. <https://doi.org/10.1007/s10811-007-9168-5>
- Ragaza, A. R., & Hurtado, A. Q. (1999). Seasonal variations in alginate yield and viscosity of *Sargassum carpophyllum* J. Agardh, *Sargassum ilicifolium* (Turner) C. Agardh and *Sargassum siliquosum* J. Agardh (Phaeophyta, Sargassaceae). *Botanica Marina*, 42, 327-331.
- Redekar, P. D., & Raje, P. C. (2000). Alginic acid and agar-agar content from the common seaweeds of Ratnagiri Coast (West Coast of India). *Seaweed Research*, 22, 41-43.
- Reyes, R., Hernández, G., Rodríguez, Y. E., & Arvizu D. L. Food grade alginates extracted from the giant kelp *Macrocystis Pyrifera* at pilot plant scale. *Revista de Investigaciones Marinas*, 26:185-192, 2005.
- Rodríguez, R. E., Van, B., & Jordán, E. (2016). Afluencia masiva de Sargazo pelágico a la costa del Caribe mexicano (2014-2015). In García E. *Florecimientos algales nocivos en México*, 352-365. Ensenada: CICESE.
- Rodríguez, Y. E., Arvizu, D. L., & Hernández, G. (2008). Seasonal variation on size and chemical constituents of *Sargassum sinicola* Setchell from Bahía de La Paz, Baja California Sur, México. *Phycology Research*, 56(4), 33-38. <http://dx.doi.org/10.1111/j.1440-1835.2008.00482.x>
- Yabur, R., Bashan, Y., & Hernández, G. (2007). Alginate from the macroalgae *Sargassum sinicola* as a novel source for microbial immobilization material in wastewater treatment and plant growth promotion. *Journal of Applied Phycology*, 19(1), 43-53.
- Zar, J. H. (1984). *Biostatistical Analysis*, 2nd ed. Englewood Cliffs, New Jersey: Prentice-Hall.

Cómo citar:

Loa-Ramírez I. A., Rodríguez-Montesinos Y. E., Arvizu-Higuera D. L., & Hernández-Carmona G. (2022). Adaptation of the alginate production process for the brown alga *Sargassum horridum* (Fucales: Sargassaceae). *Ingeniería Investigación y Tecnología*, 23 (02), 1-10. <https://doi.org/10.22201/fi.25940732e.2022.23.2.010>