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Taxonomy and systematics

Redescription of the velvet worm Oroperipatus eisenii (Onychophora: Peripatidae), through DNA sequencing, scanning electron microscopy and new collection records from Western Mexico

Gerardo A. Contreras-Félix a, *, Griselda Montiel-Parra b, Fabio G. Cupul-Magaña c, Tila M. Pérez b

a Colección Nacional de Arácnidos (CNAN), Departamento de Zoología, Instituto de Biología, Universidad Nacional Autónoma de México, 3er Circuito exterior s/n, Ciudad Universitaria, Coyoacán, 04510 Ciudad de México, Mexico
b Colección Nacional de Ácaros (CNAC), Departamento de Zoología, Instituto de Biología, Universidad Nacional Autónoma de México, 3er Circuito exterior s/n, Ciudad Universitaria, Coyoacán, 04510 Ciudad de México, Mexico
c Centro Universitario de la Costa, Universidad de Guadalajara, Av. Universidad 203, Delegación Ixtapa, 48280 Puerto Vallarta, Jalisco, Mexico

*Corresponding author: contrerasfelixga@gmail.com (G.A. Contreras-Félix)

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Abstract

Specimens of Oroperipatus eisenii (Wheeler, 1898) were re-examined, including 13 new records for the species collected in the states of Nayarit, Jalisco, and Michoacán in Mexico. Scanning electron micrographs that illustrate most of the diagnostic characters were provided allowing a redescription of the species, as well as a map of the localities where the collections were carried out. DNA sequences were obtained from the collected specimens for the cytochrome C oxidase I (COI) gene; results of the similarity analysis showed that there was no genetic divergence for COI within the sampled onychophoran populations. Natural history information of the species is included.

Keywords: Tropical forest; North America; Biodiversity; Species delimitation; Speciation

Resumen

En este trabajo se reexaminaron ejemplares de Oroperipatus eisenii (Wheeler, 1898) y se presentan 13 registros nuevos de la especie en los estados de Nayarit, Jalisco y Michoacán, en México. Se proporcionan imágenes de microscopio electrónico de barrido que ilustran la mayoría de los caracteres diagnósticos y se incluye la redescription de la especie, así como un mapa que muestra las localidades de recolecta. Se obtuvieron secuencias del gen citocromo C oxidasa I (COI) a partir de los ejemplares recolectados; los resultados del análisis de similitud muestran que no
existe diferenciación genética entre las poblaciones de *O. eisenii* estudiadas. También se presenta información sobre la historia natural de la especie.

**Palabras clave:** Bosque tropical; Norteamérica; Biodiversidad; Delimitación de especies; Especiación

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**Introduction**

Velvet worms are terrestrial invertebrates traditionally known from studies on historical biogeography, conservation biology, and evolution (Allwood et al., 2010; Murienne et al., 2014). Their populations are usually low density in most habitats, so they are labeled as “rare” (Monge-Nájera & Morera-Brenes, 2015). Velvet worms are more common in tropical and temperate forests of the Southern Hemisphere and near the equator; these animals are more diverse and better known in southern locations than in the northern counterparts (Mayer et al., 2015; Oliveira, Read et al., 2012).

Several contributions on onychophoran in Mexico, Central America, and the Caribbean have recently appeared, but the species of Costa Rica are among the most studied taxonomically and in other fields such as morphology (Monge-Nájera & Morera-Brenes, 1994; Oliveira, Franke et al., 2012); physiology (Mayer & Harzsch, 2007; Monge-Nájera & Morera-Brenes, 1994); ecology (Monge-Nájera, 1994; Monge-Nájera & Alfaro, 1995); behavior (Monge-Nájera et al., 1993); biophysics (Concha et al., 2015); biochemistry (Baer et al., 2014); genetics and genomics (Jeffery et al., 2012; Morera-Brenes et al., 1992; Podsiadlowski et al., 2008); phylogeography (Morera-Brenes & Monge-Nájera, 2010; Murienne et al., 2014); taphonomy (Monge-Nájera & Xianguang, 2002); zoaanthropology (Monge-Nájera & Morera-Brenes, 2015); conservation (Monge-Nájera, 2018; Sosa-Bartuano et al., 2018); and digital photography (Barquero-González et al., 2016).

So far, 206 species of velvet worms have been described, of which 186 are valid species. The rest have been cataloged as *nomina dubia* due to taxonomic inconsistencies (Oliveira, Read et al., 2012; Oliveira et al., 2015, 2018). The diversity of velvet worms in Mexico is poorly known; only 3 species belonging to the neotropical Peripatidae have been reported. The first description of *Oroperipatus goudoti* (Bouvier, 1899) contained imprecise data, which only mentions “Mexico” as the collection locality and has been cataloged as *nomina dubia*. The second, *Macroporipatus perrieri* (Bouvier, 1899), has an original label that does not provide details specifying if it was found in the state of “Veracruz” (sic) or the city of “Veracrus”. The third species *Oroperipatus eisenii* (Wheeler, 1898), has the best locality description from Tepec, Nayarit. There are DNA sequences from unidentified onychophorans from the states of Chiapas (Murienne et al., 2014), and Campeche (BOLD Systems, 2014). Toledo-Matus et al. (2018) reported an undescribed species of *Oroperipatus* from the deciduous forest of Tuxtla Gutiérrez, Chiapas. Monge-Nájera (2018) reports specimens unidentified from the state of Nuevo León.

The species *Oroperipatus eisenii* was described in the original paper based on 87 specimens from Tepec, Nayarit (Wheeler, 1898). Later, Rucker (1900) described the external and internal morphology of males from the same locality. The information about the deposit of type specimens is scarce in both studies. A pair of specimens labeled as syntypes is conserved in the Muséum National d’Histoire Naturelle in Paris, France (catalog number MY112, and MY113). No other information on the whereabouts of the rest of the material was published.

As part of this research, specimens of *O. eisenii* were reviewed at the American Museum of Natural History (AMNH), in New York. Several specimens had the label of Tepec (sic), Nayarit, Mexico. Knowing that William Wheeler was one of the first curators of this invertebrate collection (Parker, 1938), these specimens are presumably syntypes of the species, as they have a label by Monge-Nájera indicating so. Subsequent records of this species were reported from Rio Purus, Brazil (Fuhrmann, 1914), and El Cermeño, Panama City (Clark & Zetek, 1946), but these records require confirmation. No more information about *O. eisenii* was published until 2008. Five specimens were collected, although their sex was not determined, in a suburban area in the state of Jalisco in Western Mexico (Cupul–Magaña & Navarrete-Heredia, 2008), situated about 100 km from the type locality. Oliveira, Read et al. (2012) made an extensive checklist of the velvet worms of the world and suggested that the specimens and species required a detailed revision to discard a species misidentification. In the present work, a redescription of the species *O. eisenii* is done based on freshly collected specimens between 2014 and 2017 in several locations from the Mexican states of Jalisco, Nayarit, and Michoacán. The first scanning electron micrographs of this species are provided. Additionally, new DNA COI sequences from most localities were obtained and used in the analysis.
Materials and methods

The velvet worms were collected under scientific permits FAUT-0027 and FAUT-0029 from Secretaría del Medio Ambiente y Recursos Naturales (Semarnat) in the Mexican states of Nayarit, Jalisco, and Michoacán. All specimens were preserved in 96% ethanol and then examined morphologically. The specimens were deposited in the recently created “Onychophora Collection”, associated to the Colección Nacional de Acaros (CNAC), Universidad Nacional Autónoma de México. Others were deposited at the Centro de Estudios en Zoología (CZUG), Universidad de Guadalajara, Jalisco, Mexico; the American Museum of Natural History (AMNH), New York, USA, and the Museum of Comparative Zoology (MCZ), Harvard University, Cambridge, MA.

The classification and terminology of morphological structures followed the works of Oliveira et al. (2010), Oliveira, Franke et al., (2012). The measurements of size were taken with an ocular micrometer calibrated at 10X and are given in millimeters. Photographs of a live specimen were taken with a Nikon D5500 digital camera with AF-S Micro Nikkor 105 and ring flash Metz 15 Ms-1 digital. The microphotographs were obtained with a Leica Z16 APOA microscope and a Leica DFC490 camera; they were processed with the Leica Application Suite E Program. Other microphotographs were obtained with an Axio Zoom V16 microscope including the Zeiss AxioCamMRc5 camera; and were processed with the Zeiss Efficient Navigation program. Scanning electron microscopy examination and photography were carried out with a Hitachi S-4700 Field Emission at the American Museum of Natural History, following the methodology by Ruhberg and Daniels (2013). Pictures were edited with the Adobe Photoshop CS6 software. The geographic map was generated with ESRI ArcGIS online suite.

The genomic DNA extractions were obtained following the procedure of the Qiagen DNeasy Blood and Tissue Kit by soaking a whole segment of each velvet worm from the juvenile specimens and a small part of the body from the adult specimens. In both cases, the middle portion of the body was used to obtain the tissue. The Cytochrome C Oxidase I sequence (COI) was amplified through polymerase chain reaction (PCR) using the following combination of primers HCO: TAAACTTCAACGGTGCACA AATCA and LCO: GGTCGAACCAACTTAAAGATATGG. For PCR, the annealing temperature was 46°C. Edition of the sequences was carried out with Bioedit (Hall, 2005) and genetic distances measurements were obtained with MEGA7 (Tamura et al., 2013). DNA sequences were deposited in GenBank under accession numbers (MH454602, MH454603, MH454604, MH454605, MH454606, MH454607, MH454608, MH454609).

Redescription

Family Peripatidae Evans, 1901.
Genus Oroperipatus (Cockerell, 1908).
Oroperipatus eisenii (Wheeler, 1898).

Taxonomic summary

Synonyms: Peripatus eisenii, original name (Wheeler, 1898), Oroperipatus eiseni (sic) (Clark, 1913).

Holotype: not designated.

Type locality: Tepic, Nayarit, Mexico.

Syntype: Tepec (sic), Mexico (AMNH 416). Part of the type series described by Wheeler in 1898, without further information; probably collected by Eisen.

Notes: the species name is commonly misspelled as “eiseni” (Clark, 1913; Fuhrmann, 1914; Peck, 1975; Sampaio-Costa et al., 2009; Vasconcellos et al., 2004).


**Diagnosis:** both sexes of *O. eisenii* present a dark brown coloration on the dorsum, and a light whitish coloration on the ventral face. Each segment of the body presents 10 complete plicae; the primary papillae present 4 ranks of scales on the base and 4 ranks of longer scales on the apical portion. The antennae present 35 rings on the base and have a terminal button followed by 9 rings. The structure of the inner jaw blade presents a diastema that separates the single principal tooth and the 2 accessory teeths, from 6 to 7 denticles, the second ones are bilobated. The nephridial tubercle is on the fourth and fifth pair of legs; each leg presents 4 spinous pads. Finally, males present a prominent crural tubercles on legs 21 and 22.

**Description:** maximum body size after fixation: length: 17 to 39 mm, and width: 2 to 5 mm in adults; length: 14 to 18 mm, and width: 2 mm in young. Color pattern: coloration of dorsal integument (*in vivo*) is dark brown without any discernible pattern. The dorsal midline is darker than the rest of the integument (Fig. 1). The ventral body surface is mainly pale reddish, brighter than the dorsal integument (Fig. 2).

**Body dorsum:** the dorsomedical furrow is very well defined; without the presence of any hyaline organs. Each body segment has 10 plicae. All plicae have the same width along the body (Fig. 3). Primary and secondary papillae are present on each plicae (Fig. 4). There are never 2 primary papillae together, they always have at least 1 secondary papillae separating them. The primary papillae are aligned longitudinally, with 1 plicae between each primary papillae (Fig. 4). Both papillae occur on the top of the plicae, there is never more than a single line of papillae for each plicae. Primary papillae present a wider, rounded shape at the base, and are shorter than the apical portion (Fig. 5); the apical portion has 4 scale ranks, and these are noticeable longer than the ones on the basal portion. The sensory bristle is on the primary papillae and is larger than the scales (Fig. 5). Accessory papillae are similar in shape to primary papillae. They almost maintain a proportion of 2 to 1 with respect to the primary papillae.

**Body venter:** females have 28 or 29 pairs of legs; males and juveniles have 24 pairs of legs. The legs have 3 complete spinous pads and a smaller one which is incomplete. They are situated distally on the ventral surface of each leg (Fig. 6). The nephridial tubercle is located on the posterior part of the third spinous pad, dividing it in 2. It is present on the fourth and fifth legs (Fig. 7). Each leg has several finger-like foot papillae around the claw, and 1 papillae situated on the dorsal face, between the claws (Fig. 8). Ventral and preventral organs are absent or not evident. The genital opening is located near the penultimate pair of legs; this is a characteristic of

Figures 3-5. *Oroperipatus eisenii*. 3. Dorsal view showing the plicae of the same width all along the body. 4. Overview of the dorsal integument of female, showing primary (pp) and secondary papillae (ps) distribution pattern. 5. Detail of primary papillae showing of sensory bristle (sb), apical piece (ap) and basal (bp).
the Peripatidae family; it consists of 2 parallel lobes on each side, higher than the ventral integument (Fig. 9). Two pregenital legs have 2, big and prominent crural tubercles only found in males (Fig. 10).

Head: the arrangement of the dermal papillae on the head is irregular. The eyes are present behind the antenna and the frontal organs on the ventrolateral base of the antenna.

Antennae: Antennal tip has a terminal button followed by 9 rings equal in size; the antennal body has a total of 35 rings in both sexes. The 19 rings at the base have the presence of finger-like sensillae; 10 rings are wide and 9 are thin (Fig. 11). Frontal organ appears to begin on the eleventh ring of the antennal body to the base (Fig. 12). Mouth: opening is surrounded by lip papilla, the anterior-most is unpaired and smaller, and is followed by 7 pairs of lobes posterior ones, equal in size; all of them have small spines (Fig. 13); the spines and the lip papilla are noted by Wheeler (1898) on the drawings made in the original description. Inner and outer jaw blades, both have an enlarged principal tooth, and 1 or 2 smaller accessory teeth (Figs. 14, 15). The structure of the inner jaw blade, presents a diastema that separates the single principal tooth and the 2 accessory teeths, from 6 to 7 denticles, the second ones are bilobed (Fig. 15).

Remarks

The species was only known from its type locality (Tepic, Nayarit) as well as from a suburban area in the State of Jalisco, situated about 100 km from the type locality (Cupul-Magaña & Navarrete-Heredia, 2008). Oliveira, Read et al. (2012) suggested that the specimens of Jalisco required a revision. In this work, we provide morphological and molecular information of the specimens from Nayarit and Jalisco. Our results show that the specimens belong to the same species *O. eisenii*. Therefore, the identification carried out by Cupul-Magaña and Navarrete-Heredia (2008) was corroborated.

Geographical distribution: after our study, this species is now known to be present in 13 localities of 3 Mexican States: Jalisco, Nayarit, and Michoacán. Its distribution has a range of approximately 400 km (Fig. 16), including altitudes from the sea level to almost 1,600 m, with a preference for tropical habitats usually found close to water bodies.

Natural history: most of the specimens were found under stones, rotten logs, or leaf litter, close to mango (*Manguifera indica*), yaca (*Artocarpus heterophyllus*), and banana (*Musa paradisiaca*) plantations; usually in shady places in groups of 2 to 4 specimens (Fig. 17). Also, they were found in anthropogenic environments under bricks, stones, leaf litter, rotting wood, garbage, flowerpots, and crevices of the soil close to the roots of small plants (Fig. 18). These findings are added to the ones summarized by Monge-Nájera (2018) of peripatids occurring in urban areas of Latin America and Caribbean. The climate was between 20 °C to 23 °C and 46% to 50% relative humidity. During the fixation procedure and captivity of *O. eisenii*, it was common for females to have abortions. In some females under captivity, the parturition sequence was documented and will be reported in detail elsewhere; in short, during previous days the females showed movements of raising.

Figures 6-7. Oroperiatus eisenii. 6. Antennae lateral view, showing the antennal body (ab), antennal tip (ti) and terminal button (tb). 7. Antennae frontal view; showing the finger-like papillae (white arrow).
the anal region, the neonate came out with the antennas retracted and started moving after the fifth pair of legs was out; the newborn worms stayed with the mother for a few hours. The parturition sequence was relatively consistent requiring around 40 min. During the first weeks, the color of the juveniles was lighter than the females. There can be a difference of 3 to 14 days between each birth. The dissection of some gravid females showed up to 10 embryos in different stages of development.

Discussion

The genetic marker COI has been shown to be a reliable marker for species delimitation and differentiation of populations, in previous studies of velvet worms (Daniels et al., 2009; Lacorte et al., 2011; McDonald et al., 2012; Oliveira et al., 2011). Therefore, variations obtained within the present populations of *O. eisenii*, did not fulfill the criteria for erection of a new species of *Oroperipatus*. It is well known, that velvet worms present low vagility (Allwood et al., 2010), and the morphological and molecular similarities found among the specimens from the different sampled areas for the present study, suggested that the populations have been recently separated; due to the fact that genetic divergence between the closest populations, separated by around 10 km (Puerto Vallarta and Centro Universitario de la Costa), is the same divergence found on specimens that are separated by a much longer distance, about 170 km and 93 km (Puerto Vallarta, Acatlán de Juárez, and outside El Naranjo Cave) (Table 1).

Since the description of *O. eisenii*, 120 years ago, this species has been collected in small numbers. Regarding the finding of the specimens in household locations, it is conceivable that these populations, are present in low densities (from 2 to 26 specimens per locality) in consequence of intensive human activities; almost half of our records were close to urban areas (courtyards and vacant lots). These findings support the hypothesis proposed by Monge-Nájera (2018) that places onychophorans living in urban and suburban areas in the
category of “survivors of habitat modification.” We agree with the model he presented that explains the persistence of the onychophoran populations found in urban areas is due to nutrient abundance, artificial abiotic factors, and elimination of predators and pathogens, among others. However, the cause-effect relationship of such activities has not been quantified and requires further research. For now, onychophorans are still hard to find.

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Table 1

The p-distances recorded between the different populations of velvet worms of the species Oberperipatus eisenii, from 8 localities in 2 states of Mexico, and based on sequences of the mitochondrial COI gene. Numbers in bold are the mayor genetic distances observed in all the samples. Locality of the samples: Jalisco: 1. Puerto Vallarta, 2. Acatlán de Juárez, 9. Centro Universitario de la Costa, 10. Carretera Cihuatlán-Puerto Vallarta, 11. Unión de Tula, and 12. Tequila. Nayarit: 5. Outside El Naranjo Cave, and 16. Mesillas.

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Figures 11-12. Oberperipatus eisenii. 11. Detail of the spinous pad on the leg, showing the four segments (I – IV) composing the whole spinous pad. 12. Leg IV, ventral view, showing the nephridial tubercle (N) on the third spinous pad.

Figure 16. Geographic distribution of the Oroperipatus eisenii, illustrating the type locality (red circle), the new records for the state of Nayarit (black circles); new records for the state of Jalisco (blue circles); and a new record for the state of Michoacán (yellow circle).
Figure 17. Picture of the localities with specimens in situ of Oroperipatus eisenii. Nayarit: A) Jalcocotán, B) río Tetepozco, C) Mesillas, D) nacimiento de Agua Pantanal. Jalisco: E) Centro Universitario de la Costa, F) carretera Cihuatán-PuertoVallarta.

References


