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## Ecology

### Population traits and reproduction of the hippolytid shrimp *Tozeuma carolinense* (Decapoda: Caridea) at Laguna de Términos, Campeche, southwestern Gulf of Mexico

#### *Características poblacionales y reproducción del camarón hipolítido *Tozeuma carolinense* (Decapoda: Caridea) en la laguna de Términos, Campeche, suroeste del golfo de México*

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#### Abstract

This study explored the population biology and reproduction of *Tozeuma carolinense* in Laguna de Términos, Campeche, southwestern Gulf of Mexico, collected bimonthly from June 2009 to June 2010. The sample of 2,109 specimens analysed had a sex ratio of 0.39 males per female. On average, females were significantly larger ( $6.16 \pm 2.02$  mm carapace length, CL) than males ( $4.42 \pm 0.70$  mm CL). Recruitment was continuous, with the highest percentages of juveniles during June 2009 and June 2010. The estimated asymptotic length with the von Bertalanffy model was 10.13 mm CL for females and 6.62 mm CL for males. Longevity was 6 months for females and 5 months for males, with males having a higher mortality ( $Z = 1.08$ ) than females ( $Z = 0.85$ ). The reproduction was continuous, with highest incidence of berried specimens from August 2009 to April 2010. The estimated size at sexual maturity for females was  $6.67 \pm 1.30$  mm CL. There was a significant correlation between female size and the number of embryos, with a mean fecundity of  $268 \pm 131$  embryos. During the incubation period females lost 46% of the embryos.

**Keywords:** Hippolytidae; Growth; Sexual maturity; Fecundity; Embryo loss

#### Resumen

Se examinó la biología poblacional y reproducción de *Tozeuma carolinense* en la laguna de Términos, Campeche, suroeste del golfo de México, de especímenes recolectados bimestralmente de junio de 2009 a junio de 2010. Se

capturaron 2,109 individuos de los que se calculó una proporción de 0.39 machos por hembra. Las hembras fueron más grandes ( $6.16 \pm 2.02$  mm de longitud de caparazón, LC) que los machos ( $4.42 \pm 0.70$  mm LC). El reclutamiento fue continuo, observándose el mayor porcentaje de juveniles durante junio de 2009 y junio de 2010. La longitud asintótica estimada con el modelo de von Bertalanffy fue 10.13 mm CL para hembras y 6.62 mm CL para machos. La longevidad se estimó en 6 meses para hembras y 5 meses para machos, los cuales tuvieron mayor mortalidad ( $Z = 1.08$ ) que las hembras ( $Z = 0.85$ ). La reproducción fue continua, incrementándose entre agosto de 2009 y abril de 2010. La talla estimada de madurez sexual en hembras fue  $6.67 \pm 1.30$  mm LC. La fecundidad media fue  $268 \pm 131$  embriones y se correlacionó significativamente con la talla de las hembras. Durante la incubación, las hembras perdieron 46% de los embriones.

*Palabras clave:* Hippolytidae; Crecimiento; Madurez sexual; Fecundidad; Pérdida de embriones

## Introduction

Population traits and reproductive biology are important in studies of crustaceans since they provide an approximation of the ecology and life history of any population (Alon & Stanczyk, 1982; Corey & Reid, 1991). Sex ratio and fecundity indicate the population structure and its reproductive potential, while embryo size and volume reflect the action of selective pressures on reproductive effort and larval development (Bauer, 1991). Therefore, it is necessary to gather information about species from diverse habitats, latitudes and depths in order to distinguish between phylogenetic and environmental sources of variation in the life history of a species (Bauer, 2004).

*Tozeuma carolinense* is a caridean shrimp endemic to the western Atlantic Ocean, with records from Vineyard Sound, Massachusetts to São Paulo, Brazil, and from Bermuda to Saint Lucia Island and Curaçao (Román-Contreras & Martínez-Mayén, 2009). It is highly abundant in shallow waters among beds of the seagrasses *Syringodium filiforme*, *Thalassia testudinum*, and *Halodule wrightii* (Hays, 2005). In coastal and estuarine systems of the Gulf of Mexico and Mexican Caribbean Sea it is the second dominant species after *Hippolyte zostericola* (Barba, 2012). It feeds on algae epiphytic on the seagrass or other structures, and thereby allows the seagrass beds to grow unhindered (Hays, 2005).

In spite of its wide geographical distribution, information on the population structure and some reproductive aspects of the species has been published only by Ewald (1969) and partly by Heck (1977), Dugan (1983), and Corey and Reid (1991). Other studies have focused on diverse experimental aspects (Cournoyer & Cohen, 2011; Hays, 2005; Main, 1985), regional faunal inventories and distribution (Barba, 2012; Ledoyer, 1986; Román-Contreras & Martínez-Mayén, 2009; Rouse, 1970), and prevalence of infestation by parasitic isopods (Romero-Rodríguez & Martínez-Mayén, 2018).

To address the lack of studies of the population biology and reproduction of *T. carolinense* along the Atlantic coast

of Mexico, the present study evaluates the population structure and reproduction of this species in Laguna de Términos, Campeche, southwestern Gulf of Mexico, focusing on the size structure, sex ratio, sexual maturity, breeding season, fecundity and brood loss. Additionally, we provide data for the first time about the growth and mortality of this species; these results may provide a better understanding of the biology of this widely distributed caridean shrimp.

## Materials and methods

Shrimps were collected every 2 months from June 2009 to June 2010 at the inner margin of Isla del Carmen, Laguna de Términos, Campeche, Mexico ( $19^{\circ}10' - 18^{\circ}05' N$ ,  $92^{\circ}12' - 91^{\circ}10' W$ ) by 3 minute trawls at depth of 1 m with a Colman-Segrove sledge net (mouth aperture: 0.2 m height  $\times$  0.7 m width) equipped with a mesh size of 800  $\mu m$ ; the net was dragged from a boat over *Thalassia testudinum* meadows. The captured shrimps were preserved in 100% ethanol and then transferred into 70% ethanol. During the collections, water temperature fluctuated from 30 to 32.8  $^{\circ}C$  and the salinity from 33.9 to 38.8 psu. The characteristics of the Laguna de Términos were described in detail by Yáñez-Arancibia and Day (1982) and Raz-Guzmán and Sánchez (1996).

Samples were separated according to sex. Shrimps were classified as males or females by the presence or absence of the appendix masculina on the second pair of pleopods. The sex-ratio was estimated as the quotient between the number of males and the total number of individuals collected in each sampling (Grabowski et al., 2014). A chi-square test ( $\chi^2$ ) was used to measure deviation from the expected 1:1 male:female for the total number of individuals in each size class and each date. The body size of the individuals was established as carapace length (CL), measured from the postorbital margin of the ocular orbit to the mid-posterior end of the carapace. Carapace length data were organized into 12 size-classes of 0.75 mm intervals calculated according to Sturges (1926) in order to construct length frequency distributions for each

sex and deduce the population structure of this species. The individuals in the smallest size-class were considered to be recruits.

Growth of *T. carolinense* was calculated with the von Bertalanffy (1938) equation  $CL_t = CL_\infty (1 - e^{-k(t-t_0)})$ , where  $CL_t$  is the carapace length at time  $t$ ,  $CL_\infty$  is the asymptotic length,  $k$  is the growth coefficient, and  $t_0$  the theoretical age when size is 0. These parameters were determined with the FiSAT II software version 1.1.3 (Gayanilo et al., 1995). Growth performances were compared in terms of the growth performance index ( $\phi'$ ), which is preferred to the use of  $CL_\infty$  and  $k$  individually (Pauly & Munro, 1984); it was computed as:  $\phi' = \log_{10} k + 2 \log_{10} CL_\infty$ . The longevity of each sex was estimated by inverting the von Bertalanffy growth model (D'Incao & Fonseca, 2000), considering  $t_0 = 0$  and  $CL_t/CL_\infty = 0.99$ ; thus, longevity =  $0 - (1/k) \ln[1 - (CL_t/CL_\infty)]$  (Simões et al., 2013). *Tozeuma carolinense* is not a commercially exploited species, hence the total mortality ( $Z$ ) was considered as natural mortality (Cházaro-Olvera, 2009). Since mortality is the inverse of survival (Ricker, 1975), the following formula was used to estimate total mortality ( $Z$ ):  $\log_e(N_{yc}) = -Z_{yc} + \log_e(a)$ , where  $N_{yc}$  is the number of shrimps per year class,  $yc$  is the number of year classes and  $a$  is a constant.

All the females with sizes equal to or greater than the smallest ovigerous female found were considered mature (Alon & Stancyk, 1982; Bauer, 1989) and were used to estimate the size at sexual maturity ( $CL_{50}$ ) using a logistic curve fitted between the proportion of sexually mature females (991 specimens) in each size class ( $P$ ) and the carapace length ( $CL$ ) with the equation  $P = 1/(1 + \exp[-(a + b \times CL)])$ , where  $a$  and  $b$  represent the parameters obtained of the regression between  $P$  and  $CL$ . The  $CL_{50}$  corresponds to 50% sexually mature females as calculated from the relationship  $CL_{50} = -(a/b)$  (Oh et al., 2002).

The entire brood mass of each female was separated from the pleopods with fine forceps, and was assigned to 1 of 2 stages of embryonic development (Corey & Reid, 1991): stage I, embryos without eyespot, and stage II, eyespot fully defined and well developed appendages. Embryo volume (EV) was calculated on the basis of 120 ovigerous females (60 of each developmental stage) taking a random subsample of 10 embryos per individual to measure their individual length and width and estimate their volume with the formula  $EV = EL \times \pi \times (EW/2)^2$  as proposed by Corey and Reid (1991) for caridean shrimps.

In order to avoid errors related to embryo loss during incubation (Kuris, 1991), fecundity (total number of embryos attached to the pleopods of a female) was assessed by direct counting in females that carried a compact brood with stage I embryos. Data from females with embryos

in stage II were used to estimate the average increase in volume and the possible brood loss.

One-way Anova was applied to evaluate the changes in embryo volume between stage I and stage II. Simple linear regression of  $\log_e$  transformed data was determined for the number of embryos (stage I) on carapace length, and the parameters that describe the equation between the 2 variables were obtained with the least squares method (Corey & Reid, 1991). All statistical analyses were based on Zar (2010), and the differences were considered significant at 95% ( $p < 0.05$ ) confidence level.

## Results

The sample consisted of 2,109 individuals of *Tozeuma carolinense*, of which 1,294 were females (634 of them ovigerous) and 815 were males. The highest abundance occurred in February 2010 ( $n = 705$ ) and the minimum in October 2009 ( $n = 110$ ). The average size  $\pm$  standard deviation differed significantly between females ( $6.16 \pm 2.02$  mm) and males ( $4.42 \pm 0.70$  mm) ( $t$ -test = 89.11,  $p < 0.05$ ). Females measured from 1.37 to 10.35 mm CL and males from 2.25 to 6.13 mm (Fig. 1). The overall sex-ratio was 0.39 males: 1 female, which differed significantly from the ratio of 1:1 ( $\chi^2$  test = 108.33,  $p < 0.05$ ). Females also significantly outnumbered males in every sampling period ( $\chi^2$  test = 12.93,  $p < 0.05$ ) (Fig. 2). Recruitment took place throughout the year, but the highest percentages of juveniles were observed during June 2009 and June 2010 (Fig. 3).

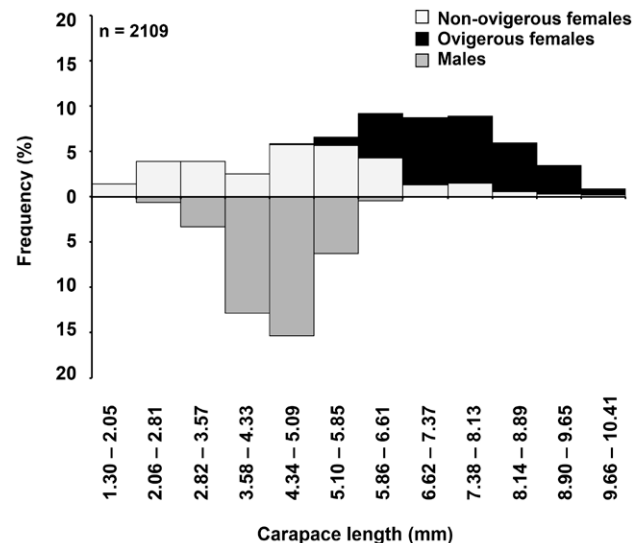


Figure 1. Variation in size frequency distributions of non-ovigerous females, ovigerous females, and males of *Tozeuma carolinense* in Laguna de Términos.

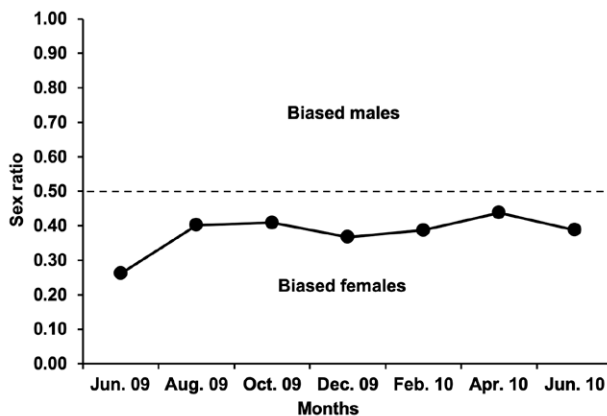


Figure 2. Sex-ratio of *Tozeuma carolinense* in Laguna de Términos during the study period.

According to the estimated parameters of the von Bertalanffy equation (Fig. 4), the asymptotic carapace length for females (10.13 mm) was greater than that in males (6.62 mm), but the growth coefficient was similar in females ( $k = 0.81 \text{ year}^{-1}$ ) and males ( $k = 0.84 \text{ year}^{-1}$ ). The growth performance index was higher in females ( $\phi' = 1.92$ ) than in males ( $\phi' = 1.57$ ). The estimated longevity was 6 months for females and 5 months for males. Mortality (Z) was 0.85 for females and 1.08 for males.

The size ( $\text{CL} \pm 95\%$  confidence interval) at which 50% of females are mature was  $6.67 \pm 1.30 \text{ mm CL}$ , a value similar to the mean population size of females. The logistic function that expressed the relationship between CL and the proportion of sexually mature females was  $P = 1/[1 + \exp(9.1385 - 1.3701 \times \text{CL})]$ .

Size of ovigerous females ranged from 4.90 to 10.25 mm CL ( $7.58 \pm 1.06 \text{ mm CL}$ ); they were collected throughout the study period, but the highest incidence of berried specimens occurred from August 2009 to April 2010 (Table 1). The mean number of embryos (stage I) produced by females was  $268 \pm 131$  and varied between 42 and 726. There was a significant relationship between the fecundity and female size ( $t = 27.55$ ,  $p < 0.05$ ), indicating that the larger females carry more embryos, although fecundity varied widely in females of similar body sizes (Fig. 5).

Embryos had a mean length of  $0.46 \pm 0.05 \text{ mm}$  at stage I, and of  $0.65 \pm 0.07 \text{ mm}$  at stage II. Their average volume varied from  $0.050 \pm 0.012 \text{ mm}^3$  in stage I to  $0.104 \pm 0.036 \text{ mm}^3$  in stage II, an increase of 108% ( $F = 1,193.31$ ,  $p < 0.05$ ). Females lost on average 46 % of their embryos during the incubation period.

## Discussion

The abundance of *Tozeuma carolinense* in Laguna de Términos throughout the year was variable, a pattern also observed in Panamá by Heck (1977) and in Florida by Dugan (1983). The last author registered the highest abundance from July to November (490-1,798 individuals) and the minimum during February, May and June (167-310 individuals); in our study, the highest and lowest abundance occurs in February 2010 and October 2009, respectively (see Fig. 3). Heck (1977) and Dugan (1983) suggested that the variability in abundance is related to changes in water salinity following the rainy seasons. During the present study, the variation of this parameter was minimal (4.9 psu), hence the notable difference in abundances possibly are given by factors such as changes in the species composition and extent of the seagrass meadows into the surrounding areas (Ewald, 1969; Heck, 1977).

The continuous recruitment of *T. carolinense* is similar to other tropical caridean shrimps that inhabit seagrass meadows (Bauer, 1992; Romero-Rodríguez & Román-Contreras, 2013). According to Bauer (1989) the variations in recruitment throughout the year can be caused by factors such as differences in the breeding intensity of the population, temporal mortality of planktonic larvae or the recruits recently settled in seagrass beds, as well as changes in currents. The latter could explain, partially, the higher number of recruits of *T. carolinense* in both June samples since changes in circulation patterns returns the planktonic larvae to seagrass meadows increasing the abundance (Bauer, 1989).

The predominance of females throughout the year in Laguna de Términos concurs with observations of *T. carolinense* in Florida, USA (Ewald, 1969), and of caridean shrimps in other regions (Bae & Oh, 2014; Nye & Copley, 2014; Oya, 1987; Oya & Oka, 1985; Torres et al., 2007; Zupo, 1994). The high proportion of females could be explained, partially, by a heterogeneous distribution of the sexes on the substratum, associated with hiding or feeding (Main, 1985), or may be attributable to a higher mortality of males (see below). Berglund (1981) pointed out that the predominance of one sex also could be determined by a sexual dimorphism in size.

The growth parameters calculated indicates that females of *T. carolinense* grew faster and reached larger sizes than males, this pattern is equal to that reported for other caridean shrimps (Bae & Oh, 2014; Cházaro-Olvera, 2009; Manjón-Cabeza et al., 2009; Paschoal et al., 2016; Romero-Rodríguez & Román-Contreras, 2013). On the other hand, despite the longevity estimated for both sexes was similar,

the larger lifespan of the females coincides with the results obtained in other crustacean studies, e.g., 12 months for females and 8 months for males of *Hippolyte inermis* (Manjón-Cabeza et al., 2009), 16 months for females and 13 months for males of *Palaemon argentinus* (as *Palaemonetes argentinus*) (Schuldt et al., 1988), and 10.2 months for females and 8.9 months for males of *Hippolyte zostericola* (Romero-Rodríguez & Román-Contreras, 2013). Thus,

both observed patterns in *T. carolinense* could be related with the mating system known as pure searching reported in many gonochoristic carideans with females larger than males (Bauer, 2004), in which males are more vulnerable to predation because they have to move constantly in order to find and fertilize as many females as possible (Correa & Thiel, 2003; Bauer, 2004), which also explain that they presenting a higher mortality respect to the females.

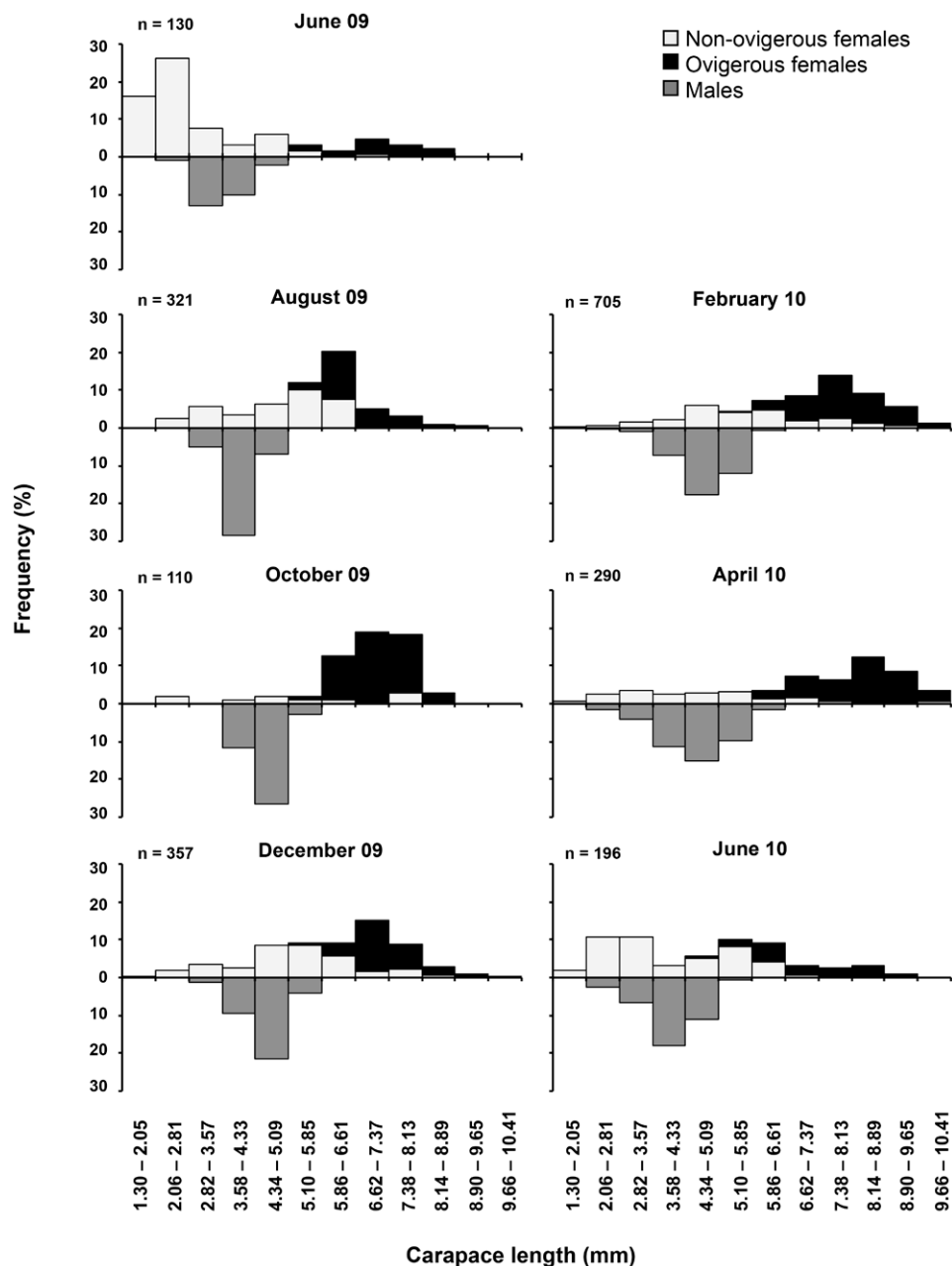


Figure 3. Bimonthly length frequency distribution of *Tozeuma carolinense* in Laguna de Términos.

In the tropics, the majority of the species breed for a long period of time, but a seasonal increase in the number of ovigerous females may occur, as was observed in *T. carolinense* in the study area, this could be related to rise of food resources in the lagoon during some months (Contreras et al., 1996) creating better conditions for larval development and settlement of the individuals. Continuous reproduction in this species has also been recorded from Bear Cut, southeast waters of Florida (Ewald, 1969) and on the Caribbean coast of Panama (Heck, 1977). In contrast, in Apalachee Bay, Florida, Dugan (1983) collected berried females only from March to October when water temperatures were greater than 15°C. Continuous reproduction has also been reported for *H. zostericola* in the area of the present study (Romero-Rodríguez & Román-Contreras, 2013), and for other caridean shrimp species in Puerto Rico (Bauer, 1989) such as *Hippolyte obliquimanus* (as *Hippolyte curacaoensis*), *Latreutes fucorum*, *Latreutes parvulus*, and *Thor manningi*.

In decapod crustaceans with a continuous reproductive pattern, sexual maturity of females is reached at a relatively small size, thus the size of the smallest ovigerous female has been set as the size of sexual maturity or size at onset of sexual maturity (Bauer, 1989; Anger & Moreira, 1998) because it represents the stage where the specimens are able to generate new individuals. In our study area, *T. carolinense* starts to produce embryos at a minor size (4.90 mm CL) than that reported for the same species in Florida, ~ 6.9 mm CL (Ewald, 1969), 5.76 mm CL (Corey & Reid 1991), and 35 mm total length (~5.75 mm CL) (Dugan, 1983). High water temperatures (28-33 °C) would favour rapid gonadal development, sexual maturity and spawning, as well as continuous reproduction (Bauer, 1989). However, temperature cannot be the sole controlling factor, since temperatures in Laguna de Términos were similar to those reported for Florida (Mason & Zengel, 1996). Therefore, this discrepancy in size at the onset sexual maturity in other regions may indicate adaptations to local environmental conditions including quality and quantity of food sources, as well as population density and predation pressure (Bas et al., 2007; Hines, 1989).

Fecundity of caridean shrimps is variable and highly dependent upon the body size of the female. The positive relationship between carapace length and number of embryos in the present study corroborated other fecundity studies in tropical caridean groups including *T. carolinense* (Anger & Moreira, 1998; Bauer, 1991; Corey & Reid, 1991; Ewald, 1969). The average fecundity calculated for *T. carolinense* differs from the value reported by Corey and Reid (1991) in the same species. It is also higher than other caridean shrimp species, except for *Exhyppolysmata oplophoroides*, *Lysmata boggessi*, and *Trachycaris restricta* (Table 2). The clutch size is highly plastic and

Table 1

Number and percentage of ovigerous females of *Tozeuma carolinense* collected between June 2009 and June 2010.

Month	Number of individuals		
	Females	Ovigerous	(%)
June 2009	96	16	16.66
August 2009	192	79	41.14
October 2009	65	55	84.61
December 2009	226	98	43.36
February 2010	432	246	56.94
April 2010	163	107	65.64
June 2010	120	33	27.5
Total	1,294	634	48.99

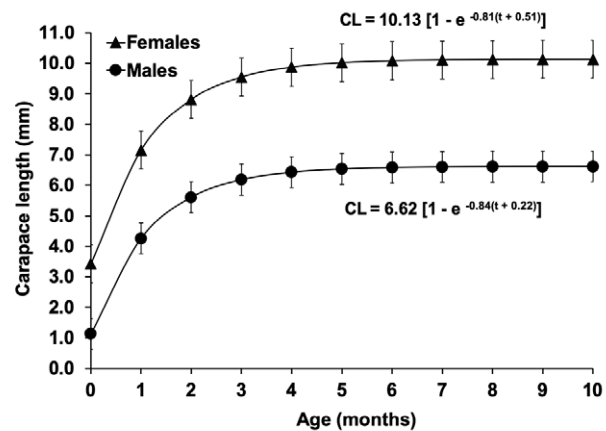


Figure 4. Growth curve estimated for females and males of *Tozeuma carolinense* with the von Bertalanffy growth model.

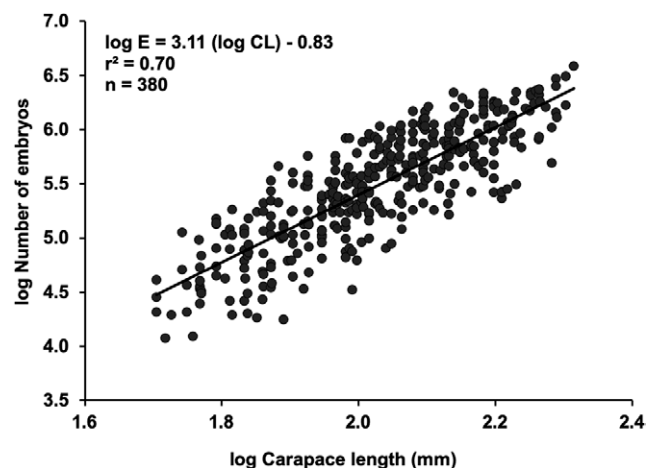


Figure 5. Relationship between log-transformed data of carapace length (CL) versus log-transformed number of embryos (E) for stage I of *Tozeuma carolinense*.

may vary from one geographical area to another even in the same species (Bas et al., 2007). This variation has been attributed to many factors such as embryo size and genetic features of the populations (Martínez-Mayén & Romero-Rodríguez, 2018), exhaustion of females due to production of successive clutches during a season (Palacios et al., 1999), embryo mortality (Oh et al., 2002), reproductive senescence (Pandian, 2016), small sample size and the method of measurement used. However, the difference in fecundity recorded here could also reflect the smaller size of the females analysed in previous studies (Table 2).

Embryo size determines the reproductive output and correlates with the development pattern of organisms. *Tozeuma carolinense* produces small embryos (0.46 mm)

that generate an extended larval development (10 larval stages, see Ewald, 1969). The average volume recorded herein agrees with the values reported for other caridean shrimp species except *L. boggei* and *T. restricta*, which produce larger embryos (Table 2). Embryo volume in this study (0.050 mm<sup>3</sup>) is practically the same as that calculated by Corey and Reid (1991) in Florida (0.057 mm<sup>3</sup>), presumably reflecting a similar investment of energy in the production of embryos (Clarke, 1993).

The 108% increase in embryo volume during ontogeny was higher than the value of 84% calculated by Corey and Reid (1991) for this species. An increase of more than twice the volume has also been recorded for other caridean shrimp species from the Americas (Corey & Reid, 1991;

Table 2

Average carapace length of ovigerous females, fecundity and embryo volume in stage I for 12 caridean shrimps species.

Species	Carapace length (mm)	Fecundity	Embryo volume (mm <sup>3</sup> )	Locality	Reference
<i>Exhippolysmata oplophoroides</i>	11.51	2,742	0.015	Ubatuba, São Paulo, Brazil	Chacur & Negreiros-Franozo (1999)
<i>Hippolyte nicholsoni</i>	0.96	25	0.018	Florida Middle Ground, USA	Corey & Reid (1991)
<i>Hippolyte obliquimanus</i>	2.4	153	0.005	Dorado, Puerto Rico	Bauer (1991) (as <i>Hippolyte curacaoensis</i> )
	2.43	187	0.012	Limón, Costa Rica	Terossi et al. (2010)
	2.09	141	0.015	Ubatuba, São Paulo, Brazil	
<i>Hippolyte zostericola</i>	2.77	147	0.037	Indian River, Florida, USA	Corey & Reid (1991)
	2.41	144	0.015	Laguna de Términos, Campeche, México	Romero-Rodríguez & Román-Contreras (2013)
<i>Latreutes fucorum</i>	2.3	57	0.012	Dorado, Puerto Rico	Bauer (1991)
	3.16	127	0.030	Florida, USA, and Virgin Islands	Corey & Reid (1991)
	2.62	96	0.013	Bahía de la Ascensión, Quintana Roo, Mexico	Martínez-Mayén & Román-Contreras (2011)
<i>Latreutes parvulus</i>	2.0	59	0.012	Dorado, Puerto Rico	Bauer (1991)
<i>Lysmata boggei</i>	7.5-8.24	554-829	0.0862-0.1017	West central coast of Florida, USA	Baeza et al. (2014)
<i>Lysmata vittata</i>	6.38	3.93	0.038	Sergipe, Brazil	Almeida et al. (2018)
<i>Nauticaris magellanica</i>	NA	NA	0.031-0.038	Guanqueros and Metri, Chile	Wehrmann & Kattner (1998)
<i>Thor manningi</i>	1.62	37	0.036	Indian River, Florida, USA	Corey & Reid (1991)
<i>Trachycaris restricta</i>	6.10	431	0.220	Virgin Gorda, Virgin Islands	Corey & Reid (1991)
<i>Tozeuma carolinense</i>	6.79	190	0.057	Indian River, Florida, USA	Corey & Reid (1991)
	7.63	268	0.050	Laguna de Términos, Campeche, Mexico	Present study

NA, data not available



Terossi et al., 2010; Wehrtmann & Kattner, 1998). The increase is a consequence of gradual water absorption by the embryo, changes in biochemical composition during embryonic development (Wehrtmann & Kattner, 1998) and embryo growth.

Brood loss during incubation diminishes reproductive potential but is common among embryo-carrying crustaceans (Oh & Hartnoll, 1999); the reduction of the clutch size may favour the survival of the remaining embryos because there is more space in the abdomen for a better accommodation of the embryos and a better oxygenation of the ovigerous mass (Nazari et al., 2003). *Tozeuma carolinense* here lost 46% of the embryos during the incubation period, a value higher than that reported previously (31–41%) in the same species (Corey & Reid, 1991), but that fits well with the percentages of brood loss tabulated (12–74%) for various caridean shrimps (Oh & Hartnoll, 1999). Factors that may influence embryo loss during development include increase in embryo volume during development, parental care, mechanical stress, and parasites (for a review, see Kuris, 1991). In *T. carolinense*, the major reason appears to be the increase in embryo volume, because this can strain and break the attachment structures.

The present study is the first regarding biological aspects of *T. carolinense* for the Atlantic coast of Mexico. Our results increase current understanding, but additional studies along the geographic distribution are required, in order to elucidate the possible differences among populations of the same species.

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