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Artículo original

Phytochemical prospection and biological activities of the floral extract Ceiba pubiflora a. St. – hill. (Malvaceae)

Prospección fitoquímica y actividades biológicas del extracto floral *Ceiba* pubiflora a. St. – hill. (Malvaceae)

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ABSTRACT

The genus *Ceiba* has species such as *Ceiba pubiflora* circumscribed in the Malvaceae family where it has annual flowering in the Cerrado domain of Brasil. Flowers of *C. pubiflora* were collected, and then the extract was produced. Phytochemical analyses, thin-layer

chromatography, yield, pH, refractive index and density were performed. Bioactive compounds were determined by the reduction of DPPH free radicals and total phenols. The extract yield was 6,88 %, several classes of compounds present as alkaloids, flavonoids and saponins. The best eluents were acetone and ethyl acetate in thin layer chromatography, the antioxidant activity showed an inhibition concentration (IC₅₀) of 217,4 mg L⁻¹ and total phenolic content of 7,26 mg expressed as gallic acid (GAE 100 g⁻¹). This study allowed, based on the unprecedented results of *C. pubiflora*, further studies to be carried out, due to the numerous results of great importance, which allow investigating in greater detail the pharmacological, chemical and bioactive characteristics.

Keywords: phytochemical screening; *Ceiba* genus; antioxidant activity; total phenols.

RESUMEN

El género *Ceiba* tiene especies como *Ceiba pubiflora* circunscrita en la familia Malvaceae, que florece anualmente en el dominio Cerrado de Brasil. Se recolectaron flores de *C. pubiflora*, luego se produjo el extracto. Se realizaron análisis fitoquímicos, cromatografía en capa fina, rendimiento, pH, índice de refracción y densidad. Los compuestos bioactivos se determinaron mediante la reducción de radicales libres DPPH y fenoles totales. El rendimiento del extracto fue de 6,88 %; hubo presencia de varias clases de compuestos como alcaloides, flavonoides y saponinas. Los mejores eluyentes fueron acetona y acetato de etilo en cromatografía en capa fina; la actividad antioxidante mostró una concentración de inhibición (CI₅₀) de 217,4 mg L⁻¹ y contenido fenólico total de 7,26 mg expresado en ácido gálico (EAG 100 g⁻¹). Este estudio permitió, en base a los resultados sin precedentes de *Ceiba pubiflora*, que se realicen más estudios, debido a los numerosos resultados de gran importancia, que permiten investigar con mayor detalle las características farmacológicas, químicas y bioactivas.

Palabras clave: tamizaje fitoquímico; género Ceiba; actividad antioxidante; fenoles totales.

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Introduction

Cerrado domain is a Neotropical savannah, being this strategic environment with rich

variability in species of Brazilian and world flora. In addition, it is considered the second

largest in natural area, occupying an area of about 2 million km², or 22 % of the National

territory. (1) In this environment a great diversity is found of plant families such as Malvaceae,

subfamily Bombacoideae and Bombaceae tribe, which includes the Ceiba Miller genus, with

about 17 species of Neotropical distribution. In Brasil 11 species of Ceiba are registered,

Ceiba pubiflora being a species native to the Midwest and also to the Northeast and

Southeast. (2)

C. pubiflora occurs mainly in semideciduous forests, and particularly in calcareous soils, and

is popularly known as "potbellied or white paineira", with height between 15-25 m with

canopy; gray stem with conical spines; compound leaves; fruits in capsule, oblong, woody,

and brown seeds with ovoid shape. In addition to the Cerrado C. pubiflora is inserted in the

flora of the Atlantic Forest and Caatinga biomes. In the Midwest region of Brasil the period of

flowering and fruiting is associated by farmers during the rainy dry season. The flowers of C.

pubiflora are large, pink color and have a light aroma which is why it is used in

ornamentation and landscape projects in cities. (3, 4)

Numerous extracts produced from the species of the Malvaceae family, present

phytocompounds of great phytotherapeutic interest, for the food industries during

conservation, in the pharmaceutical industry, in the production of topical and internal

medicines, and in the agricultural industries such as larvicide, molluscicide and insecticide,

and biotechnology mainly in the control of microorganisms. (5-7)

Genus Ceiba is described for various therapeutic, nutritional and economic purposes, in the

form of floristic inventories and ethnobotanical studies based on the gum extracted from the

stem bark and roots with aphrodisiac properties. (4, 8) However in relation to the phytochemical

composition of the floral organ, little is known about the classes of phytocompounds and their

335

possible phytotherapeutic effects requiring previous studies on phytochemistry that can be obtained from secondary metabolism.

Considering the scarcity of phytochemical studies and biological activities, this work aimed to evaluate the floral hydroethanolic extract of *Ceiba pubiflora*.

Material and methods

Plant material

Flowers were collected at the Instituto Federal Goiano, Rio Verde, Goiás State, Brasil (17°48′39.4″S and 50°53′57.5″W), at 8 pm on 25st March 2020. The plant material was identified and samples were deposited as voucher specimens in the Herbarium at the Instituto Federal Goiano, Rio Verde, Goiás State, Brasil (identification number HRV 12637).

Preparation of hydroethanolic extract

Extraction was carried out with 500 g flowers and 100 mL 70 % (v/v) hydroethanolic solution; it was kept under constant magnetic agitation for 12 h. Contact between solvent and raw material was kept for nine days at room temperature (25 °C), in the dark. It was manually agitated on a daily basis. The mixture that resulted from the extraction was separated by filtration (Unifil, C42), followed by solvent evaporation which was carried out by a rotary evaporator at reduced pressure. Then, the extract was lyophilized in a lyophilizer (Liotop, Mod. L101) with negative pressure until constant mass.

Phytochemical analysis

The phytochemical tests to detect the presence of cardiac glycosides, cyanogenic glycosides, alkaloids, organic acids, reducing sugars and no-reducing sugars, coumarins, foamy saponins and hemolytic saponins, polysaccharides, phenols, tannins, flavonoids, purines, resins, catechins, auronas and chalcones, depsides and depsidons, cyanogenic heterosides, benzoquinones, naphthoquinones and phenanthraquinones, anthraquinones, steroids and

triterpenoids, sesquiterpenolactones, proteins and amino acids, leucoanthocyanidins, anthocyanins, azulenes, and oxidation time.⁽⁹⁾ These tests were based on the visual observation of color modification or precipitate formation after the addition of specific reagents. The results were expressed by the cross-testing (-) negative, (+) weak positive, (++) moderate positive and (+++) strong positive.

Physicochemical analysis

The yield of the dry extract was determined by calculating the ratio of its weight on a digital analytical scale (Marte, Mod. W220) with respect to the weight of the plant drug, expressed as a percentage according to equation (1).⁽¹⁰⁾

$$Yield \% = [(gDE * 100)/gVD] \tag{1}$$

Where: DE = dry extraction, and VD = Vegetable drug.

The following chemical and physical-chemical parameters were performed as described by Valdés et al. (11) and Domínguez et al. (12) The pH was performed using a digital pH meter (Lucadena, Mod. 210-P) (IAL, 2008), the total solids content in an oven at 105 °C (Nova Ethics, Mod. 400-3ND) (IAL, 2008), and the refractive index in a digital refractometer (Hanna Instruments, Mod. HI96800) with a refraction range between 1,333 0 and 1,508 0 np. The relative density was determined in a 10 mL graduated cylinder (Pyrex), and expressed in g mL⁻¹ at 20 °C. Chromatography determination was performed by thin-layer chromatography (TLC) for the extract using chromatoplates (DC-Fertigfolien Alugram® Xtra SIL G/UV₂₅₄) (Macherey-Nagel). Approximately 10 µL of the extract was deposited on the chromatoplates cut into strips of (2 x 10 cm in length). The eluents used were (ethyl alcohol, chloroform, ethyl acetate and acetone), and the following developers used (sulfuric vanillin, ultraviolet light, long and short wavelengths, and ferric chloride) were used. The retention factor (Rf) was determined according to the points observed for each developer. (13) The hydroethanolic extract was scanned between 250 and 900 nm in a UV-Vis spectrophotometer (Belphotonics, Mod. M-51), the data were generated by UV-Vis professional 2 software and processed in Origin.

Antioxidant activity and total phenolic compounds

The antioxidant capacity was determined using the DPPH (2,2-diphenyl-1-picrilhydrazyl) reduction method. The DPPH sequestration method was performed in microdilution in 96-well microplates (Global Plast) modified. For each well, 100 μL of an ethanolic solution with DPPH at the concentration of 0,06 mMol L⁻¹ and 100 μL of a hydroethanolic extract solution at different concentrations. The microplate was kept in a place protected from light and heat for 1 h. Then, the microplate spectrophotometer (Scientific Hexis, SpectraMax Plus, Mod. 384) was read at a wavelength of 517 nm. The percentage of antioxidant capacity was determined according to equation (2). The inhibition concentration (CI₅₀) was determined in (mg L⁻¹) in the sequestration of 50 % of the initial concentration of the standard solution of DPPH 0.06mMol L⁻¹.

$$\% Red = \left(\frac{CSabs - ESabs}{CSabs}\right) * 100$$
 (2)

Where: CSabs = absorption of the control solution, and ESabs = absorption of the extract solution containing the radical DPPH.

In order to quantify total phenolic compounds, 1,9 mL *Folin-Ciocalteu* reagent in distilled water (1:10, v/v) was added to 200 μ L extract hydroethanolic floral. To neutralize the mixture, 1,9 mL aqueous solution of sodium carbonate (60 g L⁻¹) was used. The reaction was kept in the dark at room temperature for 120 min. Then, absorbance was measured at 725 nm. Calculation was carried out with the use of the standard curve and results were GAE expressed as mg gallic acid 100 g⁻¹ dry extract (DE).⁽¹³⁾

Results and Discussion

C. pubiflora floral period in the collection area in the municipality of Rio Verde, Goiás State, Brasil in 2020 was between the months of March to May, with a high intensity of flowering in

May. The (figure 1) shows a flower of *C. pubiflora* used in the production of hydroethanolic floral extract of this study.



Fig. 1- *Ceiba pubiflora* flower used in the hydroethanolic floral extract, collected in Rio Verde, Goiás State, Brasil in 2020. Bar: 10 cm. Source: Authors, 2020

C. pubiflora extract floral presented transparency, crystallinity, pink color, and light aroma. The percentage of dry extract, yield equal to 6,88 % (*w/w*). The phytochemical prospection indicated the presence of different classes of secondary metabolites, except foam saponins, polysaccharides, resins, benzoquinones, anthraquinones, proteins and amino acids. Table 1 shows the results of the phytochemical screening carried out on the hydroethanolic floral extract.

Table 1- Qualitative phytochemical prospection of the floral hydroethanolic extract of *Ceiba pubiflora*

Classes	Results
Cardiac glycosides	+++
Cyanogenic glycosides	++
Alkaloids	+++
Organic acids	+++
Reducing sugars	+++
No-reducing sugars	++
Coumarins	+
Foamy saponins	-
Hemolytic saponins	+++
Polysaccharides	-
Phenols	+++
Tannins	Gr
Flavonoids	+++
Purines	+
Resins	-
Catechins	++
Auronas and Chalcones	++
Depsides and depsidons	+++
Cyanogenic heterosides	+++
Benzoquinones, naphthoquinones and phenanthraquinones	-
Anthraquinones	-
Steroids and triterpenoids	++
Sesquiterpenolactones	+
Proteins and amino acids	-
Leucoanthocyanidins	+++
Anthocyanins	+++
Azulenes	++
Oxidation time	5 sec.

Note: (-) negative. (+) weak positive. (++) moderate positive. (+++) strong positive. Tannins: Green (Gr) condensed or catechic. Blue (Bl) hydrolyzable or gallic. Source: Authors, 2020.

The extract yield was high, although it is not possible to compare it with other floral extracts of the *Ceiba* genus due to lack of study. Through qualitative phytochemical prospection of the floral extract of *C. pubiflora*, it was possible to determine the presence of special metabolites classes that present in several studies a wide variety of biological activities such as antimicrobial, antioxidant, antitumor, antiophidic, photoprotective and photosensitizing. (15, 16)

Tannins and flavonoids have antimicrobial properties, ⁽¹⁷⁾ saponins have cytotoxic action ⁽¹⁸⁾ and depsides and depsidons have anti-inflammatory properties. ⁽¹⁹⁾ Although some reagents may reveal the presence of more than one phytoclass, this is only a preliminary result, requiring further testing using quantitative results that can be obtained by high performance liquid chromatography and compared to classical standards. The expressive content of phenolics in the floral extract of *C. pubiflora*, as well as in several other studies, has reported important biological actions. According to Oliveira *et al.*⁽²⁰⁾ Haslam, ⁽²¹⁾ and Dixon *et al.*⁽²²⁾

phenolic compounds inhibit lipid peroxidation and lipoxygenase *in vitro*. The oxidation time was slightly fast, suggesting that it is a potent extract with antioxidant characteristics.

Qualitative hemolytic assay on human erythrocytes, demonstrated potential cytotoxicity with a minimum of 5 min, and complete hemolysis with 10 min. After five minutes, deformation of the human erythrocytes can be observed, followed by rupture and worsening of hemoglobin. Figure 2 shows in the hemolysis assay on human erythrocytes from the floral extract of *C. pubiflora*.

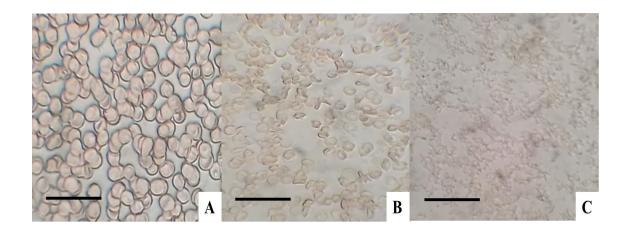


Fig. 2- Hemolysis assay in 5 % blood cells concentrate. In (**A**) normocytic blood cells, in (**B**) hemolysis with 5 min, and (**C**) hemolysis with 10 min. Bars: (**A**) 150 μ m, (**B**) 100 μ m and (**C**) 100 μ m. Source: authors, 2020

The pH was $6,41 \pm 0,02$, total solids of $1,12 \pm 0,09$, refractive index of $1,536.9 \pm 0,13$, and relative density of $0,906.1 \pm 0,01.9$ g mL⁻¹ to 20 °C. According to Menezes Filho *et al.*, ⁽¹³⁾ the acid or alkaline pH in extracts, may be related to the classes of phytocompounds of a phenolic or saponinic nature. The content of total soluble solids varies according to the plant extracts and types of solvents used. Soluble solids represent the volatile organic and inorganic material content of the extract solution.

Acetone and ethyl acetate were the eluents that had the highest number of Rf (12), respectively. The presence of sugars reducing, glucose and fructose, and no-reducing sucrose was observed by thin-layer chromatography analysis in the floral extract of C. publiflora (table 2).

Table 2- Results of thin-layer chromatography of the floral extract of Ceiba pubiflora

Eluents	UV	UV	Sulfuric vanillin	FeCl ₃	\mathbf{I}_2	KMnO ₄	C21H14Br4O5S	H ₂ CrO ₄
	254 mm	365 mm						
C_3H_6O	0,69	0,45 0,30	0,75 0,26	0,33 0,08	0,35	0,11	0,51 0,42	0,23
CHCl ₃	-	0,63	0,65	0,57 0,45 0,15	0,45	0,54 0,25	0,75	0,75
C ₄ H ₈ O ₂	0,09	0,19 0,15	0,70 0,62	0,65	0,71	0,65	0,72	0,75 0,69 0,05
C ₂ H ₅ OH	-	0,65	0,75	0,12	0,75 0,15	0,55 0,35 0,26	0,35	0,05

Note: Results in Rf (Retention factor), determined in millimeters (mm). Source: Authors, 2020.

The chromatographic assay, a large amount of Rf is observed for the four eluents evaluated and their respective developers. Only polar eluents were used, observing that the compounds of the flower extract showed to be less polar and were more easily eluted. The elotropic sequence of the eluents was: CHCl₃ (10) $Rf > C_4H_8O_2$ (12) $Rf > C_3H_6O$ (12) $Rf > C_2H_5OH$ (10) Rf, the Rf showed high values in mm, which confirms that they have a greater amount of apolar compounds in the extract. Especially acetone and ethyl acetate proved to be the best eluents.

The developers UV₂₅₄ nm light shows phytochemicals that absorb light, often they are conjugated substances and aromatic systems and for UV₃₆₅ nm light the compounds naturally fluoresce; sulfuric vanillin is especially sensitive to the presence of alcohols and terpenoids; the ferric chloride complex with phenols and enolizable compounds; complex I₂ with structures of amino acids, indoles, alkaloids, steroids, purines and lipids; potassium permanganate easily stains oxidizable compounds, olefins, alkynes and aromatics; bromocresol green shows organic acid compounds; and chromic acid difficult substances.^(23, 24)

The results obtained for both qualitative tests and for TLC showed that both are suitable for a preliminary analysis of phytochemicals, presenting speed, economy, safety and easy handling.⁽²⁴⁾ The presence of reducing sugars was more intense both in the qualitative test (table 1), and in the TLC, with two reducing sugars (glucose and fructose) and one non-

reducing sugar (sucrose). Table 2 was the results of the thin-layer chromatography of the floral extract of *C. pubiflora* in different eluents.

The UV-*Vis* spectrophotometric analysis, between 850-250 nm short and intense bands were observed, suggesting the presence of phenolic and flavonoid compounds that absorb in UV energy sources between 200-300 nm and between 400-500 nm, respectively. Figure 3 shows the UV-*Vis* spectrophotometry scan of the floral hydroethanol extract of *C. pubiflora* between 850-250 nm.

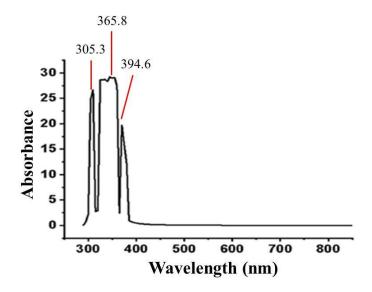


Fig. 3- UV-*Vis* spectrophotometry of the *Ceiba pubiflora* floral hydroethanolic extract between 850-250 nm. Source: Authors, 2020

The antioxidant activity of the floral extract of *C. pubiflora* presented IC₅₀ = 217.4 ± 0.19 mg L⁻¹, and total phenols content of 7.26 ± 0.16 mg GAE 100g⁻¹ in (DE). Comparing the phytochemical qualitative and quantitative methods, it is observed that in (table 1), the groups of phenolics, tannins and flavonoids evaluated by colorimetric and precipitation methods showed superior results by the cross test, when compared to the quantitative tests in the radical reduction DPPH and total phenolic content, where the first is visually observed and the second more refined, is performed through standard curves. According to Menezes Filho *et al.*, ⁽¹³⁾ Degáspari and Waszczynsyj ⁽²⁶⁾ and Camacho-Campos *et al.* ⁽²⁷⁾ antioxidants, and total phenols content are complex groups found mainly in vegetables that have functions such

as delaying or inhibiting the oxidation of lipids or other biomolecules, thus preventing the onset of oxidation chain reactions.

Conclusions

The floral extract of *Ceiba pubiflora* exhibited in this study, a great diversity of phytochemical groups present in the qualitative methodology. In addition, the extract showed potential antioxidant activity and expressive content of total phenolic compounds. Future studies should be carried out evaluating the phytochemical profile by high performance liquid chromatography, other antioxidant models and other biological activities.

Bibliographic references

- 1. BUENO, M. L.; OLIVEIRA-FILHO, A. T.; PONTARA, V.; POTT, A.; DAMASCENO-JUNIOR, G. A. "Flora arbórea do Cerrado de Mato Grosso do Sul". *Iheringia*. 2018; **73**, 53-64. ISSN 2446-8231.
- 2. GIBBS, P. E.; SEMIR, J. "A taxonomic revision of the genus *Ceiba* Mill. (Bombacaceae)". *Anales del Jardín Botánico de Madrid*. 2003, **60**, 259-300. ISSN 0211-1322.
- 3. SILVA, P. A. "Bird-flower interactions in na urban area: *Ceiba pubiflora* provides nectar and promotes biodiversity in the city". *Urban Forestry & Urban Greening*. 2018; **36**, 42-49. ISSN: 1618-8667.
- 4. PREVIATTO, D. M.; DAINEZI, D. I. S.; POSSO, S. R. "Exploitation of *Ceiba pubiflora* flowers by birds". *Revista Brasileira de Ornitologia*. 2016; **24** (1), 21-26. ISSN: 2662-673X.
- 5. PORTELLES, E. C. T.; CUÉLLAR, A. C.; OLIVET, E. S.; ANDREU, G. L. P. "Toxicidad aguda del extracto hidroalcohólico de *Croton wagneri* Müll. Arg. (moshquera) y su efecto irritante sobre la mucosa bucal". *Revista Cubana de Plantas Medicinales*. 2019; **24** (2), e791. ISSN 1028-4796.

- 6. SILVA, T. P. *et al.* "Espécies vegetais utilizadas no bloqueio da atividade hemorrágica induzida pelos venenos de serpentes do gênero *Bothrops* sp.: uma revisão da literatura". *Scientia Amazonia*. 2017; **6** (2): 36-57.ISSN 2238-1910.
- 7. MEHMOOD, S. I.; MAJEED, S.; JANNAT, Z.; HABIB, T. "Imaging based Ethno botanical studies of district Poonch, Zad Jammu and Kashmir". *International Journal of Herbal Medicine*. 2018; **6** (6): 81-91. ISSN 2321-2187.
- 8. BRAGA, P. M. S. *et al.* "Análise fitoquímica, toxicidade, potencial antioxidante e atividade antibacteriana da *Ceiba speciosa* (A. ST. –Hil.) Ravenna". *Revista Fitos.* 2019; **13** (1): 9-21. ISSN 2446-4775.
- 9. SHAIKH, J. R.; PATIL, M. K. "Qualitative tests for preliminary phytochemical screening: An overview". *International Journal of Chemical Studies*. 2020; **8** (2), 603-608. ISSN 2349-8528.
- 10. ALVES, M. M; PEREIRA, M. A. S.; PEREIRA, O. S.; FRANÇA, S. C.; BERTONI, B. W. "Caracterização química de tinturas e extratos secos de plantas medicinais do Cerrado por cromatografia em camada delgada". *Scientia Plena*. 2011; **7** (12), 1-8. ISSN 1808-2793.
- 11. VALDÉS, L. M. *et al.* "Determinación de saponinas y otros metabolitos secundarios en extractos acuosos de *Sapindus saponaria* L. (jaboncillo)". *Revista Cubana de Plantas Medicinales*. 2015; **20** (1). ISSN 1028-4796.
- 12. DOMÍNGUEZ, O. G. P.; PESANTES, K. E. B.; MARTÍNEZ, M. M.; GAITÉN, Y. G. "Estudio químico y evaluación biológica del extracto etanólico de *Allium schoenoprasum* L. Regel & Tiling (Cebollín)". *Revista Cubana de Farmacia*. 2019; **52** (1), e98. ISSN 1561-2988.
- 13. MENEZES FILHO, A. C. P.; SANTOS, M. C.; CASTRO, C. F. S. "Prospecção fitoquímica, físico-química e biológica do extrato hidroetanólico floral de [*Tabebuia impetiginosa* (Mart. ex DC.) Standl.]". *Revista Perspectivas Online: Biológicas & Saúde*. 2021; **11**(36), 1-25. ISSN 2236-8868.
- 14. MEZZA, G. N. *et al.* "Antioxidant activity of rosemary essential oil fractions obtained by molecular distillation and their effect on oxidative stability of sunflower oil". *Food Chemistry*. 2018; **242**, 9-15. ISSN 0308-8146.

- 15. ESQUENAZI, D. et al. "Antimicrobial and antiviral activities of polyphenolics from *Cocos nucifera* Linn. (Palmae) husk fiber extract". *Research in Microbiology*. 2002; **153** (10), 647-652. ISSN 0923-2508.
- 16. BARREIROS, A. L. B. S.; DAVID, J. M. "Oxidative stress: relations between the formation of reactive species and the organism's defense". *Química Nova*. 2006; **29** (1), 113-123. ISSN 1678-7064.
- 17. MATIAS, E. F. F.; SANTOS, K. K. A.; ALMEIDA, T. S.; COSTA, J. G. M.; COUTINHO, H. D. M. "Phytochemical prospection and modulation of aminoglycoside antibiotic activity by *Croton campestris* A". *Chemotherapy*. 2011; **57**, 305-309. ISSN 1421-9794.
- 18. CRUZ, M. C. *et al.* "Antifungal activity of Brazilian medicinal plants involved in popular treatment of mycoses". *Journal of Ethnopharmacology*. 2007; **111** (2), 409-412. ISSN 0378-8741.
- 19. BUGNI, T. S. *et al.* "Biologically active components of a Papua New Guinea analgesic and anti-inflammatory lichen preparation". *Fitoterapia*. 2009; **80** (5), 270-273. ISSN 0367-326X.
- 20. OLIVEIRA, A. H. *et al.* "Atividade moduladora de extratos etanólico das folhas de *Clusia nemorosa* G. Mey. (Clusiaceae) sobre drogas antimicrobianas". *Revista Cubana de Plantas Medicinales*. 2016; **21** (1). ISSN 1028-4796.
- 21. HASLAM, E. "Natural polyphenols (vegetable tannins) as drugs: possible modes of action". *Journal of Natural Products*. 1996; **59** (2), 205-215. ISSN 0163-3864.
- 22. DIXON, R. A.; DEY, P. M.; LAMB, C. J. "Phytoalexins: enzymology and molecular biology". *Advances in Enzymology and Related Areas of Molecular Biology*. 1983; **55**, 1-69, John Wiley & Sons, Inc. ISBN 0-471-86866-4.
- 23. SHERMA, J.; FRIED, B. *Handbook of thin-layer chromatography. Chromatographic Science Series*, 3th, Edition, Revised and Expanded. Lafayette College Easton, Pennsylvania, U.S.A. 2005; **89**, 1330. ISBN 0-203-91243-8.
- 24. RODRIGUES, I. M. C.; SOUZA FILHO, A. P. S.; FERREIRA, F. A. "Estudo fitoquímico de *Senna alata* por duas metodologias". *Planta Daninha*. 2009; **27** (3), 507-513. ISSN 1806-9681.

25. MENEZES FILHO, A. C. P.; SANTOS, M. C.; CASTRO, C. F. S. "Estudo fitoquímico,

bioativo, fotoprotetor e físico-químico do extrato floral de algodãozinho do Cerrado

[Cochlospermum regium Schrank. (Pilg.)] – Bixaceae". Arquivos Científicos. 2020; 3 (2), 59-

71. ISSN 2595-4407.

26. DEGÁSPARI, C. H.; WASZCZYNSKYJ, N. "Propriedades antioxidantes de compostos

fenólicos". Visão Acadêmica. 2004; 5 (1), 33-40. ISSN 1518-8361.

27. CAMACHO-CAMPOS, C.; PÉREZ-HERNÁNDEZ, Y.; VALDIVIA-ÁVILA, A.;

RUBIO-FONTANILLS, Y.; FUENTES-ALFONSO, L. "Evaluación

antibacteriana y molusquicida de extractos de hojas de Agave spp." Revista Cubana de

Química. 2020, **32** (3), 390-405. ISSN 2224-5421.

Conflicts of interest

The authors affirm that there are no conflicts of interest.

Authors contributions

Antonio Carlos Pereira de Menezes Filho: collection of plant material, extract production,

physical-chemical and chemical analysis and final writing.

Mariana Chaves: Analysis of spectrophotometry data.

Carlos Frederico de Souza Castro: coordinator.

Josemar Gonçalves de Oliveira Filho: antioxidant analysis, total phenols, text revision and

writing of the article.

Marcela Christofoli: revision of the text and writing of the article.

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347

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