



Revista de Biología Tropical

ISSN: 0034-7744

ISSN: 0034-7744

Universidad de Costa Rica

Castrejón-Varela, Alejandra; Pérez-García, Blanca;
Mendoza-Ruiz, Aniceto; Espinosa-Matías, Silvia
Gametophyte morphology of *Acrostichum aureum* and *A. danaeifolium* (Pteridaceae)
Revista de Biología Tropical, vol. 66, no. 1, 2018, pp. 178-188
Universidad de Costa Rica

DOI: 10.15517/rbt.v66i1.27815

Available in: <http://www.redalyc.org/articulo.oa?id=44955366014>

- ▶ How to cite
- ▶ Complete issue
- ▶ More information about this article
- ▶ Journal's homepage in redalyc.org

UAEM 

Scientific Information System Redalyc

Network of Scientific Journals from Latin America and the Caribbean, Spain and Portugal

Project academic non-profit, developed under the open access initiative

Gametophyte morphology of *Acrostichum aureum* and *A. danaeifolium* (Pteridaceae)

Alejandra Castrejón-Varela¹, Blanca Pérez-García¹, Aniceto Mendoza-Ruiz¹
& Silvia Espinosa-Matías²

1. Área de Botánica Estructural y Sistemática Vegetal, Departamento de Biología, Universidad Autónoma Metropolitana-Iztapalapa, Av. San Rafael Atlixco 186, Col. Vicentina, C.P. 09340, Ciudad de México, México; alecastrejón13690@gmail.com, bpg@xanum.uam.mx, amr@xanum.uam.mx
2. Laboratorio de Microscopía Electrónica de Barrido, Facultad de Ciencias, Universidad Nacional Autónoma de México, Av. Universidad 3000, C.P. 04510, Ciudad de México, México; semfc2010@gmail.com

Received 01-VI-2017. Corrected 26-X-2017. Accepted 22-XI-2017.

Abstract: *Acrostichum* is a pantropical genus and has four species, two of which occur in the Neotropics, *A. aureum* and *A. danaeifolium*. In Mexico, *A. danaeifolium* grows further in land wet soils and is much more common than *A. aureum*, which is typically found in brackish or saline habitats near the coast, and is restricted to coastal saline mangrove communities. The purpose of this paper was to describe and compare the morphogenesis of the sexual phase of *A. aureum* and *A. danaeifolium* for systematic purposes. For this, spores of each species were sown in Petri dishes with agar, previously enriched with sterilized Thompson's medium. To avoid contamination and dehydration, the dishes were kept in transparent plastic bags under laboratory conditions. For the micro-morphological observation with SEM, the gametophyte development phases were fixed in FAA with 0.8 % sucrose for 24 h. Photomicrographs of spores, development stages of gametophytes and young sporophytes were observed with scanning electron microscope. Our results showed that the spores of both species are triletes, globose and positive photoblastic. Germination is Vittaria-type; the germinate filaments are short and uniseriate (5 to 7 cells), and prothallial development is *Ceratopteris*-type. The adult gametophytes of both species have asymmetrical wings. Adult gametophytes in culture are cordiform-spatulate. Antheridia have a broad basal cell, an annular cell, and an asymmetric opercular cell. Archegonia have short necks and four triangular cells at the mouth of the neck. The first leaf of the sporophyte is lobed, with dichotomous veins and anomocytic stomata. The gemmae are formed in adult gametophytes in both species. The development of the gametophyte of *A. aureum*, *A. danaeifolium* and *A. speciosum* share many similarities such as the development of a lateral meristem, asymmetric nature of the mature prothallus, lack of hairs on the prothallus, and undivided asymmetrical opercular antheridia morphology. The genus *Acrostichum* is the sister group of *Ceratopteris*, another genus of aquatic ferns; they differ in the antheridium morphology, *Acrostichum* has an asymmetric opercular cell and *Ceratopteris* shows an undivided cap cell, but the notable difference is the sporophyte morphology. Rev. Biol. Trop. 66(1): 178-188. Epub 2018 March 01.

Key words: development; ferns; germination; scanning electron microscope; sexual phase; spore.

Acrostichum L. is a member of the Pteridaceae family and Parkerioideae subfamily (Smith et al., 2006; PPG I, 2016); it belongs to the *Ceratopteridoid* clade and is sister to *Ceratopteris*, and represent the only aquatic genera of Pteridaceae (Schuettpelz & Pryer, 2007). *Acrostichum aureum* L. is pantropical, *A. danaeifolium* Langsd. & Fisch. neotropical and *A.*

speciosum Willd. paleotropical. The genus is typically found in brackish or saline habitats near coasts. It can form dense stands following logging of mangroves and in freshwater swamps (Mickel & Smith, 2004), but it also can grow farther in land wet soil (León, 1990).

In Mexico only *A. aureum* and *A. danaeifolium* are found, which can be easily

distinguished among them by the puberulent lower surfaces of the pinnae in *A. danaeifolium*, and glabrous in *A. aureum*. Also, the paraphyses are distinctive, being capitate with a dark and circular-lobed apex in *A. aureum*, and pale and non-capitate in *A. danaeifolium* (Adams & Tomlinson, 1979). Another difference is that in *A. aureum*, the leaves are fertile only in the five distal pinna pairs and terminal pinna, whereas in *A. danaeifolium* the fertile leaves are soriferous throughout their length. In general, *A. danaeifolium* is much more common than *A. aureum* (Mickel & Smith, 2004).

The first study of *Acrostichum* gametophyte was made by Schumann (1915). He studied and described the *A. aureum* gametophyte and sporophyte development, and compared its sporophyte with other acrostichoid species of *Leptochilus* Kaulf., *Stenosemia* C. Presl and *Stenochlaena* J. Sm., and he described the asymmetrical antheridia of *Acrostichum* as a peculiarity of this genus.

Nayar and Kazmi (1964) studied the gametophyte and young leaves of *A. aureum*. Gametophyte development is similar to the one reported by Stokey and Atkinson (1952) for *A. speciosum*. In *A. aureum* and *A. danaeifolium*, the prothallial plate expands laterally by the increased activity of some of the lateral intercalary cells of the ribbon thallus, and develops into a broad spatulate plate in which a meristem is differentiated on the side facing the posterior end of the prothallus, contrary to the condition in *A. speciosum* (Nayar & Kazmi, 1964).

Lloyd and Gregg (1975) described the gametophyte development of *A. danaeifolium* as similar to the reports made by Stokey and Atkinson (1952) and Nayar and Kazmi (1964). Eakle (1975) evaluated spore germination of *A. aureum* in experimental environmental conditions, and found that a photoperiodic quantitative long-day phenomenon occurs, with a length between 8 (dark) and 16 (light) hours per day. Lloyd (1980) described the development of the gametophyte of *A. aureum* coinciding with other authors on its morphology, and assessed sexual ontogeny. On the other hand, the juvenile leaves of the sporophyte

are generally spatulate and nearly entire. The single vein supplying the lamina is forked once or twice they are dichotomizing two or three times. All the youngest leaves possess a simple lamina with a nearly entire margin, though herbaceous, the lamina is thick, lack of hairs (Stokey & Atkinson, 1952).

Kshirsagar and Mehta (1979) studied the "in vitro" life history of the fern *A. aureum*, made by factors that influence sex expression, density of gametophytes, pH, and sucrose in the gametophytes, and examined the formation of normal and apogamous sporophytes. They found that the gametophytes were potentially bisexual, but a neutral or alkaline pH favors the expression of archegonia. Growth responses of *A. aureum* suggest it can be classified as a true halophyte, whereas those of *A. danaeifolium* suggest it is a semi-halophyte (Li & Ong, 1997a, 1997b, 1998; Sun, Li, & Ong, 1999).

The aim of this work was to study the spore morphology, spore germination, gametophyte development, and young sporophytes morphology of *A. aureum* and *A. danaeifolium*, to obtain characters for systematic comparison, and contribute to the knowledge of the ontogenetic development of the sexual phase.

MATERIAL AND METHODS

Study site and spore collection: Collections were made at Sontecomapan lagoon, Catemaco, Veracruz, Mexico, in a mangrove swamp at sea level, 18.59° N, 95.08 W. A total of 10-20 fertile pinnae of *A. aureum* (FNG293A and ACV01) were collected on 14 May 2014, dry season, and *A. danaeifolium* (FNG293B) were collected 22 June 2013, rainy season; vouchers were deposited at the UAMIZ (Universidad Autónoma Metropolitana, Iztapalapa Herbarium). Fertile pinnae were kept in paper bags envelopes and dried for eight days at room temperature to favor sporangia opening and subsequent spore liberation. The material was then sieved with a metallic mesh (with pores 0.074 mm in diameter) to eliminate traces of sporangia.

Sowing: Spores were sown in four Petri dishes (5 cm in diameter), containing agar, previously enriched with sterilized Thompson media (Klekowski, 1969). The spores were spread on the surface of solidified medium with a thin brush, with an average density of 40 for *A. aureum* and 36 spores/cm² for *A. danaeifolium*. To avoid contamination and dehydration, the dishes were kept within transparent plastic bags in growth chamber under a 12 h light/dark photoperiod (Lumistell ICP-19). To determine photoblastism, one of the dishes was kept in the dark.

Electron microscopy: Dry spores were spread with a brush and mounted on aluminum stubs using double-sided tape, in accordance with Bozzola and Russell (1998), and were coated with gold using a Denton Vacuum Desk II sputter coated. Besides, gametophytes were fixed in 0.8 % FAA-sucrose for 24 h; then, they were washed, dehydrated in a graded ethanol series, and critical-point dried by means of a Leica CPD 030 critical point dryer. After drying they were mounted in a similar manner as the spores. Photomicrographs of spores, prothalli, and the young sporophytes were made with a Jeol JSM5310-LV scanning electron microscope. Digital images were obtained in TIFF format using the Orion-Joel digital system.

RESULTS

Spore morphology: Trilete, tetrahedral-globose to nearly globose, radially symmetrical, in proximal view. Spores of both species in proximal view, measured an average of 19(23)28 µm for *A. danaeifolium* (Fig. 1A), and 19(22)25 µm for *A. aureum* (Fig. 1B). They are homosporous, hyaline with a weakly brownish color, non-chlorophyllous, with the laesura short. Differences between the spores of the two species are found in the exine. In *A. aureum*, the exine is slightly rougher with the surface papillate to tuberculate, with rodlets attached to the tubercles in proximal and distal view. Whereas the exine of *A. danaeifolium* is rugose, with strands or rods mostly associated

with coarsely papillate structure, papillate surface in proximal and distal view.

Germination and filamentous phase:

In accordance with Mendoza-Ruiz (2001), the criterion used in this study to determine spore germination is the formation of the first rhizoid and/or prothallial cells or both simultaneously. Under our culture conditions, germination took place on the 5-6 days after sowing. The germination percentage was high (90 - 95 %) for *A. danaeifolium*, but lower (70 %) for *A. aureum*. After 100 days, none of the spores grown in darkness germinated.

In both species, the basal cell second division produces a brownish rhizoid that emerges through the laesura, and forms the first initial filament cell (Fig. 1C-D). The cell wall plane of the second division is perpendicular to the cell wall plane of the first division. The preceding type of germination is defined as the *Vittaria*-type.

The filament initial cell then divides in the same plane to form a filament of 9-13 cells, in *A. aureum* (29 - 40 days), and *A. danaeifolium* (18 days) (Fig. 1D). The filament barrel-shaped have cells with numerous chloroplasts, and develop 2 or 3 hyaline and non-chlorophyllous rhizoids, which are longer than the filament. The basal cell remains in the spore wall through the subsequent development of the vegetative and adult phases of the gametophyte. This phase is very brief, because one day later, the filamentous gametophytes begin to differentiate into laminar gametophytes.

Laminar phase: According to our observations, prothallial development for *A. danaeifolium* and *A. aureum* correspond to the Ceratopteris-type (Fig. 1E-F and Fig. 2A-C). Formation of a prothallial plate is initiated by longitudinal divisions in the anterior cells (often including the terminal cell) of the germ filament. The prothallial plate is spatulate with 55 cells in *A. danaeifolium* (24 - 28 days) and near 100 cells in *A. aureum* (33 - 40 days), a pluricellular meristem is established in this non-meristic plate (Fig. 2B).

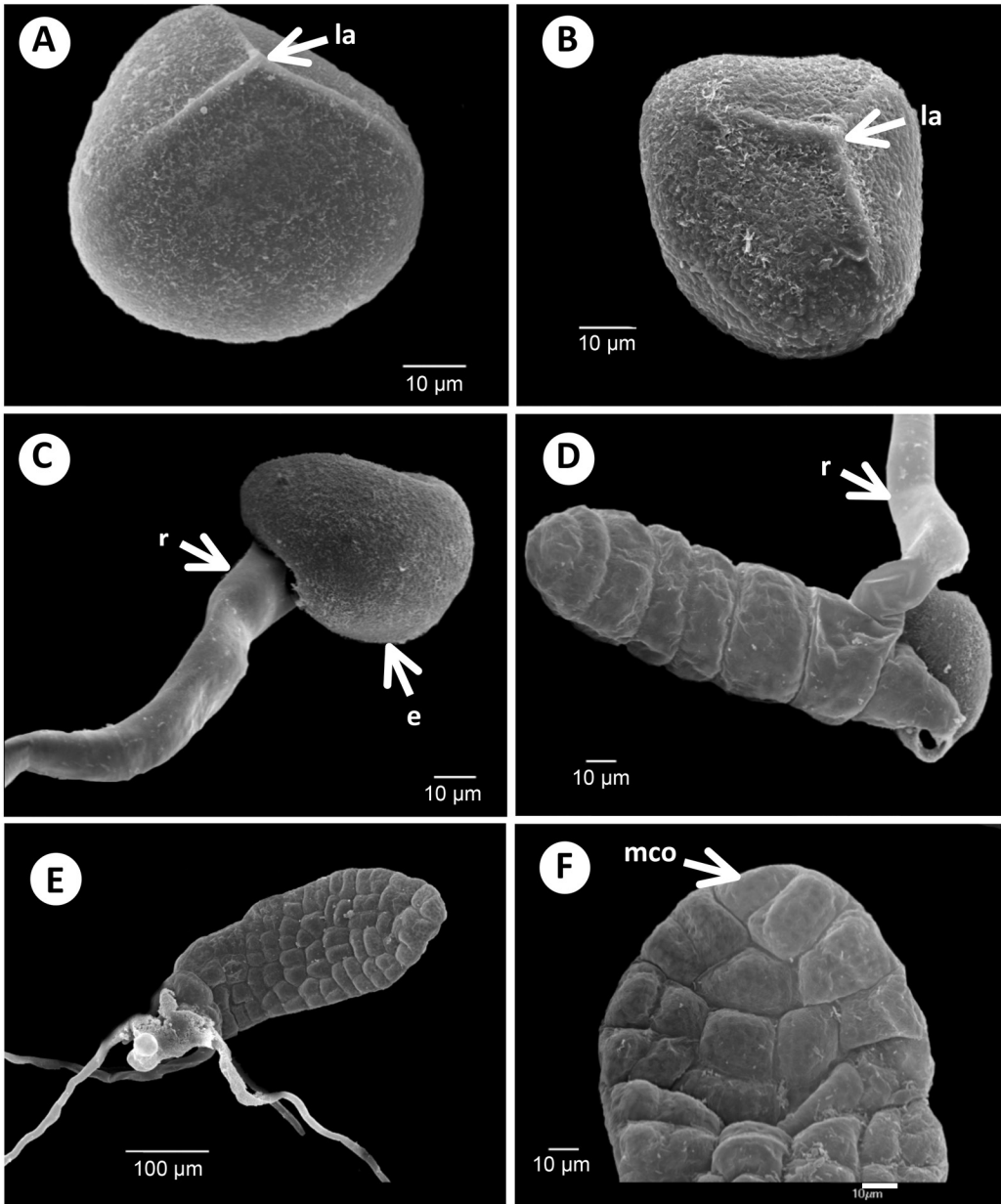


Fig. 1. Spore morphology, early germination, and young gametophytes of *Acrostichum*: A) Spore proximal view of *A. danaeifolium*. B) Spore proximal view of *A. aureum*. C) Beginning germination of *A. danaeifolium*, 18 days. D) Filament gametophyte of *A. danaeifolium*, 18 days. E) Bidimensional gametophyte of *A. danaeifolium*, 22 days. F) Close up of meristematic cell of *A. danaeifolium*, 24 days. la. Laesura. mco. Meristematic cell obconic. e. Exine. r. Rhizoid.

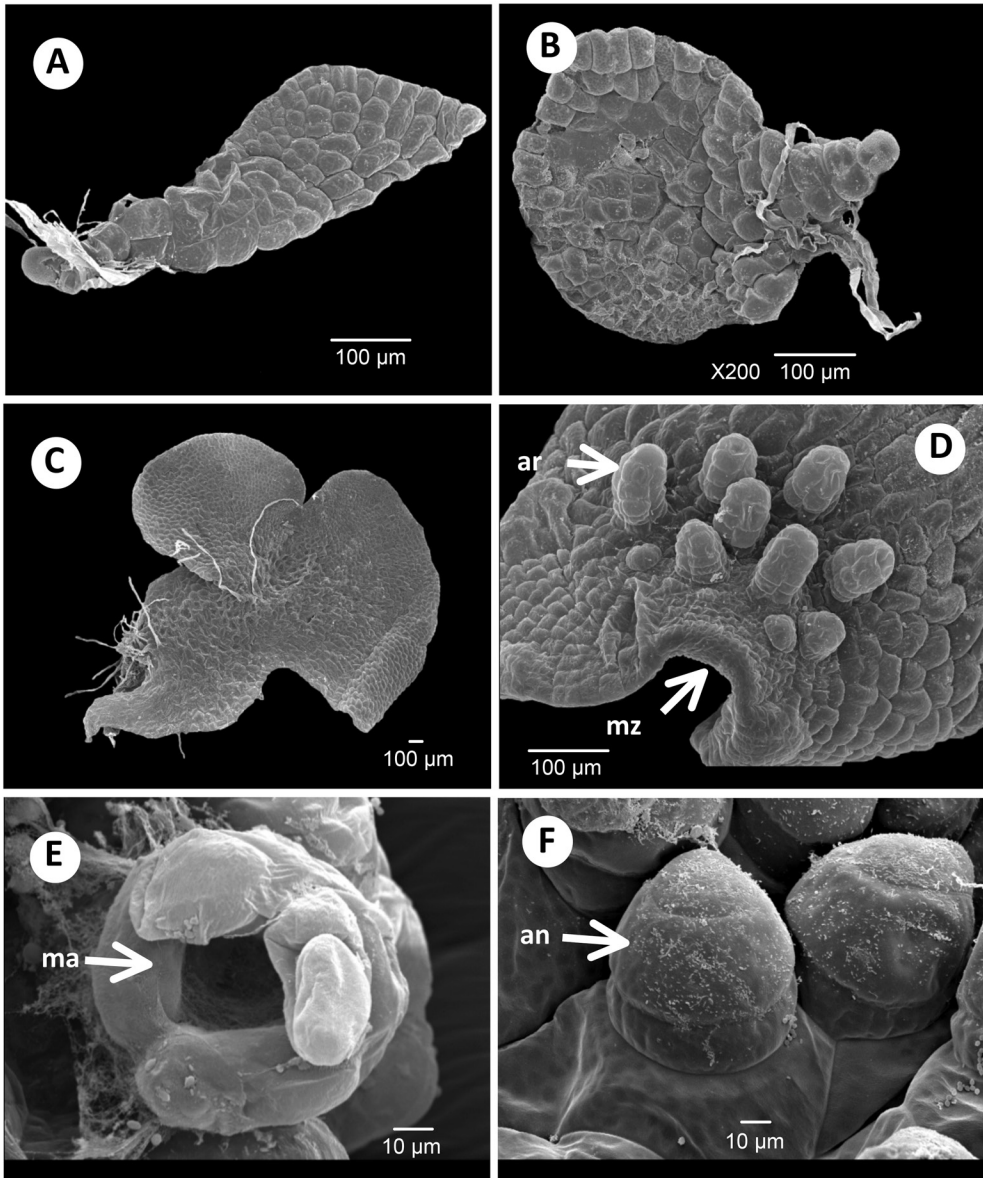


Fig. 2. Young gametophytes and gametangia of *Acrostichum*: A) Bidimensional gametophyte of *A. aureum*, 29 days. B) Bidimensional gametophyte of *A. aureum*, 40 days. C) Bidimensional gametophyte of *A. aureum*, 70 days. D) Archegonia of *A. danaeifolium*, 90 days. E) Archegonia mouth of *A. danaeifolium*, 181 days. F) Antheridia of *A. danaeifolium*, 185 days. an. Antheridia. ar. Archegonia. ma. Mouth of archegonia. mz. Meristematic zone.

The young prothallus of both species is distinctly lopsided, and the asymmetry often persists until maturity. The meristem is established laterally, more towards the basal end of the prothallial plate than towards the apex. The asymmetry of the young prothallus is only transitory, soon the meristematic region becomes apical, and a symmetrical cordate thallus is formed (Fig. 2C).

Adult gametophytes: Thirty days after sowing, the adult gametophyte in both species of *Acrostichum* is cordiform-spathulate or cordiform-reniform, and glabrous. It has broad wings, a thin cushion, and a deep notch. The wings are clearly demarcated (Fig. 2C). Gametophytes reach maturity in approximately 90 days in *A. danaeifolium* and 76 - 196 days in *A. aureum*. Gametophytes of both species are bisexual with antheridia and archegonia on the same gametophyte (Fig. 2D-F). Sex organs are of the common leptosporangiate type. Gametangia appeared only in the abaxial surface of the gametophyte with some archegonia near the notch (Fig. 2D), and many antheridia distributed mainly towards the base, mixed between rhizoids.

Archegonia appeared first in *A. aureum* (76 days) and later in *A. danaeifolium* (90 days), they may be either protandrous or protogynous. The neck of the mature archegonia is short and curved away from the notched end of the gametophyte, and it is composed of four rows each with four cells long, and slightly curved towards the apical region (notch) of the gametophyte (Fig. 2D). The mouth consists of four cells. Archegonia have an egg cell, a ventral-canal cell, and a neck-canal cell, which are surrounded by the layer of cells of the cover (Fig. 2E).

Antheridia are hemispheric to subglobose and develop on the abaxial surface of the cushion and wings of cordiform-spatulate gametophytes of *A. aureum* (196 days) and *A. danaeifolium* (135 days) (Fig. 2F and 3F). Antheridia consist of a basal cell, an annular cell, and an undivided asymmetrical opercular cell (Fig. 3A). Antheridial dehiscence was not

observed. Also, cordiform-spathulate gametophytes with abundant marginal and superficial antheridia were observed after 185 days of growth in *A. danaeifolium*.

Sporophytes: Despite the addition of droplets of sterile water to the cultures to facilitate antherozoid movement, in both species fertilization occurred after 250 days. Young sporophytes became visible on the prothalli in *A. aureum* at 250 days and in *A. danaeifolium* after 216 days, from spores sowing. These are glabrous and dichotomously branched with their first leaves depressed obovate, and with open dichotomous venation (Fig. 3C). The petiole is long terete, and leaf epidermal cells are distinctly wavy and the stomata are of the anomocytic-type, with two guard cells that have a length of approximately 30 μm , and the stomata are surrounded by epidermal undulate cells wall (Fig. 3D).

Vegetative propagation: These gemmae become visible in *A. danaeifolium* at 185 days and *A. aureum* at 196 days (Fig. 3B and 3E-F). The prothalli produced gemmae which, when shed, develop into new prothalli. The gemmae are pluricellular, dumbbell-shaped, densely chlorophyllous structures. In fact, a single prothallial cell may give rise to many gemmifers each of which generates one or more gemmae. The presented results are compared with other authors in Table 1.

DISCUSSION

Researchers such as Schumann (1915), Stokey and Atkinson (1952), Nayar and Kazmi (1964), Nayar and Kaur (1969 and 1971), Lloyd and Gregg (1975), Kshirsagar and Mehta (1979), Lloyd (1980), Lloyd and Buckley (1986) described the morphology of the gametophyte of *A. aureum*, *A. danaeifolium*, and *A. speciosum*, and our observations of gametophyte development data showed that they are similar, the difference being that the morphology is clearer with the scanning electron microscope.

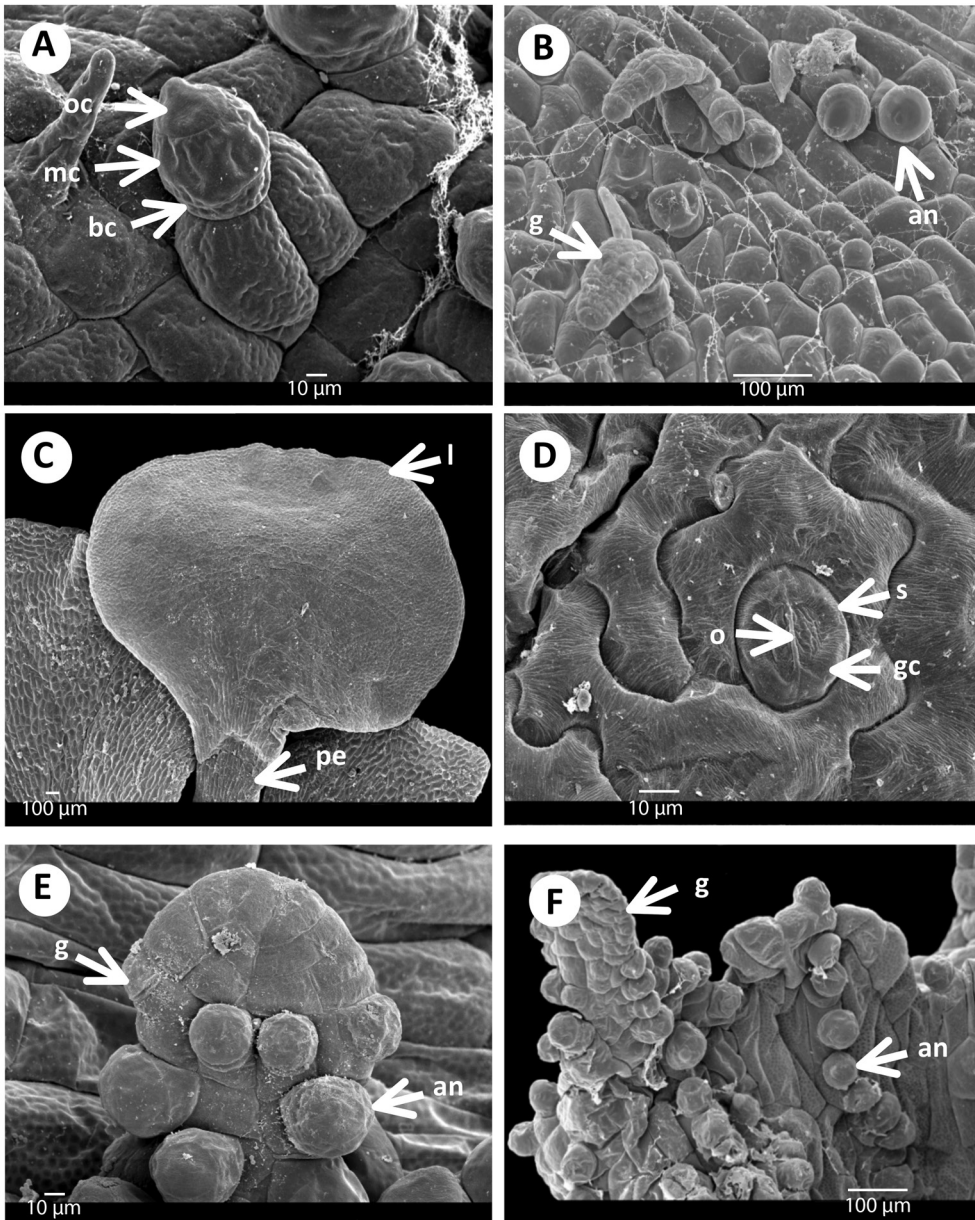


Fig. 3. Gametangia and first leaf of *Acrostichum*: A) Antheridia of *A. danaeifolium* with asymmetrical opercular cell, 185 days. B) Gametophyte of *A. aureum* with antheridia and gemmae, 196 days. C) First leaf of *A. aureum*, 250 days. D) Anomocytic stomata of *A. danaeifolium*, 216 days. E) Gemmae on the gametophyte with antheridia of *A. danaeifolium*, 185 days. F) Gemmae with antheridia of *A. aureum*, 185 days. an. Antheridia. bc. Basal cell. g. Gemmae. gc. Guard cell. l. Lamina. mc. Medial cell. o. Ostiole. oc. Opercular asymmetrical cell. pe. Petiole. s. Stomata.

TABLE 1
Morphological comparison of the characteristics of the species of the genus *Acrostichum* and the genus *Ceratopteris*

Character/taxa	<i>A. aureum</i>	<i>A. danaeifolium</i>	<i>A. speciosum</i> *	<i>Ceratopteris</i> *
Origin	Pantropical	Neotropical (American)	Paleotropical (Australasian)	Tropics
Habitat	Brackish and saline mangroves and swamps	Mangroves, brackish, saline or freshwater swamps, edges of hot springs	Mangroves, brackish and freshwater marshes	Aquatic and fresh or occasionally brackish
Sterile pinnae	Glabrous	Puberulent	Glabrous	Glabrous, succulent
Sporangia	Acrostichoid	Acrostichoid	Acrostichoid	1-4 rows along the rib
Spores: size and laesura	Trilete, tetrahedral-globose 19-25 µm	Trilete, tetrahedral-globose 19-28 µm	Trilete, tetrahedral-globose	Trilete, parallel crested 70-160 µm
Type of germination	Vittaria	Vittaria	Vittaria	Vittaria
Type of prothallial development	Ceratopteris	Ceratopteris	Ceratopteris	Ceratopteris
Young prothalli	Lopsided	Lopsided	Lopsided	Lopsided
Adult form gametophyte	Cordiform-spathulate	Cordiform-spathulate and glabrous	Cordiform-elongate	Cordiform
Antheridia	Asymmetrical	Asymmetrical	Asymmetrical	Undivided cap cell
Sporophyte (days)	250	216	Not date	Not date
Vegetative propagation (days)	196	185	Not date	Not date

* Stokey and Atkinson (1952); Tryon and Tryon (1982); Tryon and Lugardon (1990).

The difference between the two species under study lies in the ornamentation of the exine, and this was described before by Erdtman (1971), Nayar and Kaur (1971), Lloyd and Gregg (1975), García de López (1978), Adams and Tomlinson (1979), Tryon and Tryon (1982), Kubitzki (1990), Tryon and Lugardon (1992), Moran, (1995), Mickel and Smith (2004) our data support the descriptions of these authors.

In most homosporous ferns, spore germination produces a primary rhizoid, followed by a germinal multicellular filament subsequent prothallial development based on the plane of cell division, and in the direction of the growth of the rhizoid and prothallial cell, this pattern of development was observed in *A. aureum* and *A. danaeifolium*. Early in germination, there is not significant morphological variation in day (5-7 / 6-7 days germination after sowing), these data are consistent with what was mentioned by Stokey and Atkinson (1952) and Nayar and Kazmi (1964).

Filament phase which is usually 6-8 cells but sometimes 10-12 cell long, the filamentous phase are 6-12 prothallial cells with barrel form and chloroplast (Stokey & Atkinson, 1952) and correspond to Vittaria-type germination that is, the basal cell (spore cell) divides to form a rhizoid cell. The cell wall plane is parallel to the equatorial plane of the spore. The second cell division forms the first filament initial cell, and the cell wall plane is at a right angle to that of the division, or parallel to the polar axis of the spore (Stokey & Atkinson, 1952). The prothalli of *Acrostichum aureum* and *A. danaeifolium* resemble that of *A. speciosum* (Stokey & Atkinson, 1952) in morphology and development, but there are minor differences especially with regard to the development of a prothallial plate.

In *A. aureum* and *A. danaeifolium* prothallial development is of the Ceratopteris-type; in this type of development, the meristem established laterally in young gametophytes is asymmetric (lopsided), for a shorter period during its development; all species in this study shared this character.

Sex organs are of the common leptosporangiate type in species of study, adult gametophytes are cordiform and glabrous. The asymmetrical antheridia, we consider it as a synapomorphy for *Acrostichum* (Stokey & Atkinson, 1952; Nayar & Kazmi, 1964; Nayar & Kaur, 1969, 1971; Lloyd & Gregg, 1975; Lloyd, 1980; Lloyd & Buckley, 1986). This asymmetry was described by Schumann (1915) for *A. aureum*; nevertheless, the opening of the cap cell was not seen in this investigation. Antheridial dehiscence occurs through a pore in the opercular cell (Nayar & Kazmi, 1964).

Archegonia are morphologically standardized and have little significant differences in numbers of neck cells and length (Stokey & Atkinson, 1952; Nayar & Kazmi, 1964; Nayar & Kaur, 1969, 1971).

In both species the sporophyte developed after the 250 days sowing, and they have on its abaxial epidermis stomata of anomocytic type that lack of subsidiary cells as described Van Cotthem (1984).

The development and structure of the gametophytes show some similar characteristics between the sister group *Acrostichum* and *Ceratopteris*: 1) Spore germination is of the *Vittaria*-type; 2) Prothallial developments is of the *Ceratopteris*-type; 3) Young prothalli are markedly lopsided; 4) Gametangia are the common leptosporangiate type.

However *Acrostichum* and *Ceratopteris* showed different features: 1) *Acrostichum* is typically found in brackish or saline habits near coasts, true halophyte or semi-halophyte, while *Ceratopteris* inhabits fresh or occasionally brackish wetlands and rarely in terrestrial situations on poorly drained soils worldwide of tropics and subtropics (Dettmann & Clifford, 1991); 2) Spores in *Acrostichum* and *Ceratopteris* are trilete, tetrahedral-globose to nearly globose, but they differ in ornamentation, 3) Antheridia in *Acrostichum* have an asymmetric opercular cell whereas *Ceratopteris* showed undivided cap cell.

The morphological similarities found in the study of the sexual phase of *A. danaeifolium* and *A. aureum* are important data for

taxonomic purposes at a generic level, and if we add the knowledge of the sporophyte phase of the genus *Acrostichum* and *Ceratopteris*, the data supports the molecular phylogeny results, which place them as sister groups. All recent molecular data support *Ceratopteris* as a member of the Pteridaceae *s.l.* (Gastony & Rollo, 1995; 1998; Hasebe et al., 1995, Pryer, Smith, & Skog, 1995; Smith et al., 2006). Five primary clades of Pteridaceae have been resolved by molecular phylogenetic analyses (Prado, Del Nero Rodrigues, Salatino, & Salatino, 2007; Schuettpelz, Schneider, Huiet, Windham, & Pryer, 2007; Schuettpelz & Pryer, 2007). In all of these analyses, *Ceratopteris* is the sister group of *Acrostichum* (Mickel & Smith, 2004; PPG I, 2016).

The knowledge of the vegetative and reproductive morphology of gametophytes and characteristics such as spore morphology, spore germination pattern, gametophyte form, presence or absence of hairs, photosynthetic gametophytes or not, position, shape and number of antheridia and archegonia cells, number of cells of the antheridium wall and number of archegonia neck cells and gametophytes gemmae production, are diagnostic characters that will be important in future to apply in the knowledge of the phylogeny of ferns based on morphology and *rbcL* sequences (Pryer et al., 1995).

ACKNOWLEDGMENTS

This research was supported by the Universidad Autónoma Metropolitana, Unidad Iztapalapa. We thank the editor, the reviewers for valuable comments and suggestions, which substantially improved the manuscript, to Robbin C. Moran for suggestions on the manuscript, and Felipe Gómez Noguez for providing biological material.

RESUMEN

Morfología del gametofito de *Acrostichum aureum* y *Acrostichum danaeifolium* (Pteridaceae). *Acrostichum* es pantropical y tiene cuatro especies dos de las cuáles

ocurren en el Neotrópico, *A. aureum* y *A. danaeifolium*. El género se encuentra típicamente en hábitats salobres o salinos cerca de la costa. También puede crecer más lejos en tierra en suelos húmedos. *Acrostichum aureum* está restringido a las comunidades de manglares salinos costeros. *A. danaeifolium* es mucho más común que *A. aureum* en México. El propósito de este trabajo es describir y comparar la morfogénesis de la fase sexual de *A. aureum* y *A. danaeifolium*. Las esporas de cada especie fueron sembradas en cajas de Petri, en agar previamente enriquecido con medio de Thompson esterilizado. Para evitar la contaminación y la deshidratación, las cajas se mantuvieron dentro de bolsas de plástico transparentes en condiciones de laboratorio. Para la observación micro-morfológica con el MEB, las fases de desarrollo del gametofito se fijaron en FAA con sacarosa al 0.8 % durante 24 h. Se observaron las esporas, fases de desarrollo de gametofitos y esporofitos jóvenes con un microscopio electrónico de barrido. Las esporas de ambas especies son triletes, globosas y fotoblásticas positivas. La germinación es tipo-Vittaria, los filamentos germinativos son cortos y uniseriados (5 a 7 células) y el desarrollo protálico es de tipo-Ceratopteris. Los gametofitos de ambas especies tienen alas asimétricas. Los gametofitos adultos en cultivo son cordiforme-espátulados. Los anteridios tienen una célula basal amplia, una célula anular y una célula opercular asimétrica. Los arquegonios tienen cuellos cortos y cuatro células triangulares en la boca del cuello. La primera hoja del esporofito es lobulada, con venación dicotómica y estomas anomocíticos. Las yemas se forman en gametofitos adultos en ambas especies. El desarrollo del gametofito de *A. aureum*, *A. danaeifolium* y *A. speciosum* comparten características tales como el desarrollo de un meristemo lateral, el protalo maduro de naturaleza asimétrica, ausencia de pelos en el protalo y anteridio con célula opercular asimétrica no dividida. *Acrostichum* es el grupo hermano de *Ceratopteris*, otro género de helecho acuático, que difieren en la morfología del anteridio, *Acrostichum* tiene una célula opercular asimétrica y *Ceratopteris* muestran una célula opercular no dividida.

Palabras clave: desarrollo; espora; fase sexual; germinación; helechos; microscopio electrónico de barrido.

REFERENCES

Adams, S., & Tomlinson, P. B. (1979). *Acrostichum* in Florida. *American Fern Journal*, 69, 42-46.

Bozzola, J. J. & Russell, L. D. (1998). *Electron microscopy: principles and techniques for Biologists*. London, England: Jones y Bartlett Publishers.

Dettmann, M. E., & Clifford, H. T. (1991). Spore morphology of *Anemia*, *Mohria*, and *Ceratopteris* (Filicales). *American Journal of Botany*, 78, 303-325.

Eakle, T. W. (1975). Photoperiodic control of spore germination of *Acrostichum aureum*. *American Fern Journal*, 65, 94-95.

Erdtman, G. (1971). *Pollen and spore morphology. Plant taxonomy*. Stockholm, Almqvist & Wiksell, New York: Ronald Press Co.

García de López, I. (1978). Revisión del género *Acrostichum* en la República Dominicana. *Moscosoa*, 1, 64-70.

Gastony, G. J., & Rollo, D. R. (1995). Phylogeny and generic circumscriptions of cheilantheid ferns (Pteridaceae: Cheilantheoideae) inferred from *rbcL* nucleotide sequences. *American Fern Journal*, 85, 341-360.

Gastony, G. J., & Rollo, D. R. (1998). Cheilantheid fern (Pteridaceae: Cheilantheoideae) in the southwestern United States and adjacent Mexico- a molecular phylogenetic reassessment of generic lines. *Aliso*, 17, 131-144.

Hasebe, M., Wolf, P. G., Pryer, K. M., Ueda, K., Ito, M., Sano, R., Gastony, G. J., Yokoyama, J., Manhart, J. R., Murakami, N., Crane, E. H., Haufler, C. H., & Hauk, W. D. (1995 [1996]). Fern phylogeny based on *rbcL* nucleotide sequences. *American Fern Journal*, 85, 134-181.

Klekowski, E. J. Jr. (1969). Reproductive biology of the Pteridophyta III: a study of the Blechnaceae. *Botanical journal of the Linnean Society. London*, 62, 361-377.

Kshirsagar, M. K., & Mehta, A. R. (1979). *In vitro* studies of the life cycle of *Acrostichum aureum* L. In S. S. Bir (Ed.), *Recent Researches in Plants Sciences* (pp. 71-73). New Delhi: Kalyani Publ.

Kubitzki, K. (1990). *The families and genera of vascular plants*. New York: Springer-Verlag.

León, B. (1990). New localities for *Acrostichum danaeifolium* in Peru. *American Fern Journal*, 80, 113-114.

Li, X.-P., & Ong, B.-L. (1997a). Ultrastructural changes in gametophytes of *Acrostichum aureum* L. cultured in different sodium chloride concentrations. *Biologia Plantarum*, 39, 607-614.

Li, X.-P., & Ong, B.-L. (1997b). Tolerance of gametophytes of *Acrostichum aureum* (L.) to salinity and water stress. *Photosynthetica*, 34, 21-30.

Li, X.-P., & Ong, B.-L. (1998). Responses of the photosynthesis to NaCl in gametophytes of *Acrostichum aureum*. *Physiologia Plantarum*, 102, 119-127.

Lloyd, R. M. (1980). Reproductive biology and gametophyte morphology of New World populations of *Acrostichum aureum*. *American Fern Journal*, 70, 99-110.



- Lloyd, R. M., & Buckley, D. P. (1986). Effects of salinity on gametophyte growth of *Acrostichum aureum* and *A. danaeifolium*. *Fern Gazette*, 13, 97-103.
- Lloyd, R. M., & Gregg, T. L. (1975). Reproductive biology and gametophyte morphology of *Acrostichum danaeifolium* from Mexico. *American Fern Journal*, 65, 105-120.
- Mendoza-Ruiz, A. (2001). *Morfogénesis de la fase sexual de pteridofitas mexicanas, familia Dryopteridaceae*. (Tesis de Maestría) Universidad Nacional Autónoma de México, México, D. F., México.
- Mickel, J. T., & Smith, A. R. (2004). *The Pteridophytes of Mexico. Memoirs of the New York Botanical Garden*. New York, USA: The New York Botanical Garden.
- Moran, R. C. (1995). 1. *Acrostichum* L. In: R. C. Moran., & R. Ribá. (Eds.), *Flora Mesoamericana. Vol. 1, Psilotaceae a Salviniaceae* (pp. 105-106). México: Universidad Nacional Autónoma de México (Instituto de Biología) Missouri Botanical Garden & The Natural History Museum (London).
- Nayar, B. K., & Kaur, S. (1969). Types of prothallial development in homosporous ferns. *Phytomorphology*, 19, 179-188.
- Nayar, B. K., & Kaur, S. (1971). Gametophytes of homosporous ferns. *The Botanical Review*, 37, 295-396.
- Nayar, B. K., & Kazmi, F. (1964). The gametophyte of *Acrostichum aureum* L. *Proceeding Indian Academy of Science*, 59, 185-194.
- PPG I (Pteridophyte Phylogeny Group). (2016). A community-derived classification for extant lycophytes and ferns. *Journal of Systematics and Evolution*, 54, 563-603.
- Prado, J., Del Nero Rodríguez, C., Salatino, A., & Salatino, M. L. F. (2007). Phylogenetic relationships among Pteridaceae, including Brazilian species, inferred from *rbcL* sequences. *Taxon*, 56, 355-368.
- Pryer, K. M., Smith, A. R., & Skog, J. E. (1995). Phylogenetic relationship of extant ferns based on evidence from morphology and *rbcL* sequences. *American Fern Journal*, 85, 205-282.
- Schuettpelz, E., & Pryer, K. M. (2007). Fern phylogeny inferred from 400 leptosporangiate species and three plastid genes. *Taxon*, 56, 1037-1050.
- Schuettpelz, E., Schneider, H., Huiet, L., Windham, M. D., & Pryer, K. M. (2007). A molecular phylogeny of the fern family Pteridaceae: Assessing overall relationships and the affinities of previously unsampled genera. *Molecular Phylogenetics and Evolution*, 44, 1172-1185.
- Schumann, E. (1915). Die Acrosticheen und ihre Stellung im System der Farne. *Flora*, 108, 210-260.
- Smith, A. R., Pryer, V. E., Schuettpelz, E., Korall, H., Schneider, H., & Wolf, P. G. (2006). A classification for extant ferns. *Taxon*, 55, 705-731.
- Stokey, A. G., & Atkinson, L. R. (1952). The gametophyte of *Acrostichum speciosum* Willd. *Phytomorphology*, 2, 105-113.
- Sun, W.-Q., Li, X.-P., & Ong, B.-L. (1999). Preferential accumulation of D-pinitol in *Acrostichum aureum* gametophytes in response to salt stress. *Physiologia Plantarum*, 105, 51-57.
- Tryon, A. F., & Lugardon, B. (1992). *Spore of the Pteridophyta*. New York: Springer-Verlag.
- Tryon, R. M., & Tryon, A. F. (1982). *Fern and allied plants*. New York: Springer-Verlag.
- Van Cotthem, W. R. J. (1984). Stomatal types and systematics, In: A. C. Jermy., J. A. Crabbe., & B. A. Thomas. (Eds.). *The phylogeny and classification of the ferns* (pp. 59-71). Germany: Academic Press Inc.