

Odovtos International Journal of Dental Sciences

ISSN: 1659-1046 ISSN: 2215-3411

Facultad de Odontología. Universidad de Costa Rica

Reyes-Carmona, Jessie
Irrigation Protocols Effects on Radicular Dentin: Cleaning, Disinfection and Remaining Ultrastructure
Odovtos International Journal of Dental Sciences, vol. 25, no. 1, 2023, January-April, pp. 14-21
Facultad de Odontología. Universidad de Costa Rica

DOI: https://doi.org/10.15517/IJDS.2022.51869

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International Journal of Dental Sciences

https://revistas.ucr.ac.cr/index.php/Odontos | ISSN: 2215-3411

DOI: 10.15517/IJDS.2022.51869

NEW PERSPECTIVE ARTICLE

Received: 21-VI-2022

Irrigation Protocols Effects on Radicular Dentin:

Cleaning, Disinfection and Remaining Ultrastructure

Accepted: 13-VII-2022

Published Online: 19-VII-2022

Efecto del protocolo de irrigación en la dentina radicular: limpieza, desinfección y ultraestructura remanente

Jessie Reyes-Carmona DDS, MSD, PhD1

1. Professor and Researcher, Department of Restorative Sciences, Faculty of Dentistry, University of Costa Rica. San José, Costa Rica. https://orcid.org/0000-0003-2872-6623

Correspondence to: Dr. Jessie Reyes-Carmona - jessie.reyes@ucr.ac.cr

ABSTRACT: A successful endodontic treatment requires a combination of satisfactory mechanical instrumentation, adequate irrigation protocols, and three-dimensional obturation of the canal system. Irrigation is considered the most critical procedure to ensure cleaning and disinfection. To date, a large variety of irrigants has been proposed. However, sodium hypochlorite (NaOCI) remains the gold standard. In order to achieve complete cleaning and disinfection, final irrigation with EDTA and ultrasonic devices has been used as an ideal protocol. Most endodontic research focuses on the cleaning and antibacterial properties of the irrigant solutions. Recent evidence demonstrated that the irrigation protocols cause erosion, affecting the radicular dentin ultrastructure. This article aims to describe the clinical features of the present knowledge concerning the effect of irrigation protocols on radicular dentin.

KEYWORDS: EDTA; Irrigation protocol; Radicular dentin; Sodium hypochlorite.

RESUMEN: Un tratamiento de endodoncia exitoso requiere de una satisfactoria instrumentación mecánica, protocolos de irrigación adecuados y obturación tridimensional del sistema de conductos radiculares. La irrigación se considera el procedimiento más crítico para garantizar la limpieza y desinfección. Hasta la fecha, se ha propuesto una gran variedad de irrigantes. Sin embargo, el hipoclorito de sodio (NaOCl) sigue siendo el estándar de oro. Para lograr una limpieza y desinfección completa, se ha utilizado como protocolo ideal la irrigación final con EDTA y dispositivos ultrasónicos. La mayor parte de la investigación en endodoncia se enfoca en las propiedades antibacterianas y de limpieza de las soluciones de irrigación. Reciente evidencia demostró que los

protocolos de irrigación provocan erosión, afectando la ultraestructura de la dentina radicular. Este artículo tiene como objetivo describir algunas consideraciones clínicas del conocimiento actual sobre el efecto de los protocolos de irrigación en la dentina radicular.

PALABRAS CLAVE: Dentina radicular; EDTA; Protocolo de irrigación; Hipoclorito de sodio.

The American Association of Endodontists (AAE) describes that the primary objective of endodontic treatment is to prevent and intercept pulpal/periradicular pathosis and preserve the natural dentition when affected by pathosis (1). Endodontic treatment involves chemo-mechanical preparation of the root canal system to eliminate organic, inorganic, and bacterial products, in addition, to sealing the radicular space with a biocompatible material (1). Among the essential phases in endodontic treatment are the disinfection and filling of the root canal, which are primordial for the microbial reduction (2).

A vital pulp tissue maintains an immunological defense capacity. However, a microbial infection will develop in a root canal with necrotic pulpal tissue. The colonization of microorganisms within the root canal represents the main etiological factor of periradicular pathologies (Figure 1) (3).

Thus, teeth diagnosed with pulp necrosis must be considered infected, regardless of the presence or absence of a visible radiographic periapical lesion. Pulpal and periapical pathologies are usually inflammatory and of microbial etiology. Microorganisms and their products play a significant role in the induction and perpetualization of these pathologies (2,3).

The infection of endodontic origin has some characteristics that differentiate it from other infections in the organisms since it cannot be treated through systemic antibiotic therapy. As the pulp tissue is necrotic, no blood vessels can transport the cells and molecules of the immune defense system, as well as antibiotics, into the root canal. Moreover, the microorganisms are contained within the root canal system's microstructure. Thus, this type of infection can only be treated through professional intervention with endodontic treatment.

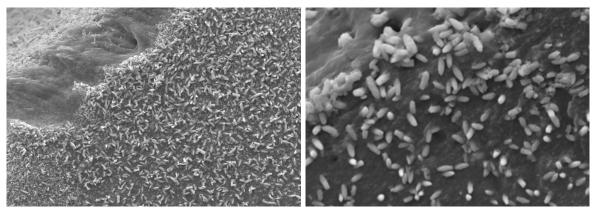


Figure 1. Representative photomicrographs of a radicular canal wall of a tooth diagnosed with pulp necrosis (unpublished data). Photomicrographs were taken with an SEM Philips XL 30 in UFSC, Brazil. A microbial infection will develop in a root canal with necrotic pulpal tissue. Teeth diagnosed with pulp necrosis must be considered infected, regardless of the presence or absence of a visible radiographic periapical lesion.

CLEANING AND DISINFECTION

The root canal treatment success depends on correct chemo-mechanical disinfection. Irrigation is a crucial part of this achievement. Irrigants facilitate the removal of microbes, necrotic and inflamed tissue, and debris (4,5). Moreover, irrigation improves the cutting efficiency of rotary and endodontic files by reducing the friction between the instruments and dentin (5,6).

Complete intracanal disinfection is challenging due to the complex anatomy that root canals generally present. Thus, different irrigating agents are used during biomechanical preparation.

Sodium hypochlorite (NaOCI) is the most worldwide used irrigant in endodontics (1-5). NaOCI is antibacterial, dissolves organic tissue, and removes biofilm due to the hypochloric acid affecting the microbial cell's vital functions, resulting in cell necrosis (5,7,8). Sodium hypochlorite in concentrations of 0.5% to 5.25% has a series of relevant properties, among them the effectiveness of dissolving organic tissue, in addition to being antimicrobial. It is ineffective against specific microorganisms at low concentrations, and at high concentrations, it has low biocompatibility, which can cause periapical inflammation (4). Although sodium hypochlorite is the irrigant of choice, it cannot dissolve inorganic tissue present in the smear layer. Evidence showed that microflora and toxins are accumulated in the smear layer, allowing the survival and reproduction of bacteria (9,10).

The combination of different irrigating solutions, which have the capacity to promote inorganic debris removals, such as 17% ethylenediaminetetraacetic acid (EDTA) and citric acid, has been proposed. Both solutions effectively remove the smear layer, and their chelating action generates the detachment of the biofilm found on the canal walls (11). However, alternating NaOCI

with EDTA during the instrumentation procedure will detriment the antibacterial activity of NaOCl and should be avoided (5). Evidence showed that the tissue previously exposed to EDTA is not effectively dissolved by NaOCl (5,12).

EDTA is a chelator that only affects the inorganic content of the smear layer. Thus, the complete removal of the smear layer can only be achieved when the final irrigation protocol must combine EDTA and NaOCI (13).

The irrigating procedure was usually performed with a syringe and needle. In recent years, several new devices have been launched to enhance the agitation of the irrigation solution to facilitate and improve the cleaning and disinfection of the root canal. Passive ultrasonic irrigation (PUI) consists of transmitting energy from an ultrasonically oscillating instrument to the irrigant solutions in the root canal to improve their antibacterial and dissolution properties (9-11). Other types of equipment introduced to the field of endodontics include the EndoActivator®, EndoVac®, and Vibringe®, among the most used. Several reports have indicated that the various devices may facilitate the cleaning and disinfection in difficult-toreach areas, such as isthmuses and secondary canals (14,15). Likewise, the sonic and ultrasonic activation were equally effective in increasing the dissolution effectiveness of NaOCI, up to over ten-fold as compared to passive irrigation (5,11,16). Moreover, PUI and some sonic devices positively influenced the push-out bond strength of epoxy resin-based root canal sealers with dentin (17) and tubular dentin sealer penetration (18).

EFFECTS OF THE IRRIGATION PROCEDURE IN THE INTRARADICULAR DENTIN

Dentin is a substrate with a complex organic and inorganic structure. This substrate is composed of an extracellular collagen matrix (ECM) with

immersed apatite crystals (19). The dentin matrix is a reservoir of non-collagenous proteins (NCPs), which display some biological properties.

Indeed, DMPI and DSPP have previously been described as active promoters and controllers of the biomineralization of dentin (20-22). Previously was demonstrated that DSPP and its cleaved products are critical for dentin mineralization, and they may function synergistically with DMPI and the cleaved by-products of both proteins (21).

Apatite deposition among collagen fibrils is a key factor in the regeneration of mineralized tissues (19,22-24). Previous data of our laboratory demonstrated that the controlled mineral nucleation on demineralized dentin involves complex molecular signaling that guides the site and rate of apatite formation (22). In a previous study, we reported the ability of a bioactive material to biomineralize dentin in a phosphate-containing fluid (25). We hypothesized that the recombinant DMPI molecules could recognize a specific molecular signature on the apatite surface, thereby guiding calcium phosphate clusters during recruitment through the collagen matrix, triggering an ion migration flux and attracting crystal precipitation in a process described as controlled biomineralization (20-22,25).

Moreover, our research group demonstrated that the remineralization of radicular dentin can

be achieved with the aid of bioactive materials and a standardized demineralization procedure to expose NCPs in the dentin matrix, allowing MMPs to convert structural matrix proteins into signaling molecules and generating peptides, which allows a specific attraction flux of apatites to guide mineral nucleation and maturation (22). Furthermore, our data revealed, for the first time in literature, the migration of the mineral precipitates from their ion source (bioactive material) to dentin, indicating the presence of an attraction flux and specific sites of nucleation in the intratubular dentin (Figure 2) (22).

Nowadays, compelling evidence shows that hypochlorite solutions affect the mechanical properties of dentin via the degradation of its organic components (19,23,24,26). The dentin substrate is exposed to solutions deposited in the root canal during the irrigation protocol. A study showed that irrigation with 5.25% NaOCI, compared to saline solution, reduces the flexural strength and the elastic modulus of dentin (27). Also, the use of NaOCI and PUI induced major changes in the substrate, with intertubular dentin erosion (Figure 3) (24). When dentin collagen is dissolved by NaOCI, the release of oxygen occurs and interfere with the polymerization of resins and adhesive materials (24). Thus, studies reported that NaOCI affected the sealing ability and adhesion of dental materials, such as resin-based cements and root canal sealers, to radicular (28,29) and coronal dentin (29,30).

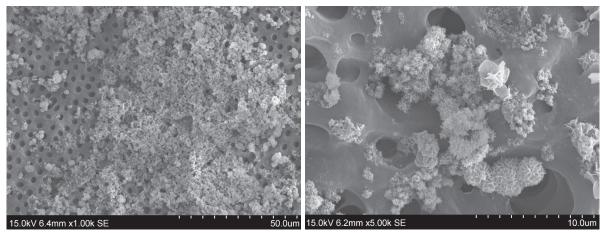


Figure 2. Representative SEM photomicrographs demonstrating the intratubular remineralization of radicular dentin in our previous study (22).

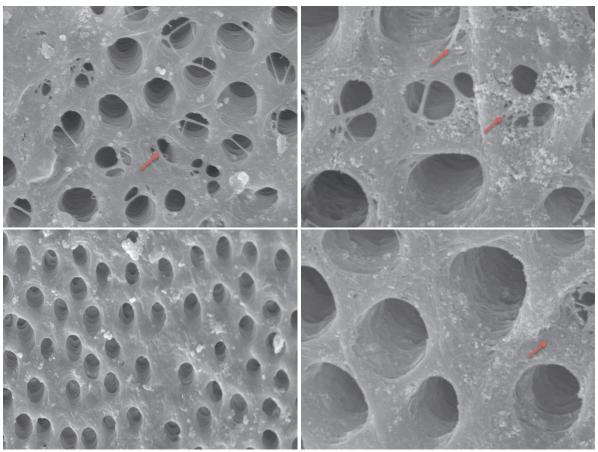


Figure 3. Representative SEM photomicrographs demonstrating the intratubular erosion, pointed out with red arrows, caused by the final irrigation protocol with NaOCl and PUI (19).

Our previous results indicated that the use of most common irrigants affects the expression of MMPs on radicular dentin. NaOCI and CHX cause lower expression of MMPs, while EDTA increases their expression in the root canal dentin (23). Moreover, we demonstrated that the ultrasonic activation of irrigants to obtain better cleaning and disinfection, caused relevant alterations on

radicular dentin composition, as well as an important proteolytic effect on DMP1-CT (Figure 4) (19). DMP1 is an essential regulator in regeneration and biomineralization (19,22). The potential loss of this signaling molecule after irrigation procedures could affect the dentin matrix as an unsuitable environment for successful regenerative procedures (19).

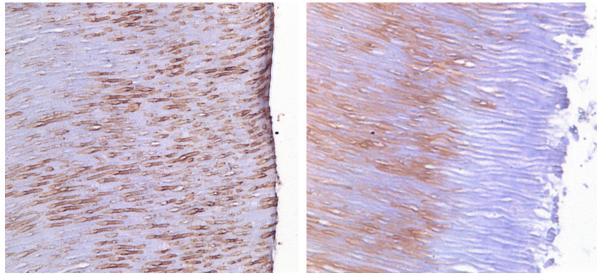


Figure 4. Representative DMP1-CT immunohistochemical photomicrographs. A. Positive control group pattern of normal DMP1-CT spatial distribution in radicular dentin. B. Effect of the final irrigation protocol with NaOCL, EDTA, and PUI showing the detrimental effect on the immunoexpression of DMP1-CT (19).

Most studies that aimed to analyzed the cleaning and disinfection ability of irrigating protocols do not contemplate the effect of the endodontic treatment on enamel and dentin substrates, and how this can be detrimental for the tooth restoration. Irrigation protocols with the most favorable cleaning and disinfection results demonstrated to have a detrimental effect on dentin ultrastructure and biochemical composition.

The clinical relevance of this article remains in alerting to the dental community the alterations that occur in dentin ultrastructure when different irrigation protocols are performed. In clinical scena-

rios in which is advocated to take advantage of the bioactivity of dentin, for regenerative endodontic treatment and to promote a biomineralization process, it is necessary to ensure that disinfectants do not counteract the function of bioactive molecules (19). For disinfection, lower concentrations of NaOCI are still effective and may preserve more of the protein components in dentin. Development of biocompatible disinfecting solutions is essential to preserve the signaling molecules and maintain a suitable environment when dentin remineralization and/or tissue regeneration are to be achieved. Future research must focus on a balance between biological and mechanical goals for endodontic treatment.

REFERENCES

- 1. AAE. Treatment Standards. aae.org. 2018. p. 1-26.
- 2. Pereira Lopes H., Siqueira Jr. J. Endodontia: Biologia e Técnica. 2nd ed. Rio de Janeiro: MEDSI; 2004. 281-288 p.
- 3. Bystrom A., Happonen R.P., Sjogren U., Sundqvist G. Healing of periapical lesions of pulpless teeth after endodontic treatment with controlled asepsis. Endod Dent Traumatol. 1987 Apr; 3 (2): 58-63.
- 4. Prada I., Micó-Muñoz P., Giner-Lluesma T., Micó-Martínez P., Muwaquet-Rodríguez S., Albero-Monteagudo A. Update of the therapeutic planning of irrigation and intracanal medication in root canal treatment. A literature review. J Clin Exp Dent. 2019 Feb; 11 (2): e185-93.
- 5. Haapasalo M., Shen Y., Wang Z., Gao Y. Irrigation in endodontics. British Dental Journal. 2014 Mar 21; 216 (6): 299-303.
- 6. Siqueira J.F., Rôças I.N., Favieri A., Lima K.C. Chemomechanical reduction of the bacterial population in the root canal after instrumentation and irrigation with 1%, 2.5%, and 5.25% sodium hypochlorite. J Endod. 2000 Jun; 26 (6): 331-4.
- 7. Barrette W.C., Hannum D.M., Wheeler W.D., Hurst JK. General mechanism for the bacterial toxicity of hypochlorous acid: abolition of ATP production. Biochemistry. 1989 Nov 14; 28 (23): 9172-8.
- 8. Harrison J.W., Hand R.E. The effect of dilution and organic matter on the anti-bacterial property of 5.25% sodium hypochlorite. J Endod. 1981 Mar; 7 (3): 128-32.
- 9. Alpizar S., Bustamante G., Calderón C., Calvo J., Gutiérrez J, Salazar P. Análisis de los diferentes protocolos de irrigación y su efecto en la permeabilidad y selle radiculary selle . [San José]: Universidad de Costa Rica; 2016.
- 10. Barbosa A.F.A., Lima C.O. de, Sassone L.M., Fares R.D., Fidalgo T.K. da S., Silva E.J.N.L.

- Effect of passive ultrasonic irrigation on hard tissue debris removal: a systematic review and meta-analysis. Braz Oral Res. 2021; 35: e123.
- 11. Orlowski N.B., Schimdt T.F., Teixeira C. da S., Garcia L. da F.R., Savaris J.M., Tay F.R., et al. Smear Layer Removal Using Passive Ultrasonic Irrigation and Different Concentrations of Sodium Hypochlorite. J Endod. 2020 Nov; 46 (11): 1738-44.
- 12. Spangberg L., Engström B., Langeland K. Biologic effects of dental materials. 3. Toxicity and antimicrobial effect of endodontic antiseptics in vitro. Oral Surg Oral Med Oral Pathol. 1973 Dec; 36 (6): 856-71.
- 13. Teixeira C.S., Felippe M.C.S., Felippe W.T. The effect of application time of EDTA and NaOCl on intracanal smear layer removal: an SEM analysis. Int Endod J. 2005 May; 38 (5): 285-90.
- 14. Klyn S.L., Kirkpatrick T.C., Rutledge R.E. In vitro comparisons of debris removal of the EndoActivator system, the F file, ultrasonic irrigation, and NaOCl irrigation alone after hand-rotary instrumentation in human mandibular molars. J Endod. 2010 Aug; 36 (8): 1367-71.
- 15. Bolles J.A., He J., Svoboda K.K.H., Schneiderman E., Glickman G.N. Comparison of Vibringe, EndoActivator, and needle irrigation on sealer penetration in extracted human teeth. J Endod. 2013 May; 39 (5): 708-11.
- 16. Stojicic S., Zivkovic S., Qian W., Zhang H., Haapasalo M. Tissue dissolution by sodium hypochlorite: effect of concentration, temperature, agitation, and surfactant. J Endod. 2010 Sep; 36 (9): 1558-62.
- 17. Augusto C.M., Cunha Neto M.A., Pinto K.P., Barbosa A.F.A., Silva E.J.N.L., dos Santos A.P.P., et al. Influence of the use of chelating agents as final irrigant on the push-out bond strength of epoxy resin-based root canal sealers: A systematic review. Aust Endod J. 2021 Sep 9.

- 18. de Souza Matos F., Rosatto C.M.P. de, Cunha T.C., Vidigal M.T.C., Blumenberg C., Paranhos LR, et al. Influence of chelating solutions on tubular dentin sealer penetration: A systematic review with network meta-analysis. Aust Endod J. 2021 Dec; 47 (3): 715-30.
- Retana-Lobo C., Ramírez-Mora T., Murillo-Gómez F., Maria Guerreiro-Tanomaru J., Tanomaru-Filho M, Reyes-Carmona J. Final irrigation protocols affect radicular dentin DMP1-CT expression, microhardness, and biochemical composition. Clin Oral Investig. 2022 May 2. doi: 10.1007/s00784-022-04516-8. Epub ahead of print.
- 20. Tay F.R., Pashley D.H. Guided tissue remineralisation of partially demineralised human dentine. Biomaterials. 2008 Mar; 29 (8): 1127-37.
- 21. Prasad M., Butler W.T., Qin C. Dentin sialophosphoprotein in biomineralization. Connect Tissue Res. 2010 Oct; 51 (5): 404-17.
- 22. Retana-Lobo C., Guerreiro-Tanomaru J.M., Tanomaru-Filho M., Mendes de Souza B.D., Reyes-Carmona J. Non-Collagenous Dentin Protein Binding Sites Control Mineral Formation during the Biomineralisation Process in Radicular Dentin. Materials (Basel). 2020 Feb 27; 13 (5): 1053.
- 23. Retana-Lobo C., Guerreiro-Tanomaru J.M., Tanomaru-Filho M., Mendes de Souza B.D., Reyes-Carmona J. Sodium Hypochlorite and Chlorhexidine Downregulate MMP Expression on Radicular Dentin. Med Princ Pract. 30 (5): 470-6.

- 24. Wagner M.H., da Rosa R.A., de Figueiredo J.A.P., Duarte M.A.H., Pereira J.R., Só M.V.R. Final irrigation protocols may affect intraradicular dentin ultrastructure. Clinical Oral Investigations. 2017 Sep 1; 21 (7): 2173-82.
- 25. Reyes-Carmona J.F., Felippe M.S., Felippe W.T. Biomineralization ability and interaction of mineral trioxide aggregate and white portland cement with dentin in a phosphate-containing fluid. J Endod. 2009 May; 35 (5): 731-6.
- 26. Sum Dr. C.P., Neo J., Kishen A. What we leave behind in root canals after endodontic treatment: Some issues and concerns. Australian Endodontic Journal. 2005 Dec; 31 (3): 94-100.
- 27. Sim T.P., Knowles J.C., Ng Y.L., Shelton J., Gulabivala K. Effect of sodium hypochlorite on mechanical properties of dentine and tooth surface strain. Int Endod J. 2001 Mar; 34 (2): 120-32.
- 28. García-Godoy F., Loushine R.J., Itthagarun A., Weller R.N., Murray P.E., Feilzer A.J., et al. Application of biologically-oriented dentin bonding principles to the use of endodontic irrigants. Am J Dent. 2005 Aug; 18 (4): 281-90.
- 29. Pascon F.M., Kantovitz K.R., Sacramento P.A., Nobre-dos-Santos M., Puppin-Rontani R.M. Effect of sodium hypochlorite on dentine mechanical properties. A review. J Dent. 2009 Dec; 37 (12): 903-8.
- 30. Perdigão J., Eiriksson S., Rosa B.T., Lopes M., Gomes G. Effect of calcium removal on dentin bond strengths. Quintessence Int. 2001 Feb; 32 (2): 142-6.

