



Darwiniana, nueva serie

ISSN: 0011-6793

ISSN: 1850-1702

Instituto de Botánica Darwinion & Museo Botánico de Córdoba

Grohar, Mariana C.; Morales, Matías; Fortunato, Renée H.; Rosenfeldt, Sonia
MICROMORPHOLOGY OF EPICUTICULAR WAXES IN GENUS *MIMOSA* (FABACEAE)

Darwiniana, nueva serie, vol. 11, no. 1, 2023, pp. 5-22
Instituto de Botánica Darwinion & Museo Botánico de Córdoba

DOI: <https://doi.org/10.14522/darwiniana.2023.111.1084>

Available in: <https://www.redalyc.org/articulo.oa?id=66976167011>

- ▶ How to cite
- ▶ Complete issue
- ▶ More information about this article
- ▶ Journal's webpage in redalyc.org

redalyc.org

Scientific Information System Redalyc

Network of Scientific Journals from Latin America and the Caribbean, Spain and Portugal

Project academic non-profit, developed under the open access initiative

MICROMORPHOLOGY OF EPICUTICULAR WAXES IN GENUS *MIMOSA* (FABACEAE)

Mariana C. Grohar^{1,2,*} , Matías Morales^{2,4} , Renée H. Fortunato³  & Sonia Rosenfeldt¹ 

¹ Departamento de Biodiversidad y Biología Experimental, Facultad de Ciencias Exactas y Naturales, Universidad de Buenos Aires. Intendente Güiraldes 2620, Ciudad Autónoma de Buenos Aires 1428, Argentina; mariana.grohar@gmail.com (author for correspondence).

² Instituto de Recursos Biológicos, Instituto Nacional de Tecnología Agropecuaria (INTA). N. Repetto y Los Reseros s/n, Hurlingham 1686. Buenos Aires, Argentina.

³ Instituto de Botánica Darwinion (CONICET/ACEFYN), Labardén 200, San Isidro B1642HYD, Buenos Aires, Argentina.

⁴ Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET). Godoy Cruz 2900, C1425FQB Ciudad Autónoma de Buenos Aires, Argentina.

Abstract. Grohar, M. C.; M. Morales, R. H. Fortunato & S. Rosenfeldt. 2023. Micromorphology of epicuticular waxes in genus *Mimosa* (Fabaceae). *Darwiniana*, nueva serie 11(1): 5-22.

Epicuticular waxes crystallize on the foliar surface with different morphologies, which can be taxonomically relevant features in many taxa. Their morphological analysis was based on scanning electron microscopy (SEM) images on herbarium specimens of selected sections of *Mimosa*. The present work is the first detailed description and classification of the epidermal micromorphology in the genus *Mimosa*, focusing on section *Calothamnus* and *Mimosa* series *Mimosa*, subseries *Dolentes* and *Brevipedes*. We also describe different wax crystallization morphologies present in these taxa, identifying three different crystallization types: film, plates, and platelets. Novel epidermal micromorphologies have been described, such as papillae in *M. lepidorepens*. Results allow grouping taxa of the studied sections, contributing to clarifying their taxonomy. SEM images of epicuticular waxes and leaf surfaces could be a new tool for performing a new taxonomic treatment of section *Mimosa* subser. *Dolentes* and subser. *Brevipedes*, and section *Calothamnus*.

Keywords. Epicuticular waxes; epidermis; Fabaceae; micromorphology; *Mimosa*.

Resumen. Grohar, M. C.; M. Morales, R. H. Fortunato & S. Rosenfeldt. 2023. Micromorfología de ceras epicuticulares en el género *Mimosa* (Fabaceae). *Darwiniana*, nueva serie 11(1): 5-22.

Las ceras epicuticulares cristalizan sobre la superficie foliar con diferentes morfologías, las cuales pueden ser de relevancia taxonómica en muchos taxones. El análisis micromorfológico fue basado en imágenes obtenidas con microscopía electrónica de barrido (MEB) de ejemplares de herbario de algunas secciones de *Mimosa*. El presente trabajo es la primera descripción y clasificación detallada de la micromorfología epidérmica en el género *Mimosa*, centrada en las secciones *Calothamnus* y *Mimosa*, subseries *Dolentes* y *Brevipedes*. También describe distintas morfologías de cristalización presentes en estos taxones, identificando tres tipos: lámina, placas y plaquetas. Se han descrito nuevas micromorfologías epidérmicas, como las papilas de *M. lepidorepens*. Estos resultados permiten agrupar los taxones de las secciones estudiadas, contribuyendo a la clarificación de su taxonomía. Las imágenes obtenidas con MEB de la superficie foliar y de las ceras epicuticulares pueden constituirse como una nueva herramienta en la elaboración de una nueva propuesta taxonómica de las secciones *Calothamnus* y *Mimosa* subser. *Dolentes* y subser. *Brevipedes*.

Palabras clave. Ceras epicuticulares; epidermis; Fabaceae; micromorfología; *Mimosa*.

INTRODUCTION

Epicuticular waxes represent the interaction surface between plants and biotic and abiotic factors of their environment, which determines their major ecologic importance. Functions of epicuticular waxes are mainly related to the restriction of transpiration water loss, although it does not seem to be the only factor that determines its rates (Bondada et al., 1996; Oliveira et al., 2003). However, other functions of epicuticular waxes have been proposed, such as protection against radiation (Barnes & Cardoso-Vilhena, 1996), herbivory, and natural pathogens (Jenks et al., 2010).

Wax biosynthesis in epidermal cell cytoplasm comprises complex enzymatic pathways starting from fatty acids (Jenks et al., 1995, 2010). The mechanism of wax deposition on the surface is not known in detail, although some interpretations include extrusion through microchannels and diffusion through the cuticle (Jenks et al., 2010).

Waxes crystallize in many different morphologies, from a smooth film, variable in thickness, to complex crystalloid structures, such as granules, plates, platelets, and rodlets, which have been described with different techniques (Barthlott et al., 1998; Ensikat et al., 2006). Morphology of these epicuticular structures varies in and between species with the chemical composition of the wax (Jeffree et al., 1975; Sen, 1987; Ensikat et al., 2006; Koch et al., 2006), the development stage of the leaf (Gülz et al., 1991; Jetter & Schaffer, 2001), environment conditions (Wallace et al., 1987; Bondada et al., 1996; Oliveira et al., 2003) and the way it is deposited on the surface (Anton et al., 1994).

Epicuticular wax micromorphology has been described in detail with scanning electron microscopy (Barthlott et al., 1998), and it has shown a great taxonomic value in different taxa, such as *Pinus* L., *Huberia* DC., *Zanthoxylum* L., *Alstonia* R. Br., *Pilocarpus* Vahl (Herbin & Sharma, 1969; Jayeola, 1998; Mimura et al., 1998; Skorupa et al., 1998; Dias-Leme et al., 2013). There are only a few extensive studies on epicuticular waxes in Fabaceae, and they are mainly focused on Caesalpinioideae genera (*Calliandra* Benth., *Acacia* Mill. s.l.), although most of them have been included in more extensive studies which analysed

various families (Maffei, 1996; Ramírez et al., 1997; Barthlott et al., 1998). Nevertheless, in the genus *Mimosa* L., which is one of the most diverse genera of Fabaceae, there are no such descriptions.

This genus comprises more than 540 species of pantropical and pan-subtropical distribution (Simon et al., 2011; Bessega & Fortunato, 2011) with relevant diversification in South America. As presently circumscribed (Barneby, 1991; Savassi-Coutinho et al., 2012; Dutra & Garcia, 2013), section *Calothamnos* Barneby comprises 28 species and eight varieties: *M. aurivillus* Mart. var. *aurivillus*, *M. aurivillus* var. *calothamnoides* Barneby, *M. aurivillus* var. *calothamnos* (Mart. ex Benth.) Barneby, *M. aurivillus* var. *sordescens* Benth., *M. aurivillus* var. *warmingii* Barneby, *M. barretoii* Hoehne, *M. bathyrrhena* Barneby, *M. berroi* Burkart, *M. bonplandii* Benth., *M. calodendron* Mart. ex Benth., *M. chrysastra* Mart. var. *chrysastra*, *M. chrysastra* var. *itambeana* Barneby, *M. cylindracea* Benth., *M. daleoides* Benth., *M. eriocarpa* Benth., *M. flocculosa* Burkart, *M. furfuracea* Benth., *M. incana* Benth., *M. involucreta* Benth., *M. lepidorepens* Burkart, *M. leprosa* J. F. Macbr. var. *leprosa*, *M. leprosa* var. *parviceps* Barneby, *M. longistipula* V.F. Dutra & F.C.P. Garcia, *M. macedoana* Burkart var. *macedoana*, *M. macedoana* var. *glabrescens* (Burkart) Barneby, *M. myuros* Barneby, *M. peduncularis* Bong. ex Benth., *M. pilulifera* Benth. var. *pilulifera*, *M. pilulifera* var. *pseudincana* (Burkart) Barneby, *M. plumosa* Micheli, *M. psittacina* Barneby, *M. rocae* Lor. & Niederl., *M. roseoalba* Sav.-Cout. & G.P. Lewis, *M. scabrella* Benth., *M. taimbensis* Burkart, and *M. urticaria* Barneby.

The subseries *Dolentes* Barneby (sect. *Mimosa*) is monotypic, comprising only *M. dolens* Vell., which is defined by the presence of determinate inflorescences and valvately dehiscent pods grouped in dense spheroid clusters. Barneby (1991) described five subspecies in this taxon: *M. dolens* ssp. *callosa* (Benth.) Barneby, *M. dolens* ssp. *rigida* (Benth.) Barneby with five varieties (var. *rigida*, var. *rigescens* (Benth.) Barneby, var. *anisitsii* (Lindm.) Barneby, var. *foliolosa* (Benth.) Barneby, var. *deterior* Barneby), *M. dolens* ssp. *acerba* (Benth.) Barneby with three varieties (var. *acerba*, var. *latifolia* (Benth.) Barneby, var. *rudis* (Benth.) Barneby), *M. dolens* ssp. *eriphylla*

(Benth.) Barneby, and *M. dolens* ssp. *dolens* and two varieties (var. *dolens* and var. *pangloea* Barneby). The subseries *Brevipedes* Barneby (sect. *Mimosa*), on the other hand, is differentiated by its indeterminate inflorescences and craspedial dehiscence of the pod, and comprises nine species: *M. brevipes* Benth., *M. diversipila* Micheli and two varieties (var. *diversipila* and var. *subglabriseta* Barneby & Fortunato emend. M. Morales), *M. fernandez-casarii* Barneby & Fortunato, *M. cryptogloea* Barneby, *M. custodis* Barneby, *M. sceptrum* Barneby, *M. longiracemosa* (Burkart) Barneby, *M. pseudopetiolearis* Barneby, *M. nitidula* Barneby, and *M. schininii* M. Morales, Grohar & Fortunato (Barneby, 1991; Morales et al., 2019).

Therefore, our aim is to contribute to the description of the morphology of epicuticular waxes in the genus *Mimosa*. Due to its high morphologic variability, traditional sections based on morphological similarities do not coincide with phylogenetic analysis (Barneby, 1991; Simon et al., 2011). This is the case of sect. *Calothamnos*, where morphologically consistent characters do not always coincide with molecular phylogeny, in which it is polyphyletic with sect. *Mimosa* (Bessegga et al., 2008; Coutinho, 2009; Bessegga & Fortunato, 2011; Simon et al., 2011; Grohar, 2020). Furthermore, in several infrageneric groups of sect. *Mimosa*, intermediate forms between specific and infraspecific taxa (Morales et al., 2010, 2014, 2015; Luna-Castro et al., 2012) generates problems to delimitate infrageneric taxa, such as in the case of the taxonomic complex which comprises *Mimosa* subseries *Dolentes* and *Mimosa* subseries *Brevipedes* (Morales et al., 2010; Grohar et al., 2016). The aim of this work is to 1) study in detail the outer epidermal micromorphology in sect. *Mimosa* (subseries *Dolentes* and *Brevipedes*) and sect. *Calothamnos*, which are groups of interest because of their complex systematics and evolutionary issues, in order to describe their natural variability, and 2) determine its relevance in the differentiation of the taxa at different levels.

MATERIALS AND METHODS

Plant material. Sixteen herbarium specimens of different taxa from sect. *Mimosa* (subseries

Dolentes and subseries *Brevipedes*) and 37 from sect. *Calothamnos*, deposited in BAB, CTES, G, HUICS, MBM, MVFA, RB, SI, and SPF (Appendix; herbarium acronyms follow Thiers, 2022) were included in the analysis. All of them were identified using Barneby (1991) taxonomic treatment.

Scanning electron microscopy (SEM). Two dehydrated mature leaflets per taxon were sampled in squares of 1 cm² from the centre of the leaflet and sputter-coated with gold-palladium for three minutes. Later, they were examined with a Zeiss Supra 40 FESEM scanning electron microscope at the Centro de Microscopía Avanzada of Universidad de Buenos Aires, or a Philips SEM 505 at the Museo de Ciencias Naturales Bernardino Rivadavia (Buenos Aires, Argentina).

Terminology. For the description of epidermis surface, we followed the terminology used by Behnke & Barthlott (1983) and Koch et al. (2006). For epicuticular waxes' crystals, we followed Barthlott et al. (1998).

RESULTS

In the studied taxa of genus *Mimosa*, the general structure of the outer surface of the leaves includes a cutin layer above the epidermis, which includes a lower cutin-pectin layer and an upper cutin-wax layer, and over the latter, a layer of epicuticular waxes (Fig. 1A-B).

Regarding the epidermis, we described two epidermal cell shapes (Table 1, Figs. 2A-B, 3A-B). Most of the taxa of both sections show polygonal cells, while *M. dolens* ssp. *acerba* var. *latifolia* and *M. dolens* ssp. *dolens* var. *pangloea* show jigsaw shape, but both only on abaxial face. On a closer analysis, we differentiate three different layers (Fig. 1C):

Primary layer. It comprises the cell outline, that is, the connecting wall between adjoining cells. In superficial view, all considered taxa showed straight outlines (Table 1, Figs. 2A-D and 3A-D). In lateral view, however, most taxa show a sunken morphology, except for *M. dolens* var. *acerba* and *M. brevipes* (sect. *Mimosa*), and *M. eriocarpa* on abaxial face, *M. plumosa* and *M. roseoalba* (sect. *Calothamnos*).

Table 1. Outer epidermis features in *Mimosa*.

Taxon	Cell shape	Primary layer		Secondary layer	Tertiary Layer		
		Superficial view	Lateral view		Type	Location	
Section <i>Mimosa</i>							
Subseries <i>Dolentes</i>							
<i>M. dolens</i> ssp. <i>callosa</i>	Polygonal	Straight	Sunken	Convex	Plates	homogenous	
<i>M. dolens</i> ssp. <i>eriophylla</i>	Polygonal	Straight	Sunken	Concave	Platelets	homogenous	
<i>M. dolens</i> ssp. <i>rigida</i>	<i>M. dolens</i> var. <i>rigescens</i>	Polygonal	Straight	Sunken	Convex	Plates	homogenous
	<i>M. dolens</i> var. <i>anisitsii</i>	Polygonal	Straight	Sunken	Convex	Platelets	homogenous
	<i>M. dolens</i> var. <i>foliolosa</i>	Polygonal	Straight	Sunken	Convex	Platelets	peristomatic
	<i>M. dolens</i> var. <i>deterior</i>	Polygonal	Straight	Sunken	Convex	Film	homogenous
<i>M. dolens</i> ssp. <i>acerba</i>	<i>M. dolens</i> var. <i>acerba</i>	Polygonal	Straight	Straight	Concave	Plates	homogenous
	<i>M. dolens</i> var. <i>latifolia</i> adaxial face	Polygonal	Straight	Sunken	Convex	Plates	homogenous
	<i>M. dolens</i> var. <i>latifolia</i> abaxial face	Jigsaw	Straight	Sunken	Convex	Plates	homogenous
	<i>M. dolens</i> var. <i>rudis</i>	Polygonal	Straight	Sunken	Convex	Platelets	homogenous
<i>M. dolens</i> ssp. <i>dolens</i>	<i>M. dolens</i> var. <i>dolens</i>	Polygonal	Straight	Sunken	Convex	Film	homogenous
	<i>M. dolens</i> var. <i>pangloea</i> adaxial face	Polygonal	Straight	Sunken	Convex	Platelets	homogenous
	<i>M. dolens</i> var. <i>pangloea</i> abaxial face	Jigsaw	Straight	Sunken	Convex	Platelets	homogenous
Subseries <i>Brevipedes</i>							
<i>M. pseudopetiolaris</i>	Polygonal	Straight	Sunken	Convex	Film	homogenous	
<i>M. custodis</i>	Polygonal	Straight	Sunken	Convex	Platelets	homogenous	
<i>M. brevipes</i>	Polygonal	Straight	Straight	Concave	Platelets	homogenous	
<i>M. sceptrum</i>	Polygonal	Straight	Sunken	Convex	Platelets	homogenous	
<i>M. cryptogloea</i>	Polygonal	Straight	Sunken	Convex	Platelets	homogenous	
<i>M. diversipilavar.subglabriseta</i>	Polygonal	Straight	Sunken	Convex	Plates	homogenous	
Section <i>Calothamnus</i>							
<i>M. aurivillus</i> var. <i>aurivillus</i>	Polygonal	Straight	Sunken	Convex	Film	homogenous	
<i>M. aurivillus</i> var. <i>calothamnoides</i>	Polygonal	Straight	Sunken	Convex	Platelets	homogenous	
<i>M. aurivillus</i> var. <i>calothamnus</i>	Polygonal	Straight	Sunken	Convex	Plates	homogenous	
<i>M. aurivillus</i> var. <i>sordescens</i>	Polygonal	Straight	Sunken	Convex	Platelets	homogenous	

Table 1. Outer epidermis features in *Mimosa*.

Taxon	Cell shape	Primary layer		Secondary layer	Tertiary Layer	
		Superficial view	Lateral view		Type	Location
<i>M. barretoii</i>	Polygonal	Straight	Sunken	Convex	Plates	homogenous
<i>M. bathyrrhena</i>	Polygonal	Straight	Sunken	Convex	Plates	homogenous
<i>M. berroi</i>	Polygonal	Straight	Sunken	Convex	Plates	homogenous
<i>M. bonplandii</i>	Polygonal	Straight	Sunken	Convex	Film	homogenous
<i>M. calodendron</i>	Polygonal	Straight	Sunken	Convex	Film	homogenous
<i>M. chrysastra</i> var. <i>chrysastra</i>	Polygonal	-	-	-	Platelets	homogenous
<i>M. chrysastra</i> var. <i>itambeana</i>	Polygonal	Straight	Sunken	Convex	Platelets	homogenous
<i>M. cylindracea</i>	Polygonal	Straight	Sunken	Convex	Platelets	homogenous
<i>M. daleoides</i>	Polygonal	Straight	Sunken	Convex	Platelets	homogenous
<i>M. eriocarpa</i> - <i>abaxial</i>	Polygonal	Straight	Straight	Concave	Plates	homogenous
<i>M. eriocarpa</i> - <i>adaxial</i>	Polygonal	Straight	Sunken	Convex	Plates	homogenous
<i>M. flocculosa</i>	Polygonal	-	-	-	Plates	homogenous
<i>M. furfuracea</i>	Polygonal	Straight	Sunken	Convex	Film	homogenous
<i>M. incana</i>	Polygonal	Straight	Sunken	Convex	Plates	homogenous
<i>M. involucrata</i>	Polygonal	Straight	Sunken	Convex	Plates	homogenous
<i>M. lepidorepens</i> - <i>abaxial</i>	Polygonal	Straight	Sunken	Convex (papillae)	Platelets	homogenous
<i>M. lepidorepens</i> - <i>adaxial</i>	Polygonal	Straight	Sunken	Convex	Film	homogenous
<i>M. leprosa</i>	Polygonal	Straight	Sunken	Convex	Platelets	homogenous
<i>M. macedoana</i>	Polygonal	Straight	Sunken	Convex	Platelets	homogenous
<i>M. myuros</i>	Polygonal	Straight	Sunken	Convex	Film	homogenous
<i>M. pilulifera</i> var. <i>pilulifera</i>	Polygonal	Straight	Sunken	Convex	Platelets	homogenous
<i>M. plumosa</i>	Polygonal	Straight	Straight	Concave	Platelets	homogenous
<i>M. rocae</i>	Polygonal	Straight	Sunken	Convex	Plates	homogenous
<i>M. roseoalba</i>	Polygonal	Straight	Straight	Concave	Plates	homogenous
<i>M. scabrella</i>	Polygonal	Straight	Sunken	Convex	Platelets	homogenous
<i>M. taimbensis</i>	Polygonal	Straight	Sunken	Convex	Film	homogenous
<i>M. urticaria</i>	Polygonal	Straight	Sunken	Convex	Plates	homogenous

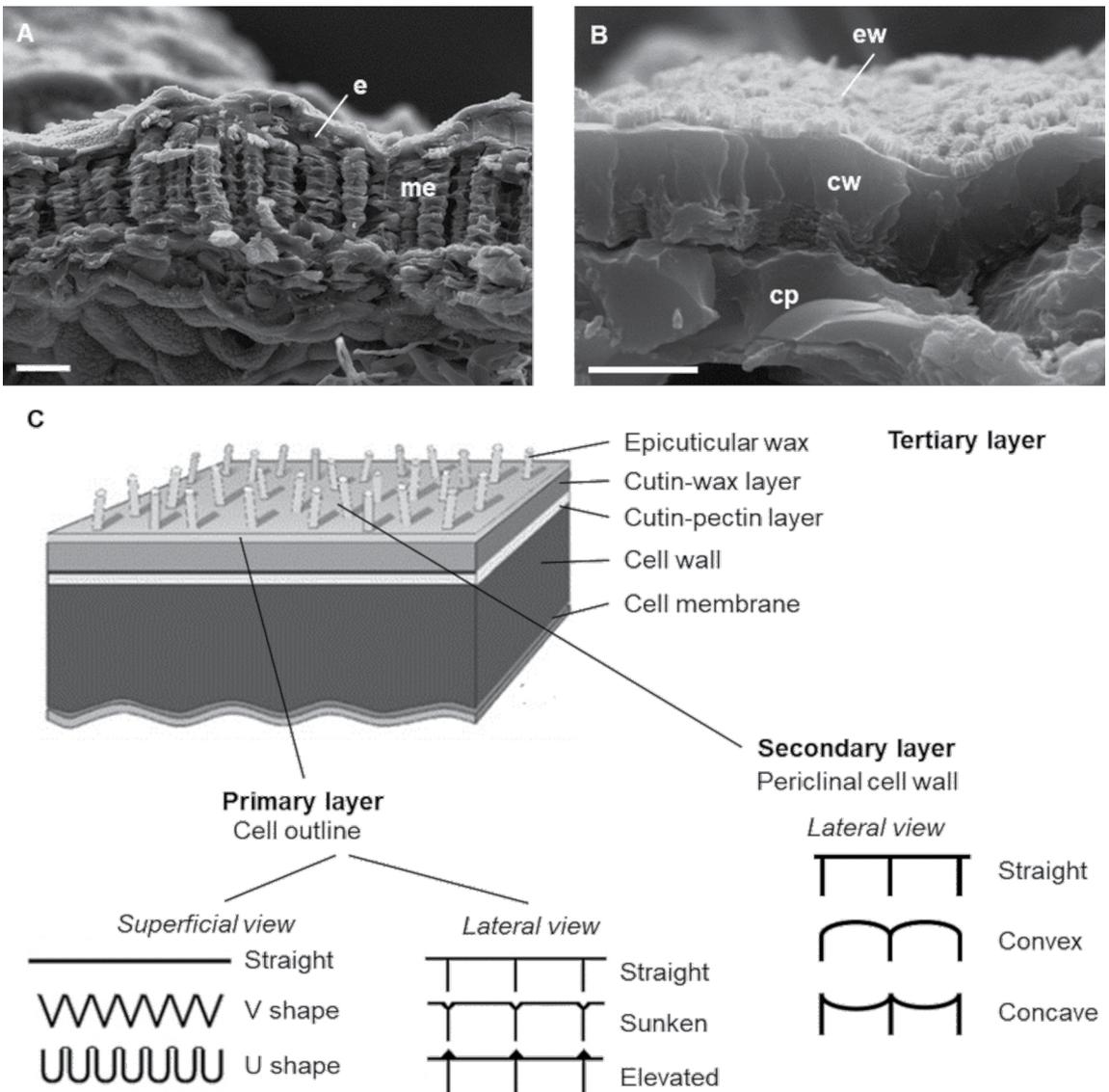


Fig. 1. Structure of leaflet epidermis in *Mimosa*. **A-B**, SEM micrographs of leaflet anatomy. **A**, *M. leprosa*, cross-section of the leaflet. **B**, *M. leprosa*, detail of the upper epidermis in cross section. **C**, structure of primary, secondary, and tertiary layers; adapted from Behnke & Barthlott (1983) and Koch et al. (2008). Abbreviations: **me**, mesophyll; **e**, epidermis; **cp**, cutin-pectin layer; **cw**, cutin-wax layer; **ew**, epicuticular waxes. The bar indicates: A = 20 μm and B = 5 μm .

Secondary layer. It comprises the outer surface of the epidermis cell, that is, its periclinal wall. Most of the taxa of both sections show convex surfaces (Table 1, Figs. 2E-F and 3E-H). However, some taxa show concave surfaces: *M. dolens* ssp. *eriphylla*, *M. dolens* ssp. *acerba* var. *acerba* and *M. brevipes* (sect. *Mimosa*),

and only on abaxial face of *M. eriocarpa*, *M. plumosa* and *M. rosealba* (sect. *Calothamnos*). Besides, *M. lepidorepens* is distinguished from all other taxa by a distinctive secondary epidermal morphology on abaxial face, as the cell wall forms prominent papillae, densely covered by wax crystals (Fig. 3G).

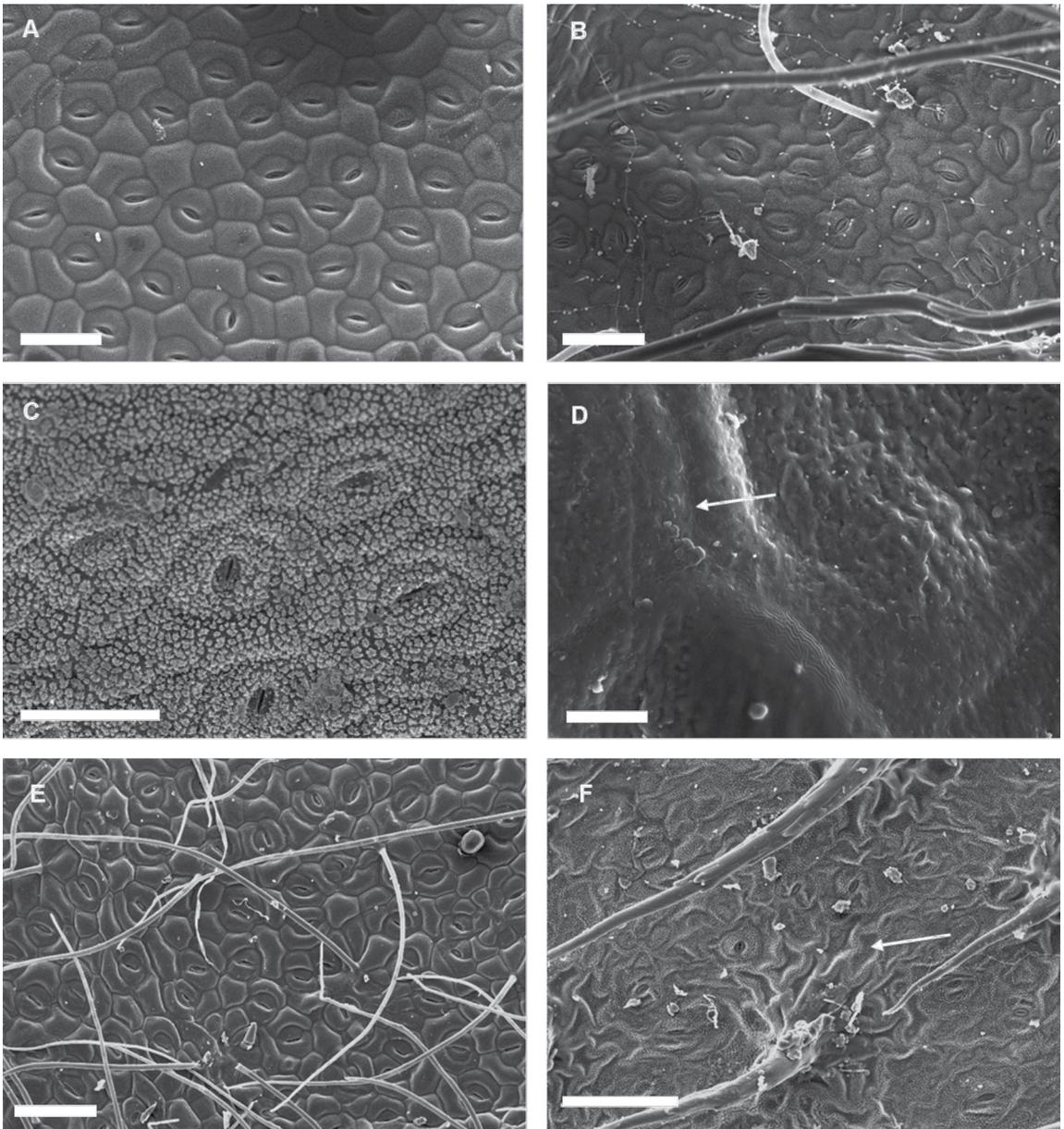


Fig. 2. SEM micrographs of primary and secondary layers in Dolentes-Brevipedes taxonomic complex. **A**, *M. dolens* var. *rigescens*. Primary layer straight in superficial view; cell shape polygonal. **B**, *M. dolens* var. *latifolia*, jigsaw cell shape. **C**, *M. dolens* var. *anisitsii*, primary layer straight in lateral view. **D**, *M. dolens* var. *dolens*, primary layer sunken in lateral view. **E**, *M. cryptogloea*, secondary layer: convex. **F**, *M. dolens* var. *acerba*, secondary layer concave. The bar indicates: A-C = 80 μ m, D = 4 μ m, E-F = 100 μ m.

Tertiary layer. It comprises epicuticular waxes and trichomes that are arranged over the thick layer of cutin on the outer surface of the cell wall. In

both sections of *Mimosa*, the tertiary layer covers the entire surface, including epidermal cells and subsidiary cells, but not stomatal cells (Fig. 2C).

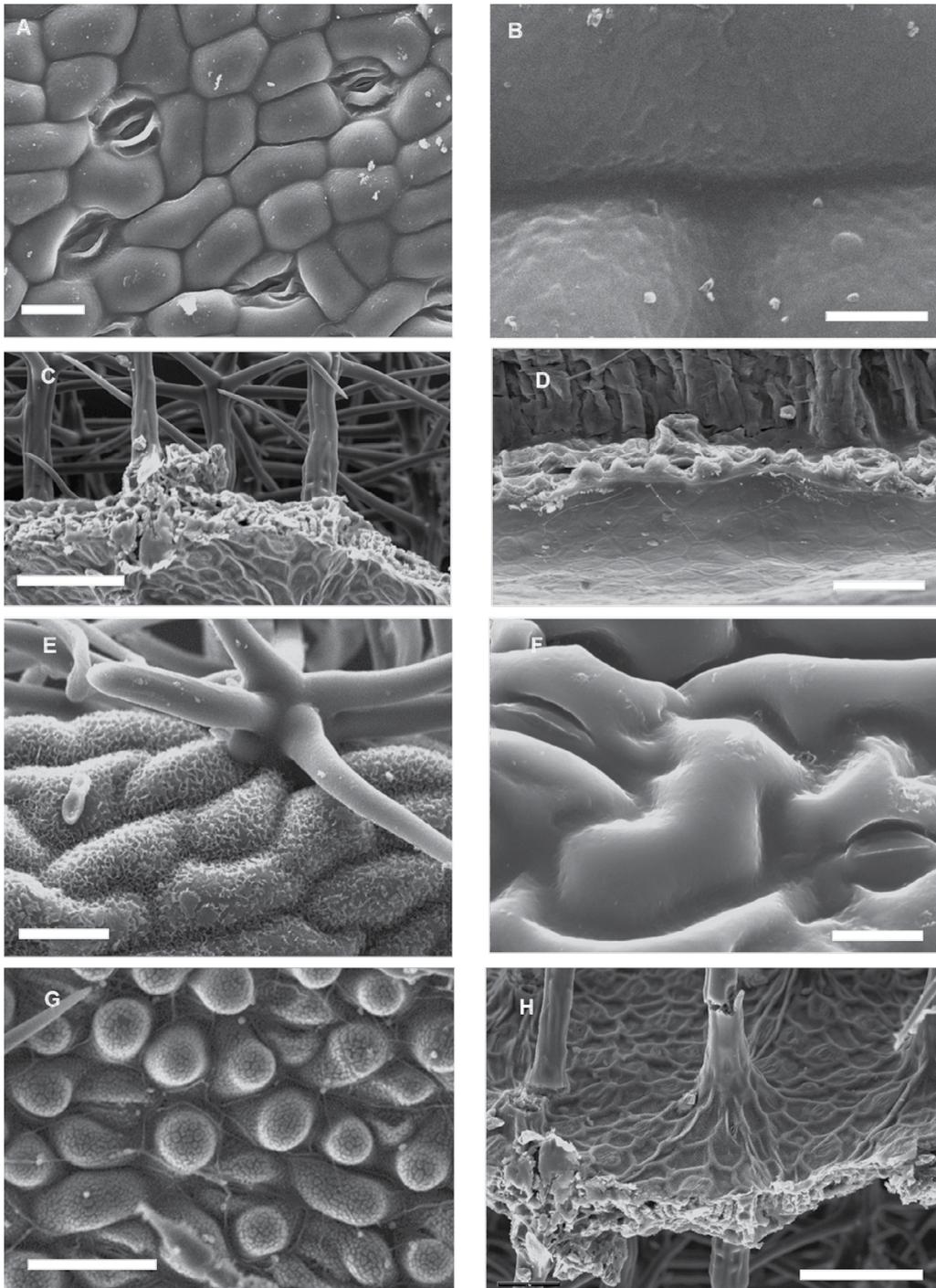


Fig. 3. SEM micrographs of primary and secondary layer in *M. sect. Calothamnion*. **A-B**, *M. myuros*. Primary layer straight in superficial view: straight; cell shape: polygonal. **C**, *M. plumosa*, primary layer in lateral view: straight. **D**, *M. calodendron*, primary layer sunken in lateral view: sunken. **E-F**, Secondary layer convex. **E**, *M. rocae*; **F**, *M. bonplandii*. **G**, *M. lepidorepens*, abaxial face, secondary layer papillae. **H**, *M. plumosa*, secondary layer concave. The bar indicates: A = 20 μm , B, E = 20 μm , C, H = 100 μm , D, G = 50 μm , F = 10 μm .

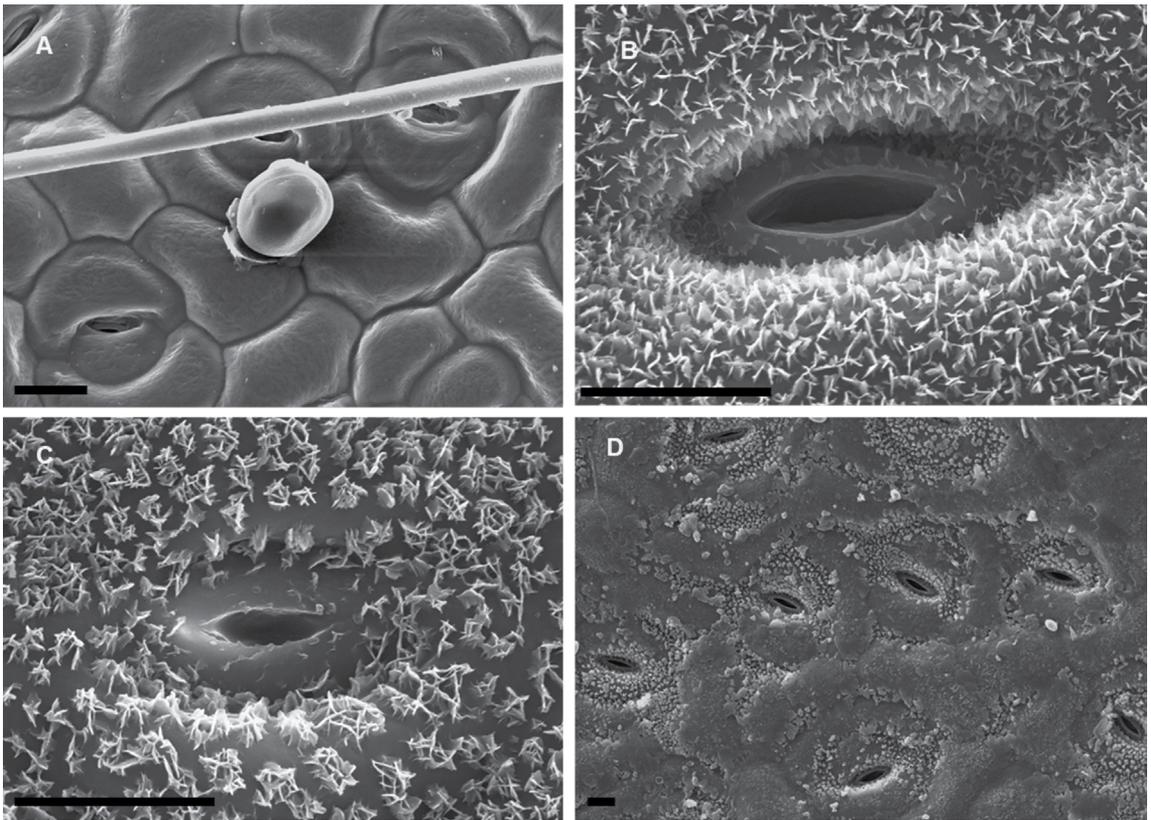


Fig. 4. SEM micrographs of epicuticular waxes in *Dolentes-Brevipedes* taxonomic complex. **A**, *M. cryptogloea*, abaxial face, homogeneous film of wax. **B**, *M. dolens* var. *acerba*, adaxial face, plate crystals. **C**, *M. custodis*, adaxial face, platelets crystals. **D**, *M. dolens* var. *foliolosa*, adaxial face, peristomatic platelets. The bar indicates 10 μm .

Besides, some variability has been observed in the shapes that epicuticular wax crystallizes on the foliar epidermis, varying from a fine homogeneous film to polyhedral crystalloids, and even concentrated in groups. Three distinct morphologies of wax deposition were found (Figs. 4-5, Table 1):

Film. Wax is deposited in a homogeneous layer which covers the whole leaflet surface. Due to wax flexibility, wax layer adjusts to cell surface, leading to sunken areas in epidermal or stomata cells borders (Figs. 4A, 5A). This deposition type was found in *M. dolens* var. *dolens*, *M. dolens* ssp. *rigida* var. *deterior*, *M. cryptogloea* and *M. pseudopetiolaris* (sect. *Mimosa*) and in *M. aurivillus* var. *aurivillus*, *M. bonplandii*, *M. calodendron*, *M. furfuracea*, on adaxial face of *M. lepidorepens*, *M. myuros* and *M. taimbensis* (sect. *Calothamnos*).

Plates. Polygonal crystalloids with distinct edges, 300-1200 nm wide, 500-920 nm long and 55-180 nm thick, are arranged in different directions, usually overlapping between them (Figs. 4B, 5B), and covering homogeneously the whole leaflet surface. The plates appear isolated, although sometimes they seem to be in circular groups. In general, wax deposition adjusts to cell surface, although there are some areas where this adjustment is not observed. In sect. *Mimosa*, this deposition morphology was found in *M. dolens* ssp. *callosa*, *M. dolens* ssp. *rigida* var. *rigescens*, *M. dolens* var. *acerba*, *M. dolens* ssp. *acerba* var. *latifolia* and *M. diversipila* var. *subglabriseta*. In sect. *Calothamnos*, it was found in *M. aurivillus* var. *calothamnos*, *M. barretoii*, *M. bathyrrhena*, *M. berroi*, *M. eriocarpa*, *M. flocculosa*, *M. incana*, *M. involucrata*, *M. rocae*, *M. roseoalba* and *M. urticaria*.

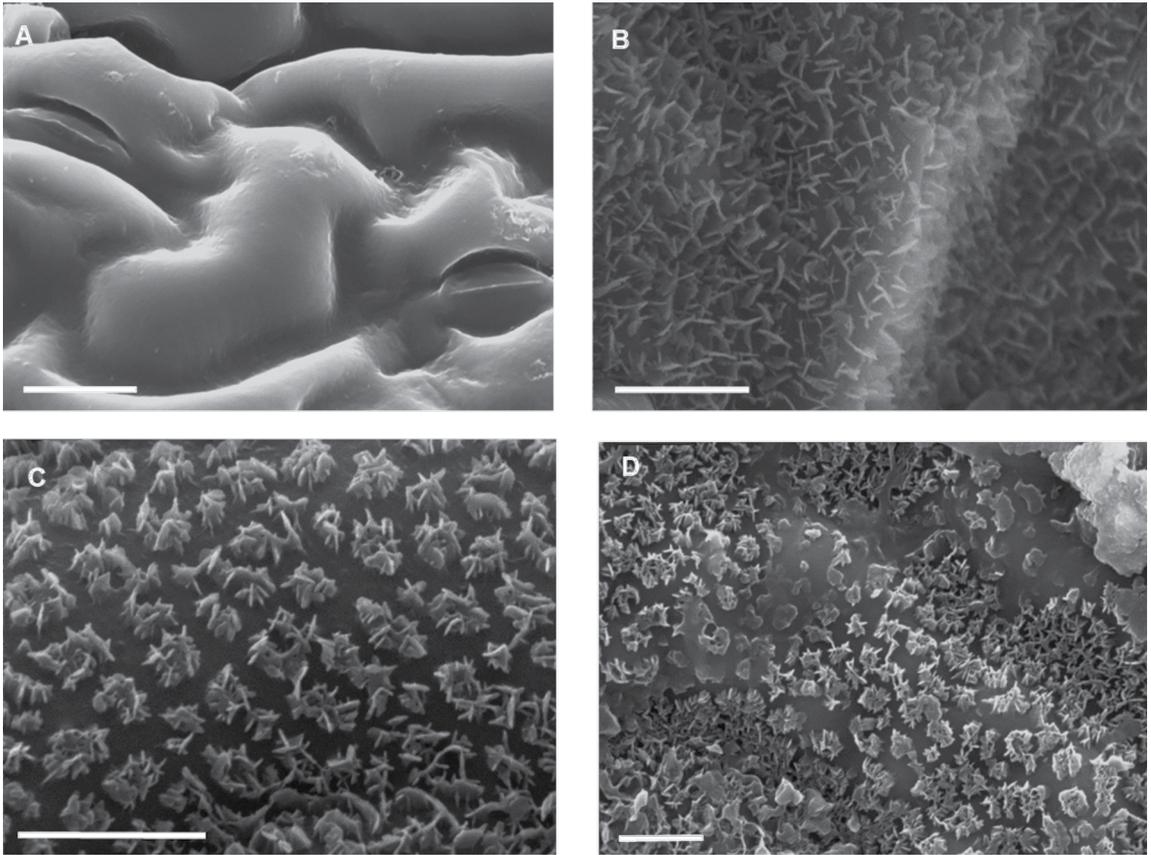


Fig. 5. SEM micrographs of epicuticular waxes in *M.* sect. *Calothamnos*. **A**, *M. bonplandii*, abaxial face, homogeneous film of wax. **B**, *M. roseoalba*, adaxial face, plates. **C**, *M. chrysastra*, adaxial face, platelet rosettes. **D**, *M. leprosa*, adaxial face, wax smoothings. The bar indicates: A = 10 μ m, B-D = 5 μ m.

Platelets rosette. Polygonal crystalloids with entire margins but indistinct edges, 460-1200 nm wide, 650-1020 nm long y 110-170 nm thick, are arranged in circular units (Figs. 4C, 5C). Crystalloids are not radially arranged around a central axe as in rosettes, but are randomly oriented. However, these clusters can be clearly identified, and they cover the whole leaflet surface. In sect. *Mimosa*, this deposition morphology was found in *M. dolens* ssp. *eriophylla*, some specimens of *M. dolens* ssp. *acerba* var. *acerba*, *M. dolens* ssp. *dolens* var. *foliolosa*, *M. dolens* ssp. *acerba* var. *rudis*, *M. dolens* var. *pangloea*, *M. brevipes*, *M. sceptrum*, *M. dolens* var. *anisitsii* and *M. custodis*. In sect. *Calothamnos* it was found in *M. aurivillus* var. *calothamnoides*, *M. aurivillus* var. *sordescens*, *M. chrysastra* var. *chrysastra*, *M. chrysastra* var. *itambeana*, *M. cylindracea*,

M. daleoides, *M. lepidorepens*, *M. leprosa*, *M. macedoana*, *M. pilulifera* var. *pilulifera*, *M. plumosa*, and *M. scabrella*.

Two distinct variations of these morphologies were also observed. In *M. dolens* ssp. *rigida* var. *foliolosa*, platelets are grouped in circular units, but are arranged only around stomata cells (Fig. 4D). In the rest of the leaflet surface, wax is deposited as a film adjusting to cell structure. On the other hand, in some taxa of sect. *Calothamnos*, epicuticular waxes crystals seem to be smoothed in some portions of the surface layer (Fig. 5D): *M. aurivillus* var. *calothamnoides*, *M. aurivillus* var. *sordescens*, *M. berroi*, *M. chrysastra* var. *chrysastra*, *M. chrysastra* var. *itambeana*, *M. cylindracea*, *M. daleoides*, *M. eriocarpa*, *M. furfuracea*, *M. leprosa*, *M. macedoana*, *M. pilulifera* var. *pilulifera*, *M. rocae* and *M. scabrella*. However, these morphologies are

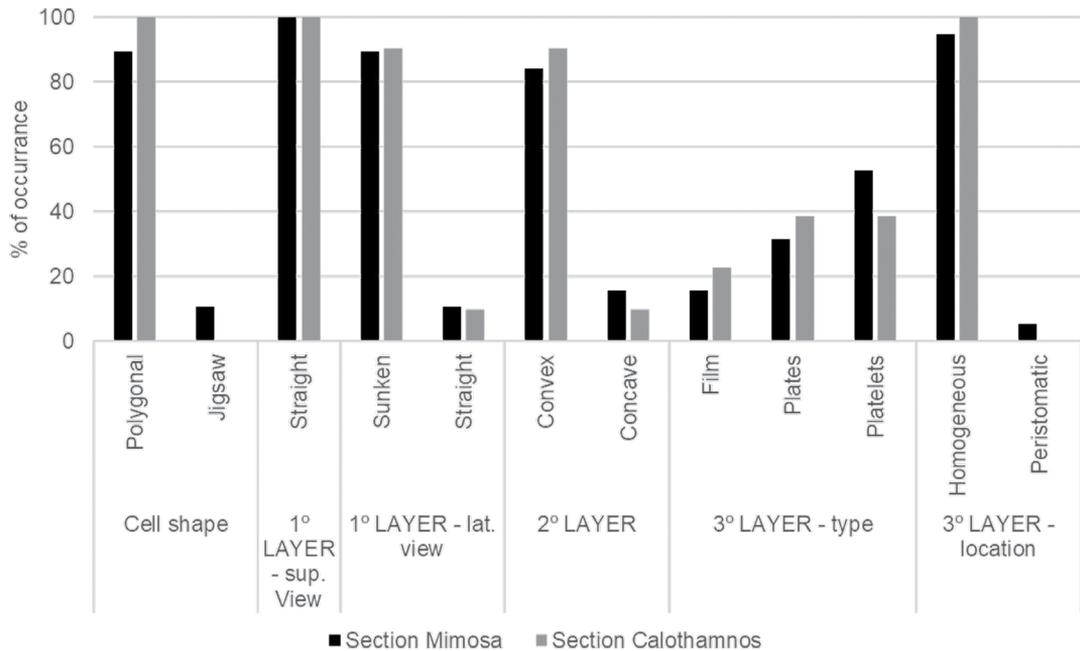


Fig. 6. Frequencies of each morphology of the primary, secondary and tertiary layers in *Mimosa*.

occasional and seem to follow no distinct pattern neither on the leaflet nor among specimens.

The frequencies of each morphology of the primary, secondary and tertiary layers vary similarly within and between sections (Fig. 6). Regarding cell shape, primary and secondary layer, always one morphology is the dominant one, that is, polygonal, straight, sunken and convex morphology, respectively, with frequencies higher than 90%. In the tertiary layer, the most frequent type of deposition in both sections is the platelet rosette, being identified in more than half of the taxa in sect. *Mimosa*, and about 40% in sect. *Calothamnos*. It is closely followed by plates (30–40% in both sections), and by the film (15–25%). In addition, some intraspecific variability is observed in sect. *Calothamnos* between faces of the leaflet (*M. eriocarpa*, *M. lepidorepens*).

DISCUSSION

Wax morphology. Cell wall morphology and its epicuticular depositions observed in *Mimosa* sect. *Mimosa* and *M.* sect. *Calothamnos* are mainly in accordance with the typical morphologies described

in angiosperms (Koch et al., 2006; Barthlott et al., 2017). The primary, secondary, and tertiary layers described for plants in general are all present in both sections (Behnke & Barthlott, 1983; Koch et al., 2006). It has been mentioned that the secondary and tertiary layers are mutually exclusive, that is, if one presents an elaborate morphology, the other presents a basic morphology, with very few exceptions (Behnke & Barthlott, 1983). Our results support this statement, since in most of the taxa of both sections the cell walls present a very uniform relief, with entire contours and a very slightly concave surface, but with elaborated epicuticular waxes (tertiary layer), which consists in crystalloids of different shapes and dispositions. In addition, we described here a novel combination for the genus of both secondary and tertiary layer with complex morphologies in *M. lepidorepens*, with prominent papillae on abaxial face of the leaflets, combined with a dense coverage of platelet rosettes. Papillae as these have already been described in genera from different plant families, such as *Euphorbia* L., *Oryza* L., and *Nelumbo* Adans. (Kochet al., 2006), and also in some related genera, for example, in *Acacia dealbata* Link (Wagner et al., 2003).

The morphology of the secondary layer, that is, the periclinal wall of the epidermis cells, can influence the interaction with water drops, determining the hydrophobicity of the surface: as the epidermis folds, the adhesion of water molecules to the surface is weaker, therefore allowing the water to drain easily (Wagner et al., 2003; Barthlott et al., 2017). Considering this, the abaxial face of *M. lepidorepens* would show the highest hydrophobicity of both sections, not only for the folded epidermal surface but also for the dense cover of wax crystals. This could be related to the distribution of this taxon in Serra do Quiriri (Santa Catarina, Brazil), where precipitation and air humidity levels are high throughout the year, as well as the fog is very frequent (De Souza et al., 2011).

The tertiary layer, which consists of epicuticular wax depositions, has been described in detail in different families (Jeffree et al., 1975; Barthlott, 1990; Barthlott et al., 1998) and has been taxonomically relevant in some genera from different plant families, such as *Pinus*, *Huberia*, *Zanthoxylum*, *Alstonia*, *Pilocarpus* and *Jasminum* L. (Herbin & Sharma, 1969; Jayeola, 1998; Mimura et al., 1998; Skorupa et al., 1998; Dias-Leme et al., 2013; Yohannan & Devipriya, 2017). Platelets have been identified as the typical wax deposition type in Fabales (Ditsch et al., 1995). Our results also show that this wax deposition type is also the most frequent in sect. *Mimosa* and sect. *Calothamos*. However, we additionally found two other different deposition morphologies: film and plates, either homogeneously arranged on the whole surface or only around stomata, being both less frequent, but not rare. Our results in plate size are similar to those reported in other families (Jeffree et al., 1975, 1976; Koch et al., 2006).

The type and density of epicuticular waxes can also influence the optical effect of the leaflet surface since it can appear either pale and greyish or bright and shiny (Baker et al., 1983; Koch et al., 2006, 2008). In *Mimosa* sect. *Calothamos*, this effect can be easily observed. On one hand, in *M. calodendron* and *M. taimbensis*, both with shiny leaflet surfaces, the wax deposition type is a film with no crystalloids, as has been described in other species (Baker et al., 1983). However, other taxa in both sections also show a wax film, but its leaflets are not shiny. Something similar was observed in *M. involucrata* or *M. lepidorepens*, which show pale and greyish

surfaces, but there seem to be no taxa-specific morphology. This indicates that the wax deposition type is not the only feature that determines the leaflet's optical properties. It has been previously suggested that more complex wax structures, like tubules, are necessary to really change the optical properties of a surface by itself (Baker et al., 1983).

Taxonomic relevance. *Mimosa* is a complex genus, and some of its sections based on both morphological traits and molecular phylogeny are not monophyletic (Bessega & Fortunato, 2011; Simon et al., 2011). Epidermal outer morphology could be a taxonomically relevant feature for some taxa with particular morphologies (e.g., papillae in *M. lepidorepens*), and are even for the distinction of varieties in some species of the genus (e.g., *M. aurivillus*). However, our results do not coincide with previous classifications (Barneby, 1991) at a supraspecific level, that is, between subseries and even the two sections considered here. In sect. *Mimosa*, other anatomical (Grohar et al., 2016; Grohar et al., 2018; Grohar et al., 2021) and also morphometric and chromosomic data (Morales et al., 2018) could help re-evaluate the supraspecific categories established only on morphological similarities (Barneby, 1991).

At a supraspecific level, the wax micromorphology by itself is not as relevant for the distinction between the sections *Calothamos* and *Mimosa* as other anatomical (Grohar et al., 2016; Grohar et al., 2018; Grohar et al., 2021) and also complex morphometric and cytogenetic analyses (Morales et al., 2018) could help reevaluate the supraspecific categories established only on morphological similarities (Barneby, 1991).

At a supraspecific level there is high variability in both of them. However, its relevance changes at inter- and intraspecific levels. In sect. *Calothamos*, wax deposition types help to differentiate the varieties of *M. aurivillus*, since *M. aurivillus* var. *sordescens* and *M. aurivillus* var. *calothamnoides* show platelet rosettes, *M. aurivillus* var. *calothamos* plates, and *M. aurivillus* var. *aurivillus* a wax film. These results, altogether with results from corolla micromorphology in the section (Grohar et al., 2021), questions the proposal of *M. aurivillus* as a single taxonomic species, which has already been suggested before based on morphological analysis (Coutinho, 2009).

In other species of the sect. *Calothamnos*, such as *M. chrysastra*, the distinction of its two varieties based on epidermal micromorphology is not observable, since they show the same deposition type, that is, platelet rosettes. Other micromorphological traits of corolla proved to be helpful in differentiating both varieties (Grohar et al., 2021).

In sect. *Mimosa*, it seems difficult to separate the subseries *Dolentes* and *Brevipedes* as it was proposed by Barneby (1991), because of the presence of the diagnostic traits in both subseries; it was analysed extensively by our previous works (Morales et al., 2018, 2019). This has also been observed in other traits, such as trichome micromorphology (Grohar et al., 2016). Although high variability of stomata morphology has been described at an intraspecific level in *Mimosa* (Montaño-Arias et al., 2018), in subser. *Dolentes* it is possible to associate types of wax deposition with some subspecies of *M. dolens*. In *M. dolens* ssp. *callosa* and *M. dolens* ssp. *eriophylla*, only one morphology is found: plates and platelets, respectively. In the other subspecies, it is not possible to define each one solely on one morphology.

However, distinction of varieties within the ssp. *rigida* and *dolens* is possible, as all varieties can be clearly differentiated by the wax deposition types. In *M. dolens* ssp. *rigida*, *M. dolens* var. *rigescens* is the only one with plates and *M. dolens* var. *deterior* is the only one with a wax film. Although *M. dolens* var. *foliolosa* and *M. dolens* var. *anisitsii* both have platelets, they also differ, as in *M. dolens* var. *foliolosa* the crystals are deposited only around the stomata guard cells, and in *M. dolens* ssp. *rigida* var. *anisitsii* crystals cover homogeneously the whole leaflet surface. Similarly, in *M. dolens* ssp. *dolens*, both varieties of *M. dolens* are clearly separated by wax deposition types, as *M. dolens* var. *dolens* shows a wax film, and in *M. dolens* var. *pangloea*, wax crystals have a platelet shape. In *M. dolens* ssp. *acerba*, there are many possible morphologies among the varieties, which fail to differentiate them. In subseries *Brevipedes*, we found that the main taxa are similar in wax morphology, since there is a trend to show a film or platelets in all studied taxa of the subseries, except for *M. diversipila* var. *subglabriseta*.

In addition to the wax morphology, location and epicuticular wax deposition type vary between taxa. Regarding wax spatial deposition, *M. dolens* ssp.

rigida var. *foliolosa* clearly distinguishes itself from all other taxa, as it is the only taxon in the complex in which platelets are arranged only around stomata cells. In all the other taxa, epicuticular waxes cover regularly the whole leaflet surface, independently of the morphology.

Regarding crystalloid types of the tertiary layer, the most frequent morphology in both subseries is the platelet. In the subseries *Dolentes*, it appears only in some varieties from each subspecies, while in subseries *Brevipedes*, it is found only in *M. custodis*, *M. brevipes* and *M. sceptrum*. Other deposition types, although less frequent, are also observed only in only a few taxa. These results could be useful as taxonomically relevant features at specific and infraspecific levels, as it was described in other anatomic traits, such as stomata micromorphology (Grohar et al., 2021). In highly variable taxonomic complexes such as *Dolentes-Brevipedes*, the micromorphological traits could provide new relevant data for the distinction of new species, as it was recently described in *M. schininii* (Morales et al., 2019).

CONCLUSIONS

The analysis of epicuticular morphology in Fabaceae has been scarce at an infrageneric level, and therefore underused as a taxonomic marker. The present study represents the most detailed analysis of epicuticular morphology in the genus *Mimosa*, and accounts for a huge variation in micromorphological features of epicuticular waxes within *Mimosa* sect. *Calothamnos* and *Mimosa* sect. *Mimosa* (subseries *Dolentes* and *Brevipedes*). These data could be a potential tool of great taxonomic value within the complex, as they clearly separate individual taxa or groups at a specific and infraspecific level. For instance, *M. lepidorepens* shows a unique micromorphology of papillae, covered with platelets, on the abaxial face of the leaflets. This type of analysis is particularly helpful in the differentiation of varieties of *M. aurivillus*, *M. dolens* ssp. *rigida* and *M. dolens* ssp. *dolens*. However, it is not relevant at a supraspecific level by itself; at that level, it is necessary to solve taxonomic conflicts with other tools. This problem leads to the requirement of multidisciplinary studies including exo-morphological, anatomical, chromosome and molecular data.

ACKNOWLEDGMENTS

We are grateful to all the staff of Instituto de Recursos Biológicos (CIRN, INTA) for their assistance during our visits to the BAB herbarium. We are also grateful to the curators of CTES, G, HUCS, MBM, MVFA, RB, SPF, and SI herbaria for easing access to the material. We acknowledge financial support from Universidad de Buenos Aires, Argentina (UBACyT Project 2013-2016 GC 20020120100056BA), Universidad de Morón, Morón, Argentina (PID UM 06-002-16), Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET, PIP 112-20110100250) and Agencia Nacional de Promoción Científica y Tecnológica, Argentina (PICT 2016-0021).

BIBLIOGRAPHY

- Anton, L.; F. W. Ewers, R. Hammerschmidt & K. L. Klomparens. 1994. Mechanisms of deposition of epicuticular wax in leaves of broccoli, *Brassica oleracea* var. *capitata* L. *New Phytologist* 126: 505-510.
- Baker, E. A.; G. M. Hunt & P. J. G. Stevens. 1983. Studies of plant cuticle and spray droplet interactions: a fresh approach. *Pesticide Science* 14: 645-658.
- Barneby, R. C. 1991. *Sensitivae Censitae. A Description of the Genus Mimosa Linnaeus (Mimosaceae) in the New World. Memoirs of the New York Botanical Garden* 65. Stuttgart: Schweizerbart.
- Barnes, J. D. & J. Cardoso-Vilhena. 1996. Interactions between electromagnetic radiation and the plant cuticle. *Plant Cuticles: An Integrated Functional Approach* 157. BIOS Scientific Publishers, Oxford, UK: 170.
- Barthlott, W. 1990. Scanning Electron Microscopy of the epidermal surface in plants. *Scanning Electron Microscopy in Taxonomy and Functional Morphology* 41. Clarendon Press Oxford. Pp 69-94.
- Barthlott, W.; M. Mail, B. Bhushan & K. Koch. 2017. Plant surfaces: structures and functions for biomimetic innovations. *Nano-Micro Letters* 9: 1-40.
- Barthlott, W.; C. Neinhuis, D. Cutler, F. Ditsch, I. Meusel, I. Theisen & H. Wilhelmi. 1998. Classification and terminology of plant epicuticular waxes. *Botanical Journal of the Linnean Society* 126: 237-60.
- Behnke, H. D. & W. Barthlott. 1983. New evidence from the ultrastructural and micromorphological fields in Angiosperm classification. *Nordic Journal of Botany* 3: 43-66.
- Bentham, G. 1875. VII. Revision of the Suborder Mimosaeae. *Transactions of the Linnean Society of London* 3: 335-664.
- Bessegga, C. & R. H. Fortunato. 2011. Section *Mimadenia*: Its phylogenetic relationships within the genus *Mimosa* (Leguminosae, Mimosoideae) using plastid TrnL-F sequence data. *Australian Systematic Botany* 24: 104-110.
- Bessegga, C.; H. E. Hopp & R. H. Fortunato. 2008. Toward a phylogeny of *Mimosa* (Leguminosae: Mimosoideae): A preliminary analysis of southern South American species based on chloroplast DNA sequence. *Annals of the Missouri Botanical Garden* 95(4): 567-579.
- Bondada, B. R.; D. M. Oosterhuis, J. B. Murphy & K. S. Kim. 1996. Effect of water stress on the epicuticular wax composition and ultrastructure of cotton (*Gossypium hirsutum* L.) leaf, bract, and boll. *Environmental and Experimental Botany* 36: 61-69.
- Coutinho, A. P. S. 2009. Revisão taxonômica e estudos filogenéticos de *Mimosa* L. sect. *Calothamnos* Barneby (Leguminosae-Mimosoideae). Doctoral dissertation, Universidade de São Paulo.
- Dias-Leme, C. L.; S. V. A. Hohlemwerger, J. Armando, M. L. S. Guedes, M. Edijane & S. Vellozo Eudes. 2013. Leaflet anatomy and epicuticular wax composition of *Zanthoxylum tingoassuiba* (Rutaceae). *Latin American Journal of Pharmacy* 32: 425-430.
- Ditsch, F.; H. Patha & W. Barthlott. 1995. Micromorphology of epicuticular waxes in Fabales sl and its systematic significance. *Beiträge zur Biologie der Pflanzen* 68: 297-310.
- Dutra, V. F. & F. C. P. Garcia. 2013. Three new species of *Mimosa* (Leguminosae) from Minas Gerais, Brazil. *Systematic Botany* 38: 398-405.
- Ensikat, H. J.; M. Boese, W. Mader, W. Barthlott & Kerstin Koch. 2006. Crystallinity of plant epicuticular waxes: Electron and X-Ray diffraction studies. *Chemistry and Physics of Lipids* 144: 45-59.
- Grohar, M. C. 2020. *Mimosa* Sección *Calothamnos* (Fabaceae): Estudios morfológicos, citogenéticos y de distribución, e inferencias taxonómicas. PhD Thesis. Ciudad de Buenos Aires: Universidad de Buenos Aires.
- Grohar, M. C.; S. Rosenfeldt, R. H. Fortunato & M. Morales. 2021. Comparative floral micromorphology in *Mimosa* Sect. *Calothamnos* (Fabaceae). *Annals of the Missouri Botanical Garden* 106: 271-291.
- Grohar, M.; S. Rosenfeldt & M. Morales. 2016. Leaflet trichome micromorphology in the Dolentes-Brevipedes taxonomic complex (*Mimosa* L., Mimosoideae). *Turkish Journal of Botany* 40: 45-58.
- Grohar, M. C.; S. Rosenfeldt & M. Morales. 2018. Venation patterns in *Mimosa* Subseries *Dolentes* and *Brevipedes* (Leguminosae) and their taxonomical inferences. *Systematic Botany* 43: 532-543.

- Grohar, M. C.; S. Rosenfeldt & M. Morales. 2021. Stomatal micromorphology in a complex of *Mimosa* Section *Mimosa* (Fabaceae). *Protoplasma* 259(1): 203-215.
- Gülz, P. G.; E. Müller & R. B. N. Prasad. 1991. Developmental and seasonal variations in the epicuticular waxes of *Tilia tomentosa* leaves. *Phytochemistry* 30: 769-773.
- Herbin, G. A. & K. Sharma. 1969. Studies on plant cuticular waxes—V. The wax coatings of pine needles: A taxonomic survey. *Phytochemistry* 8: 151-160.
- Jayeola, A. A. 1998. Surface sculpturing of *Alstonia booneii* DE WILD. and *A. congensis* ENGL. (Apocynaceae) and its importance on their taxonomy. *Feddes Repertorium* 109: 429-433.
- Jeffree, C. E.; E. A. Baker & P. J. Holloway. 1975. Ultrastructure and recrystallization of plant epicuticular waxes. *New Phytologist* 75: 539-549.
- Jeffree, C. E.; E. A. Baker & P. J. Holloway. 1976. Origins of the fine structure of plant epicuticular waxes. In: Microbiology of aerial plant surfaces, 119-158. London: Academic press.
- Jenks, M. A.; E. N. Ashworth & J. Janick. 2010. Plant epicuticular waxes: function, production, and genetics. *Horticultural Reviews* 23.
- Jenks, M. A.; H. A. Tuttle, S. D. Eigenbrode & K. A. Feldmann. 1995. Leaf epicuticular waxes of the eceriferum mutants in *Arabidopsis*. *Plant Physiology* 108: 369-377.
- Jetter, R. & S. Schaffer. 2001. Chemical composition of the *Prunus laurocerasus* leaf surface. Dynamic changes of the epicuticular wax film during leaf development. *Plant Physiology* 126: 1725-1737.
- Koch, K.; B. Bhushan & W. Barthlott. 2008. Diversity of structure, morphology and wetting of plant surfaces. *Soft Matter* 4: 1943-1963.
- Koch, K.; A. Dommissie & W. Barthlott. 2006. Chemistry and crystal growth of plant wax tubules of Lotus (*Nelumbo nucifera*) and Nasturtium (*Tropaeolum majus*) leaves on technical substrates. *Crystal Growth & Design* 6: 2571-2578.
- Luna-Castro J.; M. Morales & R. H. Fortunato. 2012. *Mimosa diversipila* var. *subglabriseta* (Fabaceae), a new record for the Flora of Paraguay. *Boletín de la Sociedad Argentina de Botánica* 47: 457-460.
- Maffei, M. 1996. Chemotaxonomic significance of leaf wax N-Alkanes in the Umbelliferae, Cruciferae and Leguminosae (Subf. Papilionoideae). *Biochemical Systematics and Ecology* 24: 531-545.
- Mimura, M. R. M.; M. L. F. Salatino, A. Salatino & J. F. A. Baumgratz. 1998. Alkanes from foliar epicuticular waxes of *Huberia* species: taxonomic implications. *Biochemical Systematics and Ecology* 26: 581-588.
- Montaño-Arias, S. A.; S. L. Camargo-Ricalde & R. Grether. 2018. Stomatal complexes of twenty-four species of *Mimosa* (Leguminosae) occurring in Mexico. *Acta Botánica Mexicana* 122: 97-107.
- Morales, M.; L. Arenas, M. I. Remis, A. F. Wulff, L. Poggio & R. H. Fortunato. 2014. Morphometric and cytogenetic studies in *Mimosa diversipila* (Mimosoideae, Leguminosae) and their taxonomic and evolutionary inferences. *Systematic Botany* 39: 875-883.
- Morales, M.; M. Fradkin, C. Bessega, L. Poggio & R. H. Fortunato. 2018. Cytogeography and morphological characterisation of a taxonomic, polyploid complex of *Mimosa* (Leguminosae) from subtropical South America. *Australian Systematic Botany* 31: 190-208.
- Morales, M.; M. C. Grohar, S. Rosenfeldt & R. H. Fortunato. 2019. A new species of *Mimosa* Section *Mimosa* (Leguminosae) from the Paraguayan Cerrado. *Phytotaxa* 401: 24-32.
- Morales, M.; O. S. Ribas & R. H. Fortunato. 2015. Increasing reports in *Mimosa* (Mimosoideae, Leguminosae) for the Brazilian Flora. *Boletín de la Sociedad Argentina de Botánica* 50: 221-233.
- Morales, M.; A. F. Wulff, R. H. Fortunato & L. Poggio. 2010. Chromosome and morphological studies in the *Mimosa debilis* Complex (Mimosoideae, Leguminosae) from southern South America. *Australian Journal of Botany* 58: 12-22.
- Oliveira, A. F. M.; S. T. Meirelles & A. Salatino. 2003. Epicuticular waxes from Caatinga and Cerrado species and their efficiency against water loss. *Anais da Academia Brasileira de Ciências* 75: 431-439.
- Ramírez, R. G.; J. L. Hernandez-Pineiro & R. K. Maiti. 1997. Nutritional profile and leaf surface structure of some native shrubs consumed by small ruminants in semiarid regions of Northeastern Mexico. *Journal of Applied Animal Research* 11: 145-156.
- Savassi-Coutinho, A. P.; G. P. Lewis & V. C. Souza. 2012. *Mimosa roseoalba* (Leguminosae: Mimosoideae), a new species from Mato Grosso do Sul, Brazil. *Kew Bulletin* 67: 827-831.
- Sen, A. 1987. Chemical composition and morphology of epicuticular waxes from leaves of *Solanum tuberosum*. *Zeitschrift Für Naturforschung C* 42: 1153-1158.
- Simon, M. F.; R. Grether, L. P. Queiroz, T. E. Särkinen, V. F. Dutra & C. E. Hughes. 2011. The evolutionary history of *Mimosa* (Leguminosae): Toward a phylogeny of the sensitive plants. *American Journal of Botany* 98: 1201-1221.
- Skorupa, L. A.; M. L. F. Salatino & A. Salatino. 1998. Hydrocarbons of leaf epicuticular waxes of *Pilocarpus*

- (Rutaceae): Taxonomic meaning. *Biochemical Systematics and Ecology* 26: 655-662.
- Souza Werneck, M.; M. E. Guerra Sobral, C. T. Vieira Rocha, E. C. Landau & J. R. Stehmann. 2011. Distribution and endemism of Angiosperms in the Atlantic Forest. *Nature Conservation* 9: 188-193.
- Thiers, B. [continuously updated, accessed 2022]. Index Herbariorum: a global directory of public herbaria and associated staff. New York Botanical Garden's Virtual Herbarium, <http://sweetgum.nybg.org/science/ih/>
- Wagner, P.; R. Fürstner, W. Barthlott & C. Neinhuis. 2003. Quantitative assessment to the structural basis of water repellency in natural and technical surfaces. *Journal of Experimental Botany* 54: 1295-1303.
- Wallace, L. L.; P. Timpano, & P. Durgin. 1987. Leaf folding in *Mimosa pudica* (Fabaceae): A nutrient conservation mechanism? *American Journal of Botany* 74: 132-135.
- Yohannan, R. & V. Devipriya. 2017. Analysis of the epicuticular wax composition of *Jasminum* employing SEM-EDX. *International Journal of Advanced Research* 5: 820-823.

Appendix. Herbarium specimens examined

Mimosa section *Mimosa*

- M. brevipes*. BRASIL. **Mato Grosso do Sul**. Rod. BR-267, próximo do trevo para Bela Vista, 11-III-2003, *Hatschbach et al.* 74574 (MBM 323494).
- M. dolens* ssp. *callosa*. ARGENTINA. **Corrientes**. Santo Tomé, Ruta Nacional 12, entre Posadas e Ituzaingó, 18-III-2013, *Morales et al.* 1357 (BAB).
- M. cryptogloea*. PARAGUAY. **San Pedro**. 36 km N de San Estanislao, 18-V-1969, *Krapovickas et al.* 13955 (CTES).
- M. custodis*. PARAGUAY. **Amambay**. Pedro Juan Caballero, 14-II-1951, *Schwarz 11900* (CTES91920).
- M. diversipila* var. *subglabriseta*. PARAGUAY. **Itapúa**. Ayolas, Refugio, 01-II-1982, *Bordas 3019* (CTES91820).
- M. dolens* ssp. *eriophylla*. BRASIL. **Mato Grosso do Sul**. Antonio João, Rodovia Amambay–Antonio João, 07-V-2009, *Morales et al.* 772 (BAB).
- M. dolens* var. *acerba*. BRASIL. **Paraná**. Guarapuava. Camino en dirección al Cristo, 18-IV-2009, *Fortunato et al.* 9479 (BAB). **São Paulo**. Ipiranga, 07-II-1908, *Swederwaldt 3* (BAB). Palmeira. **São Mateus do Sul**, 25-IV-2009, *Fortunato 9612* (BAB).
- M. dolens* var. *anisitsii* (Lindman) Barneby. PARAGUAY. **San Pedro**. San Pedro, a 1 km N Del Barrio San Pedro, en dirección a Pedro Caballero por ruta 3, 07-III-2009, *Fortunato et al.* 9217 (BAB).
- M. dolens* var. *deterior* Barneby. BRASIL. **São Paulo**. Itararé: estrada Itararé-BomSucesso, próximo a entrada da Fda. São Nicolau, 21-VI-1993, *Souza et al.* 2167 (MBM).
- M. dolens* var. *foliolosa* (Benth) Barneby. ARGENTINA. **Corrientes**. San Miguel, Ruta Provincial 118, a 54 km de la intersección con Ruta Nacional 12, 07-III-2007, *Morales & Seijo 277* (BAB).
- M. dolens* var. *latifolia* (Benth) Barneby. BRASIL. **Paraná**. Palmeira: Recanto dos Papagaios, 18-IV-2004, *Fortunato et al.* 9514 (BAB).
- M. dolens* var. *pangloea* Barneby. BRASIL. **Paraná**. Palmeira: BR277, 15 km al W de Colonia Quero-Quero, 25-IV-2009, *Fortunato et al.* 9597 (BAB).
- M. dolens* var. *rudis* (Benth) Barneby. PARAGUAY. **Amambay**. Colonia Fortuna Guazú, 09-III-2008, *Fortunato et al.* 9279 (BAB).
- M. sceptrum* Barneby. PARAGUAY. **Amambay**. Colonia Fortuna Guazú, 09-III-2008, *Fortunato et al.* 9282 (BAB).
- M. pseudopetiolaris*. PARAGUAY. **Paraguari**. Ybicuri, 14-XII-1989, *Zardini 8688* (MO1518284).
- M. dolens* var. *dolens* Vellozo. BRASIL. **São Paulo**. Jabaquara, 05-II-1968, *Handro 2013* (SPF83076).
- M. dolens* var. *rigescens* (Benth) Barneby. BRASIL. **Paraná**. Lapa, BR-476, 5 km O de Lapa, 30-XI-1986, *Ribas & Silva 201* (MBM132263).
- M. brevipes* Benth. BRASIL. **Mato Grosso do Sul**. Bonito: Rodovia Bonito a Campo dos Índios, próximo de Três Morros, 10.III.2003, *Hatschbach et al.* 74471 (MBM323491).

Mimosa* section *Calothamnus

- M. aurivillus* var. *calothamnoides*. BRASIL. **Minas Gerais**. Alto Caparaó: Parque Nacional do Caparaó, 8-III-2010, *Cordeiro & Silva 3556* (BAB). Santa Barbara: Serra do Caraça, camino a la Capelinha, SW de Catas altas, 18-II-1991, *Arbo, Menezes, Schinini & Campos 5262* (CTES);
- M. aurivillus* var. *calothamnus*. BRASIL. **São Paulo**. São José do Barreiro: Serra da Bocaina, 6-III-2010, *Cordeiro & Silva 3477* (BAB).
- M. aurivillus* var. *sordescens*. BRASIL. **Minas Gerais**. Grao Mogol: Ribeirao, 23-IV-1978, *Hatschbach 41420* (CTES).
- M. barretoii*. BRASIL. **Minas Gerais**. Santana do Riacho: Serra do Cipó, retiro no alto do Palacio, 25 km NE de Cardenal Mota camino a Conceicao de Mato Dentro, 12-II-1991, *Arbo, Menezes, Schinini & Campos 4954* (BAB).
- M. bathyrrhena*. BRASIL. **Paraná**. Ponta Grossa: Passo do Pupo, 19-X-2006, *Silva, Sampaio & Coutinho 5143* (SI).
- M. berroi*. URUGUAY. **Tacuarembó**. Tacuarembó: Alrededores de Gruta de los Cuervos, 09-IV-2009, *Morales 697* (BAB); Gruta de los Cuervos, 25-II-2018, *Morales 2063* (BAB).
- M. bonplandii*. ARGENTINA. **Corrientes**. Paso de los Libres: próximo a Laguna brava, 29°S 56°W, 7-XI-1973, *Correa 5341* (BAB).
- M. calodendron*. BRASIL. **Minas Gerais**. Belo Horizonte: Serra do Cipo, 10-II-1927, *Hoehne 18513* (SI).
- M. chrysastra* var. *chrysastra*. BRASIL. **Minas Gerais**. Sto. Antonio do Itambé: Subida ao Pico do Itambé, 05-IV-1982, *Rossi, Furlan & Henrold 3018* (BAB).
- M. chrysastra* var. *itambeana*. BRASIL. **Minas Gerais**. Sto. Antonio do Itambé: Pico do Itambé, 07-IV-1998, *Souza, Souza, Capellari, Elias y Forster 21134* (RB).
- M. cylindracea*. BRASIL. **Minas Gerais**. Caldas, 1826, *Regnell II97* (SI). **São Paulo**. Sin municipio: próximo a Interlagos, 02.IX.1947, *Hoehne NN* (MBM).
- M. daleoides*. BRASIL. **Mato Grosso do Sul**. El dorado: Fazenda Entre Ríos, río Yhovoy, 06-V-2009, *Morales 720* (BAB). URUGUAY. **Tacuarembó**. Tacuarembó: Camino a la Gruta de los Cuervos, 09-XII-1965, *Del Puerto 5587* (MVFA).
- M. eriocarpa*. BRASIL. **Rio Grande do Sul**. No division, 1814-1831, no collector (SI 18316).
- M. flocculosa*. BRASIL. **Paraná**. Curitiba: Campo Morao, Jardín Botánico Municipal de Curitiba, 16-IV-2009, *Fortunato 9476* (BAB).
- M. furfuracea*. BRASIL. **Santa Catarina**. Campo alegre: Morro do Iquererim, 5-IX-1957, *Reitz & Klein 4779* (SI).
- M. incana*. BRASIL. **Rio Grande do Sul**. São Marcos: Saída para Flores da Cunha, 31-VIII-2011, *Grizzon 44* (BAB).
- M. involucrata*. BRASIL. **Rio Grande do Sul**. Cambará do Sul: Parque Nacional dos Aparados da Serra, Canion Itaimbezinho, no date, *Bortoluzzi, Pereira & Pereira 1208* (MBM).
- M. lepidorepens*. BRASIL. **Santa Catarina**. No location: 1,5 km antes de la Posada Alto Quirirí, 23-IV-2009, *Fortunato 9567* (BAB).
- M. leprosa* var. *leprosa*. BRASIL. **Minas Gerais**. Caeté: Serra da Piedade, 1843, *Claussen 120* (SI).
- M. macedoana* var. *macedoana*. BRASIL. **Minas Gerais**. Jaboticatuba: Serra do Cipó, Fda. Palacio, 14-II-1973, *Hatschbach 31556* (SI).
- M. myuros*. BRASIL. **Minas Gerais**. Sapucaí – Mirim. 6-II-1953, *Kuhlmann 2908* (SI).
- M. pilulifera* var. *pilulifera*. ARGENTINA. **Entre Ríos**. Gualaguaychú: Balneario Ñandubaysal, monte aledaño a la playa, 12-IV-2009, *Morales 698* (BAB). BRASIL. **Paraná**. Herval: Torre da Brasil Telecom, 09-XI-2009, *Barbosa, Cordeiro, y Silva 2310* (BAB, HUCS).
- M. plumosa*. PARAGUAY. **Cordillera**. Ybycu'í: Parque nacional Ybycu'í, área NE, 1 km al sur del arroyo Corrientes, 10-II-1993, *Zardini & Guerrero 34948* (BAB).
- M. rocae*. ARGENTINA. **Buenos Aires**. Saavedra: Sa. de Cura-malal, 21-XI-1972, *Gómez, Leguizamón & Martínez 11921* (BAB).

- M. roseoalba*. BRASIL. **Paraná**. Mundo Novo: Salida de Mundo Novo para Eldorado, cerca del Puesto policial, 06-V-2009, *Morales 709* (BAB).
- M. scabrella*. ARGENTINA. **Misiones**. El Dorado: 14-XI-1980, *Miranda 16* (CTES). Loreto: 11-II-1945, *Burkart 1525* (SI). BRASIL. **Santa Catarina**. Fraiburgo. 22-I-1992, *Krapovickas & Cristóbal 43981* (BAB).
- M. taimbensis*. BRASIL. **Rio Grande do Sul**. Cambará do Sul: Fortaleza, 16-IV-1994, *Hatschbach 60608* (BAB).
- M. urticaria*. BRASIL. **Paraná**. Ortigueira: Bairro dos Machados, 13-XII-1987, *Hatschbach 51709 & Manosso* (BAB); BR 376 Portal de Ortigueira, vera do riacho, 20-IV-2009, *Fortunato 9536* (BAB).