



Biotecnia

ISSN: 1665-1456

Universidad de Sonora, División de Ciencias Biológicas y de la Salud

Martinez-Morales, Flavio; Zapata-Morales, Juan Ramon; Aragon-Martinez, Othoniel Hugo
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Biotecnia, vol. 24, no. 1, 2022, January-April, pp. 69-78
Universidad de Sonora, División de Ciencias Biológicas y de la Salud

DOI: <https://doi.org/10.18633/biotecnia.v24i1.1546>

Available in: <https://www.redalyc.org/articulo.oa?id=672974928008>

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Evaluation of the antioxidant interaction between butylated hydroxytoluene and quercetin and their utility for beef patties preservation

Evaluación de la interacción antioxidante de hidroxitolueno butilado y quercetina y su utilidad para la preservación de hamburguesas de carne de res

Flavio Martinez-Morales^a, Juan Ramon Zapata-Morales^b, Othoniel Hugo Aragon-Martinez^{a,c*}

^a Departamento de Farmacología, Facultad de Medicina, Universidad Autónoma de San Luis Potosí. Avenida Venustiano Carranza 2405, Colonia Los Filtros, San Luis Potosí, S.L.P., C.P. 78210, México.

^b Departamento de Farmacia, Division de Ciencias Naturales y Exactas, Universidad de Guanajuato. Noria Alta s/n, Colonia Noria Alta, Guanajuato, Gto., C.P. 36050, Mexico.

^c LABCON Laboratorio de Compuestos Naturales, Don Alejo 436, Colonia Don Antonio, San Luis Potosí, S.L.P., C.P. 78414, México.

ABSTRACT

The present study aimed to find a mixture of butylated hydroxytoluene (BHT) and quercetin with an antioxidant synergism that would enable the use at low concentrations of both compounds and, subsequently, evaluate the preservative action of the combination selected on ground beef patties stored at 4 °C. Our results showed that quercetin possessed higher antioxidant activity than BHT, and of the five combinations tested, the 1:5 BHT-quercetin combination presented a synergistic antioxidant activity. For the refrigerated beef patties, the 1:5 combination (5.2 and 26.0 mg/kg of BHT and quercetin, respectively) produced a beneficial action on the redness, yellowness, antioxidant capacity, metmyoglobin content, and thiobarbituric acid reactive substance value, which was similar to that produced by 100 or 36 mg/kg of BHT or quercetin ($P > 0.05$, except the redness induced by quercetin). These benefits were compared with the data obtained from a preservative-free patty control sample ($P < 0.001$ to < 0.05). Our study shows, for the first time, that the use of a 1:5 BHT-quercetin combination was an effective, simple, and available method to preserve refrigerated beef patties, which can substitute the individual addition of high concentrations of these compounds by the meat industry.

Keyword: antioxidant activity; synergism; ground beef; color degradation; protein oxidation.

RESUMEN

El presente estudio tuvo como objetivo encontrar una mezcla de BHT y quercetina con un sinergismo antioxidante que permitiera usar a ambos compuestos a concentraciones bajas para posteriormente, evaluar la acción preservante de la combinación seleccionada en hamburguesas de carne de res almacenadas a 4 °C. Nuestros resultados mostraron que quercetina tuvo mayor actividad antioxidante que BHT y de las cinco combinaciones evaluadas, la combinación 1:5 de BHT y quercetina presentó una actividad antioxidante mayor. Por lo tanto, esta misma combinación 1:5 a una dosis de 5.2 y 26 mg/kg de BHT y quercetina fue empleada para las ham-

burguesas de carne, donde ésta produjo una acción benéfica en la abundancia del color rojo y amarillo y en la capacidad antioxidante y una reducción de metamioglobina y sustancias reactivas al ácido tiobarbitúrico, semejante al producido por 100 o 36 mg/kg de BHT o quercetina, en comparación a los valores observados en las hamburguesas sin conservadores (testigo). Con este estudio se muestra por primera vez que, el uso de una combinación 1:5 de BHT-quercetina es un método efectivo, sencillo y asequible para preservar hamburguesas de carne, lo cual podrá sustituir la adición individual de altas concentraciones de estos compuestos por la industria cárnica.

Palabras clave: actividad antioxidante; sinergismo; carne de res molida; degradación del color; oxidación proteica.

INTRODUCTION

For decades, beef has played a crucial role in human diet as an important source of proteins, minerals, vitamins, and other nutrients vital for human health. As ground beef constitutes 64 % of all meat consumed by humans, extending the shelf-life of this product is a major challenge for the meat industry (Ouerfelli *et al.*, 2019; Weinroth *et al.*, 2019). In this regard, a shelf-life of approximately ten days at refrigerated temperatures can enable a meat product to be distributed to retail outlets (Kapetanakou *et al.*, 2020). Meat and meat products are susceptible to microbial growth, discoloration, and processes of protein and lipid oxidation (Ouerfelli *et al.*, 2019; Kapetanakou *et al.*, 2020). Currently, one of the methods applied to preserve meat and meat products is the use of additives with antioxidant properties such as butylated hydroxytoluene (BHT) (Ouerfelli *et al.*, 2019).

Recognized as safe for use in food under United States regulations, BHT, a synthetic phenolic compound, is included in the Title 21 of the Code of Federal Regulations and the Food Additive Status List of the US Food and Drug Administration (FDA) (FDA, 2019; Electronic Code of Federal Regulations, 2021). In this way, BHT can be added singly or in combination with other synthetic phenolic antioxidants to

*Autor para correspondencia: Othoniel Hugo Aragon-Martinez
Correo electrónico: hugo.aragon@uaslp.mx

Recibido: 08 de agosto de 2021

Aceptado: 16 de octubre de 2021

fat, oils, and meat, at a concentration of 100 mg/kg (Cantú-Valdéz *et al.*, 2020; Liu and Mabury, 2020). Nevertheless, due to their ubiquity, BHT possess several human exposure pathways, including the food intake, indoor dust ingestion, and use of personal care products (Liu and Mabury, 2020). Timing and dose exposure are important factors modulating the potentially deleterious effects of BHT use (Wang and Kannan, 2019). For example, a consumption of 1.35-5 % or 0.05-0.5 % BHT over 30 days or 10 months caused a toxic nephrosis or development of liver tumors in mice (Liu and Mabury, 2020).

In order to reduce the use of synthetic additives, preservatives obtained from natural sources have been developed to ensure safety, encourage customer acceptance, and expand shelf-life for meat products (Mtibaa *et al.*, 2019; Zahid *et al.*, 2019). Although several natural preservatives have been reported in the literature, BHT continues to be used extensively by the food and cosmetic industry (Liu and Mabury, 2020). One potential natural preservative is quercetin (3,3',4',5,7-pentahydroxyflavone), which is a flavonoid found in apples, green beans, broccoli, tomatoes, onions, Ginkgo biloba, and milk thistle, among others (Bekhit *et al.*, 2003; National Institute of Diabetes and Digestive and Kidney Diseases, 2012; Andres *et al.*, 2018). A typical western diet provides an estimated daily quercetin intake of 4 to 40 mg (National Institute of Diabetes and Digestive and Kidney Diseases, 2012). There are numerous reports on the biological effects of this flavonoid, based on both in vitro experiments and animal studies including humans. The results of these studies reveal that quercetin has antioxidant, anti-inflammatory, immunoprotective, anticarcinogenic, and ergogenic effects (Andres *et al.*, 2018). Currently, quercetin is marketed as an ingredient in various dietary supplements, is generally well-tolerated, and is free of discernible adverse events (National Institute of Diabetes and Digestive and Kidney Diseases, 2012; Andres *et al.*, 2018). Quercetin is recognized as safe by the FDA, with additional mean intakes of 200 or 400 mg/day estimated for all ages or high-intake consumers, respectively (Andres *et al.*, 2018). Although quercetin is a compound obtained from natural sources, some studies show that high doses may cause enhanced nephrotoxic effects in pre-damaged kidneys, and may promote the growth of already existing cancer cells (Andres *et al.*, 2018).

The use of low concentrations of additives recognized as safe under a certain limit, is desirable to avoid adverse health effects by an accumulative exposure but maintaining their preservative effects on meat products. On the other hand, combination studies are common methods for identifying synergy among drugs, a phenomenon which can facilitate the reduction of dosages while maintaining superior or similar therapeutic efficacy to that obtained from drugs when administered individually and, moreover, a lower incidence of adverse events than that obtained from individual use (Chou, 2006; Zapata-Morales *et al.*, 2021). To our knowledge, a combination approach has never been applied to preservatives added to meat products. Therefore, the present study

aimed to identify the BHT-quercetin mixture with a synergic antioxidant action and, subsequently, evaluate its utility against degraded antioxidant capacity, color degradation, and protein and lipid oxidation in refrigerated beef patties to which either the selected BHT-quercetin combination, BHT or quercetin alone had been added.

MATERIALS AND METHODS

Reagents

Quercetin, BHT, 2,2'-diphenyl-1-picrylhydrazyl radical (DPPH), dimethyl sulfoxide (DMSO), thiobarbituric acid, trichloroacetic acid, hydrochloric acid, and 1,1,3,3-tetraethoxypropane used in the present study were obtained from Sigma-Aldrich (St. Louis, MO, USA). A food grade Spanish extra virgin olive oil (EVO) was used (Kuali® trademark, Monterrey, NL, Mexico), while methanol and ethanol were of liquid chromatography and spectrophotometry grade (Mallinckrodt Baker Inc., Mexico City, Mexico). Deionized water (MontRial, San Luis Potosi, Mexico) was used for aqueous solutions.

DPPH radical scavenging assay

A stock solution of 920 µmol/L DPPH radical was prepared in methanol, with stock solutions of 600 µg/mL quercetin and 1400 µg/mL BHT prepared in ethanol-DMSO (96:4 v/v). Subsequently, working standards for the quercetin were established between 3 and 600 µg/mL and for the BHT between 7 and 1400 µg/mL using ethanol-DMSO (96:4 v/v), while a 460 µmol/L DPPH radical solution was prepared from the stock solution of DPPH radical and methanol. For each preparation tested, the number of standards and the range of concentrations were prepared in accordance with the guidelines for an accurate IC₅₀ estimation (Sebaugh, 2011). The antioxidant assay involved the addition of fifty microliters of sample (quercetin, BHT, or solvent) to a tube containing 200 µL of DPPH radical solution (0 or 460 µmol/L), incubated at 30 °C for 20 min. Once the incubation had been completed, 150 µL of each mixture was placed in a vial insert (part number 5181-1270, Agilent Technologies, Palo Alto, CA, USA) which was kept under constant lighting at a fixed distance to a mobile phone with an integrated eight-megapixel camera (Moto e5 play, Motorola Mobility LCC, IL, USA).

The Red Green Blue (RGB) color response and reflectance spectrum were monitored for each sample, using a colorimeter application (Colorimeter version 5.5.1, Research Lab Tools, Sao Paulo, Brazil, purchased from Google Play) installed on the mobile phone. For each experimental sample, a ΔRG value was calculated by subtracting the experimental value (sample plus 460 µmol/L DPPH radical) from a blank value (sample plus 0 µmol/L DPPH radical) with a percentage of the ΔRG value then obtained using the following equation (Ravindranath *et al.*, 2018):

$$\Delta RG(\%) = \left(\frac{\Delta RG}{255} \right) \times 100$$

Subsequently, the DPPH radical inhibition percentage was calculated using the ΔRG (%) values obtained from both a second blank value (sample solvent plus 460 µmol/L DPPH

radical) and the experimental sample. Finally, the concentration that achieved 50 % DPPH inhibition (IC_{50}) was calculated for each compound tested, using GraphPad Prism 5 software (San Diego, CA, USA). The antioxidant assay conducted in the present study was following the recommendations for the colorimetric quantifications on smartphones, which are rapid, simple, sensitive, low cost, and reliable analytical techniques (Ravindranath *et al.*, 2018; Fan *et al.*, 2021).

It should be noted that we were unable to find, in the literature, a DPPH radical scavenging assay that uses the approach set out in the present technique. Consequently, before the colorimetric assay was applied on the experimental samples, the assay was validated in accordance with FDA guidelines (US Department of Health and Human Services *et al.*, 2018), with quercetin used as a reference standard, given that it corresponded to a new assay based on a previous spectrophotometric methodology (Martinez-Morales *et al.*, 2020). Blank, lower limit of quantification, and between-day evaluations were performed over three consecutive days. In order to determine the stability percentage for each quality control sample, we compared the experimental value with that obtained immediately after preparation. Finally, the low, middle, and high-quality control samples presented concentrations of 23, 60, and 110 µg/mL quercetin, respectively.

Antioxidant interaction assessment

The experimental design for the constant ratio combinations followed was set out previously (Chou, 2006; Chou, 2010). Once the IC_{50} value of each compound was obtained via the DPPH radical scavenging assay, the BHT and quercetin (BQ) combination was assessed via the same assay using five different proportions (3:1, 1:1, 1:3, 1:5, and 1:10). The data for the individual compounds and combinations were evaluated using both the Chou-Talalay method, which is based on the median-effect equation, which, in turn, is derived from the mass-action law principle (Chou, 2006), and the isobole method described by Tallarida (2002).

The Chou-Talalay theory involves the quantitative definition of synergism, additivity, and antagonism by means of a combination index (CI) value and their visual definition by means of an isobologram (Chou, 2006). In the present study, a weighted average (wa) CI value was calculated for each combination using the following formula:

$$waCI = \frac{CI_{90} + (2 \times CI_{75}) + (3 \times CI_{50}) + (4 \times CI_{30})}{10}$$

The aim of this formula was to increase the relevance of low effect levels, as the preservatives are used at low concentrations in foodstuffs (Chou, 2006; USFDA, 2019; Checkmahomed *et al.*, 2020; Electronic Code of Federal Regulations, 2021). Therefore, a waCI value of < 0.1, 0.1-0.3, 0.3-0.7, 0.7-0.85, 0.85-0.90, 0.90-1.10, 1.10-1.20, 1.20-1.45, 1.45-3.3, 3.3-10, and > 10 indicated a very strong synergism, strong synergism, synergism, moderate synergism, slight synergism, nearly additive, slight antagonism, moderate antagonism, antagonism, strong antagonism, and very strong antagonism, respectively (Chou, 2006). If the data points fell

on the hypotenuse of the isobologram, an additive effect was determined, while, if data points fell on the lower or upper left of the hypotenuse, a synergism or antagonism was determined, respectively (Chou, 2006).

The isobole method involved the calculation of theoretical and experimental IC_{50} values (Zadd and Zexp) for each combination, values used to calculate an interaction index (γ) and to undertake a statistical comparison, where both procedures defined the type of interaction (Alonso-Castro *et al.*, 2017) as described in the section of data analysis.

Preparation of beef patties

After all the experiments described above were completed and analyzed, the utility of the compounds and their combination in the food preservation started. In this way, the meat preparation was as described previously (Gallego *et al.*, 2015; Cantú-Valdéz *et al.*, 2020). In brief, fresh beef pulp, purchased from a local supermarket, was immediately transported to the laboratory and ground in a meat grinder fitted with a 4.5-mm grind plate (Tartare, Metaltex International, Molsheim Cedex, France). Five formulations were prepared: C (meat without additive); V (meat with 20 mL/kg EVO added); B (meat with 100 mg/kg BHT added); Q (meat supplemented with 36 mg/kg quercetin); and, BQ (meat with 5.2 mg/kg of BHT and 26.0 mg/kg of quercetin added, giving a 1:5 ratio).

Prior to meat supplementation, quercetin, BHT, and combination, were dissolved in EVO (20 mL/kg of meat), with the ingredients then mixed manually in a steel bowl for 2 min. Subsequently, each preparation was mixed at low speed for 2 min in a mixer (Model 64650, Hamilton Beach Brands, Inc, VA, USA) and then a 23 g portion of each meat sample was made into a 7-cm in diameter and 1-cm in height burger. Beef patties were placed in a polyethylene bag and stored in a refrigerator at 4 °C under absence of light. Finally, the samples were analyzed for color, antioxidant capacity, metmyoglobin (MetMb) content, and thiobarbituric acid reactive substance (TBARS) value on day 0 (meat samples without addition of preservatives), 1, and 10. It is important to note that the recommended BHT concentration for meat products was used (Cantú-Valdéz *et al.*, 2020), while the concentrations for both quercetin and selected BQ combination were determined on the basis of their antioxidant IC_{50} values and interaction studies, where the used concentrations presented the same antioxidant activity found in the selected BHT concentration. In addition, the volume of the vehicle (EVO) was selected based on the quantity of oily substances used for preservative proposes in ground beef (Cantú-Valdéz *et al.*, 2020).

Color evaluation

The evaluation of beef patties color was as described by Gallego *et al.* (2015). At four different points, a colorimeter (Color Muse, Variable Inc., TN, USA) was directly placed on the surface of the beef patty in areas without fat or connective tissue. For each point, the L^* , a^* , and b^* values pertaining to the CIE Lab color system were recorded to measure lightness,

redness, and yellowness, respectively. Prior to the measurements recording, the calibration of the colorimeter was using a standard white plate provided, along with the instrument, by the manufacturer. The mean value for each CIE Lab parameter was calculated for each sample.

Antioxidant capacity

A beef patty sample (0.20 to 0.58 g) was placed in a tube containing 500 µL ethanol-DMSO (96:4 v/v), mixed for 1 min, and centrifuged at 1500 xg for 10 min at 4 °C. Subsequently, the supernatant (50 µL) was processed using the DPPH radical scavenging assay as described in the section above. For these measurements, the data was expressed as the percentage radical inhibition/200 mg of meat. For some samples, a weight of > 200 mg was necessary in order to obtain a response within the analytical range of the assay. As the methodology for the radical inhibition assay had been slightly modified for these samples, a partial validation was also performed (data not shown) in accordance with FDA guidelines (US Department of Health and Human Services *et al.*, 2018).

Metmyoglobin (MetMb) content

The quantification of MetMb was according to Mtibaa *et al.* (2019). Briefly, 0.2 g of beef patty was placed in a tube containing 1 mL of cold phosphate buffer solution (40 mM at pH 6.8) and mixed for 1 min, the mixture kept at 4 °C for 1 h and centrifuged at 4500 xg for 30 min at 4 °C. Each sample supernatant was filtered through a 0.45-µm-pore-size filter unit (Millipore Corporation, Bedford, MA, USA), after which 200 µL of the filtrate was placed into one well of a 96-well plate (Costar® 3595, Corning Incorporated, NY, USA). The absorbance was recorded at 572, 565, 545, and 525 nm using a Cytation™ 3 microplate reader and Gen5™ software (Biotek Instruments Inc., Vermont, USA). For each sample, the MetMb percentage determination was using the following formula (Mtibaa *et al.*, 2019):

$$\text{MetMb (\%)} = \left[\left(-2.51 \times \frac{A_{572}}{A_{525}} \right) + \left(0.777 \times \frac{A_{565}}{A_{525}} \right) + \left(0.8 \times \frac{A_{545}}{A_{525}} \right) + 1.098 \right] \times 100$$

In the formula, the term A alongside a number denotes the absorbance and wavelength.

Thiobarbituric acid reactive substance (TBARS) value

The determination of TBARS was according to Eymard *et al.* (2005) and Mtibaa *et al.* (2019). A sample of the ground beef patty (0.12 to 0.18 g) was placed in a tube containing 1 mL of 50 g/L trichloroacetic acid and 6 µL of 1 g/L BHT and then mixed for 1 min. The mixture was centrifuged at 1900 xg for 10 min at 18 °C, with the supernatant (250 µL) then added to 150 µL of 0.8 % thiobarbituric acid. This second mixture was incubated at 75 °C for 30 min and kept at 4 °C for 3 min. Two hundred microliters of each mixture were placed in one well of a 96-well plate (Costar® 3595, Corning Incorporated, NY, USA). The absorbance was read at 532 nm using the Cytation™ 3 microplate reader and Gen5™ software (Biotek Instruments Inc., Vermont, USA). The TBARS value

was calculated using a calibration curve of malondialdehyde with a 239 µg/mL malondialdehyde stock solution and the results were expressed as µg TBARS/g of meat. For this assay, the malondialdehyde was obtained from the degradation of 1,1,3,3-tetraethoxypropane, which was induced by their exposition to 0.1 N hydrochloric acid under a boiling water bath.

Data analysis

The antioxidant IC₅₀ values of the BHT and quercetin were analyzed using an unpaired t test, using the GraphPad Prism 5 software (San Diego, CA, USA). For the combination analyses using the Chou Talalay theory, the conformity of the data with the mass-action law was evaluated using the linear correlation coefficient (r) of the median effect plot, where an r value > 0.95 was considered an acceptable conformity (Chou, 2006). All these analyses and isobolograms were performed using the Compusyn software (Chou and Martin, 2005). Data with effects of ≥ 1.0 were not included in the analyses because the Compusyn software cannot compute values with this level.

For the isobole method, the statistical comparison of the Zadd and Zexp values alongside the γ index defined the interaction as a synergism (γ index < 1.0 and P < 0.05), additivity (γ index close to 1.0 and P > 0.05), or antagonism (γ index > 1.0 and P < 0.05) (Alonso-Castro *et al.*, 2017). These statistical analyses were performed using the GraphPad Prism 5 software (San Diego, CA, USA).

The presentation of color, antioxidant capacity, MetMb, and TBARS data are as both the mean and standard deviation. The analysis of C, V, B, Q, and BQ group values were using a two-way ANOVA with a Bonferroni post-test, using the GraphPad Prism 5 software (San Diego, CA, USA). For all the statistical analyses, a P value of < 0.05 was considered statistically significant.

RESULTS AND DISCUSSION

Antioxidant activity

Calculated via the GraphPad or Compusyn software, the respective antioxidant IC₅₀ values for quercetin and BHT were 89.7 ± 4.3 and 171.7 ± 8.2 µg/mL (nine points per curve, N= 5) or 50.2 and 139.4 µg/mL (eight points per curve, N= 5), respectively. Our evaluations showed that quercetin presents a higher antioxidant activity level than BHT (P < 0.0001), a finding also observed in previous individual evaluations of the action of both compounds against the DPPH radical (Popovic-Milenkovic *et al.*, 2014; Singh *et al.*, 2018). However, the IC₅₀ values obtained in the present study were higher than those reported previously (Popovic-Milenkovic *et al.*, 2014; Singh *et al.*, 2018), a difference due to some technical conditions, such as a lower DPPH concentration, different reaction medium, and the absence of a validation procedure for the antioxidant assay (Martinez-Morales *et al.*, 2020), unlike that used in our study. Given the use of a new antioxidant assay, Table 1 shows the results obtained from the analytical validation, which measured the quercetin antioxi-

Table 1. Complete validation of the DPPH radical inhibition (%) assay with quercetin**Table 1.** Validación completa del ensayo de la inhibición del radical DPPH con quercetina

Parameter	Value
Calibration curve (N= 6)	
Range (µg/mL quercetin)	12-125
Calibration model	Second order polynomial relationship
R ² value	0.9954 ± 0.0032
Accuracy (%) of calibrators	99.0 ± 8.2
LLOQ (12 µg/mL, N= 6)	
Accuracy (%)	98.3 ± 16.4
Precision (%CV)	16.7
Blank sample (0 µg/mL quercetin plus 450 µmol/L DPPH radical, N= 6)	
Accuracy (%)	The value is not calculable
Precision (%CV) of the ΔRG (%) values	11.5
Accuracy of QC samples (% , N= 6)	
Within day evaluation	
Low QC	92.8 ± 12.3
Middle QC	88.7 ± 2.9
High QC	102.3 ± 3.7
Between day evaluation	
Low QC	95.7 ± 10.3
Middle QC	89.7 ± 4.0
High QC	99.7 ± 4.9
Precision of QC samples (%CV, N= 6)	
Within day evaluation	
Low QC	13.4
Middle QC	3.6
High QC	3.6
Between day evaluation	
Low QC	14.2
Middle QC	6.1
High QC	4.7
Stability of QC samples under different conditions (% , N= 5)	
At room temperature (45 min)	
Low QC	97.3 ± 2.5
High QC	89.0 ± 3.7
On the vial insert (15 min)	
Low QC	97.6 ± 1.7
High QC	95.5 ± 7.8
Storage at -18 °C (24 h)	
Low QC	87.2 ± 0.8
High QC	53.4 ± 4.1

Each value is the mean ± standard deviation or percentage of coefficient of variation (%CV). DPPH: 2,2'-diphenyl-1-picrylhydrazyl; LLOQ: lower limit of quantification; QC: quality control.

dant activity against the DPPH radical. In order to complete the validation process, we carried out identification tests with the reflectance spectra of the samples, modifying the spectra record by means of a color change (Figure 1). All the validation and identification tests results were within the acceptability criteria stipulated by the FDA and International Conference on Harmonization (ICH) guidelines (ICH, 2005; US Department of Health and Human Services *et al.*, 2018), except the stability results for the high QC samples stored at -18 °C for 24 h. For this reason, the preparation of stock, standard solutions and experimental samples were on the same day they of the assays. For quantitative determinations, the application of the guidelines described by the FDA and ICH guarantees the reliability and strict control on the results, as well as the suitability, accuracy, and reproducibility of the assay (Gonzalez-Rivera *et al.*, 2019).

Antioxidant interaction outcomes

To our knowledge, there are no previous reports presenting an evaluation of the antioxidant interactions between BHT and quercetin. The antioxidant IC₅₀ values for the 3:1, 1:1, 1:3, 1:5, and 1:10 BQ combinations were 97.1, 83.4, 63.1, 43.7, and 50.2 µg/mL, respectively (six points per curve, N= 5). Table 2 and Figure 2 show the data and isobolograms

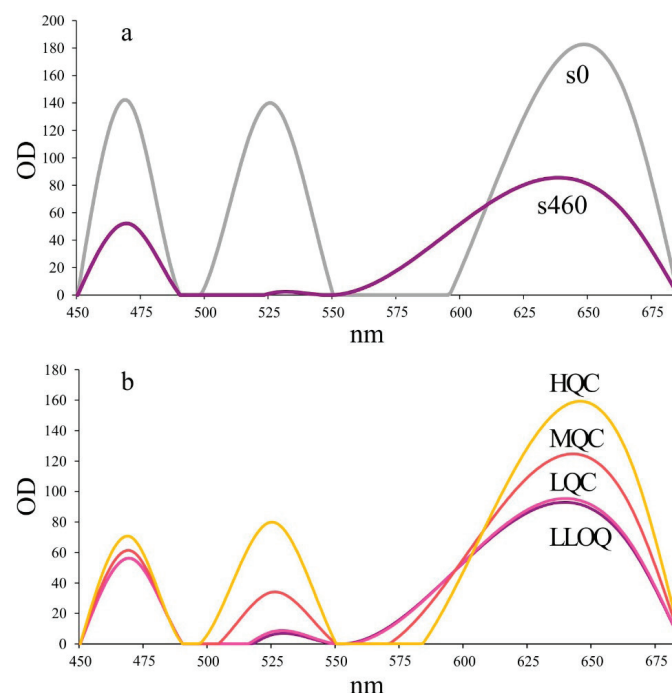


Figure 1. Representative reflectance spectra for the two blank samples (a): sample solvent plus 0 µmol/L DPPH radical (s0) and sample solvent plus 460 µmol/L DPPH radical (s460). Furthermore, the spectra records for the LLOQ and lower (L), middle (M) and high (H) QC samples are shown (b). DPPH: 2,2'-diphenyl-1-picrylhydrazyl; LLOQ: lower limit of quantification; QC: quality control.

Figura 1. Espectros de reflectancia representativos para las dos muestras de blancos (a): solvente de la muestra más radical DPPH 0 µmol/L (s0) y solvente de la muestra más radical DPPH 460 µmol/L (s460). Además, los registros de los espectros para el LLOQ y QC bajo (L), medio (M) y alto (H) son mostrados (b). DPPH: 2,2'-difeníl-1-picrilhidrazilo; LLOQ: límite más bajo de cuantificación; QC: control de calidad.

obtained from the interaction experiments conducted on the compounds. Our experiments revealed that the 1:5 BQ combination was the only combination that produced a synergistic interaction at the three levels of effects (30, 50, and 90 %). This interaction was determined using the Chou-Talalay and isobole methods, as well as a moderate level of synergy established by the Chou-Talalay theory. Consequently, we used this ratio for the utility evaluation of the BQ combination for beef patties preservation. On the other hand, the 3:1, 1:1, 1:3, and 1:10 BQ combinations presented, respectively, a nearly additive, slight antagonistic, nearly additive, and nearly additive effect, according to the Chou-Talalay method, while an antagonistic, additive, additive, and additive effect presented for the same combinations, respectively, according to the isobole method (Table 2 and Figure 2). To confirm our results, we used a second recognized methodology, such as the isobole method, since an unacceptable conformity of the data with the mass-action law was evident in the majority of the combinations when applying the Chou-Talalay approach, with the interpretation of both methods showing minor discrepancies.

Shelf-life of beef patties

Before discussing the effects of the combination selected on food evaluations, it is important to note that the 1:5 BQ combination contains much lower amounts of BHT and quercetin than the individual concentration of each compound, due to the synergism found in this combination. The meat quality parameters evaluated in beef patties at day

Tabla 2. Evaluación de las interacciones antioxidantes entre BHT y quercetina (BQ).

Table 2. Evaluation of the antioxidant interactions between BHT and quercetin (BQ).

Sample	Chou-Talalay method		Isobole method			
	waCl	r value	Zadd (µg/mL)	Zexp (µg/mL)	γ value	P value
BQ 3:1	1.001	0.9361	111.5 ± 2.4	122.0 ± 6.3	1.091	0.0081
BQ 1:1	1.125	0.9479	90.9 ± 3.8	95.2 ± 2.8	1.045	0.0761
BQ 1:3	1.051	0.9555	76.9 ± 4.6	73.9 ± 4.0	0.959	0.3064
BQ 1:5	0.774	0.9007	73.1 ± 4.8	64.0 ± 3.2	0.874	0.0075
BQ 1:10	0.936	0.9032	69.9 ± 4.9	70.4 ± 2.0	1.006	0.8571

For the Chou-Talalay or isobole method, BHT and quercetin concentrations ranged from 7.6 to 967.2 and 3.1 to 397.8 µg/mL (eight points per compound, N= 5) or from 7.6 to 1488.0 and 3.1 to 612.0 µg/mL (nine points per compound, N= 5), respectively. For both methods, a range of concentrations from 14.8 to 472.8, 8.3 to 267.0, 7.9 to 252.4, 6.7 to 215.6, and 6.1 to 196.7 µg/mL were used for the 3:1, 1:1, 1:3, 1:5, and 1:10 combinations (six points per combination, N= 5), respectively. For each comparison between the Zadd and Zexp value, their P value is shown in the latest column of the table. γ: interaction index; BHT: butylated hydroxytoluene; waCl: weighted average combination index.

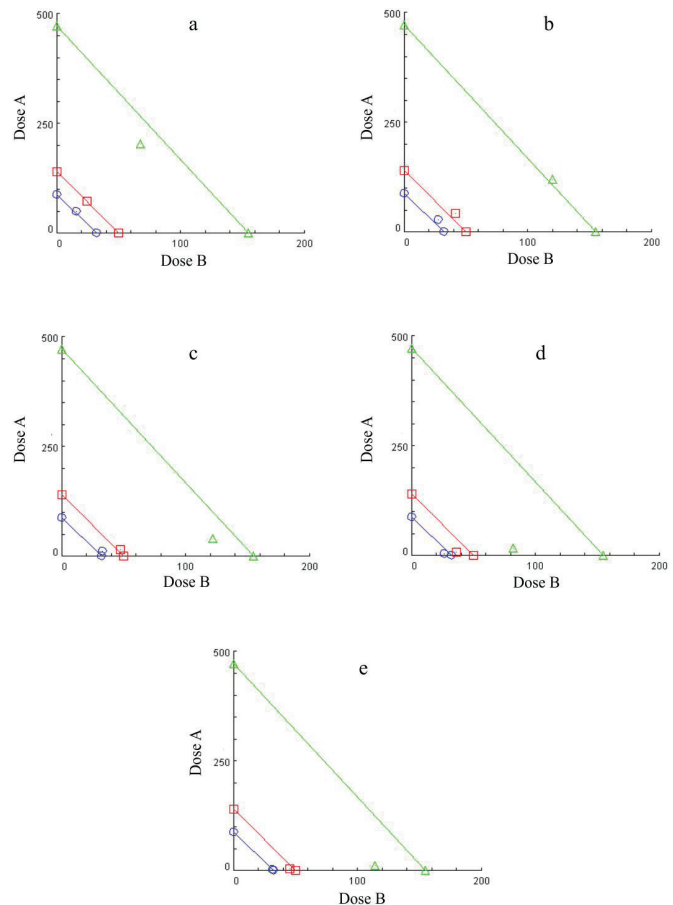


Figure 2. Isobolograms for the 30 % (blue), 50 % (red), and 90 % (green) free radical inhibition induced by the butylated hydroxytoluene and quercetin combination at 3:1 (a), 1:1 (b), 1:3 (c), 1:5 (d), and 1:10 (e). A: butylated hydroxytoluene; B: quercetin.

Figura 2. Isobogramas para el 30 % (azul), 50 % (rojo) y 90 % (verde) de inhibición del radical libre inducido por la combinación de hidroxitolueno butilado y quercetina 3:1 (a), 1:1 (b), 1:3 (c), 1:5 (d) y 1:10 (e). A: hidroxitolueno butilado; B: quercetina.

0 and 1 did not show significant changes between them ($P > 0.05$, Figure 3 and 4). Meanwhile, a decay of the parameters in beef patties was observed at day 10, in comparison with the data obtained at day 0 and 1 ($P < 0.05$, Figure 3 and 4), which was produced by the quality deterioration in meat products during their storage (Mtibaa *et al.*, 2019). For the present experiments, the period of refrigeration was close to the shelf-life required to distribute the meat products to retail outlets (Kapetanakou *et al.*, 2020). However, three parameters did not show a significant deterioration through the time as described in detail below.

Color improvement

The lightness, redness, and yellowness of the meat decrease as refrigeration time continues (Gallego *et al.*, 2015). While after ten days of storage at 4 °C, the lightness of the beef patty showed no modification by any supplementation ($P > 0.05$ for all groups at any time, N= 7, Figure 3a), the redness and yellowness of the meat added with the BHT, quercetin, or BQ combination was more intensive than that

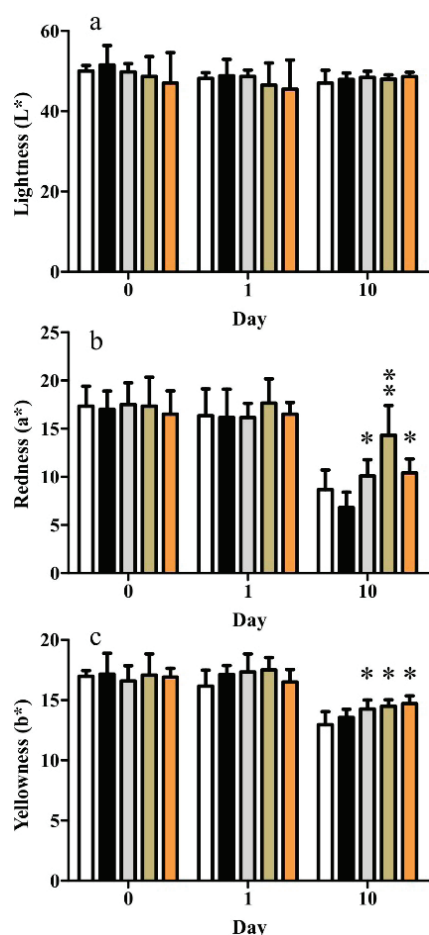


Figure 3. Lightness (a), redness (b), and yellowness (c) of the following groups studied: non-added meat (white bar) or added with the vehicle (black bar), butylated hydroxytoluene (gray bar), quercetin (brown bar), or combination (orange bar). *P value of < 0.05 versus V for redness and < 0.05 versus C for yellowness. **P value of < 0.001 versus C and V, and < 0.05 versus B and BQ.

Figura 3. Luminosidad (a) y abundancia del color rojo (b) y amarillo (c) de los siguientes grupos estudiados: carne no adicionada (barra blanca) o adicionada con el vehículo (barra negra), hidroxitolueno butilado (barra gris), quercetina (barra café) o combinación (barra anaranjada). *Valor de P de < 0.05 versus V para abundancia del color rojo y < 0.05 versus C para abundancia del color amarillo. **Valor de P de < 0.001 versus C y V, y < 0.05 versus B y BQ.

found in the non-supplemented patty or that treated with vehicle alone (N= 7, Figure 3b and c). Moreover, the quercetin produced more intense redness than the other supplements and prevented the deterioration in redness through time (P > 0.05 for Q groups on day 0, 1, and 10).

Redness is the most important color parameter for the evaluation of meat oxidation and a key predictor for the acceptability of a meat product to consumers (Gallego *et al.*, 2015). In line with our data, information presented by previous studies shows that BHT added to meat is capable of producing either higher L*, a*, and b* values, or solely higher a* values, than those observed in non-supplemented ground beef, when stored at 4 °C for seven or eight days (Kim *et al.*, 2013; Cantú-Valdéz *et al.*, 2020). Similarly, quercetin is able to produce improved redness and yellowness in beef patties,

unlike those made from non-supplemented meat, when stored at 2 °C for nine days (Bekhit *et al.*, 2003).

Antioxidant ability and inhibition of the protein and lipid oxidation

The meat added with the individual or combined compounds, showed a significant antioxidant capacity and lower MetMb and TBARS content than those values obtained from the non-supplemented meats, or those treated with vehicle solely after ten days of refrigeration (N= 7, Figure 4). However, the meat treated with vehicle also presented a beneficial effect against the presence of TBARS.

The undesirable discoloration of meat during storage is largely due to myoglobin oxidation and MetMb formation (Mtibaa *et al.*, 2019). Moreover, the percentage of MetMb is

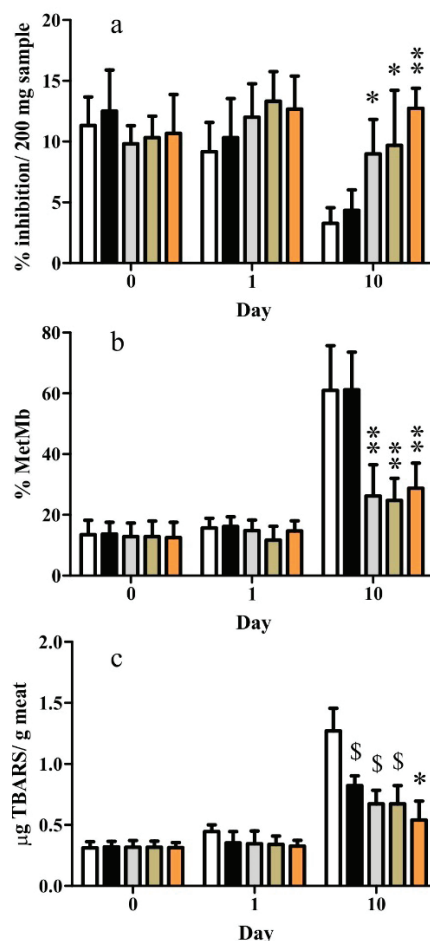


Figure 4. Antioxidant capacity (a), metmyoglobin (MetMb) content (b), and thiobarbituric acid reactive substances (TBARS) (c), found in samples obtained from the non-added meat (white bar) or added with the vehicle (black bar), butylated hydroxytoluene (gray bar), quercetin (brown bar), or the combination selected (orange bar). [§]P value of < 0.001 versus C. *P value of < 0.001 versus C and < 0.05 versus V. **P value of < 0.001 versus C and V.

Figura 4. Capacidad antioxidante (a), contenido de metamioglobina (MetMb) (b) y sustancias reactivas al ácido tiobarbitúrico (c) encontrados en las muestras obtenidas de la carne no adicionada (barra blanca) o adicionada con el vehículo (barra negra), hidroxitolueno butilado (barra gris), quercetina (barra café) o combinación seleccionada (barra anaranjada). [§]Valor de P de < 0.001 versus C. *Valor de P de < 0.001 versus C y < 0.05 versus V. **Valor de P de < 0.001 versus C y V.

a significant variable given that consumers reject meat products with levels higher than 40 % (Mtibaa *et al.*, 2019). The 1:5 BQ combination produced a level of < 40 % MetMb in the ground beef patties, as did the BHT or quercetin treatments after ten days of storage (Figure 4b). As quercetin and BHT are free radical scavengers, they directly inhibited protein oxidation due to their antioxidant properties (Mtibaa *et al.*, 2019). Previous studies have shown that BHT or quercetin maintain lower percentages of MetMb in beef than those observed in non-supplemented meat for up to ten or nine days of refrigeration, respectively. However, the effect of the quercetin was reversed after the ninth day of storage (Bekhit *et al.*, 2003; Mtibaa *et al.*, 2019), due to the method used to add quercetin to the meat, thus influencing the performance of this antioxidant (Bekhit *et al.*, 2003; Bekhit *et al.*, 2004).

Hydrogen atom transfer and single electron transfer reactions, which are the most important pathways via which an antioxidant can scavenge free radicals, are solvent and pH dependent, and in this case, our solvent for quercetin was different to the study of Borgohain *et al.* (2015). The foregoing study prepared quercetin in deionized water with an adjusted pH of 6.8, adding it at a ratio of 111.1 mL/kg of meat (Bekhit *et al.*, 2003), while the present study prepared quercetin in food grade EVO, adding it at a ratio of 20 mL/kg meat. Thus, the food grade EVO was an appropriate medium for the antioxidant function of quercetin, and its use as a solvent was to obtain a homogeneous suspension of quercetin, as it is insoluble and precipitates rapidly in water at the concentration and volume used in the present study.

Because of the radical scavenger effects of the compounds tested, the meat antioxidant capacity presented by the 1:5 BQ combination was superior to that found in the non-supplemented meat, where this variable closely resembled the capacity produced by each compound when administered independently at high concentrations (Figure 4a). Moreover, the presence of BHT, quercetin or combination avoided the deterioration of the antioxidant ability in beef patties through time ($P > 0.05$ for B, Q and BQ groups on day 0, 1 and 10). A previous study found that BHT administered at 100 mg/kg did not produce a high antioxidant capacity in the ground meat, compared to that observed in the non-supplemented meat, after seven days of storage at 4 °C (Cantú-Valdéz *et al.*, 2020). However, said study added BHT in a dry form to the meat. As explained above, for the quercetin treated samples, the EVO (solvent) was also an appropriate medium for the performance of the BHT antioxidant activity (Borgohain *et al.*, 2015). We were unable to find, in the literature, a report of an evaluation of the antioxidant capacity of ground meat or patties supplemented with quercetin. Our data shows an increase in the capacity of the meat to resist oxidative processes by the BQ mixture, a finding confirmed by our protein oxidation evaluation (MetMb content).

The TBARS results obtained supports the protection provided to ground beef patties by the 1:5 BQ combination against lipid oxidation (Figure 4c). TBARS levels increase during storage, since they are reactive aldehydes produced

by lipid peroxidation of polyunsaturated fatty acids in meat (Mtibaa *et al.*, 2019). The vehicle used in the present study, EVO, played a role in the reduction of TBARS levels in the B, Q, and BQ samples (Figure 4c), which may be due to the EVO antioxidant properties (Borges *et al.*, 2019), although the meat antioxidant capacity induced via EVO was not found to be significant (Figure 4a). Nevertheless, the antioxidant synergism of the mixture tested (BQ group) produced a greater reduction of TBARS than that observed for the vehicle (Figure 4c). Prior data shows that the addition of BHT reduces or does not affect TBARS content in meat stored at 4 °C for seven-eight days (Gallego *et al.*, 2015; Mtibaa *et al.*, 2019). Also, quercetin was not observed to significantly modify TBARS levels in meat over nine days of storage at 2 °C (Bekhit *et al.*, 2003), findings similar to those observed for the non-supplemented meats.

Limitation of the study

Despite in the present study we evaluated important meat quality parameters for beef patties, including key predictors for the acceptability of a meat product by consumers, such as the redness and MetMb levels (Gallego *et al.*, 2015; Mtibaa *et al.*, 2019), it is clear that further studies are necessary to increase the actual evidence, such as sensory evaluations. Nevertheless, the present study is the beginning for the use of synergistic combinations of antioxidants to preserve meat products.

CONCLUSIONS

For the first time, our study shows that the use of a 1:5 BQ combination (5.2 mg/kg of BHT plus 26.0 mg/kg of quercetin) was an effective, simple, and available method for the preservation of beef patties. Since this strategy produced a synergistic antioxidant interaction between BHT and quercetin, it in turn improved color, levels of protein and lipid oxidation, and antioxidant capacity of the beef patties. These beneficial changes were possible to achieve with a simple combination of these preservatives that are available in the actual market and consequently, the individual use of high concentrations of BHT and quercetin by the meat industry can be substitute for their synergistic combination.

ACKNOWLEDGEMENTS

Authors thank to Othir G. Galicia-Cruz for their technical assistance. This study was financially supported by the economical stimuli to researchers acquired from the Sistema Nacional de Investigadores (SNI) - Consejo Nacional de Ciencia y Tecnología (CONACyT).

CONFLICT OF INTEREST

The authors declare that there is no conflict of interest.

REFERENCES

- Alonso-Castro, A.J., Rangel-Velázquez, J.E., Isiordia-Espinoza, M.A., Villanueva-Solís, L.E., Aragon-Martinez, O.H. y Zapata-

- Morales, J.R. 2017. Synergism between naproxen and rutin in a mouse model of visceral pain. *Drug Development Research*. 78: 184-188. <https://doi.org/10.1002/ddr.21391>
- Andres, S., Pevny, S., Ziegenhagen, R., Bakhiya, N., Schäfer, B., Hirsch-Ernst, K.I. y Lampen, A. 2018. Safety aspects of the use of quercetin as a dietary supplement. *Molecular Nutrition & Food Research*. 62. <https://doi.org/10.1002/mnfr.201700447>
- Bekhit, A.E.D., Geesink, G.H., Ilian, M.A., Morton, J.D. y Bickerstaffe, R. 2003. The effects of natural antioxidants on oxidative processes and metmyoglobin reducing activity in beef patties. *Food Chemistry*. 81: 175-187. [https://doi.org/10.1016/S0308-8146\(02\)00410-7](https://doi.org/10.1016/S0308-8146(02)00410-7)
- Bekhit, A.E.D., Geesink, G.H., Ilian, M.A., Morton, J.D., Sedcole, J.R. y Bickerstaffe, R. 2004. Pro-oxidant activities of carnosine, rutin and quercetin in a beef model system and their effects on the metmyoglobin-reducing activity. *European Food Research and Technology*. 218: 507-514. <https://doi.org/10.1007/s00217-004-0904-7>
- Borges, T.H., Serna, A., López, L.C., Lara, L., Nieto, R. y Seiquer, I. 2019. Composition and antioxidant properties of spanish extra virgin olive oil regarding cultivar, harvest year and crop stage. *Antioxidants (Basel)*. 8: 217. <https://doi.org/10.3390/antiox8070217>
- Borghain, R., Guha, A.K., Pratihari, S. y Handique, J.G. 2015. Antioxidant activity of some phenolic aldehydes and their diimine derivatives: A DFT study. *Computational and Theoretical Chemistry*. 1060: 17-23. <https://doi.org/10.1016/j.comptc.2015.02.014>
- Cantú-Valdéz, J.A., Gutiérrez-Soto, G., Hernández-Martínez, C.A., Sinagawa-García, S.R., Quintero-Ramos, A., Hume, M.E., Herrera-Balandrano, D.D. y Méndez-Zamora, G. 2020. Mexican oregano essential oils as alternatives to butylated hydroxytoluene to improve the shelf life of ground beef. *Food Science & Nutrition*. 8: 4555-4564. <https://doi.org/10.1002/fsn3.1767>
- Checkmahomed, L., Paday, B., Pizzorno, A., Terrier, O., Rosa-Calatrava, M., Abed, Y., Baz, M. y Boivin, G. 2020. In vitro combinations of baloxavir acid and other inhibitors against seasonal influenza A viruses. *Viruses*. 12: 1139. <https://doi.org/10.3390/v12101139>
- Chou, T.C. 2010. Drug combination studies and their synergy quantification using the Chou-Talalay method. *Cancer Research*. 70: 440-446. DOI: 10.1158/0008-5472.CAN-09-1947
- Chou, T.C. 2006. Theoretical basis, experimental design, and computerized simulation of synergism and antagonism in drug combination studies. *Pharmacological Reviews*. 58: 621-681. DOI: 10.1124/pr.58.3.10
- Chou, T.C. y Martin, N. 2005. CompuSyn for drug combinations: PC software and user's guide: a computer program for quantitation of synergism and antagonism in drug combinations, and the determination of IC50 and ED50 and LD50 values. *ComboSyn*. Paramus, NJ.
- Eymard, S., Carcouët, E., Rochet, M.J., Dumay, J., Chopin, C. y Genot, C. 2005. Development of lipid oxidation during manufacturing of horse mackerel surimi. *Journal of the Science of Food and Agriculture*. 85: 1750-1756. <https://doi.org/10.1002/jsfa.2145>
- Fan, Y., Li, J., Guo, Y., Xie, L. y Zhang, G. 2021. Digital image colorimetry on smartphone for chemical analysis: A review. *Measurement*. 171, 108829. <https://doi.org/10.1016/j.measurement.2020.108829>
- Gallego, M.G., Gordon, M.H., Segovia, F.J. y Almajano, M.P. 2015. Caesalpinia decapetala extracts as inhibitors of lipid oxidation in beef patties. *Molecules*. 20: 13913-13926. <https://doi.org/10.3390/molecules200813913>
- Gonzalez-Rivera, M.L., Martinez-Morales, F., Alonso-Castro, A.J., López-Rodríguez, J.F., Aranda Romo, S., Zapata-Morales, J.R. y Aragon-Martinez, O.H. 2019. Matrix effect evaluation and validation of the 2,2'-azino-bis (3-ethylbenzothiazoline-6-sulfonic acid) radical cation scavenging assay, as well as its application using a tejate, an ancient beverage in Mexico. *Chemical Papers*. 73: 2767-2781. <https://doi.org/10.1007/s11696-019-00829-3>
- International Conference on Harmonization. 2005. International conference on harmonization of technical requirements for the registration of pharmaceuticals for human use, validation of analytical procedures: text and methodology Q2(R1). Geneva, Switzerland.
- Kapetanakou, A.E., Pateraki, G.L. y Skandamis, P.N. 2020. Developing a commercial antimicrobial active packaging system of ground beef based on "tsipouro" alcoholic distillate. *Foods*. 9: 1171. <https://doi.org/10.3390/foods9091171>
- Kim, S.J., Min, S.C., Shin, H.J., Lee, Y.J., Cho, A.R., Kim, S.Y. y Han, J. 2013. Evaluation of the antioxidant activities and nutritional properties of ten edible plant extracts and their application to fresh ground beef. *Meat Science*. 93: 715-722. <https://doi.org/10.1016/j.meatsci.2012.11.029>
- Liu, R. y Mabury, S.A. 2020. Synthetic phenolic antioxidants: a review of environmental occurrence, fate, human exposure, and toxicity. *Environmental Science & Technology*. 54: 11706-11719. <https://doi.org/10.1021/acs.est.0c05077>
- Martinez-Morales, F., Alonso-Castro, A.J., Zapata-Morales, J.R., Carranza-Álvarez, C. y Aragon-Martinez, O.H. 2020. Use of standardized units for a correct interpretation of IC50 values obtained from the inhibition of the DPPH radical by natural antioxidants. *Chemical Papers*. 74: 3325-3334. <https://doi.org/10.1007/s11696-020-01161-x>
- Mtibaa, A.C., Smaoui, S., Ben Hlima, H., Sellem, I., Ennouri, K. y Mellouli, L. 2019. Enterocin BacFL31 from a safety Enterococcus faecium FL31: natural preservative agent used alone and in combination with aqueous peel onion (*Allium cepa*) extract in ground beef meat storage. *BioMed Research International*. 2019: 4094890. <https://doi.org/10.1155/2019/4094890>
- Electronic Code of Federal Regulations. Title 21, Chapter I, Subchapter B, Part 182, Subpart D, Sec. 182.3173. Butylated hydroxytoluene. [Consultado 12 Mayo 2021] 2021. Disponible en: https://www.ecfr.gov/cgi-bin/text-idx?SID=a271652c302a4d8fb472e9e03ab4c34f&mc=true&node=pt21.3.182&rgn=div5#_top.
- National Institute of Diabetes and Digestive and Kidney Diseases (NIDDKD). 2012. Quercetin. En: *LiverTox: Clinical and research information on drug-induced liver injury* [Internet]. NIDDKD (ed.), updated 28 Mar 2020, Bethesda, MD. [Consultado 12 Mayo 2021] Disponible en: <https://www.ncbi.nlm.nih.gov/books/NBK556474/>.
- Querfelli, M., Villasante, J., Ben Kaâb, L.B. y Almajano, M. 2019. Effect of neem (*Azadirachta indica* L.) on lipid oxidation in raw chilled beef patties. *Antioxidants (Basel)*. 8: 305. <https://doi.org/10.3390/antiox8080305>
- Popovic-Milenkovic, M.T., Tomovic, M.T., Brankovic, S.R., Lujic, B.T. y Jankovic, S.M. 2014. Antioxidant and anxiolytic

- activities of *Crataegus nigra* Wald. et Kit. berries. *Acta Poloniae Pharmaceutica*. 71: 279-85.
- Ravindranath, R., Periasamy, A.P., Roy, P., Chen, Y.W. y Chang, H.T. 2018. Smart app-based on-field colorimetric quantification of mercury via analyte-induced enhancement of the photocatalytic activity of TiO₂-Au nanospheres. *Analytical and Bioanalytical Chemistry*. 410: 4555-4564. <https://doi.org/10.1007/s00216-018-1114-7>
- Sebaugh, J.L. 2011. Guidelines for accurate EC₅₀/IC₅₀ estimation. *Pharmaceutical Statistics*. 10: 128-134. <https://doi.org/10.1002/pst.426>
- Singh, D.P., Verma, S. y Prabha, R. 2018. Investigations on antioxidant potential of phenolic acids and flavonoids: the common phytochemical ingredients in plants. *Journal of Plant Biochemistry & Physiology*. 6: 1000219. DOI: 10.4172/2329-9029.1000219
- Tallarida, R.J. 2002. The interaction index: a measure of drug synergism. *Pain*. 98: 163-168. DOI: 10.1016/s0304-3959(02)00041-6
- US Department of Health and Human Services, Food and Drug Administration, Center for Drug Evaluation and Research y Center for Veterinary Medicine. 2018. Bioanalytical method validation: guidance for industry. Maryland, United States of America. [Consultado 12 Mayo 2021] Disponible en: <https://www.fda.gov/media/70858/download>.
- US Food and Drug Administration (FDA). 2019. Food additive status list. [Consultado 12 Mayo 2021] Disponible en: www.fda.gov/food/food-additives-petitions/food-additive-status-list#ftnB.
- Wang, W. y Kannan, K. 2019. Quantitative identification of and exposure to synthetic phenolic antioxidants, including butylated hydroxytoluene, in urine. *Environment International*. 128: 24-29. <https://doi.org/10.1016/j.envint.2019.04.028>
- Weinroth, M.D., Britton, B.C., McCullough, K.R., Martin, J.N., Geornaras, I., Knight, R., Belk, K.E. y Metcalf, J.L. 2019. Ground beef microbiome changes with antimicrobial decontamination interventions and product storage. *PLoS One*. 14: e0217947. <https://doi.org/10.1371/journal.pone.0217947>
- Zahid, M.A., Seo, J.K., Parvin, R., Ko, J. y Yang, H.S. 2019. Comparison of butylated hydroxytoluene, ascorbic acid, and clove extract as antioxidants in fresh beef patties at refrigerated storage. *Food Science of Animal Resources*. 39: 768-779. <https://doi.org/10.5851/kosfa.2019.e67>
- Zapata-Morales, J.R., Alonso-Castro, A.J., Muñoz-Martínez, G.S., Martínez-Rodríguez, M.M., Nambo-Arcos, M.E., Brennan-Bourdon, L.M., Aragón-Martínez, O.H. y Martínez-Morales, J.F. 2021. In vitro and in vivo synergistic interactions between the flavonoid rutin with paracetamol and non-steroidal anti-inflammatory drugs. *Archives of Medical Research*. S0188-4409(21)00078-3. <https://doi.org/10.1016/j.arcmed.2021.03.007>