

Revista Latinoamericana de Hipertensión

ISSN: 1856-4550

latinoamerican adehipertension @gmail.com

Sociedad Latinoamericana de Hipertensión

Organismo Internacional

Bermúdez, Valmore; Bermúdez, Fernando; Acosta, Guillermo; Acosta, Alejandro; Añez, Johnny; Andara, Carla; Leal, Elliuz; Cano, Clímaco; Velasco, Manuel; Hernández, Rafael; Zafar, Israili Molecular mechanisms of Endothelial Dysfunction: from the nitric oxide synthesis to ADMA inhibition Revista Latinoamericana de Hipertensión, vol. 2, núm. 3, mayo-junio, 2007, pp. 84-88

Sociedad Latinoamericana de Hipertensión

Caracas, Organismo Internacional

Available in: http://www.redalyc.org/articulo.oa?id=170216980004



Complete issue

More information about this article

Journal's homepage in redalyc.org



olecular mechanisms of Endothelial Dysfunction: from the nitric oxide synthesis to ADMA inhibition

Valmore Bermúdez. MD; PhD1., Fernando Bermúdez. MD; PhD1., Guillermo Acosta. MD1. , Alejandro Acosta. MSc1., Johnny Añez. MSc1, Carla Andara. MSc1., Elliuz Leal. MD1., Clímaco Cano. PharmD1, Velasco Manuel. MD, PhD2, Rafael Hernández. MD, PhD3, Zafar Israili. PhD4

1: Endocrine and Metabolic Diseases Research Center. University of Zulia. School of Medicine. Maracaibo, Venezuela.
2: Clinical Pharmacology Unit, Vargas Medical School, Central University of Venezuela, Caracas, Venezuela.
3: Clinical Pharmacology Unit, School of Medicine, Universidad Centroccidental Lisandro Alvarado, Barquisimeto, Venezuela
4: Emory University School of Medicine. Atlanta, GA. USA.

Short Title: MOLECULAR MECHANISM OF ENDOTHELIAL DYSFUNCTION

Address correspondence: Valmore Bermúdez, MD; PhD. La Universidad del Zulia, Facultad de Medicina, Escuela de Medicina, Centro de Investigaciones Endocrino-Metabólicas "Dr. Félix Gómez". e-mail: Vbermudez@hotmail.com; fago@medscape.com

ndothelial dysfunction symbolize several pathological conditions, including altered anticoagulant and anti-inflammatory properties of the endothelium, impaired modulation of vascular growth, and dysregulation of vascular remodeling. Nevertheless, this term has been used commonly to refer to an impairment of endothelium-dependent vasorelaxation caused by a loss of nitric oxide bioactivity.

The clinical and scientific relevance of Nitric Oxide synthesis and bioavailability in endothelial dysfunction is based on the fact that it is a common factor in the pathogenesis of cardiovascular diseases. These alterations have been demonstrated in both animal models and humans, in the scope of dangerous pathological conditions as cigarette smoking, hypertension, hypercholesterolemia, aging, diabetes and heart failure.

A decline in Nitric Oxide bioavailability may be caused by decreased expression of the endothelial NO synthase, a reduction of substrate or cofactors for this enzyme, alterations of cellular signaling, enzyme inhibition by asymmetric dimethyl arginine and, finally, accelerated Nitric Oxide degradation by reactive oxygen species.

The knowledge of the processes related to these alterations becomes of remarkable importance for the understanding and of the generation of innovating and effective therapeutic strategies for cardiovascular diseases.

Key Words: Nitric Oxide, Nitric Oxide Synthase, Endothelial Dysfunction, Cardiovascular Diseases.

Introductio

ascular endothelial cells constitute a structurally simple, but functionally complex organ which regulates a number of processes as hemostasis, fibrinolysis, inflammation, blood pressure, lipoproteins metabolism and angiogenesis, and in this way, it plays an essential role in the homeostasis of the vascular system. Alterations presented in one or more of these physiological phenomena are what it is known as endothelial dysfunction¹ (Figure 1). Even though, the association between several risk factors and cardiovascular diseases is well documented, it is often observed that some individuals who present some of these factors do not develop cardiovascular diseases, which suggests there is an "activating connector" that once affected significantly joins risk factors with cardiovascular pathologies through some anomalous processes. Due to it's strategically location and its biological properties, vascular endothelial cells constitute this "hot key" in the chain of events that ends in vascular system dysfunction². Endothelium-dependent relaxation alteration by a decrease in both, synthesis and/ nitric oxide (NO) bioavailability constitute the earlier and most important phenomenon in endothelial dysfunction³. Nitric oxide carries out important functions related to the vascular system homeostasis such as vessel tone regulation, inhibition of platelet aggregation, leukocyte's adhesion and transmigration inhibition, as well as the proliferation and migration of smooth muscle cells. Hence, the decrease of this molecule's activity constitutes a major element in the pathophysiological processes that ending in cardiovascular atherosclerotic related diseases4. However, these transformations, besides being com-

plex and diverse have not been completely clarified⁵.

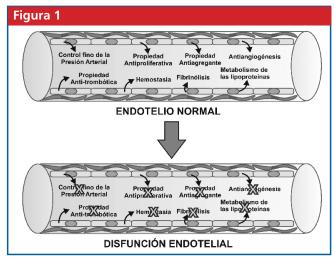


Figura 1: Disfunción Endotelial. El endotelio en condiciones normales emite múltiples señales moleculares que le conceden propiedades antiagregante plaquetaria, antitrombótica y antiaterogénica, lo cual lo hace un órgano esencial en el mantenimiento de la homeostasis del sistema vascular. Bajo condiciones patológicas se producen alteraciones en uno o más de los mecanismos de señalización molecular emitidos por éste, lo cual es conocido como disfunción endotelial.

o is a nitrogen-centered free radical produced exclusively by the nitric oxide synthase action (NOS). Three isoforms of this enzyme have been described which are highly homologue in their primary structure. Inducible nitric oxide synthase (iNOS) is expressed in the phagocytic cells by some pro-inflammatory stimuli like cytokines. The two other isoforms are constitutively expressed in nervous tissue (nNOS) and the endothelial cells (eNOS)6.eNOS synthesizes NO from Larginine through a oxidation process that involve five electrons transference by means of the intermediary NG-hidroxi-L-arginine^{7,8}. The substrates used by this enzyme are the amino acid L-arginine and molecular oxygen and the cofactors required are flavine adenine mononucleotide, tetrahydrobiopterin (BH4), flavine adenine mononucleotide (FMN) and flavine adenine dinuclotide (FAD) nicotinamide adenine dinucleotide phosphate (NADPH). Besides, the last two isoforms contain binding sites for hem group and calmodulin, being both of them essential for its activity. After calcium-calmodulin complex union to eNOS (between the COOH-terminal reductase domain and the NH2terminal oxydase domain) the electrons are yielded by the NADPH and then transported to oxigenase domain which contains the hem group, which results in citrulin and NO formation⁹ (Figure 2).

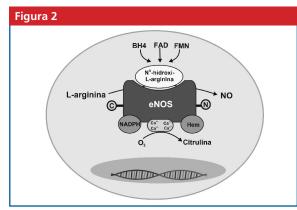


Figura 2: Biología de eNOS. El NO es producido principalmente por los leucocitos, las neuronas y las células endoteliales. En las últimas la eNOS sintetiza NO a partir de su sustrato la L-arginina mediante un paso de oxidación de 5 electrones por medio del intermediario NG-hidroxi-L-arginina.

A) Reduction of Nitric Oxide productioneNOS transcription Alteration

Molecular mechanisms of endothelial dysfunction

Although the term "inducible" has been restricted for iNOS, the expression of eNOS is also regulated by a variety of stimulus¹⁰. There is extensive evidence about factors which decrease eNOS expression, among which the tumor necrosis factor alpha (TNF-α) is included, which unstabilizes eNOS ARNm, apparently through regulatory proteins affinity increase to the 3' domain of iNOS ARNm molecule^{11,12}. Other stimuli that have been reported as ARNm stability reducers include lipopolysaccharide¹³, hypoxia¹⁴, and high concentrations of oxidized low density lipoproteins molecules (LDL_∞)¹⁵ (Figure 3).

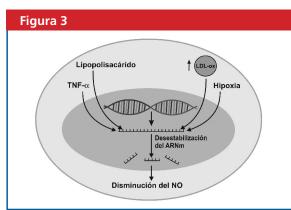


Figura 3: Disminución de la Expresión de eNOS. Diversos elementos propatogénicos como el TNF- α , el lipopolisacárido, la hipoxia y las altas concentraciones del LDL oxidada son capaces de desestabilizar la molécula de ARNm de la eNOS, ocasionando una reducción transcripcional de las concentraciones de eNOS.

Alteration in eNOS activity Decrease in BH4 intracellular concentrations and Uncoupling phenomena

Under some circumstances eNOS can generate superoxide anion instead NO as a consequence of BH4 concentration decline; this process is known as NADPH oxidation/NO synthesis uncoupling phenomenon¹⁶. Superoxide radical production is mediated through hem group in eNOS oxygenase domain when arginine and BH4 concentration is relatively low^{17,18,19}.

Mammals cells can generate BH4 through guanosine triphosphate (GTP) cyclohydrolase I (GTPCH I) enzymatic action²⁰. Physiological studies have shown a significant GTPCH I and BH4 activity decrease in various pathological states like insulinresistance, cigarette smoking, and hypercholesterolemia, probably through a LDL_{ox} increase, as well as an expression arise in some pro-inflammatory cytokines (TNF- α and the interleukin-1 β)^{21,22,23,24} (Figure 4). Furthermore, clinical and experimental studies have confirmed that BH4 acute administration improves endothelial dysfunction related to hypercholesterolemia, atherosclerosis, hypertension and smoking²⁵-²⁸. These mechanisms expose an important link between prepathogenic states involved in endothelial dysfunction development.

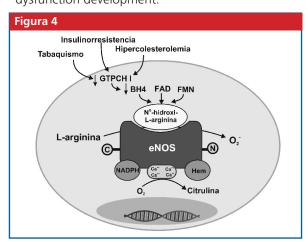


Figura 4: Desacoplamiento de eNOS. Para la síntesis del NO por parte de la eNOS son necesarias concentraciones adecuadas de BH4 y de su sustrato la L-arginina. Cuando alguno de estos factores se encuentra disminuido en cantidad suficiente, la enzima no es capaz de generar NO, produciendo en su lugar un radical libre altamente perjudicial para la biología de las células como es el anión superóxido.

Competitive inhibition of eNOS by asymmetric dimethylarginine

Vallance et al²⁹ first described in 1992 asymmetric dimethylarginine (ADMA) as a NOS endogenous inhibitor. Since then, the role of this molecule in the regulation of endothelial NO synthesis has increasingly attracted attention. This aminoacid is synthesized in endothelial cells from arginine by an enzyme belonging to arginine protein methyl transferase (PRMTs) family, specifically PRMT-130. Earlier experimental evidence showed that supplementation with L-arginine improves endothelium-dependent vasodilatation impairment in rabbits with hypercholesterolemia and atherosclerosis31,32, diminishes platelet aggregation33, inhibits monocyte adhesion³⁴ and vascular smooth muscle proliferation³⁵, which is markedly in contrast with both, experimental studies that showed a fail in L-arginine endothelium-dependent vasodilatation stimulation in isolated arterial rings, and in vitro studies showing eNOS saturation at physiological L-arginine concentrations and a failure of exogenous L-arginine administration in achieve an enzyme's activity increment, confirming a lack of L-arginine vasodilator effect in isolated arterial rings³⁶. This discrepancy between findings observed in intact animals versus in vitro assays was termed the "L-arginine paradox" that can be explained by the existence of an L-arginine competitive inhibitor now known as ADMA. Thus, administration of L-arginine at high doses should displace ADMA from the eNOS catalytic site and restore NO production to physiological levels in intact experimental models and humans beins^{36,37}.

The expression and activity of PRMT-1 and in consequence ADMA synthesis is modulated in endothelial cells by a variety of stimuli. For example, an expression increase in this enzyme had been observed in response to LDL³⁸ molecules and shear stress, conducing to intracellular ADMA concentration. On the other hand, PRMT-1 activity can be blocked by the suppression of a group of protein kinases called kappa beta inhibitors kinase (κβ-IK) that phosphorylates a group of proteins known as kappa beta inhibitors (kβ-l), which once phosphorylated are unable to retain the $k\beta$ nuclear factor ($k\beta$ -NF) in the cytoplasm, which, then becomes free to translocate to the nucleus, site where exerts its functions as transcription factor³⁸, suggesting a regulatory interplay by cytokines in the increase of its activity. Besides, multiple pathogenic factors like hypercholesterolemia, hyperglycemia, pro-inflammatory cytokines and hyperhomocysteinemia can diminish the activity of dymethilarginine dymetilaminohydrolase (DDAH). an enzyme responsible of hydrolyzes ADMA degradation, causing a significant intracellular elevation of this aminoacid³⁸ (Figure 5). Thus, the nexus between cardiovascular disease risk factors, ADMA and endothelial dysfunction are becoming evident in conjunction with the essential role of oxidative stress in atherosclerosis generation.

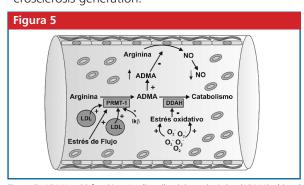


Figura 5: ADMA y Disfunción. La dimetilarginina asimétrica (ADMA) al igual que la L-arginina es un aminoácido que circula en el plasma, es excretado por la orina, y es encontrado en células y tejidos. Cuando las concentraciones de la primera (ADMA) exceden las de la segunda (L-arginina), se produce una inhibición competitiva de la síntesis de NO por parte de eNOS que conduce a la génesis de la disfunción endotelial.

Lipids and Caveolae Metabolism

Studies conducted worldwide have indicated that the eNOS location into the cell determines its enzymatic activity. A place of particular importance in the cell for the function of this enzyme is the caveolae⁴⁰. Caveolae is a structure constituted by specialized invaginations in the plasmatic membrane whose main components are cholesterol, glycoesphingolipids,

Concluding Remarks

and a structural protein called caveolin⁴¹. At present, all evidence indicates that numerous extracellular stimuli exert its signals transduction through this structure⁴². In this sense, eNOS location in caveolae determines its enzymatic activity inhibition by caveolin-1 linkage⁴³ that cause a blockade in eNOS interaction with the calmodulin when the intracellular calcium levels are low44. Also, high levels of oxidized LDL cause a decrease of cholesterol content of caveolae, resulting in caveolin-1-eNOS complex translocation to the cytoplasm, and in consequence, inhibition of its activity⁴⁵. Likewise, there is evidence that hypercholesterolemic serum and native LDL are capable to up regulate caveolin-1 concentration, increasing heterocomplexes formation between eNOS and this protein, diminishing NO production⁴⁵. Furthermore, it is known that some proatherogenic lipids, such as lysophosphatidylcholine and LDL_{ox}, interfere with signal transduction from receptors that activate eNOS^{46,48} (acetylcholine receptors, bradykinin, serotonin, histamine and others) (Figure 6). This process enlightens other mechanism which correlates prepathogenic and/or pathological conditions with the decrease of NO levels, and therefore, endothelial dysfunction.

B) Decrease in NO BioavailabilityIncrease in Arginase activity

Arginase is a key enzyme involved in arginine to ornithine (and urea) conversion. This enzyme has two isoforms, arginase I, which is constitutively expressed in the endothelial cells, and arginase II which can be induced by lipopolysacharides and interferon-y³⁹. Thus, systemic or local infectious processes could generate a significant increase on arginase levels causing an important decrease in arginine and consequently a lack in eNOS substrate bioavailability and consequently a failure in NO synthesis.

Oxidative stress

Even in presence of an adequate NO generation, some circumstances avoid this molecule to reach its biological targets due to a decrease in its bioavailability, as a consequence of the interaction with some chemical compounds⁴⁹. There is abundant experimental evidence indicating the role of NO oxidative inactivation as mediator of endothelial dysfunction and a prepathogenic vascular phenotype⁵. For example, in hyperlipidemia, excessive LDL synthesis entails a concomitant formation of LDL, which results in oxidative stress that causes NO conversion to peroxynitrite (chemical specie without the biological effects of NO) by a reaction that proceeds at 6,7 x 10⁹ M¹s^{-1,51,52}. This velocity is approximately three times higher than the reaction occurring between superoxide and superoxide dismutase (SOD). So that, in a compartment with NO, superoxide and SOD, the superoxide anion is able to react with any of the two other compounds⁵. The results of different studies support the role of the superoxide as essential element in the decrease of NO bioavailability in oxidative stress conditions. In rabbits with aortic atherosclerosis, a remarkable decrease in endothelium-related relaxation was seen (despite an NO synthesis increase up to three times greater in relation to the NO synthesis level in healthy rabbits), which was corrected by SOD treatment⁵³. Likewise, ascorbic acid infusion improves the vascular response to the acetylcholine in smokers, diabetic and patients with high blood pressure^{50,53}.

• Hyperglycemic Stress

Hyperglycemia increases oxygen-derived free radicals production via arachidonic acid metabolism arises. In human aortic endothelial cells, despite the extended exposition to high glucose concentration increases eNOS expression, a concomitantly superoxide anion elevation (probably from NADH/NADPH oxidase) result in NO inactivation. Besides, an extended hyperglycemic stress causes advanced glycosilation endproducts (AGES) accumulation, which is able to inactivate NO. In fact, the alteration in the capacity of endothelium-depending vessels relaxation in diabetic rats can be partially restablished by aminoquanidine (an AGES) administrationr^{54,56}. Thus, it is easy to deduce the importance of this mechanism as an remarcable connection between diabetes and cardiovascular diseases.

ndothelial dysfunction and more specifically, alteration in NO synthesis or action constitutes an essential step in the pathophysiology of most prevalent cardiovascular diseases. Due to essential NO functions (antiatherogenic, antithrombotic, antiproliferative agent), important changes are produced in the endothelial physiology when its biodisponibility is distorted. The multiple processes related to the reduction of synthesis and bioavailability of NO are far from being completely clarified. Although now days, a significant and growing body of information is being conformed, however, studies devoting to comprehension of biology of eNOS and its pathological modifications in the course of endothelial dysfunction are required. This fact will allow new pharmacological strategies generation in order to treat coronary heart disease since their beginnings, and consequently, preventing most feared complications in a safer and efficient approach.

References

- Drexler H. Endothelial dysfunction: clinical implications. Prog Cardiovasc Dis. 1997; 39:287–324.
- Boneti P, Lerman O, Lerman A. Endothelial Dysfunction: A Marker of Atherosclerotic Risk. Arterioscler Thromb Vasc Biol. 2003; 23:168-175.
- O'Connell B, Genest J. High- Density Lipoproteins and Endothelial Function. Circulation. 2001; 104:1978-1983.
- Kawashima S, Mitsuhiro Y. Dysfunction of Endothelial Nitric Oxide and Atherosclerosis. Arterioscler Thromb Vasc Biol. 2004; 24:998-1005.
- Harrison D. Perspective Series: Nitric Oxide And Nitric Oxide Synthases. J. Clin. Invest. 1997; 100:2153-2157.

- Govers R, Rabelink T. Cellular Regulation of Endothelial Nitric Oxide Synthase. Am J Pyshiol Renal Physiol. 2001; 280:193-206.
- Palmer RM, Ashton DS, and Moncada S. Vascular endothelial cells synthesize nitric oxide from L-arginine. Nature. 1988; 333:664–666.
- 8. Zembowicz A, Hecker M, Macarthur H, Sessa WC, and Vane JR. Nitric oxide and another potent vasodilator are formed from NG-hydroxy-L-arginine by cultured endothelial cells. Proc Natl Acad Sci USA. 1991; 88:11172–11176.
- Abu-Soud HM and Stuehr DJ. Nitric oxide synthases reveal a role for calmodulin in controlling electron transfer. Proc Natl Acad Sci USA. 1993; 90:10769–10772.
- Teichert AM, Miller TL, Tai SC, Wang Y, Bei X, Robb GB, Phillips MJ, and Marsden PA. In vivo expression profile of an endothelial nitric oxide synthase promoter-reporter transgene. Am J Physiol Heart Circ Physiol. 2000; 278:H1352–H1361.
- Nishida K, Harrison DG, Navas JP, Fisher AA, Dockery SP, Uematsu M, Nerem RM, Alexander RW, and Murphy TJ. Molecular cloning and characterization of the constitutive bovine aortic endothelial cell nitric oxide synthase. J Clin Invest. 1992; 90:2092–2096.
- Alonso J, Sanchez de Miguel L, Monton M, Casado S, and Lopez-Farre A. Endothelial cytosolic proteins bind to the 39 untranslated region of endothelial nitric oxide synthase mRNA: regulation by tumor necrosis factor alpha. Mol Cell Biol 1997; 17:5719–5726.
- Lu JL, Schmiege LM, 3rd Kuo L, and Liao JC. Downregulation of endothelial constitutive nitric oxide synthase expression by lipopolysaccharide. Biochem Biophys Res Commun. 1996: 225:1–5.
- McQuillan LP, Leung GK, Marsden PA, Kostyk SK, and Kourembanas S. Hypoxia inhibit expression of eNOS via transcriptional and posttranscriptional mechanisms. Am J Physiol Heart Circ Physiol. 1994; 267:1921–1927.
- Liao JK, Shin WS, Lee WY, and Clark SL. Oxidized lowdensity lipoprotein decreases the expression of endothelial nitric oxide synthase. J Biol Chem. 1995; 270:319–324.
- Pou S, Pou WS, Bredt DS, Snyder SH, and Rosen GM. Generation of superoxide by purified brain nitric oxide synthase. J Biol Chem. 1992; 267:24173–24176.
- Stroes E, Hijmering M, Vanzandvoort M, Wever R, Rabelink TJ, and Vanfaassen EE.
 Origin of superoxide production by endothelial nitric oxide synthase. FEBS Lett. 1998;438:161–164.
- Vasquez-Vivar J, Kalyanaraman B, Martasek P, Hogg N, Masters BSS, Karoui H, Tordo P, and Pritchard KA. Superoxide generation by endothelial nitric oxide synthase: the influence of cofactors. Proc Natl Acad Sci USA.1998: 95:9220–9225.
- Wever RMF, van Dam T, van Rijn HJ, de Groot F, and Rabelink TJ. Tetrahydrobiopterin regulates superoxide and nitric oxide generation by recombinant endothelial nitric oxide synthase. Biochem Biophys Res Commun. 1997; 37:340–344.
- Kojima, S., S. Ona, I. lizuka, T. Arai, H. Mori, and K. Kubota. Antioxidative activity of 5,6,7,8-tetrahydrobiopterin and its inhibitory effect on paraquat-induced cell toxicity in cultured rat hepatocytes. Free Rad. Res. 1995; 23:419–430.
- Pieper, G.M. Acute amelioration of diabetic endothelial dysfunction with a derivative of the nitric oxide synthase cofactor, tetrahydrobiopterin. Cardiovasc. Pharmacol. 1997;29:8–15.
- Stroes, E., J. Kastelein, F. Cosentino, W. Erkelens, R. Wever, H. Koomans, T. Luscher, and T. Rabelink. Tetrahydrobiopterin restores endothelial function in hypercholesterolemia. J. Clin. Invest. 1997; 99:41–46.
- Vann LR, Payne SG, Edsall LC, Twitty S, Spiegel S, Milstien S. Involvement of sphingosine kinase in TNF-alpha-stimulated tetrahydrobiopterin biosynthesis in C6 glioma cells. J Biol Chem. 2002; 277:12649–12656.
- Dulak J, Polus M, Guevara I, Polus A, Hartwich J, Dembinska-Kiec A. Regulation of inducible nitric oxide synthase (iNOS) and GTP cyclohydrolase I (GTP-CH I) gene expression by ox-LDL in rat vascular smooth muscle cells. J Physiol Pharmacol. 1997; 48:689–697.
- Cosentino F, Patton S, d'Uscio LV, Werner ER, Werner-Felmayer G, Moreau P, Malinski T, Luscher TF. Tetrahydrobiopterin alters superoxide and nitric oxide release in prehypertensive rats. J Clin Invest. 1998; 101:1530–1537.
- Stroes, E, Kastelein J, Cosentino F, Erkelens W, Wever R, Koomans H, Luscher T, and Rabelink Tetrahydrobiopterin restores endothelial function in hypercholesterolemia. J Clin Invest. 1997;99:41–46.
- Setoguchi S, Mohri M, Shimokawa H, Takeshita A. Tetrahydrobiopterin improves endothelial dysfunction in coronary microcirculation in patients without epicardial coronary artery disease. J Am Coll Cardiol. 2001;38:493

 –498.
- Heitzer T, Brockhoff C, Mayer B, Warnholtz A, Mollnau H, Henne S, Meinertz T, Munzel T. Tetrahydrobiopterin improves endothelium dependent vasodilation in chronic smokers: evidence for a dysfunctional nitric oxide synthase. Circ Res. 2000:86:E36–E41.
- Vallance, P., Leone, A., Calver, A., Collier, J. & Moncada, S. (1992) Accumulation of an endogenous inhibitor of NO synthesis in chronic renal failure. Lancet 339:572-575.
- Leiper JM, Santa Maria J, Chubb A, et al. Identification of two human dimethylarginine dimethylaminohydrolases with distinct tissue distributions and homology with microbial arginine deaminases. Biochem J. 1999; 343:209–14.
- Cooke, J. P., Andon, N. A., Girerd, X. J., Hirsch, A. T. & Creager, M. A. (1991) Arginine restores cholinergic relaxation of hypercholesterolemic rabbit thoracic aorta.

- Circulation 83:1057-1062.[Abstract/Free Full Text]
- **32.** Böger, R. H., Bode-Böger, S. M., Mügge, A., Kienke, S., Brandes, R., Dwenger, A. & Frölich, J. C. (1995) Supplementation of hypercholesterolaemic rabbits with L-arginine reduces the vascular release of superoxide anions and restores NO production. Atherosclerosis 117:273-284.[Medline]
- Böger, R. H., Bode-Böger, S. M., Phivthong-ngam, L., Böhme, M., Brandes, R. P., Mügge, A. & Frölich, J. C. (1997) Dietary L-arginine slows the progression of atherosclerosis in cholesterol-fed rabbits—comparison with lovastatin. Circulation 96:1282-1290.[Abstract/Free Full Text]
- Candipan, R. C., Wang, B. Y., Buitrago, R., Tsao, P. S. & Cooke, J. P. (1996) Regression or progression. Dependency on vascular nitric oxide. Arterioscler. Thromb. Vasc. Biol. 16:44-50.[Abstract/Free Full Text]
- **35.** Tsao, P. S., Theilmeier, G., Singer, A. H., Leung, L. L. & Cooke, J. P. (1994) L-Arginine attenuates platelet reactivity in hypercholesterolemic rabbits. Arterioscler. Thromb. 14:1529-1533.
- Böger, R. H., Bode-Böger, S. M., Kienke, S., Nafe, R., Stan, A. C. & Frölich, J. C. (1998) Dietary L-arginine decreases myointimal cell proliferation and vascular leukocyte accumulation in cholesterol-fed rabbits. Atherosclerosis 136:67-77.
- Mügge, A. & Harrison, D. G. (1991) L-Arginine does not restore endothelial dysfunction in atherosclerotic rabbit aorta in vitro. Blood Vessels 28:354-357.
- 38. Clarke S. Protein methylation. Curr Opin Cell Biol. 1993; 5:977–983.
- Buga, G.M., R. Singh, S. Pervin, N.E. Rogers, D.A. Schmitz, C.P. Jenkinson, S.D. Cederbaum, and L.J. Ignarro.. Arginase activity in endothelial cells: inhibition by NG-hydroxy-L-arginine during high-output NO production. Am. J. Physiol. 1996; 271:1988–1998.
- Parton RG. Caveolae and caveolins. Curr Opin Cell Biol. 1996; 8:542–548. Smart EJ, Graf GA, McNiven MA, Sessa WC, Engelman JA, Scherer PE, Okamoto T, and Lisanti MP. Caveolins, liquid-ordered domains, and signal transduction. Mol Cell Biol. 1999; 19:7289–7304.
- Ju H, Zou R, Venema VJ, and Venema RC. Direct interaction of endothelial nitric-oxide synthase and caveolin-1 inhibits synthase activity. J Biol Chem. 1997; 272:18522–18525.
- Michel JB, Feron O, Sacks D, and Michel T. Reciprocal regulation of endothelial nitricoxide synthase by Ca-calmodulin and caveolin. J Biol Chem. 1997; 272:15583–15586.
- 43. Blair A, Shaul PW, Yuhanna IS, Conrad PA, and Smart EJ. Oxidized low density lipoprotein displaces endothelial nitric-oxide synthase (eNOS) from plasmalemmal caveolae and impairs eNOS activation. J Biol Chem. 1999; 274:32512–32519.
- Feron O, Dessy C, Desager JP, Balligand JL. Hydroxy-methylglutarylcoenzyme A reductase inhibition promotes endothelial nitric oxide synthase activation through a decrease in caveolin abundance. Circulation. 2001; 103:113–118.
- Feron O, Dessy C, Moniotte S, Desager JP, Balligand JL. Hypercholesterolemia decreases nitric oxide production by promoting the interaction of caveolin and endothelial nitric oxide synthase. J Clin Invest. 1999; 103:897–905.
- 46. Hirata K, Akita H, Yokoyama M. Oxidized low density lipoprotein inhibits bradykinin-induced phosphoinositide hydrolysis in cultured bovine aortic endothelial cells. FEBS Lett. 1991; 287:181–184.
- 47. Inoue N, Hirata K, Yamada M, Hamamori Y, Matsuda Y, Akita H, Yokoyama M. Lysophosphatidylcholine inhibits bradykinin-induced phosphoinositide hydrolysis and calcium transients in cultured bovine aortic endothelial cells. Circ Res. 1992; 71:1410–1421.
- Miwa Y, Hirata K, Kawashima S, Akita H, Yokoyama M. Lysophosphatidylcholine inhibits receptor-mediated Ca²⁺ mobilization in intact endothelial cells of rabbit aorta. Arterioscler Thromb Vasc Biol. 1997; 17:1561–1567.
- Vallance P, Chan N. Endothelial function and nitric oxide: clinical relevance. Heart. 2001; 85:342-350.
- Thomson, L., M. Trujillo, R. Telleri, and R. Radi. Kinetics of cytochromec oxidation by peroxynitrite: implications for superoxide measurements in nitric oxide-producing biological systems. Arch Biochem. Biophys. 1995M; 319:491–497.
- Heitzer, T., H. Just, and T. Munzel. Antioxidant vitamin C improves endothelial dysfunction in chronic smokers. Circulation. 1996; 94:6–9.
- Ting, H.H., F.K. Timimi, K. Boles, S. Creager, P. Ganz, and M.A. Creager. Vitamin C acutely improves endothelium-dependent vasodilation in patients with non-insulin-dependent diabetes mellitus. Circulation. 1995.92(Suppl.1):1747. (Abstr.).
- Solzbach, U., B. Hornig, M. Jeserich, and H. Just. Vitamin C improves endothelial dysfunction of epicardial coronary arteries in hypertensive patients. Circulation.1997; 96:1513–1519.
- Hironori Nakagami, Yasufumi Kaneda, Toshio Ogihara and Ryuichi Morishita. Endothelial Dysfunction in Hyperglycemia as a Trigger of Atherosclerosis. Current Diabetes Reviews. 2005. 1, 59-63.
- Christian Rask-Madsen and George L King. Mechanisms of Disease: endothelial dysfunction in insulin resistance and diabetes. Nature Clinical Practice Endocrinology and Metabolism Jan 2007, Vol 3, 46-56.
- Brett E. Fenster, Philip S. Tsao P and Stanley G. Rockson. Endothelial dysfunction: clinical strategies for treating oxidant stress. American Heart Journal. Volume 146, Issue 2, August 2003, Pages 218-226