

Latin American Journal of Aquatic Research

E-ISSN: 0718-560X

lajar@ucv.cl

Pontificia Universidad Católica de Valparaíso Chile

Ramírez-Mosqueda, Edith; Pérez-Jiménez, Juan Carlos; Mendoza-Carranza, Manuel
Reproductive parameters of the southern stingray Dasyatis americana in southern gulf of Mexico
Latin American Journal of Aquatic Research, vol. 40, núm. 2, julio, 2012, pp. 335-344
Pontificia Universidad Católica de Valparaíso
Valparaiso, Chile

Available in: http://www.redalyc.org/articulo.oa?id=175024254008



Complete issue

More information about this article

Journal's homepage in redalyc.org



Research Article

Reproductive parameters of the southern stingray *Dasyatis americana* in southern gulf of Mexico

Edith Ramírez-Mosqueda¹, Juan Carlos Pérez-Jiménez¹ & Manuel Mendoza-Carranza²

¹Departamento de Aprovechamiento y Manejo de Recursos Acuáticos El Colegio de la Frontera Sur (ECOSUR), Unidad Campeche, Calle 10 #264, Col. Centro, Campeche, México ²Departamento de Aprovechamiento y Manejo de Recursos Acuáticos El Colegio de la Frontera Sur (ECOSUR), Unidad Villahermosa, Carretera Villahermosa-Reforma km 15.5 Ranchería Guineo, sección II, 86280 Villahermosa, México

ABSTRACT. The southern stingray *Dasyatis americana* (Hildebrand & Schroeder, 1928) is the most landed elasmobranch by small-scale fleets in southern gulf of Mexico. However, little is known of its life history parameters in this region. In this study, a total of 900 specimens were collected from February 2006 to December 2008 to determine the reproductive parameters needed for population assessments by means of ecological risk assessments or demographic analysis. Results suggested that females of *D. americana* reproduce annually, with a gestation of 7-8 months. The reproductive cycle of females is asynchronous, with ovulation and parturition occurring throughout the year. Females and males matured at 764 and 517 mm disc width (DW₅₀) respectively. *D. americana* has one of the highest fecundity among dasyatids, from 2 to 7 embryos, with a sex ratio of embryos of 1:1. A linear relationship between maternal DW and fecundity was estimated, the larger females contain more embryos. The status of the population of *D. americana* is a cause of concern in the southern gulf of Mexico due to its high frequency of capture in artisanal fisheries and its apparently low biological productivity.

Keywords: Dasyatis americana, disc width at maturity, reproductive cycle, fecundity, gulf of Mexico.

Parámetros reproductivos de la raya látigo americana *Dasyatis americana* en el sur del golfo de México

RESUMEN. La raya látigo americana *Dasyatis americana* (Hildebrand & Schroeder, 1928), conocida como balá en México, es el elasmobranquio más capturado y desembarcado por flotas artesanales en el sur del golfo de México. Sin embargo, se conoce poco de sus parámetros de historia de vida en esta región. En este estudio, fueron analizados 900 ejemplares entre febrero 2006 y diciembre 2008 para determinar los parámetros reproductivos necesarios para evaluaciones poblacionales por medio de análisis demográficos o evaluaciones de riesgo ecológico. Los resultados sugieren que las hembras de *D. americana* se reproducen anualmente, con una gestación de 7-8 meses aproximadamente. El ciclo reproductivo de las hembras es asincrónico, con la ovulación y alumbramiento ocurriendo a lo largo de todo el año. Las hembras y los machos maduran a los 764 y 517 mm de ancho de disco (AD₅₀), respectivamente. *D. americana* tiene una de las fecundidades más altas entre los dasyatidos, de 2 a 7 embriones, y con una proporción de sexos de 1:1 en los embriones. Existe relación significativa lineal entre la longitud de las hembras y la fecundidad, las hembras más grandes contenían más embriones. El estado de la población de *D. americana* es motivo de preocupación en el sur del golfo de México debido a que es muy frecuente en las capturas de pesquerías artesanales y a que su productividad biológica es aparentemente baja.

Palabras clave: Dasyatis americana, longitud de madurez, ciclo reproductivo, fecundidad, golfo de México.

INTRODUCTION

The southern stingray Dasyatis americana (Hildebrand & Schroeder, 1928) is distributed from New Jersey to Florida in United States, throughout the gulf of Mexico, Bahamas, and the Greater and Lesser Antilles, and bordering the northern coast of South America to southeastern Brazil (McEachran & de Carvalho, 2002). It inhabits shallow waters and feeds on bottom-dwelling invertebrates, mainly bivalves and worms (McEachran & de Carvalho, 2002). Although frequently captured near shore, this species is also found in estuaries and rivers (McEachran & Fechhelm, 1998). D. americana is abundant on sandy bottoms of the inner shelf of the gulf of Mexico (Castro-Aguirre & Espinoza-Pérez, 1996). In the western Campeche Bank, in southern gulf of Mexico, D. americana is the most frequently landed among elasmobranch species, with mean landings of 1.400 ton per year in the state of Campeche according to the statistics from the regional office of SAGARPA (Secretaría de Agricultura, Ganadería, Desarrollo Rural, Pesca y Alimentación, 2010).

This species reaches a maximum disc width of 1520 mm (Bigelow & Schroeder, 1953). Females and males mature at 750-800 and 510 mm DW, respectively, young are 170 to 180 mm DW at birth (Bigelow & Schroeder, 1953) and fecundity range from 3 to 5 (McEachran & de Carvalho, 2002). *D. americana*, as the other dasyatid rays, display aplacental viviparity with trophonemata (Hamlett *et al.*, 1996).

Reproductive parameters estimated for *D. americana* in captivity, showed a biannual reproductive cycle, a gestation period of 4.4-7.5 months, a fecundity of 2-10 with a linear relationship with maternal disc width, size at birth 200-340 mm disc width, and 1:1 sex ratio for neonates (Henningsen, 2000). However, an annual reproductive cycle has been observed on wild specimens from Florida and Virginia waters (Grubbs *et al.*, 2006).

The determination of reproductive parameters are needed for assessment of elasmobranch populations by means of demographic analysis or ecological risk assessments due to the lack of time series of catch and fishing effort by species in Mexican official catch statistics. For example, Smith *et al.* (2008) developed age-structured demographic models for *Dasyatis dipterura* from the Mexican Pacific coast, concluding that this species has low intrinsic growth potential and limited resilience to fishing pressure.

D. americana is classed as Data Deficient by the International Union for Conservation of Nature, because the status of their populations is unknown and

there is a lack of life history parameters throughout most of its range (Grubbs et al., 2006). Mexican fisheries for elasmobranchs have been managed since 2007 by the Mexican Official Standard NOM-029-PESC-2006, Responsible Fisheries of Sharks and Rays. Specifications for their Use (Poder Ejecutivo Federal, 2007) and available information from the Atlantic coast of Mexico indicates that landings of batoids have been declining since the late 1990's (Poder Ejecutivo Federal, 2006), which indicates that the intrinsic growth potential of the population of D. americana must be assessed to determine its vulnerability to fishing pressure. Objectives of the present study were to determine the reproductive parameters for D. americana in the southern gulf of Mexico needed to develop, in the near future, agestructured or stage-based demographic models.

MATERIALS AND METHODS

A total of 900 specimens were examined from February 2006 to December 2008. Rays were obtained from commercial catches of the small-scale fleet from San Pedro, Tabasco, in the southern gulf of Mexico (Fig. 1). Bottom-set longlines with 60 mm shanklength tune circle hooks were the primary method of capture. The catch depth ranged from 10 to 40 m.

Disc width (DW) was measured between the tips of the widest portion of the pectoral fins (McEachran & de Carvalho, 2002). All rays were measured and examined to quantify reproductive parameters. Maturity was assessed macroscopically for both sexes. Maturity in males was assessed by the extent of clasper calcification (Clark & Von Schmidt, 1965) and testes development (Snelson et al., 1988). Claspers were measured from the point of insertion at the cloaca to the tip of claspers. Males were assigned to one of two stages as follows: 1) immature, testes and claspers undeveloped; 2) mature, claspers fully developed, exceeding the posterior edge of the pelvic fins, presented hardened internal structure and, could be rotated toward the anterior part without bending (Clark & Von Schmidt, 1965), and had enlarged testes with prominent lobes (Snelson et al., 1988).

Maturity in females was determined by the onset of first ovulation (Walker, 2005). Follicle diameter of the largest cohort in the ovary and the width of the left uterus were measured. In uteri, the trophonemata development, presence of uterine milk, uterine eggs and embryos were recorded. Uteri were examined to determine if embryos or uterine eggs were present. When present, the sex and DW of embryos was recorded.

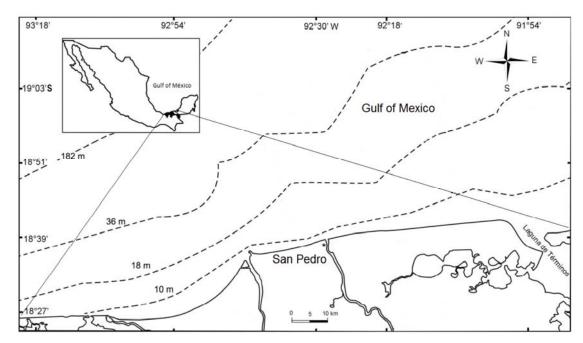


Figure 1. Location of San Pedro, Tabasco, landing site of the small-scale fleet in southern gulf of Mexico.

Figura 1. Ubicación de San Pedro, Tabasco, sitio de desembarque de la flota de pequeña escala en el sur del golfo de México.

Females were assigned to one reproductive status as follows: 1) immature, no ovarian development or if developed, there was no vitellogenic activity (follicles <25 mm diameter), uterus thin (<46 mm width) with trophonemata not well developed (<5 mm in length, thin and pale); 2) non-gravid mature, medium to large follicles (15-34 mm diameter) in the ovary, some with vitellogenic activity, wide uterus (23-47 mm), with trophonemata well developed and vascularized (>5 mm in length); 3) gravid, embryos in uterus, with medium to large follicles in the ovary (7-40 mm diameter).

Logistic models were used to estimate median disc width at maturity (DW₅₀) according to Mollet et al. (2000) by fitting the logistic model $Y = [1+e^{-(a+bX)}]^{-1}$ to the binary data of maturity (immature = 0, mature = 1). Because of the asynchronous nature of the reproductive cycle of females, the methods described by Braccinni et al. (2006) were used to determine the gestation period and the ovarian cycle. The time series of the disc width of embryos were analyzed to determine the gestation. Gravid females were assigned to one of seven categories based on the size of the embryos they carried: 1) ≤59 mm DW, 2) 60-89 mm DW, 3) 90-119 mm DW, 4) 120-149 mm DW, 5) 150-179 mm DW, 6) 180-209 mm DW and, 7) \geq 210 mm DW. The length of the gestation period was determined following the growth of embryos of different classes or cohorts through the time, based on the assumption that embryos from different cohorts have the same growth pattern (Braccinni *et al.*, 2006).

To determine if ovarian cycle and gestation were concurrent or consecutive, correlation analysis was performed to assess possible relationships between follicle diameter and embryo size. The method of Braccinni et al. (2006) may be used to determine the ovarian cycle if the gestation and the ovarian cycle run concurrently. To estimate the ovarian cycle, the time series of the follicle diameter of gravid females was analyzed. Follicle diameter from the same subset of data selected for the gestation period analysis (seven categories of gravid females based on the size of embryos) was used to estimate the length of the ovarian cycle. The length of the ovarian cycle was determined following the growth of follicles of different classes or cohorts through the time, based on the assumption that the diameter of follicles from different cohorts has the same growth pattern (Braccinni et al., 2006).

The disc width range at birth was estimated based on the disc width of the largest embryo examined and the smallest free-swimming ray recorded. To determine the fecundity, the total number of embryos was counted. The null hypothesis of a sex ratio 1:1 of embryos was tested with the chi-square procedure with Yate's correction for continuity (Zar, 1984). The relationship between maternal DW and the number of

embryos was plotted and fitted to a simple linear regression model.

RESULTS

A total of 494 females ranging from 340 to 1640 mm DW (mean = 766.5 ± 152.5) and 406 males 400-1100 mm DW (mean = 613.3 ± 69.3) were analyzed. In females, the disc width at which 50% of the population was mature was 764 mm (DW₅₀, 95% CI: 756-772 mm) (Fig. 2), with 288 mature and 206 immature females recorded. The smallest mature female measured 650 mm DW, had abundant uterine milk, follicles 32 mm diameter and uterus 34 mm width, and the largest immature female measured 960. had follicles 15 mm diameter and uterus 29 mm width. In males, the disc width at which 50% of the population was mature was 517 mm (DW₅₀, 95% CI: 510-523 mm) (Fig. 2), with 373 mature and 33 immature males recorded. The smallest mature male measured 490 mm DW (calcified claspers: 130 mm length), and the largest immature male measured 670 mm DW (partially calcified claspers: 120 mm length).

The time series of the embryos disc width (Fig. 3a) showed the asynchronous nature of the reproductive cycle of *D. americana*. The gestation lasts approximately 7-8 months. The development of embryos was traced following three cohorts through time (Figs. 4a and 4b). A first cohort could be traced from early October 2006 to late May 2007 (Fig. 4a); a second cohort from early February to late September 2008; and a third cohort from early July 2008 to late January 2009 (Fig. 4b).

The ovarian cycle could not be determined because this period does not occur concurrently with the gestation period, and the asynchronous nature of the reproductive cycle prevents it by analyzing the time series of follicle diameter of non-gravid mature females. The follicle diameter of gravid females does not show a relationship with the categories of gravid females (based on the size of the embryos they carried), as would be expected if gestation and the ovarian cycle were concurrent (Fig. 3b).

Although the correlation between the disc width of embryos and follicle diameter was significant, the degree of association is weak (r = 0.49, P < 0.001), because there exist a large variability, and only some gravid females containing full term embryos (>175 mm DW) had large follicles (Fig. 5). Therefore, the ovarian cycle and gestation were consecutive, the follicles and embryos do not develop at the same time. Apparently, however, the follicle development begins towards the end of gestation and follicles continue

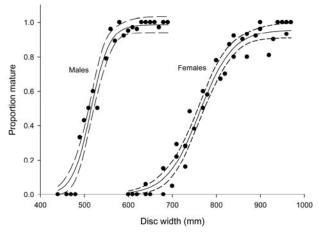
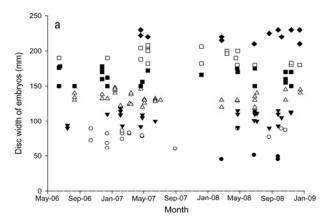


Figure 2. Female and male disc width-at-maturity ogives. Long dashed lines are 95% confidence intervals.

Figura 2. Ojivas de la longitud de madurez de hembras y machos. Las líneas punteadas son los intervalos de confianza al 95%.



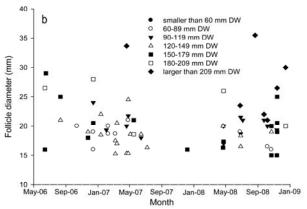


Figure 3. Time series of a) disc width of embryos and, b) average follicle diameter of gravid females of *Dasyatis americana*. Symbols descriptions apply to both graphs.

Figura 3. Series de tiempo de a) ancho del disco de los embriones, y b) el diámetro promedio de los ovocitos de hembras preñadas de *Dasyatis americana*. La descripción de los símbolos aplica para las dos gráficas.

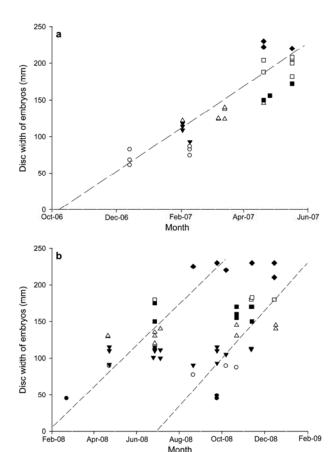


Figure 4. Time series of the growth of three cohorts of embryos (symbols descriptions in Figure 3): a) from October 2006 to May 2007, b) from February to September 2008 and from July 2008 to January 2009. Dotted lines show the cohorts of embryos.

Figura 4. Series de tiempo del crecimiento de tres cohortes de embriones (descripción de los símbolos en Figura 3): a) de octubre 2006 a mayo 2007, b) de febrero a septiembre 2008 y de julio 2008 a enero 2009. Las líneas punteadas muestran las cohortes de embriones.

growing after the 7-8 months of gestation. If it is assumed that the ovarian cycle continues several months after parturition, then it could be hypothesized that females of *D. americana* reproduce annually.

Disc width at birth was 230-340 mm. Fecundity ranged from two to seven embryos (mean = 2.5 ± 0.99). The 74.6% of gravid females carried two embryos. The most frequent number of follicles in gravid females was also two (30% of females). As the maternal DW increased, fecundity also increased (fecundity = 0.004 DW-1.41, $r^2 = 0.46$, $F_{1,59} = 46.99$, P < 0.001) (Fig. 6). In total, 77 females and 79 males embryos were recorded, resulting in a sex ratio not significantly different from 1:1 (chi-square = 0.03, P > 0.05).

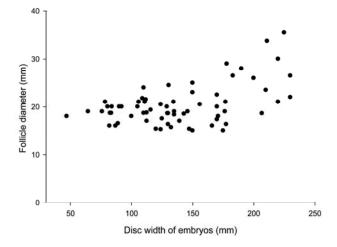


Figure 5. Correlation between the embryo disc width and the average follicle diameter (r = 0.49, P < 0.001).

Figura 5. Correlación entre el ancho de disco de los embriones y el diámetro promedio de los ovocitos (r = 0.49; P < 0.001).

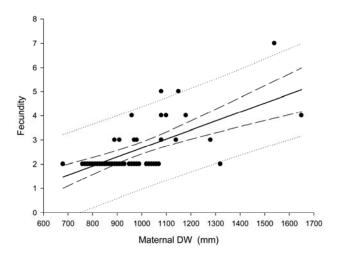


Figure 6. Relationship between maternal disc width and fecundity (fecundity = 0.004 DW-1.41; r^2 = 0.46, $F_{1,59}$ = 46.99; P < 0.001). Long dashed lines are 95% confidence intervals and dotted lines are 95% prediction intervals.

Figura 6. Relación entre el ancho de disco de las hembras y su fecundidad (fecundidad = 0,004 AD-1,41; $r^2 = 0,46$; $F_{1,59} = 46,99$; P < 0,001). Las líneas punteadas largas son los intervalos de confianza al 95% y las líneas de puntos cortos son los intervalos de predicción al 95%.

DISCUSSION

Before this study, nothing was known about the reproductive parameters of *D. americana* in the gulf of Mexico. Disc width at maturity for females and males, estimated in the present study, was similar to that documented by Bigelow & Schroeder (1953) and

MacEachran & de Carvalho (2002) for specimens collected from northwestern to southwestern Atlantic, and also similar to that estimated by Henningsen & Leaf (2010) for captive specimens (Table 1). However, because it is unclear if the criteria to determine maturity used by Bigelow & Schroeder (1953) and McEachran & de Carvalho (2002) was the same of that used in the present study, we recommend that a future study that uses the same criteria to determining the disc width at maturity among regions should be carried out to test for regional differences.

Gestation periods are slightly longer in wild (7-8 months) than in captive specimens (4.4-7.5 months) (Table 1). The difference is probably associated with environmental conditions, in particular temperature. Mahon et al. (2004) documented that gestation in captive spotted eagle ray, Aetobatos narinari, is related to water temperature, being longer at lower temperatures (11-12.5 months at 19.8-29.4°C) than at higher temperatures (6-6.2 months at 28.1-30.1°C). Temperature was not provided by Henningsen (2000), however, Henningsen & Leaf (2010) indicate that temperature was maintained at 24-25°C in their study. Surface temperatures ranges between 23.9-28.7°C in the western Campeche Bank (Cuevas-Zimbrón et al. 2011) and are closely similar to the northern Campeche Bank where they fluctuate between 23.5-29.5°C, while bottom temperatures, in the second region, ranges between 22.9-24.5°C (Piñeiro et al., 2001). If the same bottom temperatures were observed in the western Campeche Bank, specimens of D. americana are apparently exposed to slightly lower temperatures in the wild, which could explain that gestation is longer in wild specimens. Henningsen et al. (2004), indicate that temperature has an effect on gestation, decreasing with an increase in temperature, such as occurs in A. narinari (Mahon et al., 2004).

Gestation ranges from 3 to 12 months in wild specimens and from 2 to 7.5 months in captive specimens of dasyatids (Table 2). D. americana from the southern gulf of Mexico (present study) is among dasyatids (D. centroura, D. chrysonota, D. longa and D. sayi) with gestation of 8 months or longer. In addition, D. americana has the longest gestation period (4.4-7.5 months) reported for captive dasvatid rays (Table 2). According to Henningsen et al. (2004) the temperature may have a profound effect on development. So, variation in gestation could be linked to this variable, as have been reported for captive specimens (Henningsen et al., 2004; Mahon et al., 2004). Comparisons of the differences in the length of gestation among species in wild specimens is difficult because a detailed characterization of the habitat is needed for every species.

Reproductive cycle of *D. americana* differs among wild (annual) and captive specimens (biannual). Grubbs et al. (2006) suggest an annual cycle of reproduction for wild specimens of D. americana in Florida and Virginia, eastern coast of USA. An annual cycle of reproduction has also been documented for other dasyatids, such as D. sayi, D. sabina and D. chrysonota (Snelson et al., 1989; Johnson & Snelson, 1996; Ebert & Cowley, 2008). The biannual reproductive cycle for wild specimens has been suggested for two species only, D. marianae and D. guttata (Yokota & Lessa, 2007), and for captive specimens, it has been determined for three species, D. americana, D. kuhlii and Pteroplatytrygon violacea (Henningsen, 2000; Mollet et al., 2002; Janse & Schrama, 2009). An uncommon triannual cycle of reproduction among elasmobranchs has been suggested for D. marmorata in wild specimens (Capapé & Zaouali, 1995), however, the authors do not explore the possibility of an asynchronous reproductive cycle for this species, in which case, a different methodology must be used, such as the proposed by Braccinni et al. (2006).

Differences in the reproductive cycle between captive and wild specimens are well documented for elasmobranchs (Henningsen *et al.*, 2004). In captive animals, Henningsen *et al.* (2004) indicate that several species were observed to mate immediately following parturition, whereas a longer gap it is observed in the wild (as is the case of *D. americana*). In addition, the effect of some environmental variables (*e.g.* temperature) should be explored in future studies to explain for differences among species.

The reproductive cycle is synchronous in several dasyatid species, D. sayi, D. sabina, D. marianae, D. guttata and D. chrysonota (Snelson et al., 1989; Johnson & Snelson, 1996; Yokota & Lessa, 2007; Ebert & Cowley, 2008). The asynchronous reproductive cycle has been determined for D. americana (present study), D. kuhlii, D. zugei, Himantura walga (White & Dharmadi, 2007) and P. violacea (Véras et al., 2009). In sharks, females are asynchronous in some species inhabiting tropical waters, where stable conditions and abundant environmental throughout the year allow reproduction and birth to occur year around (Castro, 2009). The species of dasyatids inhabits tropical waters, which could explain that the asynchronous reproductive cycle is very common in these elasmobranchs.

Ovarian cycle and gestation were consecutive for *D. americana* in the wild (present study) and concurrent in captive environment (Henningsen, 2000). Also, in *D. sabina* has been determined that vitellogenesis and gestation was consecutive

Table 1. Comparison of reproductive parameters among southern gulf of Mexico (present study), western Atlantic and captive specimens of the southern stingray *Dasyatis americana*.

Tabla 1. Comparación de los parámetros reproductivos entre el sur del golfo de México (presente estudio), Atlántico oeste y especímenes en cautiverio de la raya látigo americana *Dasyatis americana*.

Parameters	Southern gulf of Mexico (present study)	From Chesapeake Bay to Brazil ^{1,2}	Captive specimens ^{3,4}	
DW ₅₀ at maturity of females (mm)	775 unknown		unknown	
DW ₅₀ at maturity of males (mm)	517	unknown	unknown	
Smallest mature female	650	750^{1}	750^{4}	
Largest immature female	960	800^{1}	800^4	
Smallest mature male	490	510^{1}	480^{4}	
Largest immature male	670	unknown	520^{4}	
Gestation period (months)	8	unknown	$4.4-7.5^3$	
Size at birth (mm DW)	230-340	$170-180^1$	$200-340^3$	
Fecundity	2-7	3-5 ^{1,2}	$2-10^3$	
Sex ratio of embryos	1:1	unknown	1:13	
Maternal DW vs fecundity relationship	Yes	unknown	Yes^3	
Reproductive cycle	annual	unknown	Biannual ³	

(Table 2). The relationship of these periods is an important feature in the reproductive cycle. The reproductive cycle can be defined by how often a species breeds and consist of two periods: the ovarian cycle (vitellogenesis period) and gestation (Castro, 2009). The length of the reproductive cycle depends on the duration of the ovarian cycle and gestation, and whether the two periods are concurrent or consecutive. The duration of these periods depends on how the mother can sequester energy from the environment and transfer it to the developing oocytes and embryos (Castro, 2009), and the stable conditions in captivity probably allows *D. americana* to reduce the time of gestation and the simultaneous development of oocytes and embryos.

Fecundity estimated in the present study (2-7) was smaller to that documented by Henningsen (2000) of 2-10, but higher to that documented by McEachran & de Carvalho (2002) of 3-5. The relationship between fecundity and maternal disc width was linear for *D. americana* in the present study and captivity (Henningsen, 2000). A significant linear relationship has also been estimated for *D. marmorata* (Capapé & Zaouali, 1995), and non significant for *D. sabina*, *D. sayi*, *D. centroura*, *P. violacea* and *D. chrysonota* (Snelson *et al.*, 1988; Snelson *et al.*, 1989; Capapé, 1993; Hemida *et al.*, 2003; Ebert & Cowley, 2008). This feature is important in demographic analysis, because a larger fecundity is assigned to larger females, and for management purposes the protection

of these females may have a positive effect on the population.

Fecundity in dasyatids ranges from 1 to 7 in the wild and from 1 to 10 in captivity (Table 2). *D. americana* has the highest fecundity reported among captive dasyatid rays (maximum 10), and among the highest fecundity reported in the wild for dasyatids (maximum 7). However, the average fecundity in wild specimens is very low for *D. americana* (2.5 embryos), as in other dasyatids (Table 2). Female dasyatids easily abort embryos because of stress during capture (Struhsaker, 1969; Capapé, 1993; Snelson *et al.*, 1988). It was commonly observed embryos aborted over the sides of fishing boats during landing. For this reason, it is often difficult to estimate fecundity of myliobatiform rays (Smith *et al.*, 2007).

In general, the reproductive parameters of dasyatid species are similar to that of shark species with low biological productivity (Walker, 1998). For example, Smith *et al.* (2008) determine that *D. dipterura* from the Mexican Pacific coast, has low intrinsic growth potential and limited resilience to fishing pressure, and several dasyatids around the world have been classed as Vulnerable or Endangered by the International Union of the Conservation of Nature, indicating that these batoids need more attention in countries where intensive fisheries exploits them directly or incidentally. In particular, the status of the population of *D. americana* is of concern in the southern gulf of Mexico because it is the most landed elasmobranch by

Table 2. The reproductive cycle, gestation period, and fecundity of wild and captive dasyatid rays.

Tabla 2. Ciclo reproductivo, periodo de gestación y fecundidad de dasyatidos de vida libre y cautiverio.

Species	Reproductive cycle	Gestation (month)	Gestation-ovarian cycle	Fecundity (mean)	Author
Dasyatis americana	annual	7-8	consecutive	2-7 (2.5)	Present study
Dasyatis americana	annual	4.4-7.5**	unknown	2-7 (4.2)	Grubbs et al. (2006)
Dasyatis americana*	biannual	4.4-7.5	concurrent	2-10 (4.2)	Henningsen (2000)
Dasyatis centroura	annual?	9-11	concurrent?	4-6	Struhsaker (1969)
Dasyatis centroura	unknown	at least 4	concurrent	2-6 (2.2)	Capapé (1993)
Dasyatis chrysonota	annual	9	concurrent	1-7 (2.8)	Ebert & Cowley (2008)
Dasyatis guttata	biannual	5-6	concurrent	1	Yokota & Lessa (2007)
Dasyatis kuhlii	unknown	unknown	unknown	1	White & Dharmadi (2007)
Dasyatis kuhlii*	biannual	4.5-5.5	concurrent	1	Janse & Schrama (2009)
Dasyatis longa	unknown	10-11	unknown	1-5	Villavicencio-Garayzar et al. (1994)
Dasyatis marianae	biannual	5-6	concurrent	2	Yokota & Lessa (2007)
Dasyatis marmorata	triannual?	3	concurrent	2-4 (2.9)	Capapé & Zaouali (1995)
Dasyatis sabina	annual	4-4.5	consecutive	1-4 (2.6)	Snelson et al. (1988)
Dasyatis sabina	annual	3.5-4	consecutive	1-3 (2.3)	Johnson & Snelson (1996)
Dasyatis sabina	annual	4	n/a	n/a	Tricas et al. (2000)
Dasyatis sayi	annual	12	concurrent	1-6 (3.5)	Snelson et al. (1989)
Dasyatis zugei	unknown	unknown	unknown	1-4 (2.1)	White & Dharmadi (2007)
Pteroplatytrygon violacea*	biannual	2	concurrent?	2-7	Mollet et al. (2002)
Pteroplatytrygon violacea	biannual?	aprox. 4	concurrent	2-7	Hemida et al. (2003)
Pteroplatytrygon violacea	unknown	unknown	concurrent	(3.7)	Véras et al. (2009)
Himantura walga	unknown	unknown	unknown	1-2 (1.4)	White & Dharmadi (2007)

^{*}captive specimens, **from the study of Henningsen (2000).

some small-scale fleets, and its reproductive parameters suggest that this dasyatid has low biological productivity and probably limited resilience to fishing pressure.

ACKNOWLEDGEMENTS

We thank fishermen of the artisanal fleet from San Pedro, Tabasco for allowing us to collect biological material. Special thanks to E. Segura, A. Hernández, A. Romero, W. Arevalo and I. Méndez for their help at field. Funding was provided by Bureau of Forest and Fisheries Development (SEDAFOP) of the Tabasco State Government, project "Marine small-scale fisheries of Tabasco", and also by Fondo Mixto Conacyt-Gobierno del Estado de Tabasco, project "TAB-2007-C09-74601, Biología y pesquería de tiburones y rayas en las costas de Tabasco".

REFERENCES

Bigelow, H.B. & W.C. Schroeder. 1953. Fishes of the western north Atlantic. Memoir Sears Foundation for Marine Research Number I. Part 2. Sawfishes, guitarfishes, skates and rays, 588 pp.

Braccinni, J.M., B.M. Gillanders & T.I. Walker. 2006. Determining reproductive parameters for population assessments of chondrichthyan species with asynchronous ovulation and parturition: piked spurdog (*Squalus megalopus*) as a case study. Mar. Freshw. Res., 57: 105-119.

Capapé, C. 1993. New data on the reproductive biology of the thorny stingray, *Dasyatis centroura* (Pisces: Dasyatidae) from off the Tunisian coasts. Environ. Biol. Fish., 38: 73-80.

Capapé, C. & J. Zaouali. 1995. Reproductive biology of the marbled stingray, *Dasyatis marmorata* (Steindachner, 1892) (Pisces: Dasyatidae) in Tunisian waters (Central Mediterranean). J. Aquaric. Aquat. Sci., 7: 108-119.

Castro, I.J. 2009. Observations on the reproductive cycles of some viviparous North American sharks. Aquaculture, 15: 205-222.

Castro-Aguirre, J.L. & H. Espinosa-Pérez. 1996. Listados faunísticos de México. VII. Catálogo sistemático de las rayas y especies afines de México (Chondrichthyes: Elasmobranchii: Rajiformes: Batoideiomorpha). Universidad Nacional Autónoma de México, México, 75 pp.

- Clark, E. & K. Von Schmidt. 1965. Sharks of the central gulf coast of Florida. Bull. Mar. Sci., 15: 13-83.
- Cuevas-Zimbrón, E., J.C. Pérez-Jiménez & I. Méndez-Loeza. 2011. Spatial and seasonal variation in a target fishery for spotted eagle ray *Aetobatus narinari* in the southern gulf of Mexico. Fish. Sci., 77: 723-730.
- Ebert, D.A. & P. D. Cowley. 2008. Reproduction and embryonic development of the blue stingray, *Dasyatis chrysonota*, in southern African waters. J. Mar. Biol. Assoc. U.K., 89: 809-815.
- Grubbs, D.R., F. Snelson, A. Piercy, R.S. Rosa & M. Furtado. 2006. *Dasyatis americana*. In IUCN 2009. IUCN Red list of threatened species. Version 2009.2. <www.iucnredlist.org>.
- Hamlett, W.C., J.A. Musick, A.M. Eulitt, R.L. Jarrell & M.A. Kelly. 1996. Ultrastructure of uterine trophonemata, accommodation for uterolactation, and gas exchange in the southern stingray, *Dasyatis* americana. Can. J. Zool., 74: 1417-1430.
- Hemida, F., R. Seridji, S. Ennajar, M.N. Bradaï, E. Collier, O. Guélorget & C. Capapé. 2003. New observations on the reproductive biology of the pelagic stingray, *Dasyatis violacea* Bonaparte, 1832 (Chondrichthyes: Dasyatidae) from the Mediterranean Sea. Acta Adriat., 44: 193-204.
- Henningsen, A.D. 2000. Notes on reproduction in the southern stingray, *Dasyatis americana* Chondrichthyes: Dasyatidae), in a captive environment. Copeia, 2000: 826-828.
- Heningsen, A.D., M. Smale, R. Garner & N. Kinnunen. 2004. Reproduction, embryonic development, and reproductive physiology of elasmobranchs. In: M. Smith, D. Warmolts, D. Thoney & R. Hueter (eds.). The elasmobranch husbandry manual: captive care of sharks, rays and their relatives. Biological Survey, Ohio, pp. 227-236.
- Henningsen, A.D. & R.T. Leaf 2010. Observations on the captive biology of the southern stingray. Trans. Am. Fish. Soc., 139: 783-791.
- Janse, M. & J.W. Schrama. 2009. Reproductive cycle, nutrition and growth of captive blue spotted stingray, *Dasyatis kuhlii* (Dasyatidae). J. Mar. Biol. Assoc. U.K., 90: 353-360.
- Johnson, M.R. & F.F. Snelson Jr. 1996. Reproductive life history of the Atlantic stingray, *Dasyatis sabina* (Pisces, Dasyatidae) in the freshwater St. Johns River, Florida. Bull. Mar. Sci., 59: 74-88.
- Mahon, J., F. Chua & P. Newman. 2004. Successful spotted eagle ray (*Aetobatus narinari*) breeding program and details of an assisted birth. Drum and Croaker Special Edition No. 2, High. Irreg. J. Pub. Aqua., pp. 104-107.
- McEachran, J.D. & J.D. Fechhelm. 1998. Fishes of the gulf of Mexico. Vol. 1: Myxiniformes to Gasteros-

- teiformes. University of Texas Press, Austin, 1112 pp.
- McEachran, J.D. & M.R. Carvalho. 2002. Batoid fishes. In: K.E. Carpenter (ed.). The living marine resources of the western central Atlantic. Vol. 1. Introduction, molluscs, crustaceans, hag-fishes, sharks, batoid fishes and chimaeras. FAO Species Identification Guide for Fishery Purposes and American Society of Ichthyologists and Herpetologists Special Publication No. 5. Rome, pp. 507-589.
- Mollet, H.F., G. Cliff, H.L. Pratt Jr. & J.D. Stevens. 2000. Reproductive biology of the female shortfin mako, *Isurus oxyrinchus* Rafinesque, 1810, with comments on the embryonic development of lamnoids. Fish. Bull., 98: 299-318.
- Mollet, H.F., J.M. Ezcurra & J.B. O'Sullivan. 2002. Captive biology of the pelagic stingray, *Dasyatis violacea* (Bonaparte, 1832). Mar. Freshw. Res., 53: 531-541.
- Piñeiro, R., E. Giménez, V. Moreno, R. Burgos & A. Betanzos. 2001. Características térmicas del Banco de Campeche. Cienc. Pesq., 15: 83-87.
- Poder Ejecutivo Federal. 2006. Carta Nacional Pesquera. Publicada en el Diario Oficial de la Federación el 25 de Agosto del 2006. Segunda Sección, Ciudad de México, 128 pp.
- Poder Ejecutivo Federal. 2007. NOM-029-PESC-2006, Pesca responsable de tiburones y rayas. Especificaciones para su aprovechamiento. Publicada en el Diario Oficial de la Federación el 14 de Febrero del 2007, Ciudad de México, 102 pp.
- Secretaría de Agricultura, Ganadería, Desarrollo Rural, Pesca y Alimentación (SAGARPA). 2010. Estadísticas Oficiales de Pesca, Oficina Regional de SAGARPA, Campeche.
- Smith, W.D., G.M. Cailliet & E.M. Melendez. 2007. Maturity and growth characteristics of a commercially exploited stingray, *Dasyatis dipterura*. Mar. Freshw. Res., 58: 54-66.
- Smith, W.D., G.M. Cailliet & E. Cortés. 2008. Demography and elasticity of the diamond stingray, *Dasyatis dipterura*: parameter uncertainty and resilience to fishing pressure. Mar. Freshw. Res., 59: 575-586.
- Snelson, F.F., S.E. Williams-Hooper & T.H. Schmid. 1988. Reproduction and ecology of the Atlantic stingray, *Dasyatis sabina*, in Florida coastal lagoons. Copeia, 1988: 729-739.
- Snelson, Jr.F.F., S.E. Williams-Hooper & T.H. Schmid. 1989. Biology of the bluntnose stingray, *Dasyatis sayi*, in Florida coastal lagoons. Bull. Mar. Sci., 45: 15-25.
- Struhsaker, P. 1969. Observations on the biology and distribution of the thorny stingray, *Dasyatis*

- centroura (Pisces: Dasyatidae). Bull. Mar. Sci., 19: 456-481.
- Tricas, T.C., K.P. Maruska & E. Rasmussen. 2000. Annual cycles of steroid hormone production, gonad development, and reproductive behavior in the Atlantic stingray. Gen. Comp. Endocr., 118: 209-225.
- Villavicencio-Garayzar, C.J., C.C. Downton-Hoffmann & C.M. Melendez. 1994. Tamaño y reproducción de *Dasyatis longus* (Pisces: Dasyatidae), en Bahía Almejas, Baja California Sur, México. Rev. Biol. Trop., 42: 375-377.
- Véras, D.P., I.S.L. Branco, F.H.V. Hazin, C. Wor & M.T. Tolotti. 2009. Preliminary analysis of the reproductive biology of pelagic stingray (*Pteropla-tytrygon violacea*) in the southwestern Atlantic. Collec. Vol. Sci. Pap. ICCAT, 64: 1755-1764.
- Walker, T.I. 1998. Can shark resources be harvested sustainably? A question revisited with a review of shark fisheries. Mar. Freshw. Res., 49: 553-572.

Received: 24 June 2011; Accepted: 19 May 2012

- Walker, T.I. 2005. Reproduction in fisheries science. In: W.C. Hamlett (ed.). Reproductive biology and phylogeny of chondrichthyes sharks, batoids and chimaeras. Volume 3 of Series: Reproductive biology and phylogeny. Science Publishers Enfield, pp. 81-127.
- White, W.T. & Dharmadi. 2007. Species and size composition and reproductive biology of rays (Chondrichthyes, Batoidea) caught in target and nontarget fisheries in eastern Indonesia. J. Fish Biol., 70: 1809-1837.
- Yokota, L. & R.P. Lessa. 2007. Reproductive biology of three ray species: *Gymnura micrura* (Bloch & Schneider, 1801), *Dasyatis guttata* (Bloch & Schneider, 1801) and *Dasyatis marianae* Gomes, Rosa & Gadig, 2000, caught by artisanal fisheries in northeastern Brazil. Cah. Biol. Mar., 48: 249-257.
- Zar, J.H. 1984. Biostatistical analysis. Prentice-Hall, New Jersey, pp. 718.