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## **Probiosis: Concepts and Prospects**

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### **ABSTRACT**

The use of probiotics in animal and human feeding has been a subject of increasing interest both for the academia and the industry, mostly due to their potential positive effects on health and profitability. However, the knowledge on the composition of bacterial gastrointestinal communities in humans and animals, as well as its respective nutritional requirements, is far from being elucidated. Due to the ban of the use of antibiotic growth promoters in animal feeds, there has been an increasing interest on the utilization of probiotics to improve animal intestinal health under commercial settings. However, the possibility of horizontal transference of antibiotic-resistant genes between probiotic bacteria and pathogenic species has become a concern of poultry farmers and consumers around the world. Innovative ideas have emerged, such as the addition of essential oils, spices, and other plant extracts to feeds of monogastric animals to promote intestinal health. These natural compounds are considered ecologically adequate and safe for feeding purposes. This new reality will probably change the direction of research and of the use of additives in poultry production.

### INTRODUCTION

Several infectious diseases were controlled or eradicated after the first third of the 20<sup>th</sup> century, when antibiotics were discovered. However, the overuse and misuse of these compounds in humans or animals have caused the emergence and dissemination of antibiotic-resistant bacteria. The increasing concern with the spread of pathogens from animals to men led the ban on the use of antibiotics in subtherapeutic doses in animal production. Nevertheless, according to experts, despite the general reduction of the use of in-feed antibiotics in poultry production, such ban could increase the incidence of intestinal infections, and consequently the therapeutic use of antibiotics (Castanon, 2007). Methods to control intestinal disorders in poultry and swine based on competitive exclusion, such probiotics, prebiotics or both, are considered safe (Edens, 1997; Anderson et al., 1999; Schneitz, 2005).

Probiotics are necessarily live microorganisms that, when administered to animals in adequate amounts, benefit the host's health, while prebiotics are indigestible feed components that affect the host by selectively stimulating the proliferation or the activity of desirable bacterial populations, such as Bacillus spp (Heyman & Ménard, 2002; Conly & Johnston, 2004; Saad, 2006).

In nature, the genus Bacillus can be isolated from the intestine of penaeid shrimp (Rengpipat et al., 2000), bivalve mollusks (Sugita et al., 1981) and marine fish (Sugita et al., 1998). It was demonstrated that some species of this genus have an inhibitory effect on pathogens (Rengpipat et al., 1998; Balcazar Rojas-Luna, 2007).



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Lactobacillus reuteri culture is sold as a product that allegedly promotes the competitive exclusion, but it seems to be relatively inefficient against enteropathogens (Schneitz, 2005). In broiler chickens, the concept of competitive exclusion initially aimed at reducing Salmonella infection, but it was later expanded to include other enteropathogens (Eimeria, Escherichia coli, Clostridium perfringens, Listeria, and Campylobacter) both in animals and humans (Huang et al., 2002; Dalloul et al., 2003; Rastall, 2004; Nava et al., 2007; Schneitz, 2005; De Vrese & Marteau, 2007). Competitive exclusion has also proven to be efficient in other poultry species, such as turkeys, quails and pheasants (Homma & Shinohara, 2004; Schneitz, 2005). However, it seems that most of these additives, individually or combined, are more efficient in fighting Salmonella than other potentially harmful bacteria, such as E. coli or Clostridium (Blankenship et al., 1993; Kubena et al., 2001; Chambers & Lu, 2002).

Although the term "probiotic" is accepted in Brazil to designate feed additives containing live microorganism cultures, it is not considered adequate by the US and EU legislation, as it conveys a mistaken concept with a commercial appeal. Probiotics are named by the Food and Drug Administration of the US government (FDA, 2009) and the Association of American Feed Control Officials (AAFCO, 1999) as direct-fed microbials (DFM), whereas for the ECC committee, it is simply called microorganisms. Therefore, in this article, these three terms will be indistinctively used. It must be mentioned that the FDA does not allow companies that sell DFM products to make therapeutic allegations, including the establishment of viable bacterial colonies in the intestine, enhancement of animal growth or feed intake, etc. (Alliance Animal Health).

Probiotics in dairy beverages are traditionally considered healthy because of their bacterial content. However, there are few scientific studies on their effects, particularly on healthy populations (Del Campo & Baguero, 2005). This subject remains controversial, even when well-known species, such as Lactobacillus and Bifidobacterium, are taken into account (Zhou et al., 2005). According to Abbot (2004), most of the studies on the efficacy of probiotics are poorly controlled and generate contradictory results in humans. Moreover, their effects on the stimulation of the immune system are poorly documented. Briefly, the role of intestinal bacteria seems to be teaching the immune system how to differentiate between invading bacteria and non-hostile challenges. Some clinical assays indicate that several "dietary bacteria" may aid in the recovery

from diarrheas (Guandalini et al., 2000; Cremonini, et al., 2002; D'Souza et al., 2002; Abbot, 2004) and several inflammatory intestinal diseases (Rembacken et al., 1999; Abbot, 2004). More recently, experiments aiming at developing molecular immunoassay systems for probiotics, based on food immunology, have been performed (Kitazawa et al., 2008).

### Incertitudes as to the intestinal microbiota

Volatile fatty acids production in the ceca and the occupation of the mucosa surface binding sites are the main modes of action of most probiotics (Lan et al., 2005; Donalson et al., 2008). According to Schneitz (2005), the results of studies performed with probiotics suggest that, initially, the protection is predominantly physical, rather than a phenomenon involving the synthesis of volatile fatty acids or other metabolites. In addition to probiotics, other substances that may potentially interact with the intestinal microbiota have been studied as alternatives to replace antibiotics, including enzymes (Jackson et al., 2004; McReynolds et al., 2004; Rosin et al., 2007), prebiotics (Patterson & Burkholder, 2003), and organic acids (Ricke, 2003). These additives have been tested for their modulating effects on the gastrointestinal microbiota, favoring the growth of beneficial bacteria. However, none of these alternatives has provided yet an overall solution that is efficient under different circumstances, such as the case of antibiotics.

No conclusive results have been yet achieved, as several authors have shown that the intestinal ecosystem is much more diverse than it was previously believed (Zhu et al., 2002; Józefiak et al., 2004). Moreover, microbiologists found that the traditional techniques used for microorganism isolation and culture are not able to individually isolate determined microorganisms (Abbot, 2004). In fact, most bacteria cannot be easily isolated from their habitats by the conventional methods commonly used in laboratories, because bacterial communities have specialized members that play different roles and provide vital substrates for other bacteria living in the same community. There is still relatively few information on the in-vivo activity of these organisms (Gabriel et al., 2006). Zhu et al. (2002) suggested that only 10 to 60% of the cecal bacteria can be cultivated using anaerobic techniques. Therefore, the development of strategies for the manipulation of the gastrointestinal microbiota is hindered by the lack of practical analytical instruments that monitor intestinal microorganisms as a whole. Consequently, the lack of knowledge on the role of different bacterial



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species and their growth requirements prevent the possibility of objectively manipulating the intestinal microbiota. Moreover, such prebiotics may to make some genera stronger. An example is Clostridium spp, which generates potentially carcinogenic byproducts, including nitrosamins and cresol, (Blaut & Clavel, 2007) - that may not be relevant for production animals, but that affect breeders and companion animals.

In this context, scientists have adapted techniques that were previously developed to study marine and soil bacteria to answer such questions (Abbot, 2004). Moreover, modern approaches to analyze the structure of bacterial communities using microbial DNA (Zhu et al., 2002; Zhu & Joerger, 2003) have shown that 90% of the gastrointestinal microbiota of broilers consists of unknown species. The genera of more than half of the 640 different bacteria found are still unknown (Zhu et al., 2002; Apajalahti et al., 2004; Gabriel et al., 2006). Even in humans, the best estimates consider that the complete description of the intestinal flora is far from being elucidated (Kullberg, 2008). Efforts to classify and understand the intestinal microbiota will open new research frontiers.

### Substrates and bacterial metabolites

Intestinal microbiota species and their frequency and metabolic activity are influenced by many factors, including environment, diet, presence of aflatoxins, as well as additive type and dosage (Gibson, 1999; Kubena et al., 2001; Smirnov et al., 2005; Blaut & Clavel, 2007; Mountzouris et al., 2007). Bacterial metabolites may be useful or harmful to the host (Rastall, 2004; Gabriel et al., 2006). Experiments performed in animals and in humans suggest that the intestinal microbiota affects the production of colonocyte substrates, such as butyrate (from fiber) and vitamins. The butyrate produced in the colon regulates the differentiation of mucosal cells and reduces apoptosis, which, in turn, controls inflammation and prevents cancer development (Tsukahara et al., 2006; Peña, 2007). Although the inhibition of cancer cells does not have a practical interest for the production of animals with a short life cycle, butyrate is important, because it promotes mucus release and the absorption of water and minerals, and consequently, influences intestinal health (Holtug et al., 1992; Shimotoyodome et al., 2000; Tsukahara et al., 2003; Tsukahara et al., 2006).

Intestinal bacteria synthesize B, E, and K vitamins, but it is believed that only folic acid is available to animals (Coates, 1980<sup>1</sup> apud Gabriel et al., 2006;). The requirements of some vitamins, such as pantothenic

acid, are higher in the presence of the intestinal microflora because bacterial metabolites need to be detoxified. Studies have shown that B-complex vitamins are little absorbed in vitro by the intestine of conventional broilers as compared to germ-free birds (with no intestinal microorganisms); however, these results were not confirmed in vivo. The microflora also has a negative impact on the absorption of fat-soluble vitamins, which requires bile salts (Gabriel et al., 2006).

Intestinal bacteria may impair lipid digestion and alter the digestion of proteins and carbohydrates (Gabriel et al., 2006). The maintenance of this microbial population requires a significant volume of substrates. Bacteria in the small intestine may utilize between 10 and 20% of carbohydrates and amino acids that could be used by the host (Apajalathi et al., 2004). This partially explains the effectiveness of antibiotic growth promoters, which eliminate part of the intestinal microbial population.

Probiotics may be as efficient as antibiotics provided their growth requirements are supplied. For instance, Bacillus spp are transient and do not colonize the intestinal tract. They are vegetative cells, and therefore, they must be regularly supplied to animals. Different bacterial species present different substrate preferences and growth requirements, and the chemical composition and structure of the digesta have a strong influence on the distribution of species in the gastrointestinal microbiota. The knowledge on the nutritional preferences of bacteria may allow shifting the composition of the intestinal microbial community to increase the number of beneficial bacteria. Several studies indicate that oligosaccharides are specific substrates for growing beneficial bacteria (Roberfroid, 2000; Donalson et al., 2008).

Since the concept of prebiotics was introduced, it has attracted scientific and commercial interest. The main characteristic of prebiotics is the selective stimulation of intestinal bacteria growth, contributing to the hosts' health and welfare. This requires prebiotics to be resistant to gastric acidity, bile salts, and hydrolysis by the host's enzymes (Roberfroid, 2007). Moreover, since most bacteria are located in the distal portion of the ileum and large intestine, the compounds required for their growth need to escape absorption by the host (Apajalahti et al., 2004).

Symbiotics, products that contain both probiotics and specific prebiotics that promote the growth of probiotic strains, are considered a good solution in many cases

<sup>&</sup>lt;sup>1</sup> COATES, M.E. (1980) The gut microbiota and growth. In: Growth in animals, (T.L.J. Lawrence ed), pp. 175-188, London: Butterworths.



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(Apajalahti et al., 2004; Cheng et al., 2005; Bomba et al., 2006). In this sense, some scientists are considering the possibility of combining these benefits using genetic engineering (Steidler, 2003; Abbot, 2004). There are evidences that some genetically-modified bacteria may be efficient vehicles for drugs, vitamins, or vaccines, such as genetically-modified Lactococcus lactis that may secrete medicines to fight intestinal disorders (Seegers, 2002; Steidler et al., 2000; Steidler, 2003; Charng et al., 2006).

### Inhabitants of the cecum

Microbial populations of different sizes and complexity are present throughout the digestive tract. The cecal microbiota is the most important fermentative population in non-ruminant animals. The ceca contain the largest amount of bacteria in the gastrointestinal tract of broilers (Barnes et al., 1972, Barnes et al., 1973; Barnes, 1979), and most are obligatory anaerobes. The characterization of the cecal microbiota in poultry started in the beginning of the 1970s (Barnes et al., 1972). It was recently reported that only 10 to 60% of the cecal bacteria can be cultivated using anaerobic techniques (Zhu et al., 2002). As previously discussed, the impossibility of culturing many microorganisms in the laboratory has limited the advance on the knowledge on these populations.

Fecal sampling is the most significant method to study the effect of prebiotics on the microbiota in vitro because it ensures that a representative variety of bacterial species is exposed in the test material (Lifschitz et al., 1990; Flickinger et al., 2002;). Studies on changes of selected genera or species may, therefore, allow establishing whether fermentation is indeed selective. The use of excreta may provide a precise representation of events at the distal colon (Roberfroid 2007). The general problem with the use of excreta samples is the identification of the genus and species of each bacteria present. This has been traditionally accomplished by culture in different selective agars, followed by morphologic and biochemical tests.

# Additives and the intestinal microbiota: a new perspective

During several decades, studies on selection and dissemination of the antibiotic resistance were mainly concentrated in clinically-relevant species (Roe & Pillai, 2003). However, many researchers have recently suggested that commensal bacteria, including lactic bacteria (Mathur & Singh, 2005), may also act as reservoirs of the genes responsible for antibiotic

resistance similar to those found in humans pathogens. Genes of resistance to tetracyclin, erythromycin, and vancomycin were detected and characterized in Lactococcus lacti, Enterococcus, and, recently, in Lactobacillus species isolated from meat and dairy fermented products (Aslim & Beyatli, 2004; Mathur & Singh, 2005).

Several initiatives have been put forward by several organizations around the world to deal with biosafety concerns as to probiotic microorganisms. A large number of probiotic products is being systematically analyzed as to their biosafety, as well as to any possible direct toxic effect (Abbot, 2004).

Innovative ideas emerge every day. Groups around the world have invested in research on new antibiotics and on different modes of action to fight pathogenic bacteria in humans (Bexfield et al., 2008; Rasko et al., 2008; Nigam et al., 2006; Tang et al., 2009; Van Den Berg et al., 2008; Bexfield et al., 2004); however, the possibility of applying such solutions in animals is very remote, and therefore, livestock production relies on the improvement of health conditions to obtain better results (Tabes, 2008).

Low-technology solutions occasionally attract some interest. A reasonable number of vegetable compounds can be used either in synergy or as a replacement to antibiotics in non-ruminant animals (Bomba et al., 2006). Studies that classify the bactericidal activity of essential oils are still lacking. An increasing number of research studies have validated the utilization of these compounds in broilers, fulfilling the expectation of the most passionate ecologists. It is not difficult to imagine that such concerns will also influence markets that increasingly demand the so-called "natural" products, which nomenclature is often more ideological than technical.

### FINAL CONSIDERATIONS

The common problem in the evaluation of the utilization of probiotic and prebiotic products in diets is that several factors influence their physiologic and microbiologic effects on the host, including the age of animals to which there are supplied, feedstuffs included in the formulation of feeds, in addition to the fermentative characteristics and structural differences among products. Moreover, many obstacles still prevent the full understanding of their in-vivo effects. This is why the experiments on the effects of dietary supplementation of probiotics, individually or combined to other additives, have yielded different results.



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Some studies report non-significant trends of performance improvement, in spite of the significant health effects under challenge conditions. However, it is important to stress that in several experiments on the efficacy of these additives challenges are not controlled or it was not possible to determine if a challenge was indeed present. Errors in the design and performance of experiments and improper facilities, or merely careless management, may cause cross-contamination of experimental units belonging to different treatments, and this lack of control may increase the experimental error or confuse observations, making their results unreliable.

Experimental studies developed in Brazil between 1995 and 2005 were carefully reviewed to evaluate the effect of probiotics on broiler performance (Faria Filho et al., 2005). Results demonstrated that individual comparisons of commercial probiotics added to feeds as growth promoters showed trends of improvement, but no statistical significance for most evaluated parameters. However, when the analysis was performed considering data of all experiments, there were beneficial effects of the utilization of probiotics. Those authors argued that this was due to a higher number of observations, and consequently, lower confidence interval.

The optimum levels of inclusion of several of these commercial products still need to be determined, including under laboratory conditions. In the field, this is more difficult because previous health status, diet composition, and environmental factors should be taken into account. To date, the amount of these products that eventually reach the posterior segment of the intestinal tract in a viable form is not known.

The current control of foodborne health challenges posed by poultry products and the concern with the potential horizontal transference of antibiotic-resistance genes between probiotic microorganisms and pathogenic bacteria may change the focus of research and the use of "live dietary microorganisms" in animals.

Plant extracts, essential oils, and other phytogenic products have been increasingly evaluated as alternatives to the promotion of intestinal health and may prove to be safer to human health. Such studies are being performed worldwide, but bactericidal activity of these plant ingredients still needs to be further elucidated. Their effects on the gastrointestinal microbiota need to be determined in order to avoid the mistakes that occurred with antibiotics, and perhaps will give a new direction for the utilization of additives in non-ruminants diets in a near future. It must be stressed that the mode of action of these additives needs to be elucidated to

explain why they have positive effects under some circumstances and not in others.

### **REFERENCES**

Frank CP. Plant food, feed and grain division. Georgia: Official Publication Association of American Feed Control Officials; 1999.

Alliance Animal Health. Direct fed microbial: application and usage [cited 2010 jul]. Available from: http://www.admani.com/AllianceAnimalHealth/Tech%20Bulletins/Direct%20Fed%20 Microbials.htm

Abbott A. Microbiology: gut reaction. Nature 2004; 427:284-286.

Anderson RC, Stanker LH, Young CR, Buckley SA, Genovese KJ, Harvey RB, Deloach JR, Keith NK, Nisbet DJ. Effect of competitive exclusion treatment on colonization of early-weaned pigs by Salmonella serovar choleraesuis. Swine Health and Production 1999; 7(4):155-160.

Apajalahti J, Kettunen A, Graham H. Characteristics of the gastrointestinal microbial communities, with special reference to the chicken. World's Poultry Science Journal 2004; 60(2):223-232.

Aslim B, Beyatli Y. Antibiotic resistance and plasmid DNA contents of Streptococcus thermophilus strains isolated from Turkish yogurts. Turkish Journal of Veterinary and Animal Sciences 2004; 28:257-263.

Balcázar JL, Rojas-Luna T. Inhibitory activity of probiotic Bacillus subtilis UTM 126 against vibrio species confers protection against vibriosis in juvenile shrimp (Litopenaeus vannamei). Current Microbiology 2007; 55(5):409-412.

Barnes EM. The intestinal microflora of poultry and game birds during life and after storage. Journal of Applied Bacteriology 1979; 46:407-419.

Barnes EM, Mead GC, Griffiths NM. The microbiology and sensory evaluation of pheasants hung at 5, 10, and 15 °C. British Poultry Science 1973; 14:229-240.

Barnes EM, Mead GC, Barnum DA, Harry EG. The intestinal flora of the chicken in the period 2-6 weeks of age, with particular reference to the anaerobic bacteria. British Poultry Science 1972; 13:311-326.

Bexfield A, Bond AE, Roberts EC, Dudley E, Nigam Y, Thomas S, Newton RP, Ratcliff NA. The antibacterial activity against MRSA strains and other bacteria of a <500 Da fraction from maggot excretions/secretions of Lucilia sericata (Diptera: Calliphoridae). Microbes and Infection 2008; 10:325-333.

Bexfield A, Nigam Y, Thomas S, Ratcliffe NA. Detection and partial characterization of two antibacterial factors from the excretions/ secretions of the medicinal maggot Lucilia sericata and their activity against methicillin-resistant Staphylococcus aureus (MRSA). Microbes and Infection 2004; 6:1297-1304.

Blake DP, Hillman K, Fenlon DR. The use of a model ileum to investigate the effects of novel and existing antimicrobials on indigenous porcine gastrointestinal microflora: using vancomycin as



## **Probiosis: Concepts and Prospects**

an example. Animal Feed Science and Technology 2003; 103:123-139.

Blankenship LC, Bailey JS, Cox NA, Stern NJ, Brewer R, Williams O. Two-step mucosal competitive exclusion flora treatment to diminish Salmonellae in commercial broiler chickens. Poultry Science 1993; 72:1667-1672.

Blaut M, Clavel T. Metabolic diversity of the intestinal microbiota: implications for health and disease. Journal of Nutrition 2007; 137:751S-755S.

Bomba A, Jonecová Z, Košcová J, Nemcová R, Gancarcíková S, Mudronová D, Sciranková L, Buleca V, Lazar G, Pošivák J, Kaštel R, Mareková M. The improvement of probiotics efficacy by synergistically acting components of natural origin: a review. Biologia 2006; 61(6):729-734.

Castanon JIR. History of the use of antibiotic as growth promoters in European poultry feeds. Poultry Science 2007; 86:2466-2471.

Chambers JR, Lu X. Probiotics and maternal vaccination for Salmonella control in broiler chickens. Journal of Applied Poultry Research 2002; 11:320-327.

Charng Y-C, Lin C-C, HSU C-H. Inhibition of allergen-induced airway inflammation and hyperreactivity by recombinant lactic-acid bacteria. Vaccine 2006; 24:5931-5936.

Cheng IC, Shang H-F, LIN T-F, Wang T-H, Lin H-S, Lin S-H. Effect of fermented soy milk on the intestinal bacterial ecosystem. World Journal of Gastroenterology 2005; 11(8):1225-1227.

Conly JM, Johnston BL. Coming full circle: From antibiotics to probiotics and prebiotics. The Canadian Journal of Infectious Diseases & Medical Microbiology 2004; 15(3):161-163.

Cremonini F, DI Caro S, Nista EC, Bartolozzi F, Capelli G, Gasbarrini G, Gasbarrini A. Meta-analysis: the effect of probiotic administration on antibiotic-associated diarrhea. Alimentary Pharmacology & Therapeutics 2002; 16:1461-1467.

D'souza AL, Rajkumar C, Cooke J, Bulpitt CJ. Probiotics in prevention of antibiotic associated diarrhoea: meta-analysis. British Medical Journal 2002; 324:1361-1366.

Dalloul RA, Lillehoj HS, Shellem TA, Doerr JA. Enhanced mucosal immunity against Eimeria acervulina in broilers fed a lactobacillusbased probiotic. Poultry Science 2003; 82:62-66.

De Vrese M, Marteau PR. Probiotics and Prebiotics: Effects on Diarrhea. Journal of Nutrition 2007; 137:803S-811.

Del Campo R, Boquero F. Probióticos y alimentos: estudios basados en la evidencia. Revista Española de Quimioterapia 2005; 18(1):75 - 78.

Donalson LM, Kim WK, Chalova VI, Herrera P, Mcreynolds JL, Gotcheva VG, Vidanovic' D, Woodward CL, Kubena LF, Nisbet DJ, Ricke SC. In vitro fermentation response of laying hen cecal bacteria to combinations of fructooligosaccharide prebiotics with alfalfa or a layer ration. Poultry Science 2008; 87:1263-1275.

Edens FW, Parkhurst CR, Casas IA, Dobrogosz WJ. Principles of ex ovo competitive exclusion and in ovo administration of Lactobacillus reuteri. Poultry Science 1997; 76:179-196.

Faria Filho DE, Torres KAA, Faria DE, Campos DMB, Rosa PS. Probiotics for broiler chickens in Brazil:Systematic review and meta-Analysis. Brazilian Journal of Poultry Science 2006; 8(2):89-98.

Food and Drug Administration. Inspections, compliance, enforcement, and criminal investigations. In: CPG Sec. 689.100 Direct-Fed Microbial Products [cited 2010 jul]. Available from: http://www.fda.gov/ICECI/ComplianceManuals/CompliancePolicyGuidanceManual/ucm074707.htm

Flickinger EA, Hatch TF, Wofford RC, Grieshop CM, Murray SM, Fahey Jr, GC. In vitro fermentation properties of selected fructooligosaccharide-containing vegetables and in vivo colonic microbial populations are affected by the diets of healthy human infants. Journal of Nutrition 2002;132:2188-2194.

Gabriel I, Lessire M, Mallet S, Guillot JF. Microbiota of the digestive tract: critical factors and consequences for poultry. World's Poultry Science Journal 2006; 62:499-511.

Gibson GR. Dietary modulation of the human gut microbiota using the prebiotics oligofructose and inulin. Journal of Nutrition 1999; 129:438S-1441S.

Guandalini S, Pensabene L, Zikri MA, Dias JA, Casali LG, Hoekstra H, Kolacek S, Massar K, Micetic-Turk D, Papadopoulou A, Sousa JS, Sandhu B, Szajewska H, Weizman Z. Lactobacillus GG administered in oral rehydration solution to children with acute diarrhea: a multicenter European trial. Journal of Pediatric Gastroenterology and Nutrition 2000; 30:54-60.

Heyman M, Ménard S. Probiotic microorganisms: how they affect intestinal pathophysiology. Cellular and Molecular Life Sciences 2002; 59:1151-1165.

Holtug K, Rasmussen HS, Mortensen PB. An in vitro study of short-chain fatty acid concentrations, production and absorption in pig (Sus scrofa) colon. Comparative Biochemistry and Physiology 1992; 103A:189-197.

Homma H, Shinohara T. Effects of probiotic (Bacillus cereus toyoi) on abdominal fat accumulation in the Japanese quail (Coturnix japonica). Animal Science Journal 2004; 75:37-41.

Huang J, Bousvaros A, Lee JW, Diaz A, Davidson EJ. Efficacy of probiotic use in acute diarrhea in children: a meta-analysis. Digestive Diseases and Sciences 2002; 47(11):2625-2634.

Jackson ME, Geronian K, Knox A, Mcnab J, Mccartney E. A doseresponse study with the feed enzyme beta-mannanase in broilers provided with corn-soybean meal based diets in the absence of antibiotic growth promoters. Poultry Science 2004; 83:1992-1996.

Józefiak D, Rutkowski A, Martin SA. Carbohydrate fermentation in the avian ceca: a review. Animal Feed Science and Technology 2004;113(1):1-15.

Kitazawa H, Tohno M, Shimosato T, Saito T. Development of



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molecular immunoassay system for probiotics via toll-like receptors based on food immunology. Animal Science Journal 2008; 79:11-21.

Kubena LF, Bailey RH, Byrd JA, Young CR, Corrier DE, Stanker LH, Rottinghaus GE. Cecal volatile fatty acids and broiler chick susceptibility to Salmonella typhimurium colonization as affected by aflatoxins and T-2 toxin. Poultry Science 2001; 80:411-417.

Kubena LF, Byrd JA, Young CR, Corrier DE. Effects of tannic acid on cecal volatile fatty acids and susceptibility to Salmonella typhimurium colonization in broiler chicks. Poultry Science 2001; 80:1293-1298.

Kullberg MC. Soothing intestinal sugars. Nature 2008; 453:602-604.

Lan Y, Verstegen MWA, Tamminga S, Williams BA. The role of the commensal gut microbial community in broiler chickens. World's Poultry Science Journal 2005; 61:95-104.

Lee DJ, Drongowski RA, Coran AG, Harmon CM. Evaluation of probiotic treatment in a neonatal animal model. Pediatric Surgery International 2000; 16:237-242.

Lifschitz CH, Wolin MJ, Reeds PJ. Characterization of carbohydrate fermentation in feces of formula-fed and breast-fed infants. Pediatric Research 1990; 27:165-169.

Mathur S, Singh R. Antibiotic resistance in food lactic acid bacteria: a review. International Journal of Food Microbiology 2005; 105(3): 281-295.

McReynolds JL, Byrd JA, Moore RW, Anderson RC, Poole TL, Edrington TS, Kubena LF, Nisbet DJ. Utilization of the nitrate reductase enzymatic pathway to reduce enteric pathogens in chickens. Poultry Science 2004; 83:1857-1860.

Mountzouris KC, Tsirtsikos P, Kalamara E, Nitsch S, Schatzmayr G, Fegeros K. Evaluation of the efficacy of a probiotic containing Lactobacillus, Bifidobacterium, Enterococcus and Pediococcus strains in promoting broiler performance and modulating cecal microflora composition and metabolic activities. Poultry Science 2007; 86:309-317.

Nava GM, Bielke LR, Callaway TR, Castaneda MP. Probiotic alternatives to reduce gastrointestinal infections: the poultry experience. Animal Health Research Reviews 2007; 6(1):105-118.

Nigam Y, Bexfield A, Thomas S, Ratcliffe NA. Maggot therapy: the science and implication for Cam part I-history and bacterial resistance. eCAM 2006; 3:223-227.

Patterson JA, Burkholder KM. Application of prebiotics and probiotics in poultry production. Poultry Science 2003; 82:627-631.

Peña AS. Intestinal flora, probiotics, prebiotics, synbiotics and novel foods. Revista Española de Enfermedades Digestivas 2007; 99(11): 653-658.

Rasko DA, Moreira CG, Li DR, Reading NC, Ritchie JM, Waldor MK, Williams N, Taussig R, Wei S, Roth M, Hughes DT, Huntley JF, Fina MG, Falck JR, Sperandio V. Targeting OseC signaling and virulence for antibiotic development. Science 2008; 321(5892):1078-1080.

Rastall RA. Bacteria in the gut: friends and foes and how to alter the balance. Journal of Nutrition 2004; 134:2022S-2026S.

Rembacken BJ, Snelling AM, Hawkey PM, Chalmers DM, Axon AT. Non-pathogenic Escherichia coli versus mesalazine for the treatment of ulcerative colitis: a randomised trial. Lancet 1999; 354:635-639.

Rengpipat S, Phianphak W, Piyatiratitivorakul S, Menasaveta P. Effects of a probiotic bacterium in black tiger shrimp Penaeus monodon survival and growth. Aquaculture 1998; 167:301-313.

Rengpipat S, Rukpratanporn S, Piyatiratitivorakul S, Menasaveta P. Immunity enhancement in black tiger shrimp (Penaeus monodon) by a probiont bacterium (Bacillus S11). Aquaculture 2000; 191:271-288.

Ricke SC. Perspectives on the use of organic acids and short chain fatty acids as antimicrobials. Poultry Science 2003; 82:632-639.

Roberfroid M. Prebiotics and probiotics: are they functional foods? American Journal of Clinical Nutrition 2000; 71(suppl):1682S-7S.

Roberfroid M. Prebiotics: the concept revisited. Journal of Nutrition 2007; 137:830S-837S.

Roe MT, Pillai SD. Monitoring and identifying antibiotic resistance mechanisms in bacteria. Poultry Science 2003; 82:622-626.

Rosin EA, Blank G, Slominski BA, Holley RA. Enzyme supplements in broiler chicken diets: in vitro and in vivo effects on bacterial growth. Journal of the Science of Food and Agriculture 2007; 87(6): 1009-1020.

Saad SMI. Probióticos e prebióticos: o estado da arte. Brazilian Journal of Pharmaceutical Sciences 2006; 42(1):1-16.

Schneitz C. Competitive exclusion in poultry-30 years of research. Food Control 2005; 16(8):657-667.

Seegers JFML. Lactobacilli as live vaccine delivery vectors: progress and prospects. Trends in Biotechnology 2002; 20(12):508-515.

Shimotoyodome A, Meguro S, Hase T, Tokimitsu I, Sakata T. Short chain fatty acids but not lactate or succinate stimulate mucus release in the rat colon. Comparative Biochemistry and Physiology 2000; 125A:525-531.

Steidler L. Genetically engineered probiotics. Best Practice & Research Clinical Gastroenterology 2003; 17(5):861-876.

Steidler L, Hans W, Schotte L, Neirynck S, Obermeier F, Falk W, Fiers W, Remaut E. Treatment of murine colitis by Lactococcus lactis secreting interleukin-10. Science 2000; 289(5483): 1352-1355.

Sugita H, Hirose Y, Matsuo N, Deguchi Y. Production of the antibacterial substance by Bacillus sp. strain NM12, an intestinal bacterium of Japanese coastal fish. Aquaculture 1998; 165:269-280.

Sugita H, Tanaami H, Kobashi T, Deguchi Y. Bacterial flora of coastal bivalves. Bulletin of the Japanese Society of Scientific Fisheries 1981; 47:655-661.

Sun Y, Liu Z, Zeng J, Zhou Y, Yu X, Jia J. Helicobacter pylori protein



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response to human bile stress. Journal of Medical Microbiology 2008; 57:151-158.

Tabes G. The bacteria fight back. Science 2008; 321(5887):356-361.

Tang Y-L, Shia Y-H, Zhaoa W, Haoa G, Le G-W. Discovery of a novel antimicrobial peptide using membrane binding-based approach. Food Control 2009; 20(2):149-156

Tsukahara T, Hashizume K, Koyama H, Ushida K. Stimulation of butyrate production through the metabolic interaction among lactic acid bacteria, Lactobacillus acidophilus, and lactic acid-utilizing bacteria, Megasphaera elsdenii, in porcine cecal digesta. Animal Science Journal 2006; 77:454-461.

Tsukahara T, Iwasaki Y, Nakayama K, Ushida K. Stimulation of butyrate production in the large intestine of weaning piglets by dietary fructooligosaccharides and its influence on the histological variables of the large intestinal mucosa. Journal of Nutritional Science and Vitaminology 2003; 49:414-421.

Van Den Berg MA, Albang R, Albermann K, Badger JH, Daran J-M, Driessen AJM. et al. Genome sequencing and analysis of the filamentous fungus Penicillium chrysogenum. Nature Biotechnology 2008; 26:1161-1168.

White DG, Zhao S, Singh R, Mcdermot PF. Antimicrobial resistance among gram-negative foodborne bacterial pathogens associated with foods of animal origin. Foodborne Pathogens and Disease 2004; 1(3):137-152.

Zhou JS, Pillidge CJ, Gopal PK, Gill HS. Antibiotic susceptibility profiles of new probiotic Lactobacillus and Bifidobacterium strains. International Journal of Food Microbiology 2005; 98:211-217.

Zhu XY, Joerger, RD. Composition of microbiota in content and mucus from cecae of broiler chickens as measured by fluorescent in situ hybridization with group-specific, 16S rRNA-targeted oligonucleotide probes. Poultry Science 2003; 82:1242-1249.

Zhu XY, Zhong T, Pandya Y, Joerger RD. 16S rRNA-based analysis of microbiota from the cecum of broiler chickens. Applied and Environmental Microbiology 2002; 68(1):124-137.