



Revista Brasileira de Ciência Avícola

ISSN: 1516-635X

revista@facta.org.br

Fundação APINCO de Ciência e
Tecnologia Avícolas
Brasil

Sousa, JPL de; Albino, LFT; Vaz, RGMV; Rodrigues, KF; Da Silva, GF; Renno, LN;
Barros, VRSM; Kaneko, IN
The Effect of Dietary Phytase on Broiler Performance and Digestive, Bone, and Blood
Biochemistry Characteristics
Revista Brasileira de Ciência Avícola, vol. 17, núm. 1, enero-marzo, 2015, pp. 69-76
Fundação APINCO de Ciência e Tecnologia Avícolas
Campinas, Brasil

Disponível em: <http://www.redalyc.org/articulo.oa?id=179738404011>

- Como citar el artículo
- Número completo
- Más información del artículo
- Página de la revista en redalyc.org

redalyc.org

Sistema de Información Científica
Red de Revistas Científicas de América Latina, el Caribe, España y Portugal
Proyecto académico sin fines de lucro, desarrollado bajo la iniciativa de acceso abierto



■ Author(s)

Sousa JPL de^I
Albino LFT^{II}
Vaz RGMV^{III}
Rodrigues KF^{III}
Da Silva GF^{III}
Renno LN^{II}
Barros VRSM^{II}
Kaneko IN^{IV}

^I Instituto Federal de Educação, Ciência e Tecnologia do Tocantins, Dianópolis, Tocantins, Brazil

^{II} Universidade Federal de Viçosa, Department of Animal Science, Viçosa, Minas Gerais, Brazil

^{III} Universidade Federal do Tocantins, School of Veterinary Medicine and Animal Science, Department of Animal Science, Araguaína, Tocantins, Brazil

^{IV} Universidade Estadual de Maringá, Department of Animal Science, Maringá, Paraná, Brazil

Part of the Ph.D. thesis of the first author

■ Mail Address

Corresponding author e-mail address
Sousa JPL de
Rua 132A, Qd45, lote 05, Setor Sul, CEP
74093-220, Goiânia, GO
E-mail: jp-zoot24@yahoo.com.br

■ Keywords

Antagonism, digestible amino acid, egg components, Haugh units.

The Effect of Dietary Phytase on Broiler Performance and Digestive, Bone, and Blood Biochemistry Characteristics

ABSTRACT

The dietary inclusion of phytase increases nutrient and energy bioavailability for broilers. The effect of phytase increases nutrients and energy bioavailability for either the objective of this experiment was to evaluate the influence of nutrient and energy reduction in diets supplemented with phytase on the performance, gastrointestinal pH, organ and bone composition, and blood biochemistry of broilers between eight and 21 days of age. In the study, 1.120 male Cobb 500® broilers, with 161±1g average weight, were used. At eight days of age, birds were distributed according to a completely randomized experimental design with seven treatments in a 3x2+1 factorial arrangement with eight replicates of 20 broiler each. Treatments corresponded to reduction of calcium (Ca) and phosphorus (P), amino acids and energy, or reduction of Ca, P, amino acids and energy; supplementation or not of phytase; and a positive control treatment. Broiler fed the diet with reduced Ca and P levels and phytase supplementation presented the best performance of all groups. The diet with reduced amino acid and energy levels and phytase addition reduced gizzard and proventriculus pH. Dietary Ca and P reduction increased relative liver and heart weights, as well as albumin blood levels. The bones of broilers fed phytase-supplemented diets presented higher ash content.

Key words: poultry production, enzymes, intestinal pH, bone ash, calcium, available phosphorus, extra-phosphoric effect

INTRODUCTION

The increasing broiler production around the world has led to the application of nutritional strategies that improve nutrient utilization. The supplementation of exogenous enzymes to corn- and soybean meal-based broiler diets allows supplying nutrient deficiencies and to reduce endogenous losses, thereby optimizing performance, particularly during early life.

The dietary supplementation of the exogenous phytase reduces nutrient variability of feedstuffs, and counteracts the antinutritional effects of phytate, increasing the accuracy of feed formulation.

Exogenous phytase is included in feed formulations not only to reduce phosphorus supplementation, but also to release minerals, particularly calcium, as well as amino acids and carbohydrates by the hydrolysis of phytate, improving nutrient utilization (Oluyinka *et al.*, 2007; Slominski, 2011). However, nutrient utilization can also be affected by other factors, including dietary calcium, phosphorus, protein, and energy levels, intestinal pH, environmental temperature, etc.

Dietary calcium content may negatively influence phosphorus utilization, particularly phytic phosphorus, due to the formation of insoluble complexes with calcium in the digestive tract, counteracting



the effect of phytase, which is not able to hydrolyze such complexes (Leeson *et al.*, 1996). Nelson *et al.* (1964) reported that dietary calcium levels higher than 0.70% at pH 6.0 promote the reaction of calcium with phytic acid, resulting in calcium phytate, which precipitates and cannot be broken down by phytase, consequently reducing phosphorus bioavailability. High dietary calcium levels change the pH of the upper sections of the digestive tract, inactivating phytase. Phytase activity is optimal at pH between 5.0 and 6.5, and it is reduced when pH is lower or equal to 3.0 (Casey & Walsh, 2004).

Broilers fed diets with low inorganic calcium and phosphorus levels have higher capacity to hydrolyze phytate than those fed high levels of these minerals (Denbow *et al.*, 1995). Therefore, minimal calcium and phosphorus levels need to be maintained in the diet; however, their interaction should not be overlooked, particularly considering phytase activity (Qian *et al.*, 1997).

Protein utilization and amino acid absorption may also be influenced by dietary phytic acid levels (Lehnen *et al.*, 2011; Walk *et al.*, 2012). These authors observed that phytic acid is a potent chelating agent. Its negative charges react with the positive charges of some amino acids (lysine, arginine, histidine), some proteins (including those involved in protein digestion, such as pepsin and trypsin), and carbohydrates (α -amylase), forming insoluble complexes, thereby reducing their availability and digestibility (Maenz, 2001) and consequently affecting metabolism and organ biometry.

Therefore, the objective of this study was to evaluate the supply of feeds containing reduced amino acid, energy, calcium, and phosphorus levels and supplemented or not with phytase on the performance, gastrointestinal pH, relative organ weight, bone mineral content, and blood biochemistry of broilers.

MATERIALS AND METHODS

The experiment was carried out at the Poultry Sector of the Department of Animal Science of the Federal University of Viçosa. In the study, 1,120 male Cobb 500® broilers, with 161 ± 1 g average weight and between eight and 21 days of age, were used. A completely randomized experimental design in a 3x2+1 factorial arrangement (Ca and P reduction, energy and amino acid reduction, and Ca, P, energy, and amino acid reduction; supplementation or not with phytase; and a positive control treatment) with eight replicates of 20 broiler each. Until eight days of age, all broilers

were reared in a conventional poultry house according to the recommendations of the genetic company's (Manual, 2009). On day 8, broiler were individually weighed and distributed into 56 pens, measuring 1,5 m² and equipped with a nipple drinker and a tube feeder each.

The increase in nutrient and energy bioavailability obtained with phytase supplementation was considered when formulating the experimental diets (Table 1). The phytase product was derived from *Escherichia coli*, and presented 500 FTU/kg of feed. It was added at a dose of 100g/ton of feed.

Table 1 – Nutritional matrix of the bacterial phytase applied*

Nutrients and energy	500 FTU/kg feed
Crude protein, %	0.420
Metabolizable energy, kcal/kg	52.00
Calcium, %	0.165
Phosphorus, %	0.150
Lysine, %	0.017
Methionine, %	0.004
Threonine, %	0.030
Arginine, %	0.030

*Values informed by the manufacturer

The experimental diets were based on corn and soybean meal and were formulated to supply the nutritional requirements of male broilers, as recommended by Rostagno *et al.* (2011) for the starter phase.

Birds, feed offer, and feed residue were weighed per pen to calculate weight gain, feed intake, and feed conversion ratio. Mortality was also recorded to correct performance data.

On day 21, two broiler per stall, with body weight close to the average stall weight (± 100 g), were selected, identified, and had their blood collected by heart puncture. Blood was placed in test tubes, centrifuged for 5 min at 1,500 rpm, and the serum was then placed in duly identified Eppendorf tubes and immediately frozen until analyses. Serum calcium, phosphorus, albumin, and total protein were analyzed using specific commercial kits.

The selected birds were identified and sacrificed according to the norms of the Committee of Ethics for the Use of Animals of the Department of Animal Science of the Federal University of Viçosa process nº 76/2013. Broilers were stunned, sacrificed by neck dislocation, and their gastrointestinal tract (GIT) with the digesta was aseptically removed. Before removing



Table 2 – Composition of the experimental diet fed to broilers between 8 and 21 days of age (% as fed)

Ingredients	Control	Treatment with reduced levels		
		Ca and P	AA and ME	Ca, P, AA, and ME
Corn	58.81	60.3	61.39	62.89
Soybean meal	35.04	34.76	33.70	33.41
Soybean oil	2.30	1.79	1.07	0.57
Limestone	1.06	1.15	1.06	1.15
Dicalcium phosphate	1.47	0.66	1.48	0.66
Salt	0.48	0.48	0.48	0.48
L-Lysine HCL 79%	0.16	0.17	0.18	0.19
DL-Methionine 99%	0.26	0.26	0.23	0.23
L-threonine 98%	0.03	0.03	0.01	0.02
Mineral supplement ¹	0.11	0.11	0.11	0.11
Vitamin supplement ²	0.11	0.11	0.11	0.11
Choline chloride 60%	0.10	0.10	0.10	0.10
Salinomycin, 12%	0.06	0.06	0.06	0.06
Antioxidant	0.01	0.01	0.01	0.01
Total	100	100	100	100
Calculated values				
Crude protein, %	20.8	20.8	20.38	20.38
Metabolizable energy, kcal/kg	3.000	3.000	2.948	2.948
Calcium, %	0.86	0.70	0.86	0.70
Available phosphorus, %	0.38	0.23	0.38	0.23
Digestible lysine, %	1.14	1.14	1.12	1.12
Total lysine (%)	1.24	1.24	1.22	1.22
Digestible methionine + cystine (%)	0.82	0.82	0.78	0.78
Total methionine + cysteine, %	0.90	0.90	0.86	0.86
Digestible threonine (%)	0.74	0.74	0.71	0.71
Total threonine, %	0.85	0.85	0.81	0.81
Digestible arginine, %	1.31	1.31	1.28	1.28

Supplement/kg feed: ¹Manganese 77.0 mg; iron - 55.0 mg; zinc - 71.5 mg; copper - 11.0 mg; cobalt- 1.0 mg; iodine- 1.1 mg; selenium- 0.33 mg.²Vitamin A - 8250 UI; Vitamin D3 - 2090 UI; Vitamin E - 31 UI; Vitamin B1 - 2.2 mg; Vitamin B6 - 3.08 mg; pantothenic acid - 11.0 mg; biotin- 0.077 mg; Vitamin K3 - 1.65 mg; folic acid - 0.770 mg; nicotinic acid - 33.0 mg; Vitamin B12 - 0.013 mg.

the digesta, the proventriculus, gizzard, duodenum (from the caudal end of the gizzard until the end of the duodenal loop), jejunum (from the end of the duodenum until Meckel's diverticulum), ileum (from Meckel's diverticulum until the ileal-cecal junction), and the ceca were isolated to prevent the digesta from mixing. Then, the pH was measured using a digital pHmeter gently introducing the sterilized fine tip of the glass electrode in each GIT section. As recommended by Pang & Applegate (2007), pH was measured twice in each section and the average of the two measurements was used for the statistical analysis. Subsequently, the intestine, gizzard, liver, heart, bursa, and spleen were removed. The intestine was measured using a measuring tape.

The left tibia bone was collected, dehydrated in ethanol, and de-fatted in petroleum ether in Soxhlet apparatus for four hours. Bones were then burnt in a muffle at 600°C to determine ash content, according to Gardiner *et al.* (1961). Ashes were then used to prepare a mineral solution by dissolving the ashes by digestion on a heated plate for four hours at 200°C. Calcium content in the ashes was determined by atomic absorption spectrophotometry and phosphorus content was determined by colorimetric spectrophotometry, according to the methods described by Detmann.

Data were submitted to Shapiro Wilk's normality test and then to analysis of variance using the General Linear Model (PROC GLM) procedures of SAS statistical package (1998). Means were compared by orthogonal contrasts. Each contrast was independently tested by the F test a 10% probability level.

The following orthogonal contrasts were analyzed: contrast C₁, comparing diets with reduced calcium (Ca) and phosphorus (P) levels supplemented or not with phytase; contrast C₂, comparing diets with reduced amino acid (AA) and energy (ME) contents supplemented or not with phytase; contrast C₃, comparing diets with reduced AA, ME, Ca and P levels supplemented or not with phytase; contrast C₄, comparing the positive-control diet and the diet with reduced AA, ME, Ca and P levels and not supplemented with phytase;

contrast C₅, comparing the positive-control diet and the diet with reduced AA, ME, Ca and P levels and supplemented with phytase, and contrast C₆, comparing the diets with reduced nutrient levels supplemented or not with phytase.

RESULTS

Contrast C₁ shows that the performance of broilers fed the diet with reduced Ca and P levels and supplemented with phytase was better (P<0.01) than those fed the same diet with no phytase supplementation, with improvements of 4.40, 11.04, and 7.14% in feed intake (FI), weight gain (WG), and feed conversion ratio (FCR), respectively (Table 3).



Table 3 – Performance of broilers fed starter diets (8-21 days of age) with reduced nutrient and energy levels and supplemented or not with phytase.

Parameter	Treatment with reduced levels							(CV ⁵ (%))	Orthogonal contrasts ⁶					
	CP ¹	Ca and P ²		AA and ME ³		Ca, P, AA, and ME ⁴								
		W/o phytase	With phytase	W/o phytase	With phytase	W/o phytase	With phytase							
									C ₁	C ₂	C ₃	C ₄	C ₅	C ₆
FI (g)	1085	1064	1113	1100	1094	1064	1110	3.01	*	NS	*	NS	NS	NS
WG (g)	782	725	815	765	763	684	741	4.18	*	NS	*	*	NS	*
FCR (g/g)	1.390	1.471	1.366	1.439	1.437	1.559	1.503	4.46	*	NS	***	*	NS	*

¹Positive control = 3,000 kcal/kg ME, 20.8% CP, 0.38% P and 0.86% Ca; ²Ca and P = 3,000 kcal/kg ME, 20.8% CP, 0.23% P and 0.69% Ca; ³AA and ME = 2,948 kcal/kg ME, 20.38% CP, 0.38% P and 0.86% Ca; ⁴Ca, P, AA and ME = 2,948 kcal/kg ME, 20.38% CP, 0.23% P and 0.69% Ca; ⁵coefficient of variation; ⁶orthogonal contrasts: C₁ = reduced Ca and P vs. reduced Ca and P + phytase; C₂ = reduced AA and ME vs. reduced AA and ME + phytase; C₃ = reduced Ca, P, AA and ME vs. reduced Ca, P, AA and ME + phytase; C₄ = positive control diet vs. treatments with no phytase; C₅ = positive control diet vs. treatments with phytase; C₆ = treatment with phytase vs. treatment with no phytase compared by the F test (P<0.05). NS = not significant at 10% probability level; * p<0.01; **p<0.05; ***p<0.10.

The contrast comparing the supplementation or not of phytase of diets with reduced Ca, P, AA, and ME level (Contrast C₃) indicated that broilers receiving phytase presented better FI (p<0.01), WG (p<0.01), and FCR (p<0.10), which improved 4.14, 7.69, and 3.59%, respectively. Interestingly, the improvement in performance with phytase supplementation was more evident in broilers fed reduced Ca and P levels (contrasts C₁ and C₃).

When the positive-control diet was compared with the diets supplemented with phytase (contrast C₅), it was observed that the dietary reduction of nutrients and energy did not influence broiler performance (p>0.05). On the other hand, broilers fed the positive-control diet presented better FI and WG (p<0.01) compared with those fed the diet with reduced nutrient and energy levels and no phytase supplementation (contrast C₄).

When the diets with reduced nutrient and energy levels and supplemented or not with phytase were compared (contrast C₆), phytase supplementation improved WG and FCR (p<0.01).

The addition of phytase to the reduced AA and ME diet (contrast C₂) reduced the pH of the proventriculus (p<0.10) and of the gizzard (p<0.01) in 11.41 and 19.54%, respectively (Table 4).

The pH of the proventriculus (contrast C₄) and of the gizzard (contrast C₆) increased (p<0.10) when phytase was added to the diets.

There was no influence of treatments on gizzard relative weight (Table 5). However, when the phytase supplementation was evaluated in reduced Ca and P diets (contrast C₁), liver (p<0.10) and heart (P<0.01) relative weights increased in 9.06 and 13.33% in broilers fed the diet with no phytase addition.

Table 4 – Gastrointestinal tract pH of broilers fed starter diets (8-21 days of age) with reduced nutrient and energy levels and supplemented or not with phytase.

Parameter	Treatment with reduced levels							(CV ⁵ (%))	Orthogonal contrasts ⁶					
	CP ¹	Ca and P ²		AA and ME ³		Ca, P, AA, and 3 ⁴								
		W/o phytase	With phytase	W/o phytase	With phytase	W/o phytase	With phytase							
									C ₁	C ₂	C ₃	C ₄	C ₅	C ₆
Proventriculus	3.79	4.13	4.23	4.47	3.96	4.21	4.09	13.96	NS	***	NS	**	NS	NS
Gizzard	3.87	3.93	3.83	4.40	3.54	4.26	4.25	16.01	NS	*	NS	NS	NS	***
Duodenum	5.68	5.69	5.47	5.78	5.66	5.36	5.45	9.10	NS	NS	NS	NS	NS	NS
Jejunum	5.92	6.10	5.78	6.03	6.04	6.10	5.89	7.13	NS	NS	NS	NS	NS	NS
Ileum	6.09	5.95	5.74	5.84	6.01	5.90	5.73	8.18	NS	NS	NS	NS	NS	NS
Cecum	5.94	5.56	5.56	6.18	6.06	6.09	5.66	9.28	NS	NS	NS	NS	NS	NS

¹Positive control = 3,000 kcal/kg ME, 20.8% CP, 0.38% P and 0.86% Ca; ²Ca and P = 3,000 kcal/kg ME, 20.8% CP, 0.23% P and 0.69% Ca; ³AA and ME = 2,948 kcal/kg ME, 20.38% CP, 0.38% P and 0.86% Ca; ⁴Ca, P, AA and ME = 2,948 kcal/kg ME, 20.38% CP, 0.23% P and 0.69% Ca; ⁵coefficient of variation; ⁶orthogonal contrasts: C₁ = reduced Ca and P vs. reduced Ca and P + phytase; C₂ = reduced AA and ME vs. reduced AA and ME + phytase; C₃ = reduced Ca, P, AA and ME vs. reduced Ca, P, AA and ME + phytase; C₄ = positive control diet vs. treatments with no phytase; C₅ = positive control diet vs. treatments with phytase; C₆ = treatment with phytase vs. treatment with no phytase compared by the F test (p<0.05). NS = not significant at 10% probability level; * p<0.01; **p<0.05; ***p<0.10.



Table 5 – Organ biometrics of broilers fed starter diets (8-21 days of age) with reduced nutrient and energy levels and supplemented or not with phytase.

Parameter	Treatment with reduced levels							(CV ⁵ (%))	Orthogonal contrasts ⁶					
	CP ¹	Ca and P ²		AA and ME ³		Ca, P, AA, and ME ⁴								
		W/o phytase	With phytase	W/o phytase	With phytase	W/o phytase	With phytase							
									C ₁	C ₂	C ₃	C ₄	C ₅	C ₆
Gizzard (%)	1.44	1.52	1.49	1.47	1.43	1.51	1.47	9.80	NS	NS	NS	NS	NS	NS
Liver (%)	2.78	2.98	2.71	2.68	2.67	2.83	2.76	9.87	**	NS	NS	NS	NS	NS
Heart (%)	0.62	0.75	0.65	0.66	0.61	0.67	0.63	8.78	*	NS	NS	*	NS	*
Length Small intestine (m)	1.40	1.35	1.32	1.41	1.37	1.36	1.30	5.98	NS	NS	NS	NS	***	NS

¹Positive control = 3,000 kcal/kg ME, 20.8% CP, 0.38% P and 0.86% Ca; 2Ca and P = 3,000 kcal/kg ME, 20.8% CP, 0.23% P and 0.69% Ca; 3 AA and ME = 2,948 kcal/kg ME, 20.38% CP, 0.38% P and 0.86% Ca; 4Ca, P, AA and ME = 2,948 kcal/kg ME, 20.38% CP, 0.23% P and 0.69% Ca; 5 coefficient of variation; ⁶orthogonal contrasts: C₁ = reduced Ca and P vs. reduced Ca and P + phytase; C₂ = reduced AA and ME vs. reduced AA and ME + phytase; C₃ = reduced Ca, P, AA and ME vs. reduced Ca, P, AA and ME + phytase; C₄ = positive control diet vs. treatments with no phytase; C₅ = positive control diet vs. treatments with phytase; C₆ = treatment with phytase vs. treatment with no phytase compared by the F test (p<0.05). NS = not significant at 10% probability level; * p<0.01; **p<0.05; ***p<0.10.

Heart relative weight increased (p<0.01) when broilers were fed the reduced nutrient diets with no phytase additions (contrasts C₄ and C₆).

Intestinal length was different (p<0.10) in broiler fed diets containing phytase, as shown in contrast C₅.

Tibial ash content (p<0.01, % DM) and phosphorus deposition (p<0.10; % DM) increased 9.53 and 13.56%, respectively, when broilers were fed the diet with reduced Ca and P levels supplemented with phytase (contrast C₁; Table 6).

Tibial ash content (%DM) was significantly different (p<0.05), as shown by contrast C₃. Broilers were fed the reduced nutrient diets containing phytase than those fed the positive-control diet (contrasts C₄ and C₅) presented higher tibial ash content (% DM). In addition, when the positive-control diet was compared

with the reduced Ca and P diet containing phytase, there was higher Ca and P deposition (% DM), with 5.32 and 13.77%, respectively. Contrast C₅ did not reveal any differences (p>0.05) in tibial Ca or P content (% DM).

As determined by contrast C₆, tibial Ca and P deposition (% DM) and Ca to P ratio were higher (p<0.05) when the diets were supplemented with phytase.

Blood albumin levels (g/dL) were higher (p<0.10) in birds in the treatments with no phytase supplementation, as shown by contrasts C₁ and C₂ (Table 7).

Blood P levels (mg/dL) were different (p<0.10) as determined by contrasts C₃ and C₄, as well as Ca/P ratio detected by C₁ (p<0.01), C₃ (p<0.01) and C₄ (p<0.05).

Table 6 – Bone mineral content of broilers fed starter diets (8-21 days of age) with reduced nutrient and energy levels and supplemented or not with phytase.

Parameter	Treatment with reduced levels							(CV ⁵ (%))	Orthogonal contrasts ⁶					
	CP ¹	Ca and P ²		AA and ME ³		Ca, P, AA, and ME ⁴								
		W/o phytase	With phytase	W/o phytase	With phytase	W/o phytase	With phytase							
									C ₁	C ₂	C ₃	C ₄	C ₅	C ₆
Dry matter (%)	94.33	93.54	93.97	91.98	92.60	92.36	92.18	1.56	**	NS	NS	*	**	NS
Ashes (% DM)	51.22	46.48	50.92	44.94	46.87	46.96	49.85	4.86	*	NS	*	*	*	NS
P (% DM)	12.02	12.25	13.94	10.83	10.80	10.69	10.44	5.68	***	NS	NS	NS	NS	*
Ca (% DM)	26.66	27.31	28.16	26.94	26.94	25.48	26.21	4.78	NS	NS	NS	NS	NS	*
Ca/P (DM)	2.22	2.23	2.02	2.49	2.49	2.38	2.51	7.10	NS	NS	NS	NS	NS	**

¹Positive control = 3,000 kcal/kg ME, 20.8% CP, 0.38% P and 0.86% Ca; 2Ca and P = 3,000 kcal/kg ME, 20.8% CP, 0.23% P and 0.69% Ca; 3 AA and ME = 2,948 kcal/kg ME, 20.38% CP, 0.38% P and 0.86% Ca; 4Ca, P, AA and ME = 2,948 kcal/kg ME, 20.38% CP, 0.23% P and 0.69% Ca; 5 coefficient of variation; ⁶orthogonal contrasts: C₁ = reduced Ca and P vs. reduced Ca and P + phytase; C₂ = reduced AA and ME vs. reduced AA and ME + phytase; C₃ = reduced Ca, P, AA and ME vs. reduced Ca, P, AA and ME + phytase; C₄ = positive control diet vs. treatments with no phytase; C₅ = positive control diet vs. treatments with phytase; C₆ = treatment with phytase vs. treatment with no phytase compared by the F test (p<0.05). NS = not significant at 10% probability level; * p<0.01; **p<0.05; ***p<0.10.



Table 7 – Blood biochemistry of broilers fed starter diets (8-21 days of age) with reduced nutrient and energy levels and supplemented or not with phytase.

Parameter	Treatment with reduced levels							(CV ⁵ (%)	Orthogonal contrasts ⁶					
	CP ¹	Ca and P ²		AA and ME ³		Ca, P, AA, and ME ⁴								
		W/o phytase	With phytase	W/o phytase	With phytase	W/o phytase	With phytase							
									C ₁	C ₂	C ₃	C ₄	C ₅	C ₆
Total proteins (g/dL)	3.07	3.08	2.93	2.94	3.10	3.11	2.88	13.45	NS	NS	NS	NS	NS	NS
Albumin (g/DL)	1.58	1.64	1.48	1.54	1.56	1.61	1.47	13.98	**	NS	***	NS	NS	NS
Globulins (g/dL)	1.49	1.43	1.45	1.40	1.53	1.50	1.41	17.39	NS	NS	NS	NS	NS	NS
Alb/glob	1.09	1.20	1.05	1.12	1.02	1.09	1.06	16.85	**	NS	NS	NS	NS	***
Ca (mg/dL)	9.23	9.50	8.60	7.41	7.71	8.68	9.01	30.93	NS	NS	NS	NS	NS	NS
P (mg/dL)	7.84	6.96	7.45	7.03	7.04	6.21	7.50	27.32	NS	NS	***	***	NS	NS
Ca/P ratio (%)	1.18	1.41	1.13	1.05	1.10	1.42	1.20	16.11	*	NS	*	**	NS	NS

¹Positive control = 3,000 kcal/kg ME, 20.8% CP, 0.38% P and 0.86% Ca; 2Ca and P = 3,000 kcal/kg ME, 20.8% CP, 0.23% P and 0.69% Ca; 3 AA and ME = 2,948 kcal/kg ME, 20.38% CP, 0.38% P and 0.86% Ca; 4Ca, P, AA and ME = 2,948 kcal/kg ME, 20.38% CP, 0.23% P and 0.69% Ca; 5 coefficient of variation;

⁶ Orthogonal contrasts: C₁ = reduced Ca and P vs. reduced Ca and P + phytase; C₂ = reduced AA and ME vs. reduced AA and ME + phytase; C₃ = reduced Ca, P, AA and ME vs. reduced Ca, P, AA and ME + phytase; C₄ = positive control diet vs. treatments with no phytase; C₅ = positive control diet vs. treatments with phytase; C₆ = treatment with phytase vs. treatment with no phytase compared by the F test (p<0.05). NS = not significant at 10% probability level; * p<0.01; **p<0.05; ***p<0.10.

DISCUSSION

Literature studies show differences in the capacity of phytase of releasing P from phytate due to influence of dietary Ca content. The better performance of broilers fed reduced Ca and P levels and phytase obtained in the present study suggest that phytase promoted better Ca and P utilization. This is consistent with the results of Powell *et al.* (2011), who found that it phytase is used more efficiently when dietary Ca levels are reduced.

These results also confirm that the use of enzymes in corn- and soybean meal-based broiler diets promote better performance (p<0.01) by breaking down the antinutritional factors present in the cell wall of plants (Graham *et al.*, 2002), and hence, better nutrient absorption and animal performance (Costa Leite *et al.*, 2011; Powell *et al.*, 2011; Walk *et al.*, 2012).

On the other hand, Alvarenga *et al.* (2011) obtained lower broiler weight gain when the diets contained phytase. Also, in his review, Slominsk (2011) found little effect of phytase supplementation on performance and the utilization of energy and amino acid utilization in broilers. These differences may be related to the different source of phytase currently available in the market (Elkhalil *et al.*, 2007). The positive results obtained in the present study may be possibly attributed to a better efficiency phytase derived from *E. coli*.

Another possible cause of the improved responses may have been the lower dietary Ca levels. Inadequate dietary Ca levels may impair Ca, P, Zn, and Mg absorption. However, Ca reduction in phytase-supplemented diets must be proportional do total phosphorus (tP) reduction in order to maintain the required Ca:P ratio. According to Pereira *et al.* (2012), the inclusion of phytase in feed formulation matrices provides economic and environmental benefits in broiler production.

The lower pH values observed in the gizzard (p<0.01) and the proventriculus (p<0.10) of broilers fed the reduced nutrient and energy level and no phytase can be attributed to the attempt of birds to balance nutrient intake and energy utilization to supply their requirements.

Phytate is hydrolyzed mainly in the upper gastric tract (proventriculus and gizzard), where the pH favors the action of phytase and its substrate is more water soluble (Selle & Ravidran, 2007). According to Penz *et al.* (1993), GIT pH reduction enhances enzyme activity, thereby increasing dietary nutrient absorption and improving animal performance.

The observed increase in liver (p<0.10) and heart (p<0.01) relative weights in the broilers fed diets with reduced Ca and P levels and no phytase supplementation is probably due to low phosphorus supply. Heart hypertrophy (increase of heart muscle mass beyond the usual limit) may be caused by



hypophosphatemia due to low phosphorus supply (Rosário *et al.*, 2004; Temprano *et al.*, 2004). Hearts with hypertrophy usually present reduced contraction due to reduced myosin ATPase activity and lower cyclic adenosine monophosphate (cAMP), which may result from hypophosphatemia. This may lead to heart failure, reducing the systolic volume and increasing myocardial fiber diameter (Riedesel, 2006).

Low ATP production stimulates the synthesis of liver enzymes, increasing liver volume (Reis *et al.*, 1999). Oliveira *et al.* (2009) observed reduced liver and heart relative weights when broilers were fed diets with 70% reduction in non-phytic phosphorus level.

The increase in tibial ash ($p < 0.01$, % DM) and phosphorus deposition ($p < 0.10$; % DM) determined in broilers fed the diet with phytase addition reduced Ca and P levels and in ash content ($p < 0.05$) in broilers fed the reduced Ca, P, AA, and ME diet with phytase supplementation was expected, because phytase increases the release of phosphorus and other minerals from the feedstuffs. According to Nelson & Walker (1964) and Pereira *et al.* (2012), bone ash content is the most efficient parameter to estimate the amount of phosphorus released by phytase in corn- and soybean meal-based diets.

Geraldo *et al.* (2006) found 1.88% reduction in tibial ash content for each 1% increase in dietary calcium levels. The dietary supplementation of phytase allows reducing dietary available phosphorus in 0.15% and Ca in 0.15% (Gomide *et al.*, 2011). Consistent with the present study, Schoulten *et al.* (2003) observed that reduced P diets supplemented with phytase allowed reducing Ca requirements to lower values than those present in nutritional tables, but recommended that dietary Ca reduction should be proportional to total P reduction, maintaining adequate Ca to P ratio, as also mentioned by Qian *et al.* (1997).

In the present experiment, average total protein levels in the blood ranged between 2.88 (reduced Ca, P, AA, and ME and no phytase) and 3.11 mg/dL (reduced Ca, P, AA, and ME and with phytase) are below those considered normal (3.6 mg/dL). According to Viveiros *et al.* (2002), total protein level in the serum may be reduced in case of malnutrition or low intestinal absorption of nutrients, but the treatments applied in the present study did not cause any of these conditions.

Blood Ca level (mg/dL) was not influenced ($p > 0.10$) by the applied treatments, possibly because Ca homeostasis in the extracellular fluid is maintained by the joint effects of the parathyroid hormone (PTH),

calcitonin, and 1,25 di-hydroxy vitamin D ($1,25(\text{OH})_2\text{D}_3$) in the intestines, kidneys, and bone. Those hormones regulate Ca flow between these organs and the extracellular fluid, resulting in less than 5% variation in Ca blood levels under normal circumstances, and only occurs when hormonal mechanisms fail, secondarily to reduced intestinal absorption (Macari *et al.*, 2002). In the present study, mean Ca serum values were lower than those obtained by Vieites *et al.* (2011), of 6.72 and 6.89 mg/dL.

The higher albumin values ($p < 0.10$) found in broilers fed reduced nutrient and energy levels and no phytase supplementation are considered normal (1.6-2.0 g/dL; Reece & Swenson, 2006).

Albumins are synthesized in the liver and are very abundant in blood plasma, where they play an essential role in the maintenance of the osmotic pressure (Junqueira & Carneiro, 2011). Differences in serum albumin levels, according to Macari *et al.* (2002) are probably due to acid-base imbalance (metabolic acidosis or alkalosis), because of the buffering action of these proteins that capture or donate H^+ ions by their carboxylic terminals (COOH or COO^-) attempting to reestablish normal blood pH. This was possibly the case in the present study, and this hypothesis is corroborated by the increase in liver relative weight,

Consistent with this result, the Ca/P ratios presented in contrasts C1, C3, and C4 show higher concentration of P relative to Ca, causing osmotic pressure imbalance (Reece & Swenson, 2006). This difference in the Ca/P ratio is mainly due to P variation (mg/dL), which was different in contrast C3 and C4 ($p < 0.10$). According to Vargas Jr. *et al.* (2003), the Ca to P ratio must be constant; any imbalance may affect body functions.

The difference in the glob/alb ratios obtained in contrasts C1 and C6 may be explained by the reduction of this ratio in the treatments with phytase, as shown in the contrasts between treatments with or without phytase and Ca and P reduction (contrast C1). This was caused by variations in the albumin fraction, demonstrating the effect of Ca and P reduction on the metabolism of the liver, which is the only site of albumin synthesis.

The diet with reduced amino acid and energy levels and phytase addition reduced gizzard and proventriculus pH. Dietary Ca and P reduction increased relative liver and heart weights, as well as albumin blood levels. The bones of broilers fed phytase-supplemented diets presented higher ash content. The results of the present study suggest that broiler nutritional requirements need to be reevaluated when the diet is supplemented with phytase.



REFERENCES

- Alvarenga RR, Nagata AK, Rodrigues PB, Zangeronimo MG, Pucci LEA, Hespanhol R. Adição de fitase em rações com diferentes níveis de energia metabolizável, proteína bruta e fósforo disponível para frangos de corte de 1 a 21 dias. *Ciência Animal Brasileira* 2011;12(4):602-609.
- Casey, A.; Walsh, G. Identification and characterization of a phytase of potential commercial interest. *Journal of Biotechnology* 2004;110(3):313-322.
- Costa Leite PRS, Leandro NSM, Stringhini JH, Café, MB, Gomes NA, Jardim Filho M. Desempenho de frangos de corte e digestibilidade de rações com sorgo ou milho e complexo enzimático. *Pesquisa agropecuária Brasileira* 2011;46(3):280-286.
- Denbow DM, Ravindran V, Kornegay YIZ, Hulet RM. Improving phosphorus availability in soybean meal for broilers by supplemental phytase. *Poultry Science* 1995;74(11):1831-1842.
- Detmann E, Souza MA, Valadares Filho SC, Queiroz AC, Beterchielli TT, Saliba EOS, Cabral LS, Santos Pina D, Ladeira MM, Azevedo JAG. Métodos para análise de alimentos. Visconde do Rio Branco: Universidade Federal de Viçosa; 2012.
- Elkhalil EAI, Manner K, Borris R, Simon O. In vitro and in vivo characteristics of bacterial phytases and their efficacy in broiler chickens. *British Poultry Science* 2007;48(1):64-70.
- Geraldo A, Bertechini AG, De Brito JAG, Kato RK, Fassani EJ. Níveis de cálcio e granulometria do calcário para frangos de reposição no período de 3 a 12 semanas de idade. *Revista Brasileira de Zootecnia* 2006;36(1):113-118.
- Gomide ME, Rodrigues PB, Beterchini AG, Freitas TF, Fassani EJ, Reis MP, Rodrigues NEB, Almeida EC. Rações com níveis reduzidos de proteína bruta, cálcio e fósforo com fitase e aminoácidos para frangos de corte. *Revista Brasileira de Zootecnia* 2011;40(11):2405-2414.
- Graham KK, Kerley DM, Firman JD, Allee G.L. The effect of enzyme treatment of soybean meal on oligosaccharide disappearance and chick growth performance. *Poultry Science* 2002;81:1014-1019.
- Junqueira LC, Carneiro J. Histologia básica. 11th ed. Rio de Janeiro: Guanabara Koogan; 2011. p. 221-253.
- Leeson S, Caston L, Summers JD. Broiler response to energy or energy and protein dilution in the finisher diet. *Poultry Science* 1996;75:522-528.
- Lehnen CR, Lovatto PA, Andretta I, Kipper M, Hauschild L, Rossi CA. Meta-análise da digestibilidade ileal de aminoácidos e minerais em suínos alimentados com rações contendo enzimas. *Pesquisa Agropecuária Brasileira* 2011;46(4): 438-445.
- Macari M, Furlan RL, Gonzáles E. Ingestão de alimentos: Mecanismos regulatórios. In: Macari M, Furlan RL, Gonzáles E. Fisiologia aviária aplicada a frangos de corte. Jaboticabal: Ed. Funep; 2002. v.2, p.187-192.
- Maenz DD. Enzymatic characteristics of phytases as they relate to their use in animals feeds. In: Bedford MR, Partridge GG, editor. *Enzymes in farm animal nutrition*. Wallingford: Publishing; 2001. p.60-84.
- Manual de manejo de Frangos de corte. Guapiaçu: Cobb Vantrans; 2009. 70p.
- Nelson TS, Walker AC. The biological evaluation of phosphorus compounds: A summary. *Poultry Science* 1964;43:94-98.
- Oliveira MC, Marques RH, Gravena RA, Traldi AB, Godoy CR, Moraes VMB. Fitase em rações com níveis reduzidos de fósforo não-fítico para frangos de corte. *Biotemas* 2009;22(4):169-176.
- Oluyinka AO, Cowieson AJ, Adeola O. Energy utilization and growth performance of broilers receiving diets supplemented with enzymes containing carbohydrase or phytase activity individually or in combination. *British Journal of Nutrition* 2007;99:682-690.
- Pang Y, Applegate TJ. Effects of dietary copper supplementation and copper source on digesta pH, calcium, zinc, and copper complex size in the gastrointestinal tract of the broiler chicken. *Poultry Science* 2007; 86:531-537.
- Penz Jr AM, Silva AB, Rodrigues O. Ácidos orgânicos na alimentação de aves. Anais da Conferência Apinco'93 de Ciência e Tecnologias Avícolas; 1993; Santos, São Paulo. Brasil. Santos: Facta; 1993. p.111-119
- Pereira R, Menten JFM, Romano GG, Silva CLS, Zavarize KC, Barbosa NAA. Eficiência de uma fitase bacteriana na liberação de fósforo fítico em rações de frangos de corte. *Arquivo Brasileiro de Medicina Veterinária e Zootecnia* 2012;64(1):137-144.
- Powell S, Bidner TD, Southern LL. Phytase supplementation improved growth performance and bone characteristics in broilers fed varying levels of dietary calcium. *Poultry Science* 2011;90: 604-608.
- Qian H, Kornegay ET, Denbow DM. Utilization of phytate phosphorus and calcium as influenced by microbial phytase, cholecalciferol, and the calcium: total phosphorus ratio in broiler diets. *Poultry Science* 1997;76:37-46.
- Reece WO, Swenson MJ. 2006. Composição e funções do sangue. In: Swenson MJ, Reece WO. *DUKES fisiologia dos animais domésticos*. 12th ed. Rio de Janeiro: Guanabara Koogan; 2006. p. 24 – 48.
- Reis CVS, Penna FJ, Oliveira MC, Roquete MLV. Glicogenose tipo I. *Jornal de Pediatria* 1999;75(4): 227-236.
- Riedesel DH. O coração: Estrutura macroscópica e propriedades básicas. In: Dukes, Reece, WO, editor. *Fisiologia dos animais domésticos* 12.ed: Rio de Janeiro: Guanabara Koogan; 2006. p. 173-186.
- Rosário MF, Da Silva MAN, Coelho AAD, Savino VJM. Síndrome ascite em frangos de corte: uma revisão sobre a fisiologia, avaliação e perspectiva. *Ciência Rural* 2004;34(6):1987-1996.
- Schouten NA, Teixeira AS, Conte AJ, Silva HO, Bertechini AG, Fialho ET. Efeito dos níveis de cálcio da ração suplementada com fitase sobre a deposição de minerais na tíbia de frangos de corte de 22 a 42 dias de idade. *Ciência e Agrotecnologia*. 2003;27:(1) 206 – 210.
- Rostagno HS, Albino LFT, Donzele JL, Gomes PC, Oliveira RF, Lopes DC, Ferreira AS, Barreto LST, Euclides RF. Tabelas brasileiras para aves e suínos: composição de alimentos e exigências nutricionais. 3ªed. Viçosa: Universidade Federal de Viçosa; 2011. 252p.
- Selle PH, Ravindran V. Microbial phytase in poultry nutrition. *Animal Feed Science Technology* 2007;135:1-41.
- Slominski BA. Recent advances in research on enzymes for poultry diets (Review). *Poultry Science* 2011;90: 2013-2023.
- Steiner T, Mosenthin R, Fundis A, Jakob S. Influence of feeding on apparent total tract digestibility of phosphorus and calcium in pigs fed low-phosphorus diets supplemented with microbial or wheat phytase. *Livestock Science* 2006;102:1 -10.
- Temprano JL, Bretón I, Zugasti A, Cuerda C, Cambor M, Pérez-Díaz MD, García P. Hipofosfatemia grave tras el inicio de nutrición parenteral en una paciente con fistula intestinal. *Nutrición Hospitalaria* 2004;19(4):243-247.
- Vieites FM, Fraga AL, Moraes GHK, Vargas Junior JG, Nalon RP, Correa GSS, Nunes RV. Cálcio, fósforo e proteína total no sangue de frangos de corte em função de níveis de balanço eletrolítico da ração. *Arquivo Brasileiro de Medicina Veterinária e Zootecnia*. 2011;63:(4) 887-894
- Viveros A, Brenes A, Arija I, Centeno C. Effects of microbial phytase supplementation on mineral utilization and serum enzyme activities in broiler chicks fed different levels of phosphorus. *Poultry Science* 2002;81:1172-1183.
- Walk CL, Addo-Chidie EK, Redford MR, Adeola O. Evaluation of a highly soluble calcium source and phytase in the diets of broiler chickens. *Poultry Science* 2012;91:2255-2263.