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## Characterization and Digestibility of Detoxified Castor Oil Meal for Japanese Quails

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Alternative food, poultry, cytotoxicity, digestibility, energy.

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### **ABSTRACT**

These experiments were performed to determine the chemical composition, coefficients of nutrient and energy metabolizability, amino acid composition, and cytotoxicity of different castor oil meals subjected to different detoxification processes and added to the diet of Japanese quails. In the trial, 180 46-d-old female Japanese quails were distributed according to a completely randomized design into five treatments and with replicates of six bird each. The treatments consisted of following detoxification methods of castor oil meal: Castor oil meal A (CMA) recovery in alcohol at 80 °C for 20 minutes and drying at 80 °C; castor oil meal B (CMB) and C (CMC) - recovery in alcohol at 80 °C for 6 minutes, neutralization with 5% NaOH, and drying under direct sunlight sun for two days (CMB) or pelleted (CMC); castor oil meal D (CMD) - recovery in alcohol at 110 °C for 15 minutes and drying at 110 °C. Castor oil meal was added replacing 20% of the reference diet. There was slight chemical composition variation (1.21% in crude protein, 6% in dry matter, 2.2% in ether extract and 64 kcal/kg in gross energy) among the castor oil meals submitted to the different treatments. The castor oil meal submitted to treatment C showed the highest amino acid values. In the cytotoxicity test, treatment D presented lower ricin activity. Castor oil meals A, C, and D may be included in Japanese quail diets; however, castor oil meal D is recommended due to the simplicity its industrial process, its low toxicity, and metabolizability coefficients obtained.

### **INTRODUCTION**

Japanese quail production stands out in the Brazilian market as an excellent activity because it requires low initial investment costs as well small areas, providing fast return on the capital invested, in addition of presenting low labor costs (Ton *et al.*, 2011).

Feed represents a major cost in poultry production. Considering the increasing prices of corn and soybean meal, which are typically the main ingredients of poultry diets, alternative feed stuffs have been sought to replace those costly ingredients, which production also competes with human food. Studies on alternative feedstuffs have allowed considerable reduction in production costs (Albuquerque *et al.*, 2014) as the partial or total replacement of expensive ingredients contributes for the economic viability of animal production (Santos *et al.*, 2013).

The castor oil meal has been the object of research because of its high protein value. However, its protein content may vary due to seed characteristics (Azevedo &Lima, 2007), as well as its metabolizable energy (ME) level, which is influenced by the industrial methods applied for castor oil extraction for biodiesel production. In Brazil, the National Council for Energy Policy (CNPE) established in Act No. 11.097/2005 the mandatory blending of 2% biodiesel in conventional diesel by 2008 and



5% by2013, and therefore, castor bean by-products are highly available in the Brazilian market.

Abdalla et al. (2008) reported that the crude protein (CP) of castor oil meal ranges between 39 and 43%; however, its use is limited because of its toxicity (ricin and ricinin) and allergenicity (2S albumins) (Anandan et al., 2005). Ricin is the main toxic component of the castor oil plant (Ricinus communis) and it consists of a water-soluble protein found only in the endosperm of the castor oil plant seeds (Severino, 2005). Ricin belongs to the family of lectins A-B, i.e., it is composed of two subunits: one with enzymatic activity and the other with a specific binding site for galactose. Its toxicity mechanism is exerted through the inactivation of ribosomes. The subunit A inactivates specifically and irreversibly the eukaryotic ribosomes, preventing protein synthesis by removing adenine residues in 28S ribosomal RNA (Oliveira et al., 2006). The subunit B is bound to the cell wall and the sub-unit A through the disulfide bonds, allowing the latter to enter the cytosol by endocytosis. Therefore, if the bonds between the two subunits are broken, the resulting parts are not toxic (Audi et al., 2005).

After being detoxified, castor oil meal can be provided to animals as protein concentrate in partial replacement of soybean meal. There are different castor oil meal detoxification that allow its inclusion in the feeds of monogastric animals. Anandan *et al.* (2005) concluded that submitting the castor bean meal to autoclave (15 psi, 60 min) or treatment with calcium hydroxide (40 g/kg castor oil meal) completely denatured ricin. Other methods, such as autoclave (15 psi for 90 minutes) or calcium hydroxide or calcium oxide diluted in water (1:10) at a dose of 60 g/kg of castor oil meal, have proven effective to denature the ricin (Oliveira *et al.*, 2006).

Due to the lack of information on the use of castor oil meal in the diet of Japanese quails, these experiments were performed to determine the chemical composition and coefficients of metabolizability of nutrients and energy, and the cytotoxicity of different castor oil meals submitted to different processing methods.

### **MATERIAL AND METHODS**

The experiment was conducted at the Department of Animal Science, Federal Rural University of Pernambuco (UFRPE), Recife, Pernambuco, Brazil. The project was approved by the Ethics Committee on Animal Use (CEUA-UFRPE).

The castor bean cultivar used was the BRS Nordestina. Oil was extracted from the seeds using the

standard method of cooking a batch for 30 minutes at 80 °C and, subsequently, two presses were used for oil extraction, producing the castor bean pie at a pilot scale in the Biodiesel Plan, municipality of Pesqueira, Pernambuco, Brazil.

The castor bean pie was subjected to additional processing at industrial scale for the production of different castor bean meal types, called A, B, C and D. Castor oil meal A (CMA) was subjected to recovery in ethanol at 80 °C for 20 minutes and dried at 80 °C; castor oil meal (CMB) was submitted to the same ethanol recovery and temperature as CMA, but for six minutes, and then to neutralization with 5% sodium hydroxide (NaOH), and drying under direct exposure to sunlight for two days; castor oil meal C was submitted to the same processing procedures as CMB, except for drying, which was replaced by pelleting; castor oil meal D (CMD) was recovered in alcohol at 110 °C for 15 minutes and dried at 110 °C.

The chemical composition of the evaluated meals was analyzed at the Laboratory of Animal Nutrition (LANA), UFRPE. Analyses of dry matter (DM), CP, ash, ether extract (EE) and crude fiber (CF) were carried out according to methods described by Silva & Queiroz (2006), and neutral detergent insoluble fiber was determined according to the method of Van Soest *et al.* (1991). Gross energy (GE) analysis was performed at the Laboratory of Physical and Chemical Analysis of Brazilian Company of Agricultural Research Swine and Poultry (EMBRAPA).

Total amino acids profiles of processed castor oil meals were determined by HPLC (High Performance Liquid Chromatography) at the Laboratory of Applied Technology of EVONIK Animal Nutrition, in Hanau, Germany. The cytotoxicity of processed castor oil meal corresponds to the percentage of specific standardized cells (Vero cells) that are destroyed using a standard quantitative cell counting and measurement of lactate dehydrogenase activity. The limit of detection of the Vero cell assay was 10 ng/mL using a concentration of 1.6 x 10(5) cells/well. The procedure was according to Fernandes et al. (2012) and analysis was performed at the Laboratory of Chemistry and Function of Proteins and Peptides of Bioscience and Biotechnology Center of Universidade Estadual do Norte Fluminense Darcy Ribeiro. Values of cytotoxicity are expressed as percentage comparing with the castor bean cake with no further processing management considered as reference relative 100% cytotoxicity.

The digestibility assay of castor oil meal was performed in the Laboratory of Non-Ruminant Digestibility, UFRPE. One hundred and eighty46-d-old



Japanese quails were acquired from Fujikura farm. Birds were distributed in a completely randomized design (CRD) with five treatments and six replicates of six birds each, and housed in two cages. Each cage was divided in five overlapping floors with three divisions each(33 cm x 25 cm x 20 cm), totaling 15 divisions per cage. The cages were equipped with feeders and trough drinkers. Trays lined with plastic canvas were fit under the cages for excreta collection. Birds were distributed according to a completely randomized experimental design (CRD) with five treatments and six replicates of six birds each. The animal use protocol was approved by the institutional animal care guidelines the Federal Rural University of Pernambuco.

The reference diet (Table 1) was formulated using the nutritional values of feedstuffs proposed by Rostagno et al. (2005) and the nutritional requirements obtained by Silva & Costa (2009).

**Table 1** – Composition of basal diet

Ingredients <sup>1</sup>	%
Corn Grain	57.06
Soybean meal	32.61
Limestone	6.76
Dicalcium phosphate	1.36
Soybean oil	1.33
DL-methionine	0.35
Common salt	0.33
Vitamin/Mineralpremix <sup>2</sup>	0.15
L-lysine	0.05
total	100.0

<sup>&</sup>lt;sup>1</sup> Nutritional composition: metabolizable energy 2800 (kcal/kg), crude protein 20%, calcium 3.05%, available phosphorus (%) 0.28%, methionine+cysteine 0.78%, methionine 0.42%, lysine 1.15%, threonine 0.23%, sodium 0.79%.

The treatments included REF - reference feed based on corn and soybean meal, RCMA - 80% of the REF diet + 20% of CMA, RCMB - 80% of the REF diet + 20% of CMB, RCMC - 80% of the REF diet + 20% of CMC and RCMD - 80% of the REF diet + 20% CMD. Feed and water were supplied ad libitum throughout the experimental period, which lasted ten days, with five days of adaptation to the cages and the experimental diet and five days of total excreta collection. Feed intake and excreta output volumes were recorded. In the experiment, on the first and last days of total excreta collection, the external marker ferric oxide (Fe<sub>2</sub>O<sub>2</sub>) was added to the diets at2%. Excreta were collected once daily (12:00 hours), placed in duly identified plastic bags, and stored in a freezer at -20 °C until the end of the collection period.

The samples were thawed, homogenized per experimental unit, weighed, and submitted to the laboratory for analyses. In the lab, samples were pre-dried in a ventilated oven at 65 °C for 72 hours. Dried samples were then weighed, ground to 1-mm particle size, and submitted to the Laboratory of Animal Nutrition (LANA), UFRPE, for DM and nitrogen (N) analyses by the method described by Silva & Queiroz (2006). The GE analysis was performed at the Laboratory of Physical and Chemical Analysis of Embrapa Swine and Poultry.

The obtained were submitted to analysis of variance and means were compared by the Tukey's test at 5% probability level, using the SAS statistical program (2000).

### **RESULTS**

The processes applied to detoxify castor oil meal resulted in variations in the nutritional composition values assessed (Table 2).

**Table 2** – Chemical composition of different processed castor oil meals, expressed on natural matter basis.

Chemical composition	Processed castor oil meal				
(%)	CMA	CMB	CMC	CMD	
Dry matter	90.80	90.47	89.43	91.10	
Crude protein	32.04	32.13	32.65	33.25	
Ether extract	9.32	7.21	9.22	9.42	
Crude fiber	40.55	36.70	39.34	38.33	
Neutral detergent fiber	46.00	47.97	42.78	47.26	
Ash	5.69	6.13	6.12	5.30	
Calcium	0.60	0.65	0.61	0.64	
Phosphorus	0.90	1.05	0.90	0.88	
Gross energy (kcal/kg)	4748	4684	4723	4720	

 $<sup>^{\</sup>rm l}$  CMA - Recovery in ethanol at 80°C for 20 min, drying at 80°C; CMB - Recovery in ethanol at 80°C for 6 min, neutralization with sodium hydroxide, and drying under direct sunlight sun for two days, CMC - Recovery in ethanol at 80°C for 6 min, neutralization with sodium hydroxide, and pelleting; CMD - Recovery in alcohol at 110°C for 15 minutes and drying at 110 °C.

Maximum absolute values differences of 1.6% DM, 1.21% CP, 2.2% EE, 3.9% CF 3%, 0.83% ash, 0.05% calcium, and 0.17% phosphorus were obtained among processed castor oil meal samples.

A GE difference of 64 kcal/kg was determined between CMB and CMA. The lowest GE level, which was obtained in CMB, is compatible with the reduction in EE absolute value (7.21%) in this meal.

The different processed castor oil meal types (Table 3) presented different total amino acid composition, as a function of processing method. The processing method used for CMA resulted in reduced levels of all essential amino acids.

<sup>&</sup>lt;sup>2</sup> Composition per kg of product: Fe, 20000mg; Co, 200mg, 4000mg Cu, Mn, 75000mg, Zn, 50000mg; If, 250mg; I, 1500mg, Antioxidant, 100000mg; Folic Acid, 200mg, Pantothenic acid, 5350 mg, Niacin, 19,900 mg; - Vit. A, 8000000 IU, Vit. D3, 2 million IU, vit k3, 2000 mg, Vit. B2, 4000 mg, Vit. B6, 1000 mg, B12, 10000 mcg, Vit E, 15000mg.

**Table 3** – Total amino acid composition of different processed castor oil meals, expressed on natural matter basis.

Aminoacids 1	Processed castor oil meal				
	CMA	CMB	CMC	CMD	
Methionine (%)	0.48	0.52	0.53	0.50	
Cystine(%)	0.64	0.68	0.72	0.66	
Met + cys (%)	1.13	1.20	1.24	1.16	
Lysine (%)	0.94	0.96	1.00	0.96	
Threonine (%)	0.95	1.03	1.03	0.99	
Arginine (%)	3.04	3.19	3.31	3.04	
Isoleucine (%)	1.15	1.26	1.26	1.21	
Leucine (%)	1.73	1.86	1.88	1.78	
Valine (%)	1.45	1.56	1.59	1.51	
Histidine (%)	0.59	0.61	0.64	0.59	
Phenylalanine (%)	1.13	1.19	1.20	1.15	
Glycine (%)	1.26	1.34	1.35	1.29	
Serine (%)	1.51	1.59	1.63	1.51	
Proline (%)	1.00	1.12	1.19	1.08	
Alanine (%)	1.20	1.29	1.33	1.23	
Aspartic acid (%)	2.56	2.76	2.77	2.64	
Glutamic acid (%)	5.29	5.57	5.36	5.28	

<sup>&</sup>lt;sup>1</sup> Analyses conducted by the company EVONIK.CMA - Recovery in ethanol at 80°C for 20 min, drying at 80°C; CMB - Recovery in ethanol at 80°C for 6 min, neutralization with sodium hydroxide, and drying under direct sunlight sun for two days, CMC - Recovery in ethanol at 80°C for 6 min, neutralization with sodium hydroxide, and pelleting; CMD - Recovery in alcohol at 110°C for 15 minutes and drying at 110 °C.

Figure 1 shows the cytotoxicity of processed castor oil meals. The results indicate that the treatment CMD presented the lowest ricin toxic activity. Numbers on the bars represent the relative cytotoxicity when comparing to the castor bean cake with no further processing management (considered the reference relative 100% cytotoxicity).

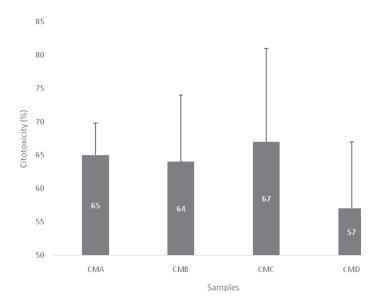


Figure 1 - Evaluation of the cytotoxicity generated byprocessedcastor oil mealsamples Laboratory of Chemistry and Function of Proteins and Peptides of the Center for Bioscience and Biotechnology. CMA - Recovery in ethanol at 80°C for 20 min, drying at 80°C; CMB - Recovery in ethanol at 80°C for 6 min, neutralization with sodium hydroxide, and drying under direct sunlight sun for two days, CMC - Recovery in ethanol at 80°C for 6 min, neutralization with sodium hydroxide, and pelleting; CMD - Recovery in alcohol at 110°C for 15 minutes and drying at 110 °C.

The digestibility assay results (Table 4) show that the birds fed the diet containing castor oil meal B (CMBF) presented low feed intake, which led to amino acid imbalance, resulting in insufficient N and energy supply to meet the birds requirement. As a consequence, low metabolizability coefficients of DM, N, and GE, and reduced dietary AME and AMEn were determined.

The metabolizability coefficients of the different castor oil meals evaluated are shown in Table 5. Castor oil meals C and D promoted greater MCGE and N balance than CMA and CMB. Relative to energy use (AME and AMEn), CMA was not significantly different from CMC and CMD.

### DISCUSSION

Faria Filho et al. (2010), evaluating broiler diets with castor oil meal detoxified with 60 g of micro-processed calcium oxide diluted in water at 1:10, obtained CP (33.1%), EE (1.9%), and CF (24.8%) values, which are lower than those found in the present study in treatments A, B, C and D. This may be due to differences in the efficiency of the mechanical press during processing, and in the methods used for castor oil meal analyses.

The castor oil meal composition (89.40% DM, 39.20% CP, 1.55% EE, 18.50%, on natural matter basis) in the Brazilian Tables for Poultry and Swine (Rostagno *et al.*, 2011) presents similar DM and as hand lower in CF and EE values than those found in



**Table 4** – Average values of feed intake (FI), and of metabolizability coefficients of dry matter (MCDM), nitrogen (MCN) and gross energy (MCGE), nitrogen balance (NB), apparent metabolizable energy (AME) and apparent metabolizable energy corrected for nitrogen retention (AMEn) of Japanese quails fed the experimental diets.

Feeds	FI (g)	MCDM (%)	MCN (%)	MCGE (%)	NB (g/day)	AME¹ (kcal/kg)	AMEn <sup>1</sup> (kcal/kg)
CMAF	17.58 b	67.94 b	43.88 bc	70.21 b	7.54 b	2555 a	2500 a
CMBF	16.95 c	65.77 c	43.34 с	67.82 c	8.86 ab	2409 b	2344 b
CMCF	18.95 a	70.68 a	45.13 a	73.53 a	9.11 a	2580 a	2513 a
CMDF	18.75 a	68.36 b	44.70 ab	69.71 b	9.55 a	2544 a	2474 a
LSD	0.26	1.00	1.13	1.23	1.52	63	56
CV, %	0.83	0.90	1.58	1.08	10.7	1.54	1.41

<sup>&</sup>lt;sup>1</sup> Energy values expressed on natural matter basis; RCMA, RCMB, RCMC and RCMD: diets containing 20% of CMA, CMB, CMC and CMD, respectively. Means in the same column followed by different letters differ at 5% by Tukey's test, LSD - least significant difference (= 0.05) and CV - Coefficient of variation.

the present study. However, as previously mentioned, this may result from differences in the methods of oil extraction. Gomes *et al.* (2009) analyzed castor oil meal submitted to the detoxification process proposed by Anandan *et al.* (2005), obtaining 89.8% DM, 56.8%CP, 3.67% EE, 12.80% ash, 47.32% NDF levels on natural matter basis, which are different from the values observed in the current study.

Variations in temperature, pressure, and chemical agents used for the extraction of castor bean oil at industrial scale account for differences in castor oil meal protein levels and quality (Silva et al., 2010), directly influencing the efficiency of nutrient utilization by poultry. In the present study, essential amino acid contents differed as a function of processing method. In particular, CMA presented lower concentrations of essential amino acids compared to the other evaluated meals. This effect was probably due to long time (20 minutes) of recovery in ethanol, which may have caused protein denaturation, thereby affecting amino acid availability. The best total amino acids results were obtained in CMB and CMC, both of which were submitted to rapid recovery in ethanol at 80°C for 6 minutes, neutralization, and differed only as to drying method, with CMB dried by sun exposure and

CMC by pelleting. The average essential amino acid contents obtained were lower than those reported by Rostagno *et al.* (2011) and Benesi (1979), probably due to the detoxification methods of castor bean meal. When compared with the commercial castor oil meal evaluated by Vilhjalmsdottir & Fisher (1971), which was submitted to crushing for oil extraction, followed by4-5 washings with solvent. The resulting meal was boiled for four hours, dried and ground for detoxification process, the values were higher than those found in the present study.

Diets with amino acid imbalance may induce poultry to increase their feed intake to meet their physiological needs. On the other hand, birds may reduce their feed intake in an attempt to protect themselves from possible pathological lesions caused by amino acid excess in the body. Japanese quails fed the diets containing CMC and CMD were possibly able to reduce the effect of unbalanced diets, as shown by their better N and energy utilization compared with those fed the CMB diet.

The lowest GE levels obtained in CMB is compatible with its lowest absolute EE value (7.21%) relative to the other meals. This reduction EE content was possibly due to the drying method applied (two days

**Table 5** – Average values of feed intake (FI), and of metabolizability coefficients of dry matter (MCDM), nitrogen (MCN) and gross energy (MCGE), nitrogen balance (NB), apparent metabolizable energy (AME) and apparent metabolizable energy corrected for nitrogen retention (AMEn) castor oil meals submitted to different processing methods

	•			•		
Treatment	MCDM (%)	MCN (%)	MCGE (%)	NB (g/d)	AME¹ (kcal/kg)	AMEn¹ (kcal/kg)
CMA	52.17 b	41.27 bc	56.53 a	5.40 b	2684 a	2736 a
CMB	41.31c	38.54 с	40.37 b	6.45 b	1891 b	1894 b
CMC	65.86 a	47.51 a	57.94 a	7.16 a	2736 a	2730 a
CMD	54.27 b	45.35 ab	53.96 a	7.26 a	2547 a	2524 a
LSD	4.99	5.64	6.76	1.38	319	282
CV, %	5.78	8.09	8.01	10.1	8.00	7.09

<sup>&</sup>lt;sup>1</sup> Energy values expressed on natural matter basis; RCMA, RCMB, RCMC and RCMD: diets containing 20% of CMA, CMB, CMC and CMD, respectively. Means in the same column followed by different letters differ at 5% by Tukey's test, LSD - least significant difference (= 0.05) and CV - Coefficient of variation.



of sun exposure), allowing the longest exposure time to the catalyst, which in turn promoted a continuation of the saponification reaction or the transesterification of residual ethanol with the catalyst. These reactions occur simultaneously.

The metabolizability coefficient of gross energy is influenced by several factors, including the quality of the feedstuff, because nutrients such as carbohydrates, proteins, and lipids are not used efficiently by poultry, when the efficiency of energy is reduced as a function of diet(Sakomura & Rostagno, 2007). Other factors may influence energy value of feedstuffs, such as feedstuff processing, bird age and species, and experimental procedures (Penz Jr. et al., 1999). The efficiency of energy utilization by the Japanese quails in the present study was reduced by the high CF content of 38.73% (average of all treatments). The castor oil meal C and D promoted greater MCGE and nitrogen balance than CMA and CMB. Relative to energy utilization (AME and AMEn), CMA was not significantly difference from CMC or CMD.

CMB presented the lowest metabolizability coefficients of DM, N, GE, and the lowest AME and AMEn values, as well as the lowest feed intake when added to the reference diet. These results may also be attributed to processing method (ethanol, recovery at 80 °C for 6 min, neutralization with NaOH, and drying by exposure to the sun for two days), which caused long exposure of the meal to NaOH, possibly resulting in undesirable reactions, and therefore compromising the availability of nutrients for poultry. It is should be noted that CMB also presented the lowest EE value (7.21%), and consequently lower GE value (4684 kcal/kg).

Faria Filho *et al.* (2010), evaluating broiler diets containing detoxified castor oil meal with 60g of micro-processed calcium oxide diluted in water at the ratio of 1:10, observed an AME value of 1,829 kcal/kg), which was lower than those obtained with treatments CMB and CMC and similar to CMD. The AME (2032 kcal/kg), AMEn (1829 kcal/kg), and MCDM (57.8%) values obtained by Junior *et al.* (2011) with broilers fed detoxified castor bean meal compared with those obtained with Japanese quails in the present experiment demonstrate that there are species differences.

#### CONCLUSION

The castor oil meal submitted to treatments A, C, and D showed satisfactory results for Japanese quails. However, considering the simplicity its industrial process, its low toxicity, and metabolizability coefficients

obtained, the castor oil meal obtained by recovery in ethanol at 110 °C for 15 minutes (CMD) is the most recommended for inclusion in Japanese quail diets.

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