



Revista Brasileira de Ciência Avícola

ISSN: 1516-635X

revista@facta.org.br

Fundação APINCO de Ciência e
Tecnologia Avícolas
Brasil

Göze, I; Göze, ÖF; Yelkovan, I; Çetinus, A; Saygin, H; Ercan, N
The Review of Certain In Vivo Antioxidant Effects on Essential Oils of Origanum
Minutiflorum O Schwarz-Ph Davis, Juniperus Excelsa Bieb.subsp. Excelsa and
Histopathologic Changes
Revista Brasileira de Ciência Avícola, vol. 19, núm. 2, abril-junio, 2017, pp. 333-338
Fundação APINCO de Ciência e Tecnologia Avícolas
Campinas, Brasil

Available in: <http://www.redalyc.org/articulo.oa?id=179751953021>

- How to cite
- Complete issue
- More information about this article
- Journal's homepage in redalyc.org

redalyc.org

Scientific Information System

Network of Scientific Journals from Latin America, the Caribbean, Spain and Portugal

Non-profit academic project, developed under the open access initiative



■Author(s)

Göze I^I
Göze ÖF^{II}
Yelkovan I^{III}
Çetinus ŞA^{IV}
Saygin H^V
Ercan N^{VI}

^I Göze Pharmacy, Çarşıbaşı Street, No 7, 5800

^{II} Cumhuriyet University Faculty of Medicine Department of Pathology

^{III} Department of Biology

^{IV} Faculty of Science Department of Biochemistry

^V Sivas Numune Hospital Department of Urology

^{VI} Cumhuriyet University Faculty of Veterinary Medicine Department of Biochemistry Sivas

■Mail Address

Corresponding author e-mail address
Nazlı Ercan

Cumhuriyet University Faculty of Veterinary
Medicine Department of Biochemistry
Sivas, Turkey. 58140
Tel: (90-346) 2191010-2575
Email: nazliercan@yahoo.com

■Keywords

Brain, histopathology, antioxidant enzymes, *Juniperus excelsa*, kidney, liver, *Origanum minutiflorum*.

The Review of Certain In Vivo Antioxidant Effects on Essential Oils of *Origanum Minutiflorum* O Schwarz-Ph Davis, *Juniperus Excelsa* Bieb.subsp. *Excelsa* and Histopathologic Changes

ABSTRACT

Essential oil of plants called *Juniperus excelsa* Bieb. (JE), *Origanum minutiflorum* O. Schwarz and P.H. Davis (OM) were used in this study. In order to determine experimental doses, LD₅₀ values of essential oils were determined on mice. Taking into consideration the LD₃₀ range, the experimental toxic doses were calculated for each rat (rat/kg). The toxic dosages thus determined were adapted to rats for active substances (rat/kg). Using commercially available pure virgin olive oil (VOO) as the solvent and diluting agent, OM oil (n=10), JE fruitoil (n=10), carvacrol (CRV) (n=10), VOO (n=10) and normal saline SF (n=8) were administered on the basis of 12 days into intraperitoneal (IP). Enzyme activities of Glucose-6-Phosphate dehydrogenase (G6PDH), malate dehydrogenase (MDH), Superoxide Dismutase (SOD), Glutathione-S-transferase (GST), Adenosine-deaminase (ADA) and Catalase were studied in isolates of kidney, brain and liver tissues. The data was statistically analyzed through Kruskal Wallis variance analysis. Elevated levels of GST and catalase have been found statistically important, as have both essential oil activities of OM and JE in the kidney tissue (p<0.005). All of the enzymes except the levels of ADA and SOD led to a statistically significant change in the brain and liver. There was sinusoidal hyperemia and capsular adhesion in the liver as histopathological were found to be statistically significant (p<0.005). It did not observe any important changes in the other organs. Findings were scored and analyzed by using χ^2 (chi-square) test and Fisher's definite variance analysis.

INTRODUCTION

There are about 70 kinds of *Juniperus* L. (Cupressaceae) in the world. The type of *Juniperus* is represented by 10 taxons with seven sub types in Turkey. *Juniperus excelsa* Bieb (gray tall juniper), (Cupressaceae) which is an evergreen and coniferous tree, covers 82% of juniper forests in Turkey. It has disseminated to all countries within Iranian-Turan climate zone. The fruits of this tree are registered as juniper berries in Turkish codex (Demirhan, 2011); and the researches conveyed show that it involves alpha-pinene as active ingredient in its compound (Adams *et al.*, 2013).

The fruits of *Juniperus excelsa* Bieb (J.E.) are being used as diuretic, stimulant, antiseptic and for treatment of injuries by Anatolians since the ancient ages. It is known that, its extracts bear the effects of being anti-inflammatory, anti-microbial, insecticide, anti-termite, hypotensive, diuretic; and also used as antiseptic for urinal system (Adams 1990a; Adams 1990b; Adams 2011; Demirhan 2001; Adams *et al.*, 2013). Also



it is used as flavor or fragrance material in some foods and beverages, by distilling the wood and leafs of some kinds of juniper, the juniper essence which is produced is used in perfumery and medicine industry (Adams *et al.*, 2013; Yesenofski 1996). In various studies activities of antioxidant, antibacterial, antispasmodic and chemical composition of JE were found (Dadalioğlu & Evrendilek, 2004; Sokovic *et al.*, 2004; Topçu *et al.*, 2005; Asilli *et al.*, 2008; Moein *et al.*, 2010; Emami *et al.*, 2011; Orhan *et al.*, 2011; Taviano *et al.*, 2011; Ataş *et al.*, 2012; Ehsani *et al.*, 2012; Khan *et al.*, 2012; Orhan *et al.*, 2012; Karapandzova *et al.*, 2014; Moein, 2014). The type of *Origanum* is represented by 23 kinds 32 taxons in Turkey and 41 kinds 52 taxons in the world (Baser *et al.*, 1993; Baser, 2002).

Thyme *Origanum minutiflorum* O. Schwarz and P.H. Davis (O.M.), which is known as 'Dairy thyme' and 'Tote thyme' in the thyme world market, is an endemic kind that has disseminated only at Dairy (Sütçüler) region in Isparta city and contains 80 % of carvacrol and 4.0 & p-cimen (Baser *et al.*, 1993; Baser, 2002; Dadalioğlu & Evrendilek, 2004; Goze *et al.*, 2010).

Some kinds of *Origanum* are also used as spice in food, and in medicine among people against indigestion, inappetency and cough. It bears the features such as being antiseptic, sedative, expectorant and cramp healing. It is used in cosmetics, alcoholic or alcohol free beverages. Due to its anti-bacterial effect on bacteria which causes food spoil and food poisoning, it is a most wanted medicinal plant. It is generally known for its rich essential oils, antioxidant, antibacterial, antiviral and antifungal activities (Baser *et al.*, 1993; Davis *et al.*, 1998; Baytop 1999; Baser, 2002; Dadalioğlu & Evrendilek, 2004; Baydar, 2005; Baser, 2008; Goze *et al.*, 2010; Oke 2010; Kılıçgün & Korkmaz 2014). Owing to this fact, for the natural active ingredients to be defined in plants, which are used for medication or food, it is important to convey researches that examine the biological activity, toxicological and pharmacological dose-response relation.

This study aims to determine the in vivo antioxidant activity of O.M. and J.E. on kidneys which are used orally and pursuit possible tissue changes histopathologically. Although there are many studies about in vitro antioxidant and antibacterial activities of active ingredients of O.M. and J.E., there are not many studies of in vivo. Therefore, the research findings are the first one in this field.

MATERIAL AND METHOD

JE fruit and OM were harvested from Isparta, Sütçüler flora. The samples of the plant were registered under JE (CUFH Voucher No. ED11003) and OM (CUFH Voucher No. ED 1101) numbers at the Herbarium of Cumhuriyet University Faculty of Science and Literature Department of Biology.

Plant samples which were dried in shade and pulverized, were held in Clevenger apparatus for approximately 3 hours being exposed to hydro distillation. The oil yield of 500gr dried plant was determined as 2.1 ml for JE and 10 ml for OM.

In the study, 40 healthy 180gr and approximately 4 weeks old adult albino Wistar rats (*Rattus Norvegicus* var. Albino) were used; and in order to determine the LD₅₀ dose, again, 40 healthy 25-30gr, 4 weeks old rats (*Mus Musculus* var. Albino) were used. The experimental animals were provided from Experimentary Animals Laboratory at Cumhuriyet University. Experimental Animals Ethical Committee's approval was gained (approval date and no: 1/9/2005-116). The animals were preserved under standard laboratory circumstances, with free feed and water at room temperature (21 ± 3 °C).

During literature searches no studies about OM and JE, LD₅₀ doses were found. Therefore, LD₅₀ dose of oils was determined by using rats. The toxic dose was calculated as rat/kg for each and found as LD₃₀ (for JE -LD₃₀:160 ul). The animals which were tried on with Pure OM and pure carvacrol died at all the levels of doses given. Thus, OM and carvacrol (CRV) were mixed into olive oil and a depot/store solution was prepared (1 OM+3 Olive oil); every rat was given 100u (LD₃₀) i.p. from this solution and this dose was determined as LD₃₀ for OM. In the same way, by applying 100ul to each rat from the store solution which was prepared by using carvacrol (1 carvacrol +6 olive oil), LD₃₀ value was found for the carvacrol.

This experiment was made on rats. The rats were divided into 5 groups. To the 1st group of rats JE (160ul), to the 2nd group only olive oil (40 ul), to the 3rd group OM (100ul), to the 4th group carvacrol (100ul) were injected for 12 days with i.p. and 10 rats were used as control group (SF). At the end of 12 days the rats were killed with dislocation and their organs were taken into cooled boxes.

Preparation of Homogenates: Tissues were taken and washed with 0.15 KCl. These pieces of tissue were cut on a glass; kidney, brain and liver were homogenized by using rotating 1000 rpm/min via glass homogenizer (B. Braun) after added 0.15 M KCl ratio of 1:3(w/u)



per 1 gram. The homogenates were centrifuged (Beckman Model J2) for 15 min at 4800 rpm. Glucose 6 phosphate dehydrogenase (G₆PDH), (Beutler 1971); malate dehydrogenase (MDH), (Warburg & Christian 1931); Glutathione S-transferase (GST), (Habig *et al.*, 1974); Superoxide dismutase (SOD), (Sun *et al.*, 1988); Adenosine deaminase (ADA), (Giuisti, 1974); and catalase enzymes activities were studied in tissue as spectrophotometrically (Spectro UV-VIS Double Beam PC Scanning), (Aebi, 1983). The data was statistically analyzed through Kruskal Wallis variance analysis by SPSS (Ver: 13.0) program. Standard error level is taken as 0.05 (Akgül, 2005).

The tissues samples which were sent to a pathology laboratory for routine follow up in automatic tracking device via handon Path Centre (Thermo Electron Corporation, UK). It observed adhesions among the intestine kidney, brain and liver of rats which were given JE in the macroscopic examination. After scored of findings analyzed with X² test and precise analysis of variance Fisher (Akgül, 2005).

RESULTS

Results of kidney homogenates

The activities of GST and catalase were found important as a statistics for OM and JE ($p < 0.005$).

Table1 – Analyzes of enzymes activities in kidney homogenates

	G6PDH	GST	MDH	ADA	SOD	CATALASE
OM	0.000165±0.0001	0.00150±0.0012	0.00023±0.0003	0.00047±0.0003	97.80±0.57	97.58±32.02
JE	0.00043±0.0003	0.00027±0.0002	0.00021±0.0001	0.00037±0.0000	97.96±0.84	117.95±21.80
CRV	0.00064±0.0004	0.00308±0.0019	0.00023±0.0001	0.00035±0.0001	98.13±0.45	117.75±34.76
VOO	0.00040±0.000	0.00230±0.0004	0.00022±0.0001	0.0027±0.0010	97.77±0.26	171.51±17.44
SF	0.00125±0.00012	0.00136±0.0001	0.00024±0.0001	0.00027±0.0001	98±0.60	129.48±32.87
Kw	10.19	22.69	4.35	4.44	3.34	18.20
p Value	0.037	0.000	0.003	0.366	0.502	0.001

G6PDH U/g pro; GST U/mg pro; MDH U/mg pro; ADA U/mg pro; SOD U/g pro; CATALASE U/mg pro

Table2 – Analyzes of enzymes activities in brain homogenates

	G6PDH	GST	MDH	ADA	SOD	CATALASE
OM	0.00124±0.0012	0.00167±0.0015	0.0001±0.0001	0.00038±0.0007	97.45±0.56	2.07±0.75
JE	0.00032±0.0002	0.00096±0.0004	0.00025±0.0001	0.00017±0.0001	97.52±0.58	2.79±0.81
CRV	0.00088±0.0002	0.00470±0.0050	0.00013±0.0001	0.00005±0.0001	97.13±0.15	2.62±0.75
VOO	0.0019±0.0004	0.0027±0.0002	0.00014±0.0001	0.00006±0.0000	97.67±0.67	9.76±0.60
SF	0.00151±0.0003	0.00276±0.0003	0.00016±0.000	0.00026±0.0002	96.10±2.60	10.38±0.44
Kw	19.21	18.270	17.01	11.73	5.81	14.89
p Value	0.001	0.001	0.002	0.009	0.213	0.005

G6PDH U/g pro; GST U/mg pro; MDH U/mg pro; ADA U/mg pro; SOD U/g pro; CATALASE U/mg pro

The enzymes of G6PDH, MDH, ADA and SOD did not show significant differences between the groups compared in pairs ($p > 0.005$) (Table3).

The difference between groups was not significant as G₆PDH, MDH, ADA and SOD enzymes values compared with in pairs ($p > 0.05$) Apart from these, there were significant difference in values ($p < 0.05$). Between groups of OM and CRV, JE and CRV, JE and olive found significant differences to pairwise comparisons are made in terms of value in GST ($p < 0.005$).

When catalase activity compared in pairs of groups OM and CRV, OM and VOO, JE and CRV, JE and olive oil values difference was found statistically significant as shown in Table1 and Table 2.

Results of brain homogenates

The level of G6PDH between groups of VOO /JE and JE/SF ($p < 0.005$); the level of GST between groups of JE-CRV ($p < 0.005$); the level of MDH between groups of OM-JE and CRV-JE ($p < 0.005$); the level of catalase between groups of OM- VOO and JE-VOO-SF, CRV-JE and SF were found important if statistically compared in pairs (Table 2).

Results of liver homogenates

The activity of G6PDH in between groups of OM-SF and JE-VOO and CRV/VOO/SF; the activity of GST between groups of JE-CRV and JE-VOO and JE-SF; the activity of MDH between groups of OM and VOO, the activity of catalase between groups of OM/VOO and SF were found statistically important.

In histopathological examination, a finding was not seen in kidney tubule epithelia cell except cloudy swelling ($p > 0.05$). After scored of findings analyzed



Table 3 – Analyzes of enzymes activities in liver homogenates

	G6PDH	GST	MDH	ADA	SOD	CATALASE
OM	0.00099±0.00016	0.0017±0.0016	0.00032±0.00025	0.00006±0.0001	90.81±4.44	116.17±10.2
JE	0.0021±0.0015	0.00335±0.0023	0.00024±0.0001	0.0001±0.0001	93.52±0.89	98.13±8.49
CRV	0.0034±0.0004	0.00020±0.0001	0.00016±0.000	0.0001±0.00001	93.11±1.72	103±20.4
VOO	0.0039±0.0025	0.00021±0.0003	0.00001±0.000	0.00001±0.0000	92.61±2.68	90.21±2.465
SF	0.00448±0.001	0.0001±0000	0.00014±0.0011	0.00017±0.0002	91.83±2.07	90±3.368
Kw	18.87	20.12	12.89	19.94	4.32	14.89
p Value	0.000	0.000	0.012	0.364	0.364	0.005

GP6DH U/g pro; GST U/mg pro; MDH U/mg pro; ADA U/mg pro; SOD U/g pro; CATALASE U/mg pro

with X² test and precise analysis of variance Fisher. Fuzzy swelling (FS) in surface of tissue sample according to where tissue is turned off as semi quantitative (%), focal necrosis (FN) and the group of cell necrosis as a number (present or absent), Kupffer cell hyperplasia (present or absent), periportal inflammation (present or absent), sinusoidal hyperemia (SHR) (present or absent) were evaluated (Table 4). The sinusoidal hyperemia (SH) and capsular adhesion (CA) in liver were found statistically important (p<0.05) (Figure1). There are not significant changes which in other organs were observed.

Table 4 – Evaluation of tissue samples in statistically

GROUPS		OM		JE		RESULTS
		S	%	s	%	
SCN	0	11	68.8	3	100	P=0.530
	1	5	31.3	-	-	
KOL	0	15	93.8	3	100	P=1.00
	1	1	6.3	-	-	
PPI	0	2	12.5	-	-	X2=3.20 P=0.361
	1	7	43.8	3	100	
	2	5	31.3	-	-	
	3	2	12.5	-	-	
CA	0	7	43.8	2	66.7	X2=6.69 P=0.025
	1	9	56.0	7	33.3	
FS	0	5	31.3	3	100	X2=4.89 P=0.179
	1	1	6.3	-	-	
	2	2	12.5	-	-	
	3	8	50.0	-	-	
SH	0	10	62.5	2	66.6	X2=7.71 P=0.021
	1	6	37.5	1	33.3	
	2	-	-	-	-	

SCN (Single Cell Necrosis), PPI (Periportal inflammation), KOL (Kollaps), FS (Fuzzy swelling), SH (Sinusoidal hyperemia), CA (Capsular Adhesion)

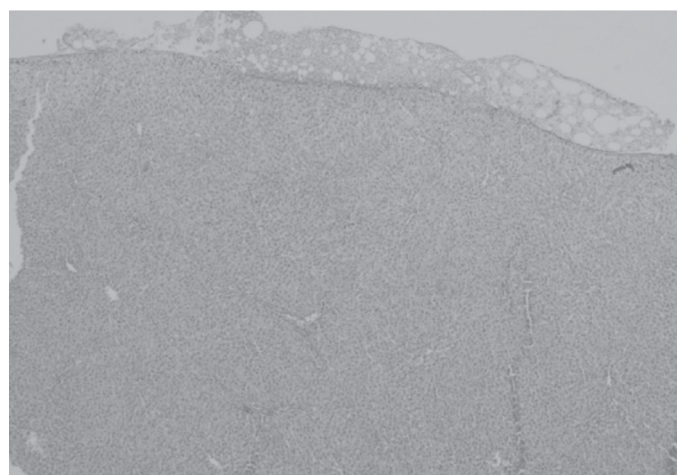


Figure1 – A general view of adhesions in the liver capsule (HEEx40).

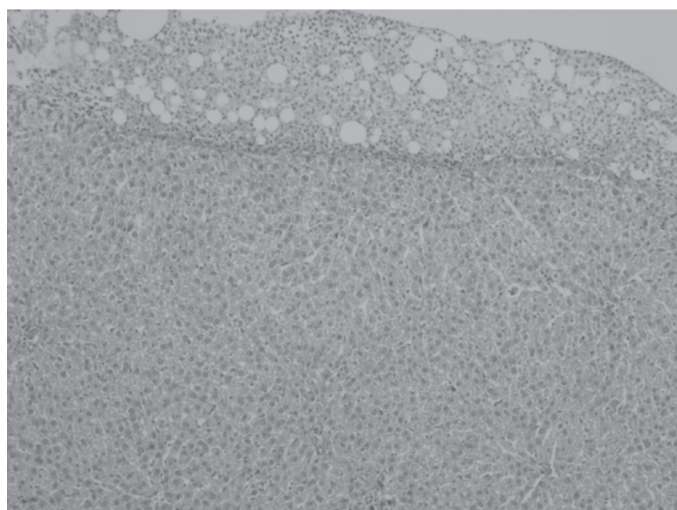


Figure 2 – The breakdown in the liver capsule adhesions: fibroadipose hipogralunom to similar tissue infiltration (HEEx100).

DISCUSSION

Serum enzyme activities are widely used in determining of organ functions in mammals. Also these are very important for environmental biology, searching for the effects of environmental pollutants on living organisms, the toxic effects of orally consumed plants and for bioavailability in recent years. These tests have been of great importance in recent

years, particularly in the context of bioavailability, to explore the aquatic effects of environmental biology and environmental pollutants and the toxic effects of orally consumed plants.

The changes that may occur in the activities of these important enzymes working in the energy metabolic pathways may affect the formation of energy by the



organism and affect the functioning of other metabolic pathways.

Superoxide dismutase (SOD), catalase (CAT), Glutathione reductase (GST) and malate dehydrogenase (MDH) are enzymatic compounds. Superoxide dismutase (EC 1.15.1.1, EC-SOD), as a superoxide anion is a form that part of the cellular antioxidant defense system by catalyzing the conversion in hydrogen peroxide to molecular oxygen. It plays an important role in controlling cellular superoxide levels in the compartment. It consists in high concentrations in the brain, liver, heart, kidney and erythrocytes (Fridovich, 1989).

In this study, SOD was found stable in all of the tissues. The mature erythrocytes don't include ribosome, mitochondria and nucleus. Enzymes of SOD, catalase, GSH-Px and pentose phosphate pathway protected against O_2 radicals. It can consider that OM and JE don't cause inhibition of the metabolic pathway.

Glucose-6-phosphate dehydrogenase (G-6-PD) (D-glucose-6-phosphate: NADP+ 1-oxidoreductase, E.C. 1.1.1.49) is the first and control enzyme of the pentose phosphate pathway. G₆PDH; it is an enzyme that plays a key role in the pentose, phosphate and glycolysis (Scott 1975). Also it has an anti-oxidant activity. G₆PDH enzyme activity was found to decrease in homogenates of all tissues. According to these results, it can be observed that essential oils may generate inhibiting change.

Adenosine deaminase is purine metabolism enzyme which is responsible for immune mechanism (Wilson *et al.*, 1991). Significant changes weren't observed in ADA activity in all organs except for the brain. In this case, the tested substances aren't effective in purine metabolism and suggest no creating damage.

MDH; is taken office in the maintenance of cell membrane integrity and the regulation of enzyme activity (Musrati *et al.*, 1998). In this research, the levels of MDH was high in the liver and in the brain but was stabile in the kidney.

GST, glutathione plays an important role against sourced interior and exterior toxic chemicals in the cellular defense system (Armstrong, 1990). Reversible damage may have been created during the period of JE and OM application by various mechanisms in all tissues. In this case, it can be considered to change their enzyme activations to maintain the redox state of cells and especially repairing damage membranes by keeping to a minimum damage. It is very important for all of tissues.

Catalase (H₂O₂: H₂O₂ oxidoreductase, EC 1.11.1.6) has 4 units with a group hemoprotein and localized in intracellular organelles as peroxisomes. Its task is to destroy hydrogen peroxide to oxygen and water (Kono & Fridovich, 1982). This is quite low compared to the catalase activity in the kidneys of the control group that can cause essential oils erythrocyte catalase activity by inhibiting the accumulation of H₂O₂ and consequently hemolysis which is very important for all of tissues.

As a result, the activity of catalase was very meaningful in all tissues while the level of SOD was stable. The levels of GST and catalase were in kidney, ADA and SOD were important in liver. Statistical differences were found in all enzymes except for SOD. It can be observed that these types of plants consumed as a food of toxic effects should have more comprehensive studies. Additionally, the LD value for each study which uses the plant's essential oils, need to be determinate. Because the composition of the plant essential oils vary depending on many reasons such as seasonal, droughts and temperature difference.

ACKNOWLEDGE

This study was supported by The Scientific Research Project Fund of Cumhuriyet University under project number SHMYO-5.

REFERENCES

- Adams RP. The volatile leaf oils of *Juniperus excelsa* M-Bieb. From a native stand in Greece versus *J. Excelsa* cultivated at Kew, London. Journal Essential of Oil Research 1990a;2:45-48.
- Adams RP. *Juniperus procera* of east Africa: volatile leaf oil composition and putative relationship to *J. excelsa*. Biochemical Systematic and Ecology 1990b;18:207-210.
- Adams RP. Junipers of the world, the genus *Juniperus*. 3rd ed. Bloomington: Trafford Publishing, 2011. p. 428.
- Adams RP, Tashev AN, Baser KHK, Christou AK. Geographic variation in volatile leaf oils of *Juniperus excelsa* M. Bieb. Phytologia 2013;95(4):279-285.
- Aebi HE. Catalase. In: Bergmeyer UH, Bergmeyer J., Grassl M. Method of enzymatic analysis. Michigan: Verlag Chemie; 1983;3: 273-286.
- Akgül A. Biyoistatistik. 3. baskı. Ankara: Emek Ofset; 2005.
- Armstrong RN. Glutathione s-transferases: reaction mechanism, structure and function. Chemical Research in Toxicology 1990;4(2):131-140.
- Asili, J, Emami SA, Rahimizadeh M, Fazly-Bazzaz BS, Hassanzadeh MK. Chemical and antimicrobial studies of *Juniperus excelsa* subsp. *Excelsa* and *Juniperus excelsa* subsp. *Polycarpus* essential oils. Journal of Essential Oil Bearing Plants 2008;11(3):292-302.
- Atas AD, Goze I, Alim A, Cetinus SA, Durmus N, Vural N, et al. chemical composition, antioxidant, antimicrobial and antispasmodic activities of the essential oil of *Juniperus excelsa* subsp. *excelsa*. Journal of Essential Oil Bearing Plants 2012;15(3):476-483.



- Baser KHC. The turkish origanum species. In: Kintzios SE, editor. Oregano. The genera origanum and lippia. London: Taylor and Francis; 2002. p.108-126
- Baser KH. Biological and pharmacological activities of carvacrol and carvacrol bearing essential oils. Current Pharmaceutical Design 2008;14:3106-3119.
- Başer KHC, Özek T, Tümen G, Sezik E. Composition of the essential oils of Turkish *Origanum Specie* with Commercial Importance. Journal Essential of Oil Research 1993;5(6):619-23.
- Baydar H. The effects of different harvest dates on essential oil content and essential oil composition in *Origanum minutiflorum* Schwarz. Akdeniz Üniversitesi Ziraat Fakültesi Dergisi 2005;18:175-178.
- Baytop T. Therapy with medical plants in Turkey, past and present). Istanbul: Nobel Tıp Kitapevleri; 1999.
- Beutler E. Red cell metabolism manual of biochemical methods. London: Academic Press; 1971.
- Dadalioğlu I, Evrendilek GA. Chemical compositions and antibacterial effects of essential oils of Turkish oregano (*Origanum minutiflorum*), bay laurel (*Laurus nobilis*), Spanish lavender (*Lavandula stoechas*L), and fennel (*Foeniculum vulgare*) on common food borne pathogens. Journal of Agricultural and Food Chemistry 2004;29/52(26):8255-8260.
- Davis PH, Mil RR, Tan K. Flora of Turkey land the east Aegean islands. Edinburgh: University Press; 1998. v.10, suppl 1.
- Demirhan Erdemir A. Şifalı bitkiler doğal ilaçlarla gelen tedaviler. Istanbul: Alfa Basım Yayım Dağıtım; 2001.
- Ehsani E, Akbari K, Teimouri M, Khadem A. Chemical composition and antibacterial activity of two *Juniperus* species essential oils. African Journal Microbiology Research 2012;6(38):6704-10.
- Emami SA, Abedindo BF, Hassanzadeh-Khayyat M. Antioxidant activity of the essential oils of different parts of *Juniperus excelsa* M. Bieb. subsp. *Excelsa* and *J. excelsa* M. Bieb. subsp. *polycarpus* (K. Koch) Takhtajan (Cupressaceae). Iranian Journal of Pharmaceutical Research 2011;10(4):799-810.
- Fridovich I. Superoxide dismutases. An adaptation to a paramagnetic gas. Journal of Biological Chemistry 1989;264:7761-7764.
- Goze I, Cetin A, Goze A Investigation of effects of essential oils of *Origanum minutiflorum* O Schwarz pH Davis and *Cyclotrichium niveum* (Labiatae) plants on angiogenesis in shell-less chick embryo culture. African Journal of Biotechnology 2010;9(14):2156-2160.
- Giuisti G. Enzyme activities. In: Bergmeyer U, ed. Methods of Enzymatic Analysis. Gmbh Weinheim, Bergest: Verlag Chemie; 1974;1092-1098.
- Habig WA, Tabst MS, Jacoby WV. Glutathion S transferase Biochem. 1974; 7130-7139.
- Karapandzova M, Stefkov G, Cvetkovikj I, Sela F, Panovska TK, Kulevanova S. Chemical characterization and radical scavenging activity of leaves of *Juniperus foetidissima*, *J. Excelsa* and *J. Communis* from Macedonian flora. Macedonian Pharmaceutical Bulletin 2014;60(2):29-37.
- Khan M, Khan A, Rehman N, Gilani A.H. Pharmacological explanation for the medicinal use of *Juniperus excelsa* in hepatoprotective gastrointestinal and respiratory disorders. Journal Natural of Medicines 2012;66:292-301.
- Kılıçgün H, Korkmaz M. Hepatoprotective and antidiabetic activity of *Origanum minutiflorum* grown wild in Turkey. Bothalia Journal 2014;44:3.
- Kono Y, Fridovich I. Superoxide radicals inhibit catalase. Journal of Biological Chemistry 1982;257:5751-5754.
- Moein MR, Ghasemi Y, Moein S, Nejati M. Analysis of antimicrobial, antifungal and antioxidant activities of *Juniperus excelsa* M. B. subsp. *polycarpus* (K. Koch) Taghtajan essential oil. Pharmacognosy Research 2010;2(3):128-31.
- Moein MR, Moein S, Mousavi F. Study on relationship between antioxidant potential and phenolic contents of *Juniperus excelsa* fruit. International Journal of Pharmacy and Pharmaceutical Science 2014;6(7):192-194.
- Musrati RA, Kollárová M, Mernik N, Mikulášová D. "Malate dehydrogenase: distribution, function and properties". General Physiology and Biophysics 1998;17(3):193-210.
- Oke F. Biological potentials and cytotoxicity of various extracts from endemic *Origanum minutiflorum* O. Schwarz & P.H. Davis. Food and Chemical Toxicology 2010;48:1728-1733.
- Orhan N, Akkol E, Ergun F. Evaluation of anti inflammatory and antinociceptive effects of some *Juniperus* species growing in Turkey. Turkish Journal of Biology 2012;36:719-726.
- Orhan N, Orhan, IE, Ergun F. Insights into cholinesterase inhibitory and antioxidant activities of five *Juniperus* species. Food and Chemical Toxicology 2011;49(9):2305-2312.
- Scott WA. Glucose-6-phosphate dehydrogenase from *Neurospora crassa*. Methods Enzymology 1975;41:177-182.
- Sokovic MD, Ristic M, Grubisic D. Chemical composition and antifungal activity of the essential oil from *Juniperus excelsa* berries. Pharmaceutical Biology 2004;42:328-331.
- Sun YI, Oberley LW, Li Y. A simple method for clinical assay of superoxide dismutase. Clinical Chemistry 1988;34(3):497-500.
- Taviano MF, Marino A, Trovato A, Bellinghieri V, La Barbera TM, Guvenc A, et al. Antioxidant and antimicrobial activities of branches extracts of five *Juniperus* species from Turkey. Pharmaceutical Biology 2011;49:1014-102
- Topçu G, Gören AC, Bilsel, G, Bilsel M, Çakmak O, Schilling J, et al. Cytotoxic Activity and Essential Oil Composition of Leaves and Berries of *Juniperus excelsa*. Pharmaceutical Biology 2005;43(2):125-8.
- Warburg O, Christian H (1931a). Aktivierung von Kohlehydrat in roten Blutzellen. Biochem. Z. 238:131.
- Wilson DK, Rudolph FB, Quirocho FA. Atomic structure of adenosine deaminase complexed with a transition-state analog: understanding catalysis and immunodeficiency mutations. Science 1991;252(5010):1278-1284.
- Yesenofski J. Juniper oil distillation and marketing project: western juniper commercialization program. Oregon: The Confederated Tribes of the Warm Springs Reservation; 1996. (Final Report).