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Fishes from the Jaru Biological Reserve, Machado River drainage, Madeira River basin, Rondônia State, northern Brazil

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Abstract: This work assessed freshwater fishes collected at 12 stations located along the Tarumã River, middle Machado River, Madeira River basin. The fieldwork took place in May and September 2015 during the high and low water seasons, respectively. We gathered 1,482 specimens representing seven orders, 30 families, 54 genera and 74 species using seine, gill, and hand nets. The family Characidae was the most representative, exhibiting the highest number of species captured. The species with the greatest abundance were *Hemigrammus vorderwinkleri*, *Hyphessobrycon bentosi*, *Hemigrammus* cf. *bellottii*, *Bryconella pallidifrons*, and *Apistogramma resticulosa*. Two species that remained unidentified are probably a new species. This study represents the third fish survey totally conducted at a Conservation Unit in the Rondônia State, and will certainly provide valuable information for future investigations on biodiversity conservation in the Machado River.

Keywords: Amazon, freshwater, conservation, ichthyofauna, inventory.

Peixes da Reserva Biológica do Jaru, drenagem do rio Machado, bacia do rio Madeira, Estado de Rondônia, norte do Brasil

Resumo: Este trabalho avaliou os peixes de água doce coletados em 12 estações localizadas ao longo do rio Tarumã, médio rio Machado, bacia do rio Madeira. O trabalho de campo ocorreu em maio e em setembro de 2015 durante os períodos de cheia e seca, respectivamente. Um total de 1.482 exemplares, sete ordens, 30 famílias, 54 gêneros e 74 espécies foram amostrados com malhadeiras, picaré e puça. Characidae foi a família com maior número de espécies capturadas. As espécies com maior abundância foram *Hemigrammus vorderwinkleri*, *Hyphessobrycon bentosi*, *Hemigrammus* cf. *bellottii*, *Bryconella pallidifrons* e *Apistogramma resticulosa*. Duas espécies não foram identificadas e provavelmente podem ser novas espécies. Este estudo representa a terceira pesquisa de peixes totalmente realizada em uma Unidade de Conservação no Estado Rondônia, e certamente fornecerá informações valiosas para futuros estudos sobre a conservação da biodiversidade do rio Machado.

Palavras-chave: Amazônia, água doce, conservação, ictiofauna, inventário.

Introduction

Among the 33,984 fish species described around the world (Eschmeyer & Fong 2016), at least 13,000 varieties are unique to freshwater environments (Nelson et al. 2016). The Neotropical realm (South and Central America) encompasses more than 4,000 valid fish species (Reis 2013), with the Amazon River basin harboring the world's greatest diversity of freshwater fish (Freitas et al. 2010, Winemiller et al. 2016, Reis et al. 2016). To date, most studies of the Amazonian ichthyofaunal diversity have concentrated in the floodplains adjacent to large rivers (cf. Lowe-McConnell 1999) and next to urban areas (Mendonça et al. 2005, Souza et al. 2016). Notwithstanding, there are few reports on the ichthyofauna inhabiting environmental areas of high conservation value (Camargo & Giarrizzo 2007, Oliveira et al. 2009, Pedroza et al. 2012, Vieira et al. 2016).

Despite concerns about the effectiveness of conservation units in the tropics, there is growing evidence that they have been surprisingly useful tools for curbing deforestation (e.g. Jaru Biological Reserve) (Bruner et al. 2001) or conservation of fish species (Frederico et al. 2016). Historically, the majority of the current protected areas in Brazil were created with disregard for the aquatic environment (Agostinho et al. 2005, Abell et al. 2007). Nevertheless, it is uncontested that they can protect many water bodies and, thus, play a highly important role in the conservation of freshwater organisms (Agostinho et al. 2005).

This study presents a survey of the ichthyofauna of the Tarumã River, a right-bank tributary of the Machado River, located within the Jaru Biological Reserve. This area displays virtually unchanged habitats in respect to its external environment. As the Tarumã River flows within a legally protected area, the objective of this study is to provide a first

ichthyofaunal inventory, which may eventually support future studies on fish biology and conservation.

Material and Methods

1. Study area

The Jaru Biological Reserve (Rebio Jaru) was established on July 11, 1979, under Federal Decree-law number 83,716, and is managed by the Instituto Chico Mendes de Conservação da Biodiversidade/Ministério do Meio Ambiente (ICMBio/MMA). Marked by the high degree of conservation, the rain forest of the Rebio Jaru is practically intact. The reserve has a humid tropical climate with temperatures varying between 23°C and 26°C, and the average annual rainfall ranges from 1,700 to 2,400 mm. The dry season occurs between May and October (Justina 2009).

The Rebio Jaru hydrographic network is part of the Machado River Basin located in eastern Rondônia State, northern Brazil. The Tarumã River, the main sub-basin of the Rebio Jaru, runs almost entirely (99%) within the Rebio Jaru. The average depth of the Tarumã River during the dry season was 2.8 ± 0.9 m; the average width, 32.8 ± 7.8 m; and the average water speed, 0.4 ms^{-1} . On the other hand, the average depth, width, and water flow values are 5.6 ± 1.2 m; 41.9 ± 4.0 m; and $0.3 \pm 0.1 \text{ ms}^{-1}$ during the wet season, respectively. The Tarumã River has many rapids flowing across the granitic formation of the Serra da Providência and Jamari complex (Justina 2009). The high transparency of the water (average transparency_{dry} = 1.2 ± 2.1 m; and average transparency_{wet} = 1.1 ± 0.4 m) due to the low amount of sediment, characterizes the Tarumã as a clear water river.

2. Data collection

We performed collections of freshwater fish in the Tarumã River in May and September 2015. Each expedition lasted four days. Our samplings comprised 12 sites in two different aquatic environments: five in the main channel and seven in the small stream channels (igarapés) of the Tarumã River (Table 1, Figure 1).

The physical conditions of some sampling stations in the Tarumã River basin are shown in Figure 2.

Stream 4 (S4) – igarapé, 1.81 m wide and 0.26 m deep, preserved riparian vegetation, swift current, sand, pebbles, and dead leaves at the bottom (Figure 2a).

Stream 6 (S6) – igarapé, 1.39 m wide and 0.21m deep, preserved riparian vegetation, slow current, sand, pebbles, large branches and trunks, and dead leaves at the bottom (Figure 2b).

River 1 (R1) - stretch, 112 m wide and 6.2 m deep located near the mouth of the Tarumã River, preserved riparian vegetation and slow current (Figure 2c).

Stream 5 (S5) – igarapé, 0.70 m wide and 0.25 m deep, preserved riparian vegetation, slow current, sand, pebbles, large branches and trunks at the bottom (Figure 2d).

Stream 3 (S3) – igarapé, 1.00 m wide and 0.21 m deep, preserved riparian vegetation, swift current, sand, bare ravine, large branches and trunks at the bottom (Figure 2e).

River 4 (R4) - stretch, 30 m wide and 4.5 m deep located near the head of the Tarumã River, preserved riparian vegetation, slow current (Figure 2f).

We accomplished the collections in the river channels using a total of eight gill nets with standard size meshes of 2×20 m, and fishing nets with mesh sizes varying from 30 to 100 mm (between opposite knots). The fishing nets were set at each sampling site during the morning, from 8:00 am to 12:00 pm, and at night, from 8:00 pm to 5:00 am. For the same period, we used a trotline with four 5/0 hooks with ends tied either to the bank vegetation or to mooring spikes. We used some pieces of piranha, *Serrasalmus rhombeus* Linnaeus 1766, as baits attached to the trotline hooks.

In small streams, the fish collection in a stretch of 80 m lasted one hour during the daytime. Three collectors used hand gathering techniques with a seine net (1.5×2 m, 2 mm mesh) and a hand net (0.5×0.8 m, 2 mm mesh) along the entire stretch, selecting the best technique for each environment. Before the collections, the ends of the sampling sections were blocked with fine-mesh nets (5 mm between opposite knots) to prevent fish escapes, regardless of the capture method used. Abiotic data, such as depth, width, and soil were observed *in situ*. We sacrificed the specimens in a solution of clove oil (Eugenol, 2 drops per liter; cf. American Veterinary Medical Association 2001). After that, the fish were fixed in 10% formalin solution and then preserved in 70% ethanol. For species identifications, we consulted the most currently accepted taxonomic literature and identification keys (Queiroz et al. 2013b). The classification followed Nelson et al. (2016). The specimens were deposited in the Coleção de Peixes, Universidade Federal do Mato Grosso, Cuiabá, MT, Brazil (CPUFMT); Laboratório de Ictiologia de Ribeirão Preto da Universidade de São Paulo, Ribeirão Preto, SP, Brazil (LIRP) and Museu de Zoologia da Universidade de São Paulo, São Paulo, SP, Brazil (MZUSP) (Appendix 1). The fish sampling was authorized by the Instituto Chico Mendes de Conservação da Biodiversidade (ICMBio, License: 48723–2/2015).

Results

Our sampling comprised 1,482 specimens representing seven orders, 33 families, 54 genera, and 74 species. A total of 1,263 specimens representing six orders, 20 families, and 50 species were collected in the streams and 219 specimens; five orders, 13 families, and 24 species were collected in the river (Table 2). Characiformes, Siluriformes, and Cichliformes represented 59% (47 species), 21% (17 species), and 10%

Table 1. Sampling sites in the Tarumã River basin with environmental classification, geographic coordinates, and altitude.

Site	Environments	Geographic coordinates	Altitude (m)
R1	River	9°27'19" S, 61°40'43" W	75
R2	River	9°32'15" S, 61°40'13" W	90
R3	River	9°42'46" S, 61°39'42" W	111
R4	River	9°46'23" S, 61°38'45" W	119
R5	River	9°47'04" S, 61°40'19" W	141
S1	Stream	9°27'28" S, 61°40'34" W	80
S2	Stream	9°27'51" S, 61°40'8" W	82
S3	Stream	9°31'44" S, 61°40'13" W	90
S4	Stream	9°32'31" S, 61°38'54" W	181
S5	Stream	9°41'47" S, 61°39'47" W	97
S6	Stream	9°44'05" S, 61°39'31" W	126
S7	Stream	9°46'58" S, 61°40'11" W	130

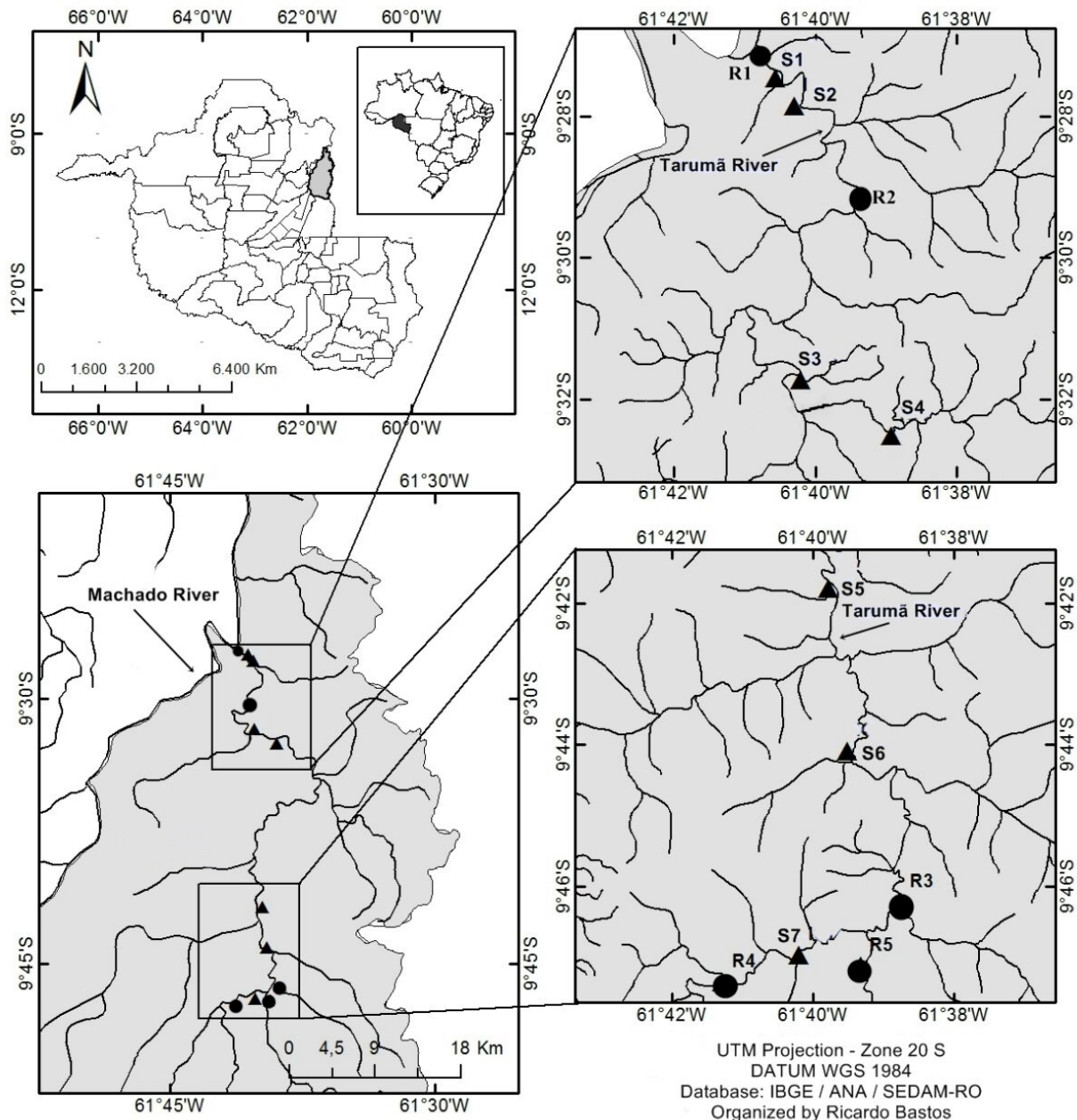


Figure 1. Map of the study area showing the collection stations in the drainage systems in the Jaru Biological Reserve (shaded area), Rondônia, Brazil. Triangles represent streams and circles represent collection points in the river channels of the Tarumã River.

(eight species) of all species, respectively. Nevertheless, Cichliformes were the second most dominant regarding the abundance of capture (6%, $n = 101$). The Myliobatiformes, Beloniformes, and Synbranchiformes orders showed richness and abundance lower than 4% ($n = 6$) and 2% ($n = 6$), respectively (Figure 3). In the streams the Characiformes, Siluriformes, and Cichliformes represented 52% (26 species), 28% (14 species), and 10% (five species) of all species, respectively. Nevertheless, Cichliformes were the second most dominant regarding the abundance of capture (7%, $n = 97$). In the river the results were similar to streams, where the Characiformes, Siluriformes, and Cichliformes represented 75% (18 species), 8% (two species), and 8% (two species) of all species, respectively. The other orders showed richness and abundance lower than 6% (Table 2). The families

with the highest richness and abundance were Characidae (25 species, 31%; $n = 874$, 58%), Serrasalminidae (six species, 8%; $n = 129$, 9%), and Cichlidae (seven species, 9%; $n = 100$, 7%) (Figure 4). In the stream environment the families with the highest richness and abundance were Characidae (18 species, 36%; $n = 1085$, 78%), Cichlidae (five species, 10%; $n = 97$, 7%) and Crenuchidae (four species, 8%; $n = 92$, 6%), for the river environment the families Characidae (20 species, 5%; $n = 208$, 50%), Serrasalminidae (21 species, 5%; $n = 127$, 31%), and Cynodontidae (two species, 8%; $n = 23$, 5%) were the most representative in richness and abundance. The other families showed richness and abundance lower than 8% (Table 2). *Hemigrammus vorderwinkleri* Géry 1963 ($n = 202$), *Hyphessobrycon bentosi* Durbin 1908 ($n = 196$), *Hemigrammus cf. bellottii*



Figure 2. Habitats of some collection stations in the drainage systems in the Jarú Biological Reserve, Tarumã River Basin, Rondônia, Brazil. (a) stream 4; (b) stream 6; (c) stretch 1 of the river channels; (d) stream 5; (e) stream 3; (f) stretch 4 of the river channels.

Steindachner 1882 ($n = 185$), *Bryconella pallidifrons* Fowler 1946 ($n = 147$) ($n = 100$), and *Apistogramma resticulosa* Kullander 1980 ($n = 86$) were the most abundant species in the total of specimens collected and these species also were predominant in the stream environment. The species *Serrasalmus rhombeus* ($n = 45$), *Serrasalmus compressus* ($n = 30$) and *Myloplus lobatus* ($n = 24$) were the most abundant in the river environment.

Discussion

Most of the several studies conducted in different portions of the Madeira River Basin in the Brazilian territory have focused either on a relatively small area or on specific tributaries. These investigations identified 122 species from the Jamari River (Santos 1996), 133 from the Marmelos River (Camargo & Giarrizzo 2007), 447 from the Aripuanã and middle Madeira rivers (Py-Daniel et al. 2007), 74 from the Belmont Stream (Araújo et al. 2009), 160 from the Guariba and Roosevelt Rivers (Pedroza et al. 2012), 189 from the Cuniã Lake (Queiroz et al. 2013a), and 174 from the middle Madeira River (Torrente-Vilares et al. 2011). A more comprehensive inventory identified 820 species along the Madeira River Basin (Queiroz et al. 2013b). More specifically, in the Machado River Basin, 48 species were recorded near the urban area of the Cacoal town (Perin et al. 2007), and 140 species in different tributaries (Casatti et al. 2013). However, contrary to the previously mentioned works, only the

present study and the assays of Queiroz et al. (2013a) and Vieira et al. (2016) were conducted entirely in a Conservation Unit in the Rondônia State, resulting in the identification of 74, 189, and 141 species, respectively. The current work contributed 24 species to the general inventory of the Machado River and 26 species to the inventory conducted in Conservation Units in the Rondônia State. Additionally, the material deposited in the Brazilian collections enabled the accomplishment of recent taxonomic studies, which contributed to enhancing the knowledge of the ichthyofauna of both the Madeira River (Rocha et al. 2008a, Rocha et al. 2008b, Zanata & Ohara 2009, Ribeiro et al. 2011, Ohara 2012, Marinho & Ohara 2013, Zanata & Ohara 2015, Ohara & Lima 2015a, Tencatt & Ohara 2016a, Ohara & Neuhaus 2016, Tencatt & Ohara 2016b, Ohara et al. 2016a, Pastana & Ohara 2016), and the Machado River (Ohara & Lima 2015b, Ohara & Marinho 2016, Ohara et al. 2016b).

Among the sampled species from the Tarumã River, almost 44% (35 species) appear on the list of ornamental fish (Brasil 2012). Some of those species, such as *Apistogramma resticulosa*, *Hemigrammus ocellifer*, *Hemigrammus vorderwinkleri*, *Hemigrammus cf. bellottii*, *Hyphessobrycon bentosi*, *H. copelandi*, *Elachocharax pulcher*, and *Serrasalmus rhombeus* (Table 2) were well represented in our survey, suggesting a relatively high local abundance. However, according to the Environmental Crimes Law number 9605, of February 12, 1998, the capture of ornamental fish in the Rebio Jarú is prohibited (ICMBio 2010).

Table 2. Fish captured in the Tarumã River, Rondônia, in May and September 2015. Method of collection: T – Trotline, G – Gillnets and S/H – Seine nets (*picarés*) and hand net (*puçã*). * Potential for the ornamental fish trade (Brasil 2012), ¹ species additional to species list to studied watershed. Systematic positions were based on Nelson et al. (2016).

Class/Order/Family/Species	Streams	River	Method
CLASS CHONDRICHTHYES			
MYLIOBATIFORMES			
Potamotrygonidae			
<i>Potamotrygon falkneri</i> Castex & Maciel 1963 ¹	-	1	T
CLASS OSTEICHTHYES			
CHARACIFORMES			
Acestrorhynchidae			
<i>Acestrorhynchus falcatus</i> Bloch 1794*	2	-	S/H
<i>Acestrorhynchus falcirostris</i> Cuvier 1819*	-	2	G
Alestidae			
<i>Chalceus guaporensis</i> Zanata & Toledo-Piza 2004	-	1	G
Characidae			
<i>Axelrodia stigmatias</i> Fowler 1913	7	-	S/H
<i>Bario steindachneri</i> Eigenmann 1893	1	-	S/H
<i>Bryconella pallidifrons</i> Fowler 1946	147	-	S/H
<i>Brycon amazonicus</i> Spix & Agassiz 1829	-	2	G/T
<i>Brycon</i> cf. <i>pesu</i> Müller & Troschel 1845 ¹	-	3	G
<i>Brycon falcatus</i> Müller & Troschel 1844 ¹	-	2	G
<i>Creagrutus anary</i> Fowler 1913 ¹	6	-	S/H
<i>Hemigrammus analis</i> Durbin 1909	25	-	S/H
<i>Hemigrammus</i> cf. <i>bellottii</i> Steindachner 1882*	185	-	S/H
<i>Hemigrammus ocellifer</i> Steindachner 1882*	45	-	S/H
<i>Hemigrammus melanochrous</i> Fowler 1913	51	-	S/H
<i>Hemigrammus vorderwinkleri</i> Géry 1963*	202	-	S/H
<i>Hyphessobrycon agulha</i> Fowler 1913*	26	-	S/H
<i>Hyphessobrycon copelandi</i> Durbin 1908*	42	-	S/H
<i>Hyphessobrycon bentosi</i> Durbin 1908*	196	-	S/H
<i>Hyphessobrycon sweglesi</i> Géry 1961	2	-	S/H
<i>Jupiaba zonata</i> Eigenmann 1908* ¹	2	-	S/H
<i>Moenkhausia collettii</i> Steindachner 1882*	5	-	S/H
<i>Moenkhausia oligolepis</i> Günther 1864*	10	-	S/H
<i>Moenkhausia mikia</i> Marinho & Langeani 2010	1	-	S/H
<i>Phenacogaster</i> cf. <i>retropinnus</i> Lucena & Malabarba 2010 ¹	3	-	S/H
<i>Roeboides affinis</i> Günther 1868 ¹	-	4	G
<i>Triportheus albus</i> Cope 1872*	-	4	G
Gasteropelecidae			
<i>Carnegiella strigata</i> Günther 1864*	25	-	S/H
Crenuchidae			
<i>Characidium</i> cf. <i>pellucidum</i> Eigenmann 1909	20	-	S/H
<i>Characidium</i> aff. <i>zebra</i> Eigenmann 1909 ¹	2	-	S/H
<i>Elachocharax pulcher</i> Myers 1927*	68	-	S/H
<i>Microcharacidium</i> sp. ¹	2	-	S/H
Erythrinidae			
<i>Hoplias malabaricus</i> Bloch 1794*	9	-	S/H
Lebiasinidae			
<i>Pyrrhulina</i> cf. <i>brevis</i> Steindachner, 1876	3	-	S/H
Ctenolucidae			
<i>Boulengerella cuvieri</i> Spix & Agassiz 1829	-	16	G
Hemiodontidae			
<i>Hemiodus unimaculatus</i> Bloch 1794 *	-	1	G
Anostomidae			
<i>Leporinus friderici</i> Bloch 1794	-	5	G
Cynodontidae			
<i>Hydrolycus armatus</i> Jardine 1841 ¹	-	17	G/T
<i>Hydrolycus tatauaia</i> Toledo-Piza, Menezes & Santos, 1999* ¹	-	6	G
Serrasalminidae			

Table 2. Continued...

Class/Order/Family/Species	Streams	River	Method
<i>Myloplus lobatus</i> Valenciennes 1850 ¹	-	24	G
<i>Myloplus rubripinnis</i> Müller & Troschel 1844*	-	24	G
<i>Pygocentrus nattereri</i> Kner 1858*	-	4	G
<i>Serrasalmus compressus</i> Jégu, Leão & Santos, 1991 ¹	-	30	G
<i>Serrasalmus rhombeus</i> Linnaeus 1766*	-	45	G
Prochilodontidae			
<i>Prochilodus nigricans</i> Agassiz 1829	-	21	G
SILURIFORMES			
Aspredinidae			
<i>Bunocephalus coracoideus</i> Cope 1874* ¹	9	-	S/H
Trichomycteridae			
<i>Ituglanis amazonicus</i> Steindachner 1882	1	-	S/H
Callichthyidae			
<i>Corydoras</i> cf. <i>trilineatus</i> Cope 1872 ¹	1	-	S/H
<i>Corydoras</i> cf. <i>armatus</i> Günther 1868 ¹	5	-	S/H
Cetopsidae			
<i>Helogenes marmoratus</i> Günther 1863*	5	-	S/H
Heptapteridae			
<i>Mastiglanis asopos</i> Bockmann 1994	1	-	S/H
<i>Nemuroglanis furcatus</i> Ribeiro, Pedroza & Rapp Py-Daniel, 2011 ¹	16	-	S/H
<i>Pimelodella howesi</i> Fowler 1940	4	-	S/H
<i>Pimelodella</i> cf. <i>steindachneri</i> Eigenmann 1917 ¹	1	-	S/H
Pseudopimelodidae			
<i>Microglanis poecilus</i> Eigenmann 1912* ¹	1	-	S/H
Doradidae			
<i>Acanthodoras spinosissimus</i> Eigenmann & Eigenmann 1888* ¹	1	-	S/H
<i>Physopyxis lyra</i> Cope 1871	1	-	S/H
Loricariidae			
<i>Ancistrus</i> sp.	1	-	S/H
<i>Rineloricaria lanceolata</i> Günther 1868*	2	-	S/H
Pimelodidae			
<i>Pimelodus ornatus</i> Kner 1858* ¹	-	2	T
<i>Platynemichthys notatus</i> Jardine 1841 ¹	-	1	T
GYMNOTIFORMES			
Hypopomidae			
<i>Hypopygus lepturus</i> Hoedeman 1962*	15	-	S/H
Rhamphichthyidae			
<i>Gymnorhamphichthys rondoni</i> Miranda-Ribeiro 1920*	2	-	S/H
Gymnotidae			
<i>Gymnotus coropinae</i> Hoedeman 1962	8	-	S/H
BELONIFORMES			
Belonidae			
<i>Potamorhaphis guianensis</i> Jardine 1843*	3	-	S/H
SYNBRANCHIFORMES			
Synbranchidae			
<i>Synbranchus madeirae</i> Rosen & Rumney 1972	2	-	S/H
CICHLIFORMES			
Cichlidae			
<i>Aequidens</i> cf. <i>tetramerus</i> Heckel 1840*	4	-	S/H
<i>Apistogramma resticulosa</i> Kullander 1980*	86	-	S/H
<i>Crenicara punctulatum</i> Günther 1863*	3	-	S/H
<i>Crenicichla regani</i> Ploeg 1989*	2	-	S/H
<i>Crenicichla santosi</i> Ploeg 1991 ¹	2	-	S/H
<i>Cichla pleiozona</i> Kullander & Ferreira 2006	-	2	G
<i>Satanoperca jurupari</i> Heckel 1840*	-	1	G
ACANTHURIFORMES			
Sciaenidae			
<i>Pettilipinnis grunniens</i> Jardine & Schomburgk 1843 ¹	-	1	G

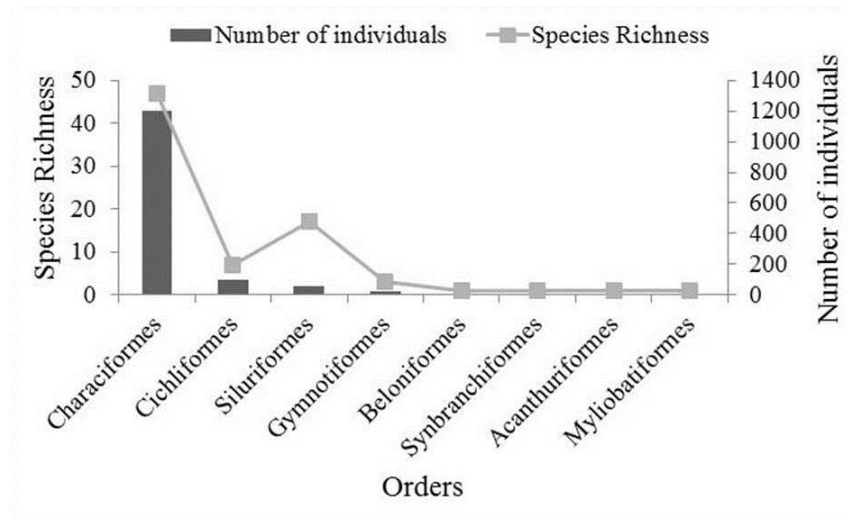


Figure 3. Number of individuals and richness of species of the fish orders collected in the Tarumã River Basin, Rondônia, Brazil.

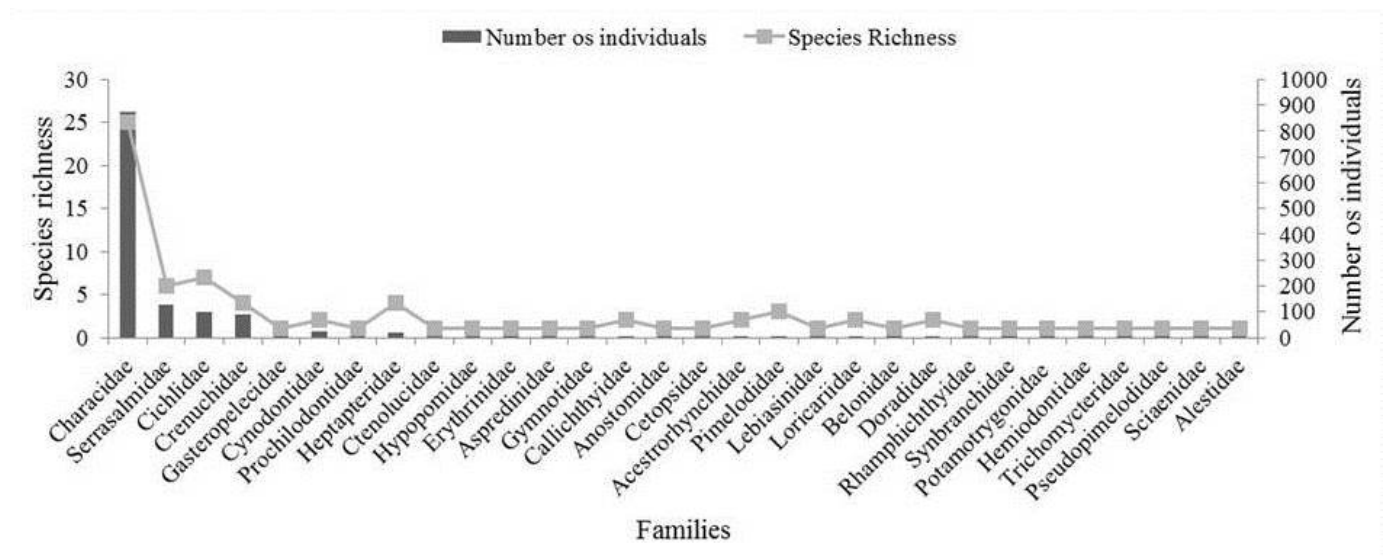


Figure 4. Number of individuals and richness of species of the fish families collected in the Tarumã River Basin, Rondônia, Brazil.

The presence of migratory species in the Tarumã River indicates that this area can be used for reproduction and/or feeding. (*Brycon amazonicus* Spix & Agassiz 1829, *B. falcatus* Müller & Troschel 1844, *Triportheus albus* Cope 1872, *Prochilodus nigricans* Agassiz 1829, *Myloplus lobatus* Valenciennes 1850, *Myloplus rubripinnis* Müller & Troschel 1844, and *Platynemichthys notatus* Jardine 1841).

Some rare species, considered as such due to their absence in most major ichthyological collections, were recorded in the Tarumã River. These included the Characiformes *Axelorodia stigmatias* Fowler 1913, *Bario steindachneri* Eigenmann 1893, *Chalceus guaporensis* Zanata & Toledo-Piza 2004, *Creagrutus anary* Fowler 1913, and *Hemigrammus melanochrous* Fowler 1913, the Siluriformes, *Nemuroglanis furcatus* Ribeiro, Pedroza & Rapp Py-Daniel, 2011, and the Cichliformes, *Crenicichla santosi* Ploeg 1991. Two taxa were provisionally identified, due to their uncertain taxonomic status. They may be records of new species, such as *Microcharacidium* sp. (Crenuchidae) and *Ancistrus* sp. (Loricariidae). We emphasize that none of the sampled species are on the IUCN Red List.

Several species were discriminated with the use of “cf” or “aff”, indicating that the number of new species may be higher. For example, the species identified herein as *Brycon* cf. *pesu* Müller & Troschel 1845 belongs to a complex of species, where new species are undergoing a description process (Zanata & Lima pers. comm.). Several species, namely, *Hemigrammus* cf. *bellottii*, *Phenacogaster* cf. *retropinnus* Lucena & Malabarba 2010, *Characidium* cf. *pellucidum* Eigenmann 1909, *Characidium* aff. *zebra* Eigenmann 1909, *Corydoras* cf. *armatus* Günther 1868, and *Corydoras* cf. *trilineatus* Cope 1872, belong either to poorly known taxonomic groups or represent still undescribed species, meaning that further taxonomic studies will be necessary.

Our work highlights the importance of conducting studies within protected areas and strengthens the role of territorial spaces with relevant environmental characteristics in a context where only three inventories within 14 Conservation Units in Rondônia have taken place. The presence of protected areas may help mitigate environmental impacts and maintain the biological integrity of a region surrounded by a long history of anthropogenic

disturbances (deforestation, gold mining, the construction of the BR 364 road and, more recently, the building of large hydroelectric dams).

Appendix 1

Voucher specimens.

MYLIOBATIFORMES: *Potamotrygon falkneri**; **CHARACIFORMES:** *Acestrorhynchus falcatus* (MZUSP 118769, MZUSP 118812), *Acestrorhynchus falcirostris* (UFRO-I 5523, UFRO-I 18313), *Axelrodia stigmatias* (MZUSP 118772, MZUSP 118786), *Bario steindachneri* (MZUSP 118734), *Bryconella pallidifrons* (MZUSP 118744, MZUSP 118791, MZUSP 118724, MZUSP 118759), *Brycon amazonicus* (MZUSP 14017), *Brycon* cf. *pesu* (LIRP 11773, UFRO-I 14213, UFRO-I 15506), *Brycon falcatus* (LIRP 13045, 10269), *Chalceus guaporensis* (UFRO-I 4321, UFRO-I 17473), *Creagrutus anary* (MZUSP 118745, MZUSP 118779), *Hemigrammus ocellifer* (MZUSP 118738, MZUSP 118767, MZUSP 118798, MZUSP 118725), *Hemigrammus* cf. *bellottii* (MZUSP 118765, MZUSP 118741, MZUSP 118790, MZUSP 118733), *Hemigrammus melanocephalus* (MZUSP 118743, MZUSP 118802, MZUSP 118726), *Hemigrammus analis* (MZUSP 118758, MZUSP 118788), *Hemigrammus vorderwinkleri* (MZUSP 118766, MZUSP 118794), *Hyphessobrycon agulha* (MZUSP 118785, MZUSP 118742, MZUSP 118797), *Hyphessobrycon copelandi* (MZUSP 118763), *Hyphessobrycon bentosi* (MZUSP 118764, MZUSP 118792), *Hyphessobrycon sweglesi* (MZUSP 118732), *Jupiaba zonata* (MZUSP 118762), *Moenkhausia oligolepis* (MZUSP 118753), *Moenkhausia mikia* (MZUSP 118755), *Moenkhausia collettii* (MZUSP 118770), *Phenacogaster* cf. *retropinnus* (MZUSP 118754, MZUSP 118789), *Roeboides affinis* (CPUFMT 3393), *Triporthus albus* (CPUFMT 3398), *Carnegiella strigata* (MZUSP 118760), *Characidium* cf. *pellucidum* (MZUSP 118757, MZUSP 118801, MZUSP 118727), *Characidium* aff. *zebra* (MZUSP 118774, MZUSP 118793), *Elachocharax pulcher* (MZUSP 118756, MZUSP 118796, MZUSP 118719), *Microcharacidium* sp. 2 (MZUSP 118781, MZUSP 118778), *Hoplias malabaricus* (MZUSP 118751, MZUSP 118768, MZUSP 118776, MZUSP 118813), *Pyrrhulina* cf. *brevis* (MZUSP 118740, MZUSP 118730), *Boulengerella cuvieri* (CPUFMT 3392), *Hemiodus unimaculatus* (UFRO-I 12750, UFRO-I 14109), *Leporinus friderici* (CPUFMT 3400), *Hydrolycus armatus* (CPUFMT 3390), *Hydrolycus tatauaia* (LIRP 10293, 10298), *Myloplus lobatus* (CPUFMT 3394), *Myloplus rubripinnis* (CPUFMT 3397), *Pygocentrus nattereri* (CPUFMT 3401), *Serrasalmus compressus* (CPUFMT 3399), *Serrasalmus rhombeus* (CPUFMT 3391), *Prochilodus nigricans* (CPUFMT 3396); **SILURIFORMES:** *Bunocephalus coracoideus* (MZUSP 118784, MZUSP 118747, MZUSP 118787, MZUSP 118714), *Ituglanis amazonicus* (MZUSP 118808), *Corydoras* cf. *trilineatus* (MZUSP 118748), *Corydoras* cf. *armatus* (MZUSP 118752), *Helogenes marmoratus* (MZUSP 118800, MZUSP 118715), *Mastiglanis asopos* (MZUSP 118737), *Nemuroglanis furcatus* (MZUSP 118739, MZUSP 118721, MZUSP 118805), *Pimelodella howesi* (MZUSP 118782, MZUSP 118736), *Pimelodella* cf. *steindachneri* (MZUSP 118729), *Microglanis poecilus* (MZUSP 118717), *Acanthodoras spinosissimus* (MZUSP 118810), *Physopyxis lyra* (MZUSP 118771), *Ancistrus* sp. 1 (MZUSP 118728), *Rineloricaria lanceolata* (MZUSP 118735, MZUSP 118773), *Pimelodus ornatus* (LIRP 11969, 12177), *Platynemateichthys notatus* (UFRO-I 3835); **GYMNOTIFORMES:** *Hypopygus lepturus* (MZUSP 118803, MZUSP 118723), *Gymnorhamphichthys rondoni* (MZUSP 118775, MZUSP 118799), *Gymnotus coropinae* (MZUSP 118783, MZUSP 118809, MZUSP 118722); **BELONIFORMES:** *Potamorhaphis guianensis* (MZUSP 118811); **SYNBRANCHIFORMES:** *Synbranchus madeirae* (MZUSP 118795, MZUSP 118720); **CICHLIFORMES:** *Aequidens* cf. *tetramerus* (MZUSP 118780, MZUSP 118777, MZUSP 118718, MZUSP 118814), *Apistogramma resticulosa* (MZUSP 118761, MZUSP 118806, MZUSP 118731), *Crenicara punctulatum* (MZUSP 118746, MZUSP 118804), *Crenicichla regani* (MZUSP 118749), *Crenicichla santosi* (MZUSP 118750,

MZUSP 118807), *Cichla pleiozona* (CPUFMT 3395), *Satanoperca jurupari* (UFRO-I 16652, UFRO-I 17429); **ACANTHURIFORMES:** *Petilipinnis grunniens* (UFRO-I 4883, LIRP 10405). * In process of deposit in Museu de Zoologia da Universidade de São Paulo, São Paulo, SP, Brazil (MZUSP).

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Author Contributions

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