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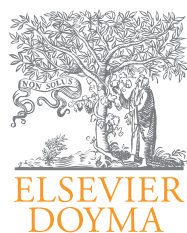
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# REVISTA ARGENTINA DE MICROBIOLOGÍA

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## BRIEF REPORT

### Natural occurrence of entomophthoroid fungi of aphid pests on *Medicago sativa* L. in Argentina

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#### KEYWORDS

Lucerne;  
Aphids;  
Entomopathogenic  
fungi

#### Abstract

Four species of entomophthoroid fungi, *Pandora neoaphidis* (Entomophthorales: Entomophthoraceae), *Zoophthora radicans* (Entomophthorales: Entomophthoraceae), *Entomophthora planchoniana* (Entomophthorales: Entomophthoraceae) and *Neozygites fresenii* (Neozygites: Neozygitaceae) were found to infect *Aphis craccivora*, *Therioaphis trifolii*, and *Acyrtosiphon pisum* and unidentified species of *Acyrtosiphon* on lucerne in Argentina. Samples were collected from five sites (Ceres, Rafaela, Sarmiento, Monte Vera and Bernardo de Irigoyen) in the province of Santa Fe. In this study, *Zoophthora radicans* was the most important pathogen and was recorded mainly on *Acyrtosiphon* sp. *Zoophthora radicans* was successfully isolated and maintained in pure cultures. This study is the first report of entomophthoroid fungi infecting lucerne (*Medicago sativa* L.) aphids in Argentina.

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#### PALABRAS CLAVE

Alfalfa;  
Áfidos;  
Hongos  
entomopatógenos

#### Ocurrencia natural de hongos entomophthorales de áfidos plaga de *Medicago sativa* L. en Argentina

#### Resumen

Se encontraron cuatro especies de hongos Entomophthorales, *Pandora neoaphidis*, *Zoophthora radicans*, *Entomophthora planchoniana* (Entomophthorales: Entomophthoraceae) y *Neozygites fresenii* (Neozygites: Neozygitaceae) infectando a *Aphis craccivora*, *Therioaphis trifolii*, *Acyrtosiphon pisum* y a especies no identificadas pertenecientes al género *Acyrtosiphon* en cultivos de alfalfa (*Medicago sativa* L.), en la Argentina. Los muestreos fueron realizados en cinco sitios (Ceres, Rafaela, Sarmiento, Monte Vera y Bernardo de Irigoyen) de la provincia de Santa Fe. *Zoophthora radicans* fue el patógeno

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más importante registrado principalmente en *Acyrtosiphon* sp. *Zoophthora radicans* fue exitosamente aislado y mantenido en cultivos puros. Este estudio documenta por primera vez en la Argentina la presencia de hongos Entomophthorales infectando áfidos en alfalfa. © 2013 Asociación Argentina de Microbiología. Publicado por Elsevier España, S.L. Todos los derechos reservados.

Aphids (Hemiptera: Aphididae) are among the most successful families of insects and many represent serious agricultural pests<sup>11</sup>. Four of the ten species of aphids in the world that infect lucerne<sup>2</sup> are considered serious pests in Argentina, these being *Aphis craccivora* Koch, *Acyrtosiphon pisum* (Harris), *Acyrtosiphon kondoi* Sinji and *Therioaphis trifolii* (Monell)<sup>13</sup>. Aphids feed on phloem sap via extremely thin maxillary stylets that penetrate phloem sieve tubes, greatly reducing the possibility of these insects to ingest viruses, bacteria or protozoa from plant surfaces. Aphids became relevant due to their capacity of transmission of several viruses like Alfalfa Mosaic Virus (AMV) and other potyviruses, which limit the performance and persistence of plants<sup>3</sup>. Entomophthoralean fungi can cause lethal infections of various aphid species and they belong to the group of most effective control agents of natural aphid colonies. The only record of Entomophthoroid fungi of aphids on lucerne in South America was found in Uruguay<sup>1</sup>. Limited research efforts have been devoted to investigating the entomopathogenic fungi as agents of natural mortality of aphids in lucerne crops in Argentina. The aim of this paper was to identify and to isolate entomophthoroid fungi of aphid pests on *M. sativa* in the Argentina Pampas. The study was not intended to provide quantitative data. The taxonomy of entomophthoroid fungi used here is in accordance with the new molecular-based classification of these fungi, including it in a newly described phylum, *Entomophthoromycota*<sup>4</sup>.

The survey covered the west of Santa Fe province, in the Argentine Pampas (situated between 28–40° S and 68–57° W). The Argentine Pampa is a vast region of 52 million ha of suitable land for agriculture and cattle production. Samplings of insects were conducted in five sites from April 2010 to June 2012 (Table 1). Surveys were occasionally carried out in Ceres, Sarmiento and in Bernardo de Irigoyen, and weekly in Rafaela and Monte Vera. Sampled areas did not exceed 500 m<sup>2</sup> per site. No insecticides or fungicides were applied to the parts of the fields where collections were made during the course of the study. Fifteen (15) lucerne stems (from 30 to 50 cm each) were collected along both diagonals of each field. Stems sustaining aphids were placed in labeled plastic bags and transported to the laboratory as described by Zumoffen et al.<sup>14</sup>. Lucerne stems were checked to evaluate the presence of healthy or infected aphids. The plants were later discarded. Samples of healthy living aphids were collected and transferred into plastic cups with lids (150 cm<sup>3</sup>) from where subsamples were transferred to microcentrifuge tubes (Eppendorf; 1.5 cm<sup>3</sup>). These subsamples were preserved in 70% ethanol for further identification to species level, according to Blackman & Eastop's keys<sup>2</sup>.

Dead aphids with evidence of external fungal growth (showing sporulation) were examined under a stereo

**Table 1** Location of study sites

Town/City	District	Latitude S	Longitude W
Ceres	San Cristobal	29°52'55,38''	61°56'25,00''
Rafaela	Castellano	31°12'3,67''	61°30'25,83''
Sarmiento	Las Colonias	31°3'24,84''	61°10'13,29''
Bernardo de Irigoyen	San Jerónimo	32°10'05,65''	61°09'19,38''
Monte Vera	La Capital	31°32'58,21''	60°41'34,74''

microscope and an optic microscope to evaluate the presence of rhizoids, cystidia, and/or spores. Dead aphids without external mycosis signs were placed in Petri dishes (60 mm diam) containing a filter paper moistened with a few drops of distilled water (humid chambers), which was maintained at 20 °C for 24–72 h to allow the development of overt mycoses. Living aphids with apparent infection signs were also disposed in humid chambers and maintained under the same conditions detailed above until they showed an infection development, finally checking that aphid mortality was caused by Entomophthoralean fungi. Fungal structures were mounted in lactophenol-aceto-orcein (LPAO) (1:1) or stained with 1% aceto-orcein plus glycerine for semipermanent mounts. Measurements of fungal structures were made to enable specific identification. Fungal species were identified according to taxonomic keys and monographs of Humber<sup>6</sup> and Keller<sup>7,8</sup>.

In order to obtain pure cultures, infected aphids were placed on a moistened piece of sterile filter paper attached with double coated tape to the lid of a sterile 60 mm Petri dish, which was then inverted over the bottom of a sterile Petri dish containing SEMA (80% Sabouraud dextrose agar + 1% yeast extract and 20% of a mixture of egg yolk and skim milk)<sup>4</sup> plus 40.000 units/ml penicillin G (Merck®, Germany) and 80.000 units/ml streptomycin (Parafarm®, Argentina). This assembly was left 12 h in the dark at 22 ± 1 °C. A sterile lid replaced the lid with the attached aphids after 12 h. All isolates were incubated at 22 ± 1 °C with a photoperiod of 16:8 (L: D).

Only one of the species of Entomophthoroid fungi was successfully isolated and maintained in pure cultures. *Zoophthora radicans* isolates were deposited in the Mycological Culture Collection at Centro de Estudios Parasitológicos y de Vectores (CEP, La Plata, Argentina) and at USDA-ARS Collection of Entomopathogenic Fungal Cultures (ARSEF, Ithaca, New York) under access numbers CEP 362 and ARSEF 11859 CEP, respectively. Herbarium materials such as dried infected specimens and microscope slides were deposited in the Mycological Culture Collection

Table 2 Entomophthoralean fungi recorded from *M. sativa* during 2010-2012

Fungal species	Host	Locality	Date of collection
<i>Entomophthora planchoniana</i> Cornu	<i>Therioaphis trifolii</i>	Rafaela	Sep 16, 2010
	<i>Acyrtosiphon</i> sp.	Monte Vera	Sep 24, 2010
<i>Pandora neoaphidis</i> (Remaudière & Hennebert) Humber	<i>Acyrtosiphon pisum</i>	Monte Vera	Apr 21, 2010
	<i>Acyrtosiphon</i> sp.	Rafaela	May 14, 2010
	<i>Aphis craccivora</i>	Bernardo de Irigoyen	May 13, 2011
		Sarmiento	May 17, 2011
			May 19, 2011
			May 26, 2011
			Jun 13, 2011
			Jul 06, 2011
			Jul 13, 2011
			May 11, 2012
<i>Zoophthora radicans</i> (Brefeld) Batko	<i>Acyrtosiphon pisum</i>	Monte Vera	May 10, 2010
	<i>Acyrtosiphon</i> sp.	Rafaela	Jun 09, 2010
	<i>Aphis craccivora</i>	Sarmiento	Jun 16, 2010
		Ceres	Jul 06, 2010
			Dec 30, 2010
			May 13, 2011
			May 18, 2011
			May 19, 2011
			May 20, 2011
			May 27, 2011
			Jun 13, 2011
			Jun 24, 2011
			Jul 06, 2011
			Jul 13, 2011
			May 27, 2012
<i>Neozygites fresenii</i> (Nowakowski) Remaudière & Keller	<i>Aphis craccivora</i>	Monte Vera	Jun 10, 2012
			Feb 28, 2012

at Centro de Estudios Parasitológicos y de Vectores (CEP, La Plata, Argentina).

Three of the species of aphids observed were infected by entomophthoroid fungi: *A. craccivora*, *A. pisum*, *T. trifolii* and unidentified species of the genus *Acyrtosiphon*. Four species of entomophthoralean fungi were identified in these aphids: *Pandora neoaphidis* (Remaudière & Hennebert) Humber, *Zoophthora radicans* (Brefeld) Batko, *Entomophthora planchoniana* Cornu (Entomophthorales: *Entomophthoraceae*), and *Neozygites fresenii* (Nowakowski) Remaudière & Keller (Neozygites: *Neozygitaceae*) (Table 2). Fungal infections occurred mainly between May and July and, to a lesser extent, during April, September and December 2010 and February 2012. Previous studies on the phenology of entomophthoroid fungi in populations of insects other than aphids recorded that fungal infections

were more common during autumn-winter in Argentina (in the Southern hemisphere, from March to September)<sup>9</sup>.

Entomophthoralean fungal infections were most frequently observed in *Acyrtosiphon* spp. than in the rest of the aphid species collected. *Entomophthora planchoniana*, *P. neoaphidis* and *Z. radicans* were identified infecting *Acyrtosiphon* spp. In our study *Z. radicans* was the most important pathogen recorded from aphid pests on *M. sativa* and it was successfully isolated from *A. pisum*. On the other hand, Alzugaray *et al.*<sup>1</sup> reported *P. neoaphidis* as the principal mortality agent of aphids in lucerne crops in Uruguay. In this study *P. neoaphidis* was secondary to *Z. radicans* in occurrence and the first was identified among three aphid pest species. Entomophthoralean fungi were reported from other *Leguminosae* plants related to lucerne, as for example *N. fresenii* that was recorded from *A. craccivora* on faba

bean plants<sup>12</sup>. *Entomophthorales* species have been recorded to infect *A. pisum* and *T. trifolii* on legumes in Australia<sup>10</sup>. In the present research *T. trifolii* was only infected by *E. planchoniana*. There are previous records of *E. planchoniana* as pathogen of *T. trifolii* and *A. kondoi* in New Zealand<sup>5</sup>.

*Pandora neoaphidis*, *Z. radicans*, *E. planchoniana* and *N. fresenii* were found to infect *A. craccivora*, *T. trifolii*, *A. pisum* and unidentified species of *Acyrtosiphon* spp. The present study is a preliminary record of Entomophthoralean fungi causing infections in natural populations of aphids on lucerne crops in the Argentina Pampas.

## Conflicts of interest

The authors declare that they have no conflicts of interest.

## Acknowledgments

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