



Revista Argentina de Microbiología

ISSN: 0325-7541

ram@aam.org.ar

Asociación Argentina de Microbiología
Argentina

Kuhar, Francisco; Castiglia, Valeria C.; Zamora, Juan C.

Detection of manganese peroxidase and other exoenzymes in four isolates of *Geastrum*
(Geastrales) in pure culture

Revista Argentina de Microbiología, vol. 48, núm. 4, octubre-diciembre, 2016, pp. 274-278

Asociación Argentina de Microbiología
Buenos Aires, Argentina

Available in: <http://www.redalyc.org/articulo.oa?id=213049175003>

- How to cite
- Complete issue
- More information about this article
- Journal's homepage in redalyc.org

redalyc.org

Scientific Information System

Network of Scientific Journals from Latin America, the Caribbean, Spain and Portugal

Non-profit academic project, developed under the open access initiative



BRIEF REPORT

Detection of manganese peroxidase and other exoenzymes in four isolates of *Geastrum* (Geastrales) in pure culture

Francisco Kuhar^{a,*}, Valeria C. Castiglia^b, Juan C. Zamora^c

^a Centro de Investigación y Extensión Forestal Andino Patagónico – CONICET, Argentina

^b Laboratorio de Micología Experimental, Departamento de Biodiversidad y Biología Experimental, Facultad de Ciencias Exactas y Naturales, Universidad de Buenos Aires, Argentina

^c Departamento de Micología, Real Jardín Botánico, CSIC, Madrid, Spain

Received 17 May 2016; accepted 7 September 2016

Available online 2 December 2016

KEYWORDS

Cellulase;
Xylanase;
Laccase;
Lipase;
Litter degradation

PALABRAS CLAVE

Celulasa;
Xilanasa;
Lacasa;
Lipasa;
Degradación de
hojarasca

Abstract Knowledge regarding the enzymatic machinery of fungi is decisive to understand their ecological role. The species of the genus *Geastrum* are known to grow extremely slowly in pure culture, which makes it difficult to evaluate physiological parameters such as enzyme activity. Qualitative assays were performed on isolates of four species of this genus, showing evidence of laccase, cellulase, pectinase, amylase and lipase activity and suggesting that a wide range of carbon sources can be exploited by these species. For the first time in this genus, quantitative assays verified manganese peroxidase activity (up to 0.6 mU/g) in 30-day old cultures, as well as laccase, β -glycosidase and β -xylosidase activities.

© 2016 Asociación Argentina de Microbiología. Published by Elsevier España, S.L.U. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

Detección de manganeso peroxidasa y otras exoenzimas en 4 aislamientos de *Geastrum* (Geastrales) en cultivo puro

Resumen El conocimiento de la maquinaria enzimática de un hongo es decisivo para entender su rol ecológico. Las especies del género *Geastrum* son conocidas por su crecimiento extremadamente lento en cultivos puros, lo que hace difícil la evaluación de parámetros fisiológicos como las actividades enzimáticas. Se realizaron ensayos cualitativos sobre aislamientos de 4 especies de este género, mostrando evidencias de actividades lacasa, celulasa, pectinasa, amilasa y

* Corresponding author.

E-mail address: fkuhar@gmail.com (F. Kuhar).

lipasa, mostrando el amplio rango de fuentes de carbono que pueden ser explotadas por estas especies. Ensayos cuantitativos verificaron por primera vez en este género la actividad manganeso peroxidasa (hasta 0,6 mU/g) en cultivos de 30 días, así como también β -glucosidasa y β -xilosidasa.

© 2016 Asociación Argentina de Microbiología. Publicado por Elsevier España, S.L.U. Este es un artículo Open Access bajo la licencia CC BY-NC-ND (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

Fungal extracellular enzymes have been a subject of study not only because of their intrinsic interest, but also due to their biotechnological applications. For this reason, most studies have been based on fast growing organisms that produce high enzyme activities, and on model species whose genetics and physiology are well known.

The finding of lignin-modifying enzymes in litter or soil fungi⁵ redirected the attention of many scientists to the adaptive role they may play in this environment. Recent studies based on genomic data provide invaluable information about the enzyme machinery of many fungal taxa in relation to their ecology¹¹ and phylogenetic placement¹; however, to date, no studies of this kind have included members of the family Geastraceae.

Since degradative fungal enzymes may have an important role in bioremediation scenarios, the study of enzyme activities is also relevant for soil and litter fungal species regardless of their possible industrial application. One of the problems in this field is the slow growth in culture of many species (e.g. ectomycorrhizal fungi), which is usually the main obstacle to their enzyme research. An example of this fact is the genus *Geastrum*, which, though entirely saprotrophic, shows an extremely slow growth rate. Many of its species are known to form extensive mycelial networks and cushions in all kinds of forests, tolerating inhibitory substrates such as *Eucalyptus* or Cupressaceae litter¹⁰.

This genus has been the subject of exhaustive taxonomic and floristic studies. Sunhede¹⁰ made a systematic revision and took thorough notes on its morphology, biology and ecology, however, at present just a few works make reference to its ecophysiology. Sunhede¹⁰ describes the growth of the isolates as "very slow, after six weeks varying from 3 to 22 mm." Ilko et al.⁴ indicate that the colony of *Geastrum pouzarii* reached 3–4 mm after six weeks and Zamora et al.,¹⁴ report isolates of *G. argentinum* achieving up to 4 mm after a week. This characteristic makes these species hard to isolate and to maintain in pure cultures, and this is the main reason why there are no reports of their exoenzyme activities. Sunhede¹⁰ reports oxidase enzymatic activity in some species by observing metachromatic staining of mycelia with some laccase and tyrosinase substrates. Zamora et al.¹³ carefully studied similar reactions in fresh basidiomes; however, no attempts to study *in vitro* enzymatic activity have been performed until now.

Four *Geastrum* species were isolated and studied, namely *G. argentinum*, *G. papinuttii*, *G. morganii*, and *G. plicatum*. Specimens were collected in the province of Buenos Aires and deposited in BAFC (Universidad de Buenos Aires), LPS (Universidad Nacional de La Plata) and MA-fungi (Real Jardín Botánico de Madrid) herbaria (Index

Herbariorum, <http://sweetgum.nybg.org/science/ih/>)¹². Isolates were obtained from the pseudoparenchymatous layer of the exoperidium in malt agar medium, and deposited in BAFC Culture Collection under the following numbers (herbarium collection numbers in parentheses): *G. argentinum* BAFC 3282 (LPS 48446), *G. plicatum* BAFC 3283 (MA-Fungi 83774), *G. morganii* BAFC 3284, *G. papinuttii* BAFC 3285 (MA-Fungi 83764). Three of the four species (*G. argentinum*, *G. plicatum*, and *G. papinuttii*) were molecularly characterized by Zamora et al.^{15,16}, and GenBank accession numbers are available there for ITS (Internal Transcribed Spacer) nrDNA (nuclear ribosomal DNA), LSU (Large Subunit) nrDNA, rpb1 (polymerase II largest subunit gene) and *atp6* (ATP synthase subunit 6 gene).

An agarized medium containing glucose and glutamic acid (GG medium)³ was utilized for the screening. Laccase (EC:1.10.3.2) activity was quantified according to Paszczynski et al.⁸ and MnP (manganese peroxidase; EC:1.11.1.13) following Paszczynski et al.⁹ with and without MnCl₂ 0.1 mM as inducer. In all cases, aliquots of supernatant were replaced by agar plugs (5 mm diameter) provided by 60-day malt agar cultures. β -xylosidase (EC:3.2.1.37) and β -glucosidase (EC:3.2.1.21) enzyme activities were determined by measuring the release of p-nitrophenol from p-nitrophenyl- β -D-xylopyranoside (pNPX) and p-nitrophenyl- β -D-glucopyranoside (pNPG). Agar plugs were incubated in 900 μ l of 1 mM substrate at 50 °C for 30 min. The reaction was stopped by adding 2 ml of Clark and Lubs buffer pH 9.8. The release of p-nitrophenol was measured using a UV-vis spectrophotometer at λ 405 nm and 1 unit of β -xylosidase activity was defined as the amount of the enzyme releasing 1 μ mol p-nitrophenol equivalent per minute under the assayed conditions. Hydrolytic enzyme activities were qualitatively evaluated according to Pardo and Forchassin⁷, replacing glucose in each assay by starch 0.1% (for amylase EC:3.2.1.1 activity), pectin 0.1% (for pectinase EC:3.2.1.15 activity), carboxymethyl-cellulose (CMC) 0.1% (for endoglucanase EC:3.2.1.4 activity) and xylan 0.1% (for xylanase EC:3.2.1.8 activity). Lipase (EC:3.1.1.3) qualitative assay was as indicated by Nikoleit et al.⁶ using Tween 80 as a carbon source but replacing the medium by basal GG medium³ without a carbon source. Laccase and LiP (lignin peroxidase) activity were also qualitatively evaluated in a GG medium by adding dimethoxyphenol 0.1 mM and Azure B. All data presented are the means of the results of triplicates and error bars indicate SE.

All isolates showed active growth in each assayed carbon source (xylan, cellulose, starch and pectin). The use of lipid esters as a carbon source was also demonstrated in *G. plicatum* through the active growth and degradation of Tween 80.

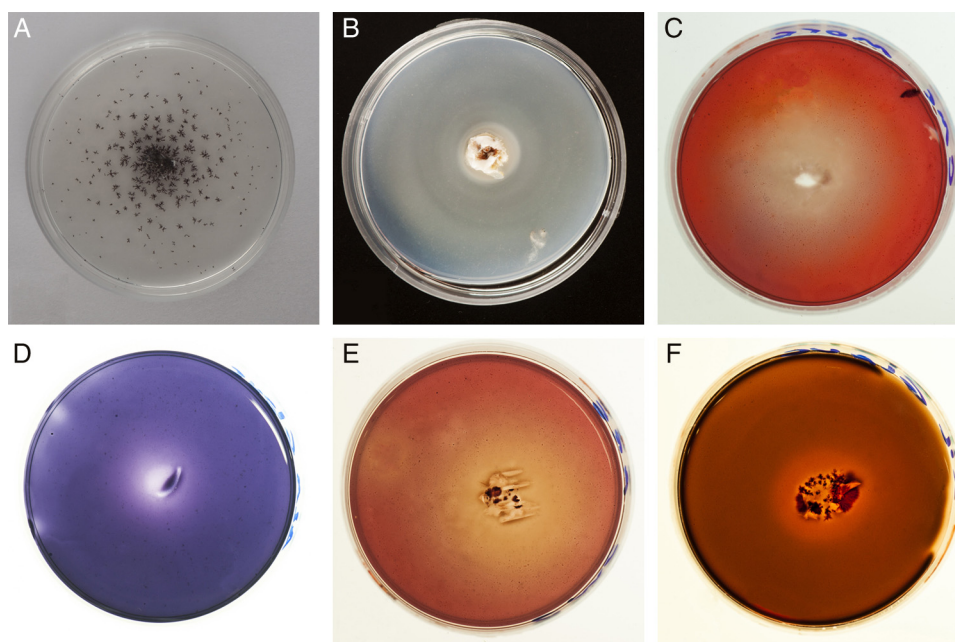


Figure 1 Selected positive reactions of qualitative assays. (A) Laccase (*G. argentinum*). (B) Lipase (*G. plicatum*). (C) Cellulase (*G. morganii*). (D) Lignin peroxidase (*G. papinuttii*). (E) Cellulase (*G. argentinum*). (F) Xylanase (*G. papinuttii*).

Enzyme activities evidenced by qualitative assays are shown in Fig. 1. Activity halos reaching higher diameter than the colony evidence the activity of exoenzymes. The degradation of carbon sources is represented in Table 1 by the diameters of degradation and growth halos corresponding to day 20. A weak decolorization halo in the cultures containing Azure B suggests LiP activity in *G. papinuttii*, while laccase activity was evidenced 24 h after inoculation in every isolate.

The values of laccase, MnP, β -glucosidase (EC:3.2.1.21) and β -xylosidase (EC:3.2.1.37) activities are shown in Fig. 2. Uninduced cultures did not show MnP activity. The results corresponding to this enzyme were measured in cultures

induced with MnCl_2 0.1 mM. The absence of degradation halos in some qualitative assays (Table 1) may reflect that the carbon source was not actively degraded by the strain, and the positive growth in those cases could be explained by the use of the amino acid, instead. Manganese peroxidase activity was only detectable after induction with MnCl_2 . Laccase, as well as β -glucosidase and β -xylosidase showed intrinsic activity in the absence of inducers.

Since the whole *Geastraceae* family forms dense mycelia, matting together organic debris¹⁰, the enzymatic machinery should be adapted to decay this substrate. In fact, Fuji et al.², suggest that laccase and MnP (which showed to be active in our isolates) are involved in the early stages of

Table 1 Diameter of degradation (D) and growth (G) halos (mm) on different carbon sources after 30 days of culture by the four isolates

	CMC*	Pectin	Starch	Xylan	Tween
<i>G. argentinum</i>					
D	28 $\pm$ 5.2	12 $\pm$ 3.7	14 $\pm$ 2.6	0 $\pm$ 0.0	0 $\pm$ 0.0
G	5 $\pm$ 1.1	1 $\pm$ 2.1	11 $\pm$ 2.0	9 $\pm$ 1.4	2 $\pm$ 0.2
<i>G. papinuttii</i>					
D	27 $\pm$ 4.0	12 $\pm$ 2.9	4 $\pm$ 0.7	0 $\pm$ 0.0	0 $\pm$ 0.0
G	3 $\pm$ 0.5	7 $\pm$ 1.1	8 $\pm$ 0.3	7 $\pm$ 1.2	6 $\pm$ 0.8
<i>G. morganii</i>					
D	29 $\pm$ 7.3	13 $\pm$ 1.2	0 $\pm$ 0.0	6 $\pm$ 0.9	0 $\pm$ 0.0
G	7 $\pm$ 0.2	6 $\pm$ 0.7	5 $\pm$ 0.9	7 $\pm$ 0.9	3 $\pm$ 0.2
<i>G. plicatum</i>					
D	21 $\pm$ 5.1	11 $\pm$ 2.4	0 $\pm$ 0.0	0 $\pm$ 0.0	25 $\pm$ 3.5
G	5 $\pm$ 2.0	6 $\pm$ 1.2	5 $\pm$ 0.7	5 $\pm$ 1.3	5 $\pm$ 1.0

Each value represents the means of three independent cultures $\pm$ standard error.

* Carboxymethyl cellulose.

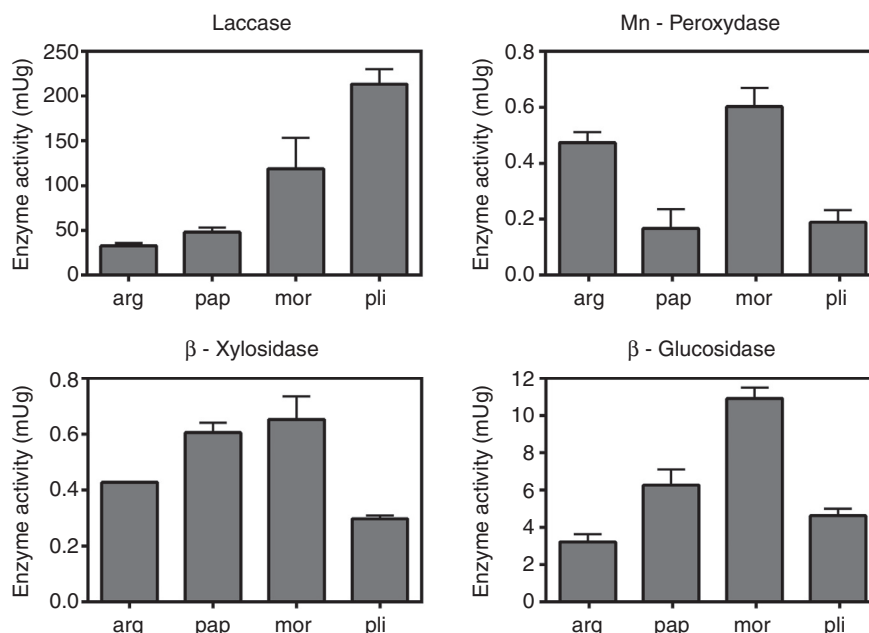


Figure 2 Quantification of enzymatic activities in agar plugs. arg, *G. argentinum*; pap, *G. papinuttii*; mor, *G. morgani*; pli, *G. plicatum*. Bars represent the mean enzyme activity of laccase, MnP, β -glucosidase and β -xylosidase per gram of agarized medium. MnP quantification corresponds to cultures induced with MnCl_2 . Uninduced cultures did not show MnP activity. Values represent means $\pm$ SE of three independent cultures.

litter decomposition. *G. argentinum* is often associated with fallen trees¹³; however, the advanced decay of the logs suggests that the lignocellulolytic activities found in our work are not correlated with an active degradation of wood matter. Sunhede¹⁰ also reports that basidiomes of *Geastrum* species are often found on moldering stumps. β -Xylosidase and β -glucosidase activities, together with CMC and pectin halos (shown by all isolates), account for their capacity of exploiting cellulosic substrates. The differential behavior of other hydrolytic enzymes such as xylanases, amylases and lipases suggests different strategies of organic matter degradation. For instance, the positive reaction of *G. plicatum* in the lipase assay may be related to the degradation of persistent waxy cuticles of leaves; however, more isolates of each species are needed to confirm these patterns. The possible interactions of *Geastrum* species with other organisms enhancing the degradation of cellulosic materials as well as the role of some *Geastrum* species in late stages of wood debris degradation need to be further investigated. The detection of MnP and laccase activities in the tested isolates suggests that they might be a promising source of enzymes with the ability to transform xenobiotics, although their extremely slow growth entails the fact that no high scale biotechnological application of these enzymes is possible without cloning them in more efficient organisms.

Ethical disclosures

Protection of human and animal subjects. The authors declare that no experiments were performed on humans or animals for this study.

Confidentiality of data. The authors declare that no patient data appear in this article.

Right to privacy and informed consent. The authors declare that no patient data appear in this article.

Conflict of interest

The authors declare that they have no conflicts of interest.

Acknowledgements

This work was supported by Consejo Nacional de Investigaciones Científicas y Técnicas, Argentina (PIP 11220120100408CO), Universidad de Buenos Aires (UBACyT, 20020120200085), Ministerio de Ciencia and Tecnología e Innovación Productiva PICT 3457/14. We also thank the Consejo Superior de Investigaciones Científicas (Spain) for supporting JCZ with the grant JAE-Pre.

References

1. Eichlerová I, Homolka L, Žiřčáková L, Lisá L, Dobiášová P, Baldrian P. Enzymatic systems involved in decomposition reflects the ecology and taxonomy of saprotrophic fungi. *Fungal Ecol.* 2015;13:10–22.
2. Fuji K, Uemura M, Hayakawa C, Funakawa S, Kosaki T. Environmental control of lignin peroxidase, manganese peroxidase, and laccase activities in forest floor layers in humid Asia. *Soil Biol Biochem.* 2013;57:109–15.
3. Galvagno MA. Ensayos de nutrición en *Ascomobolus crenulatus* P. Karst. (Fungi: Ascomycetes). *Bol Soc Arg Bot.* 1976;17: 95–118.
4. Ilko S, Ladislav H, Frantisek N. *Geastrum pouzarii* Staněk in agar culture. *Antonie van Leeuwenhoek.* 2001;79:363–4.
5. Liers C, Arnstadt T, Ullrich R, Hofrichter M. Patterns of lignin degradation and oxidative enzyme secretion by

- different wood and litter colonizing basidiomycetes and ascomycetes grown on beech wood. *FEMS Microbiol Ecol.* 2011;78:91–102.
6. Nikoleit K, Rosenstein R, Verheij HM, Götz F. Comparative biochemical and molecular analysis of the *Staphylococcus hyicus*, *Staphylococcus aureus* and a hybrid lipase. *European J Biochem.* 1995;228:732–8.
 7. Pardo AG, Forchiassin F. Influence of different cultural conditions on cellulase production by *Nectria catalinensis*. *Rev Argent Microbiol.* 1997;30:20–9.
 8. Paszczynski A, Crawford RL. Degradation of azo compounds by ligninase from *Phanerochaete chrysosporium*: involvement of veratryl alcohol. *Biochem Biophys Res Commun.* 1991;178:1056–63.
 9. Paszczynski A, Crawford RL, Huynh VB. Manganese peroxidase of *Phanerochaete chrysosporium*: purification. *Methods Enzymol.* 1988;161:264–70.
 10. Sunhede S. Geastraceae (Basidiomycotina): morphology, ecology, and systematics with special emphasis on the Northern European species. Oslo, Norway: Fungiflora; 1989.
 11. Talbot JM, Martin F, Kohler A, Henrissat B, Peay KG. Functional guild classification predicts the enzymatic role of fungi in litter and soil biogeochemistry. *Soil Biol Biochem.* 2015;88:441–56.
 12. Thiers B, [continuously updated]. Index herbariorum: a global directory of public herbaria and associated staff. New York Botanical Garden's Virtual Herbarium. <http://sweetgum.nybg.org/ih/>
 13. Zamora JC, Calonge FD, Martín MP. New sources of taxonomic information for earthstars (*Geastrum*, Geastraceae, Basidiomycota): phenol oxidases and rhizomorph crystals. *Phytotaxa.* 2013;132:1–20.
 14. Zamora JC, Kuhar F, Castiglia V, Papinutti L. On *Geastrum argentinum*, a forgotten species. *Mycoscience.* 2014;55:177–82.
 15. Zamora JC, de Diego Calonge F, Hosaka K, Martín MP. Systematics of the genus *Geastrum* (Fungi: Basidiomycota) revisited. *Taxon.* 2014;63:477–97.
 16. Zamora JC, Calonge FD, Martin MP. Integrative taxonomy reveals an unexpected diversity in *Geastrum* section *Geastrum* (Gaeastrales, Basidiomycota). *Persoonia.* 2015;34:130–65.