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Cryptococcosis by *Cryptococcus gattii* in immunocompetent goats in Spain and review of the literature

Criptococose causada por *Cryptococcus gattii* em cabras imunocompetentes na Espanha e revisão de literatura

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ABSTRACT

Cryptococcus neoformans has been described for years as a species causing spontaneous mycosis in a great variety of animals. The new species *C. gattii* has been described as an agent of animal cryptococcosis mainly in Australia, but it has been found also in many parts of the world. The main group of animals suffering those natural infections are mammals, but also birds, reptiles and some invertebrates have suffered cryptococcosis. Usually the infections are sporadic and occasional, but some epidemic outbreaks have been reported affecting a high number of animals. In 1998 the isolation of *C. gattii* was reported by the first time in Europe in 5 epidemic outbreaks of cryptococcosis in goats grazing freely in west Spain grasslands. In all outbreaks, mycological studies were possible from samples obtained on necropsy of some animals dead during the epidemic. Animals belonged to various milking breeds and were grazing with variable status of health and husbandry. Goats affected by cryptococcosis showed similar respiratory symptoms, consisting in mucopurulent nasal discharge, cough, dyspnea and progressive cachexia, causing death in a period of 2 to 4 weeks. In three outbreaks many animals also showed ataxia, midriasis, blindness and progressive paralysis. Clinical prevalence varied from 2 to 12% in the different outbreaks. It is evident that in spite of the great amplitude of geographical distribution observed for *C. gattii*, this species has a limited presence, possibly restricted to determined habitats, as that of infection of goats flocks in Spain. Veterinarians must be concerned about Cryptococcosis in grazing animals. This finding introduced new elements connected to the epidemiology and ecology of *Cryptococcus gattii*.

Key words: cryptococcosis, *Cryptococcus gattii*, goats, immunocompetent, outbreaks, Spain.

RESUMO

Cryptococcus neoformans tem sido descrito desde longa data, como uma espécie causadora de micose espontânea em uma grande variedade de animais. A nova espécie, *C. gattii*, tem sido isolada de casos de criptococose animal principalmente na Austrália, mas também em várias partes do mundo. Os mamíferos constituem o grupo de animais que mais sofrem infecção natural, mas também pássaros, répteis e alguns invertebrados têm sido acometidos por criptococose. Usualmente as infecções são esporádicas e ocasionais, mas alguns surtos epidêmicos têm sido relatados como afetando um grande número de animais. Em 1998, *C. gattii* foi isolado pela primeira vez na Europa em 5 surtos epidêmicos de criptococose em cabras que pastavam livremente em propriedades situadas no oeste da Espanha. Em todos surtos, o diagnóstico micológico foi realizado com as amostras obtidas a partir da necropsia de alguns animais mortos durante os surtos. Estes animais pertenciam a várias raças leiteiras criadas em diferentes regimes sob o ponto de vista do status sanitário e de manejo. As cabras com criptococose apresentavam sinais clínicos respiratórios similares, que consistiam de descarga nasal mucopurulenta, tosse, dispnéia e caquexia progressiva, os quais causavam a morte num período de 2 a 4 semanas. Em três surtos, muitos animais também apresentaram ataxia, midríase, cegueira e paralisia progressiva. A prevalência clínica variou de 2 a 12% nos diferentes surtos. Parece evidente que, apesar da grande amplitude da distribuição geográfica do *C. gattii*, esta espécie tem uma presença possivelmente restringida a determinados habitats, como no caso das infecções nos rebanhos de cabras na Espanha. Os veterinários devem estar atentos para a possibilidade de ocorrência de criptococose em animais criados a campo. Estes achados contribuem, com a apresentação de novos elementos, para uma melhor compreensão da epidemiologia e ecologia do *Cryptococcus gattii*.

Descritores: criptococose, *Cryptococcus gattii*, cabras, imunocompetente, surtos, Espanha.

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I. INTRODUCTION

Cryptococcus neoformans has been described for years as a species causing spontaneous mycosis in a great variety of animals. Recently *C. gattii* has been considered as a different species from *C. neoformans*, to which it was related as a variety with different biochemical and physiological characteristics and possibly with different pathogenetic traits [22,35,36].

The new species *C. gattii* has been described as an agent of animal cryptococcosis mainly in Australia, but it has been found also in many parts of the world [4].

The main group of animals suffering those natural infections are mammals, but also birds, reptiles and some invertebrates have suffered cryptococcosis [28]. Usually the infections are sporadic and occasional, but some epidemic outbreaks have been reported affecting a high number of animals [21,38].

Actually inhalation of environment yeasts may cause infections of the upper and lower respiratory tract as well as visceral dissemination, depending on the virulence of the infectious strain and the health status of the animal. Penetration through the skin or mucosae and development of local lesions, also have been described in different animals although less frequently [9].

II. EPIDEMIOLOGY

1. Habitat and source of infection

C. neoformans has been isolated everywhere from nature, varying its prevalence and abundance [5]. Less frequently and with a strict geographical distribution,

C. gattii also has been isolated mainly from detritus of trees, specially from *Eucalyptus* [14,18,24]. For those reasons it is not difficult to explain the high variety of animals that become infected by the exposition to that environment yeast (Table 1). It is very probable that another animal species, specially the wild ones, could suffer cryptococcosis without a diagnosis made until now.

In many occasions the strains of *Cryptococcus* isolated from nature have not been adequately studied in order to differentiate the two species and their serotypes.

Another aspect of interest is the finding of asymptomatic dogs and cats carriers of *C. gattii* colonizing their nasal sinus [12]. This situation seems to be a risk factor for a progressive cryptococcosis in those animals [13].

2. Factors favouring animal infection

Factors that favour infection by *Cryptococcus* have been studied mainly in dogs and cats. Previous infection by FIV (feline immunodeficiency virus), a lentivirus similar to human HIV, may explain the high susceptibility of this species mainly to *C. neoformans* [31]. Another example is provided by the co-infection of FeLV (feline leukaemia virus) and the isolation of *C. neoformans* var. *gattii* from a siamese cat with disseminated disease, in a case reported in Brazil [16].

The abundance of this yeast in soil samples may suppose a higher prevalence of cryptococcosis in animals with smelling or herbivorous habits, more expo-

Table 1. Species of animals naturally infected by *Cryptococcus neoformans*, *Cryptococcus gattii* or without specific biotype.

Mammals	Birds	Others
Cats	Columbiformes (Pigeons)	Reptiles
Dogs	Kiwi	Insects (cockroach, lepidopters)
Cows/Bufalos	Psittaciformes (<i>Charmosyna papou</i> , <i>Lorius lory</i> , <i>Trichoglossus goldiei</i> , <i>Psittacula krameri</i> , <i>Psittacus erithacus</i>)	Acarus
Goats	Anseriformes (ducks, swans)	
Horses	Cockatoo	
Pigs	Magpie	
Hamsters	Crow	
Rodents (rat, mouse)	Paroquet	
Primates (Mangabey)	Canary	
Camelids (llama)	Swallow	
Guepards (cheeta)	Krestel	
Fox	Buzzard	
Ferrets		
Algalia (civet)		
Fox of desert (<i>Fennecus zerda</i>)		
Marmots		
Koalas		
Dolphins		
Bats		

sed to yeast inhalation or by eating contaminated vegetable with cryptococcus.

Unusual high number of infections by *C. gattii* in wildlife animals, affecting different species, including dogs and cats, as well as humans, occurred in Vancouver (British Columbia, Canada) after 1999 [10]. In domestic animals the infections were related with the easiness of acquiring Cryptococcosis due to their street activity and living near crowded commercial malls [11].

III. CRYPTOCOCCOSIS IN ANIMALS

1. Domestic animals

Among domestic animals, cats and dogs seems to be the most commonly affected by cryptococcosis, although permanent and closer contact with their owners is possibly the cause of an early detection of their health deterioration, and a veterinary examination. This would explain the higher number of diagnosis of crypto-

coccosis in that group of animals. In other domestic animals like cows, horses, pigs and goats, the diagnosis is done when epidemic outbreaks occur and after necropsy and mycological search of obtained tissues [34].

Both species of *Cryptococcus* have been isolated from those animals, greatly depending on geographic area and the available veterinary facilities. On the other hand, diagnosis made in wild animals, like guepards, foxes, primates, dolphins and other more, have been done in zoological gardens or aquarium under veterinary supervision.

2. Pigeons

The relation between *Cryptococcus neoformans* and birds is very narrow since it was demonstrated in Europe in 1974 that pigeon excrements are an important reservoir of that yeast [37]. The isolation of that yeast in pigeon faeces remains a study motive to know the epidemiology of cryptococcosis around the world.

C. neoformans var. *grubii* (serotype A) was isolated in the city of Barcelona, northwest of the Iberic Peninsula, on Mediterranean Sea coast, with a high population density (316 inhabitants per hectare) and a traditional abundance of pigeons, in 17% out of 303 collected samples [29]. All isolated samples were of the molecular standard VNI [27]. It is interesting to note that after an *in vitro* sensibility study at least 4 of those environmental strains showed natural resistance to fluconazol, which implicates that primary infection in humans or animals with isolates resistant to azoles could occur.

Similar results have been found in samples from the city of Seoul (Korea) [6], where 8 from 72 dry faeces samples of pigeons were positive to *C. neoformans* var. *grubii* (serotype A), with the same molecular standard.

In the state of Goiás (Brazil) has been also reported the isolation of *C. neoformans* var. *grubii* in 20% of pigeons faeces and in 14% of *Eucalyptus* samples, though in that case no resistance to antifungi were detected, including fluconazol [20].

3. Wild birds

Abegg *et al.* in Rio Grande do Sul state (southern Brazil) [1] has been recently reported that 38 faeces samples out of 59 different birds captive in zoo were positive to *C. neoformans* but only Psittaciformes. In that case a diversity was demonstrated because 89% of isolated samples were from variety *grubii* VNI and 13% were from *C. gattii* serotype B, molecular standard VGI.

In São Paulo (Brazil) an epidemic outbreak was described in different species of Psittacides which died of disseminated cryptococcosis. In 7 studied animals, *C. gattii* serotype B, resistant to fluconazol was isolated [33].

That yeast has been isolated from other birds around the world. In Egypt *Cryptococcus neoformans* was isolated in swallow faeces [26], in Massachussets (USA) a zoonotic transmission of *Cryptococcus neoformans* has been evidenced by a cockatoo [25,30], and in Holland the same transmission was proved by a magpie (in this last case the transmission to an immunocompetent patient was described) [23]. In New Zealand, a case of disseminated cryptococcosis was reported in a kiwi [19] by *C. gattii*; in Italy, *C. neoformans* was isolated from canary faeces [8] and, in Germany, *C. neoformans* has been observed in paroquet excrements [39, 40]. Also, in New York (Manhattan), fungal diseases have been reported, including cryptococcosis in Anse-

riformes (ducks and swans) [32], while in India *Cryptococcus neoformans* was isolated in faeces of crows [17].

Recently in Southeast of Italy, *C. neoformans* var. *grubii* has been isolated from cloacae and faecal samples of birds of prey including *Falco tinnunculus* and *Buteo buteo* [3].

IV. CRYPTOCOCCUS GATTII IN GOATS

1. Antecedents

In 1998 the isolation of *C. gattii* was reported by the first time in Europe [2] in at least 5 epidemic outbreaks of cryptococcosis in goats grazing freely in west Spain grasslands. This finding introduced new elements connected to the epidemiology of animal cryptococcosis and the ecology of *C. gattii*.

Between 1991 and 1995 some epidemic outbreaks were reported affecting goats in different fields of the Province of Cáceres (Extremadura), located at the Southwest Spain, in the border of Portugal. The north zone of that province is mountainous, while the rest of the province is a fertile plain bathed by Tajo River and its affluents, with important development of agriculture and livestock.

Mycological studies were done in 5 different epidemic outbreaks affecting goats in samples obtained by necropsy. Figure 1 shows the zones where outbreaks occurred.

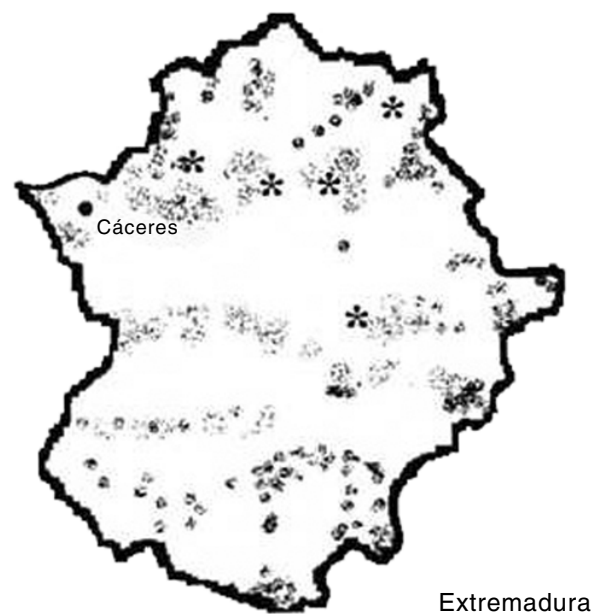


Figure 1. Map of the Province of Extremadura (southeast of Spain). Asterisks show the zones where cryptococcosis outbreaks occurred. The shadow zones correspond to Eucalyptus plantations.

The outbreak affected goats, most of them completely healthy, belonged to various milking breeds free grazing in different fields of Cáceres.

In one outbreak animals with other diseases were observed (brucellosis, paratuberculosis). Animals affected by cryptococcosis showed similar respiratory symptoms in all outbreaks, consisting in mucopurulent nasal discharge, cough, dispnea and progressive caquecsy, causing death in a period of 2 to 4 weeks. In three outbreaks many animals also showed ataxia, midriasis, blindness and progressive paralysis. Clinical prevalence varied from 2 to 12% in the different outbreaks.

All animals affected by cryptococcosis developed a serious pneumonia with signals of consumption and in many cases neurological signs (Figure 2). Total mortality in each flock is presented in Table 2, being the higher of 12%.

Various animals were submitted to necropsy taking tissue samples in 6 of them for histological analysis and mycological culture (Figure 3).

In cultures of 13 samples grew abundant colonies of yeasts identified as *Cryptococcus neoformans*. Abundant encapsulated yeasts were also found in histological sections of lung and brain (Figure 4).

Cultures with CGB (Canavanine, Glycin, Bromotimol blue) and the assimilation of proline and triptophan let classified them as *C. neoformans* var. *gattii*. The serological test¹ showed that all of the isolates corresponded to serotype B. Molecular typed by RAPD technique classified the isolates as VGNI, standard according to Meyer *et al.* [26].

Attempts trying to isolate the yeast from the environment at the zone where goats were grazing resulted unfruitful. The grazing fields characteristics were very different but plantations of *Eucalyptus* forests were present in most cases. These trees were introduced between 1955 and 1970, from France seed fields.

2. Outbreaks

The detailed and chronological description of the studied outbreaks, detected and diagnosed by the Cáceres Faculty of Veterinary is as follows:

Outbreak 1. (October 23, 1990). Location: Madrigal de la Vera, 167 km from Cáceres. Flock of 140 grazing goats. No pigeons in the farm.

The process began one year before with a high clinical prevalence according to the anamnesis, though not quantified.

Passive agglutination with latex² for detection of capsular antigen of *Cryptococcus* performed in 50 sera showed 12% of prevalence. Clinical signs were respiratory with pneumonia and caquecsy, without neurological symptoms, mortal evolution.

Two auxanographically distinct strains of *C. neoformans* were isolated in lung tissue, mediastinic ganglia and traqueal exudate, as well as in intestinal content, faeces and mesenteric ganglia from two animals. Culture of samples of CNS were negative. An environmental scanning also was negative.

Outbreak 2. (April 14, 1991). Location: Madroñera, 61 km from Cáceres. Flock of 250 grazing goats. Clinical prevalence of 2%. Symptoms: respiratory, ataxia, progressive caquecsy and death. Presence of abundant pigeons.

The process presented a one year evolution. Health of flock was poor with animals affected by brucellosis and paratuberculosis.

Cryptococcosis presented also neurological symptoms, confirming the diagnosis by microscopy and positive culture of samples from lung and CNS. Strains of one auxanographic type were isolated.

Outbreak 3. (January 20, 1994). Location: Pescueza, 78 km from Cáceres. Flock of 300 grazing goats. Clinical prevalence of 10%, subacute or chronic pneumonia, caquecsy of mortal evolution. No pigeons in the zone but presence of eucaliptus.

Microscopic observation and isolation of *Cryptococcus neoformans* in lungs. One auxanographic type detected.

Outbreak 4. (March 15, 1994). Location: Serradilla, 70 km from Cáceres. Flock of 200 grazing goats. Prevalence not determined. Pneumonia process, caquecsy and neurological symptoms (ataxia and mortal progressive paralysis). No pigeons or eucaliptus in the zone.

Microscopic observation and isolation of *Cryptococcus neoformans* in lungs and CNS. One auxanographic type detected.

Outbreak 5. (May 03, 1994). Localización: Casas de Millán, 53 km from Cáceres. Flock of 120 grazing goats. Clinical prevalence of 2,5%, presenting pneumonic process, caquecsy and neurological symptoms (ataxia, midriasis and blindness). Mortal evolution in 15-20 days. Presence of abundant pigeons in the farm.

Microscopic observation and isolation of *C. neoformans* in lungs, CNS and liver. One auxanographic type detected.

Table 2. Goats affected by lung cryptococcosis from *Cryptococcus gattii* serotype B in 5 outbreaks occurred between 1990 and 1994 in Cáceres (Extremadura, Spain).

Year of the outbreak	Location of the infection (City)	Number of animals of the flock	Clinical Prevalence (%) [*]	DNA fingerprinting
1990	Madrigal de la Vera	140	12	VGNI
1991	Madroñera	250	2	VGNI
1994	Pescueza	300	10	VGNI
1994	Serradilla	200	unknown	VGNI
1994	Casas de Millán	120	2,5	VGNI

^{*}Mortality rate of diseased animals reached 100%.



Figure 2. Goat affected by pulmonar consumptive cryptococcosis.

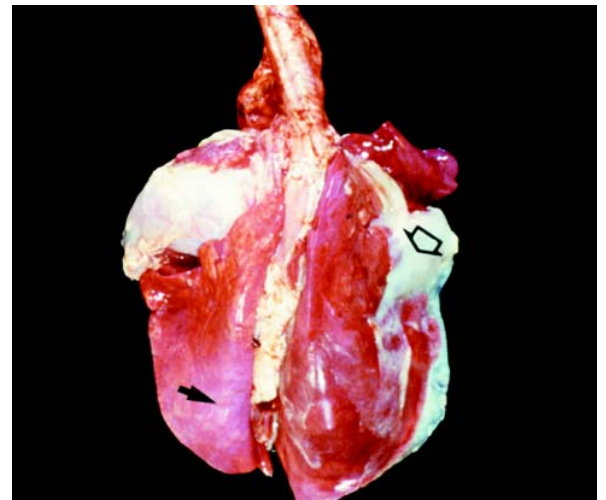


Figure 3. Anatomical piece obtained in the necropsy of a goat dead by cryptococcosis. Arrows show pleuritis zones.

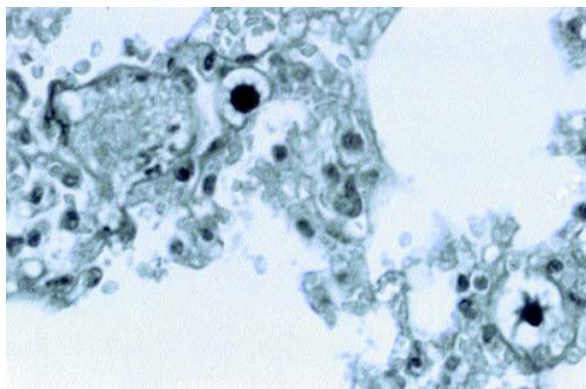


Figure 4. Histological section of a lung of a goat showing yeasts with capsules withdrawn by the dye.

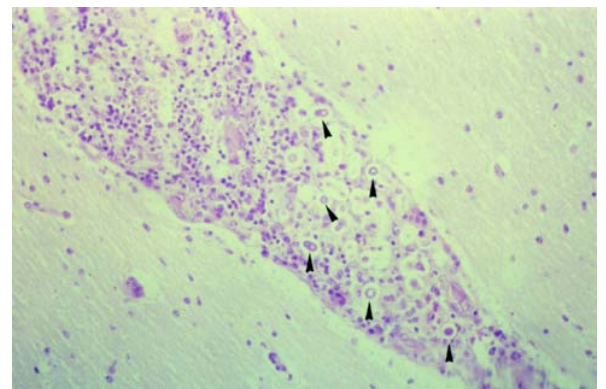


Figure 5. Histological section of a brain of a goat dead by cryptococcosis. Arrows show abundant encapsulated yeasts.

V. DISCUSSION AND CONCLUSIONS

The mycological analysis of 13 isolated strains of those outbreaks showed the variety *gattii* (*C. gattii*), serotype B [25], suggest that *C. gattii* was present in the environment of the affected animals in an endemic form. Nevertheless, the cultures of environment samples (soil, pigeon dejection, eucalyptus debries) were negative for this fungus. On the other hand, vast majority of goats with cryptococcosis seemed to be previously healthy, suggesting a high virulence of the isolated strains.

The relationship between *C. gattii* and *Eucalyptus camaldulensis* in Australia, and other species of *Eucalyptus* (*E. tereticornis*, *E. globulus*, etc) in other countries, suggests that probably the yeasts that infected the goats were in the *Eucalyptus* detritus near the animals, persisting in the environment during long time in a large zone.

The respiratory symptoms support the idea that the goats suffered an air-borne infection, probably with abundant and repetitive inocule. Although it is known that this is the main way of infection of *Cryptococcus*, mucous or digestive inoculation can not be rejected, as some animals presented digestive lesions and *C. gattii* was isolated in a sample of intestinal content of one goat.

Information obtained from the Infectious Diseases Departments of the main Hospitals of Cáceres, confirmed that human cases of cryptococcosis were rare without an increase in its frequency. The scarce isolates of *Cryptococcus* cultured from human samples were not typed.

More recently the first case of human cryptococcosis by *C. gattii* was reported in a patient resident in Spain [7], in a different and distant region, in south-east of the Peninsula (Alicante), with Mediterranean weather. In that case isolation of this species from environmental and soil samples, bird faeces or vegetal detritus (*Eucalyptus* was not present) was unfruitful.

The authors marked as uncontrolled antecedent the contact with a parrot belonging to the neighbours.

Between 1998 and 2000 in different zones of Catalunya (Northeast of the Peninsula), 132 samples of leaves, cortex, flowers and material from holes of *Eucalyptus* (*E. camaldulensis*, *E. globulus* and *E. darympleana*) were cultured without obtaining isolation of *C. gattii* or *C. neoformans*, but other no pathogenic species like *C. laurentii* and *C. albidus* were isolated [29].

In other parts of the world, different from Australia, isolation of *C. gattii* serotype B has been possible from nature linked to the presence of *Eucalyptus*, as in Cundinamarca (Colombia) where this yeast was isolated in 27,5% out of 167 samples of that tree, all of them of *mating type a* [15].

It is evident that in spite of the great amplitude of geographical distribution observed for *C. gattii*, this species has a limited presence, possibly restricted to certain habitats, as that those where the goats grazing.

In places different from those where *C. gattii* seems to be endemic, it is probable that animals with cryptococcosis become infected by *C. neoformans* from soil contaminated with a high number of yeasts.

Veterinarians must be concerned about Cryptococcosis in grazing animals. Health authorities must supply the adequate resources to perform a fast and accurate mycological diagnosis, as well as epidemiology analysis to know the actual prevalence of this disease in different geographical areas of the world. That information will help to design useful methods for controlling the disease.

SOURCES AND MANUFACTURERS

¹Iatron Labs Inc. Tokyo, Japan.

²Pastorex Crypto-latex. Bio-Rad Laboratories, SA. Alcobendas, Madrid 28109, Spain.

REFERENCES

- 1 Abegg M.A., Cella F.L. & Faganello J. 2006. *Cryptococcus neoformans* and *Cryptococcus gattii* isolated from the excreta of psittaciformes in a southern Brazilian zoological garden. *Mycopathologia*. 161: 83-91.
- 2 Baró T., Torres-Rodríguez J.M., De Mendoza M.H., Morera Y. & Alía C. 1998. First identification of autochthonous *Cryptococcus neoformans* var. *gattii* isolated from goats with predominantly severe pulmonary disease in Spain. *Journal of Clinical Microbiology*. 36: 458-461.
- 3 Cafarchia C., Romito D., Latta R., Camarda A., Montagna T. & Otranto D. 2006. Role of birds of prey as carriers and spreaders of *Cryptococcus neoformans* and other zoonotic yeasts. *Medical Mycology*. 44: 485-492.
- 4 Callejas A., Ordoñez N., Rodríguez M.C. & Castañeda E. 1998. First isolation of *Cryptococcus neoformans* var. *gattii* serotype C, from the environment in Colombia. *Medical Mycology*. 36: 341-344.

- 5 Casadevall A. & Perfect R.J. 1998. *Cryptococcus* and cryptococcosis. Washington. ASM Press. American Society of Microbiology.
- 6 Chee H.Y. & Lee K.B. 2005. Isolation of *Cryptococcus neoformans* var. *grubii* (serotype A) from pigeon droppings in Seoul, Korea. *Journal of Microbiology*. 43: 469-472.
- 7 Colom M.F., Frases S., Ferrer C., Jover A., Anfreu M., Réus S., Sánchez M. & Torres-Rodríguez J.M. 2005. First case of human cryptococcosis due to *Cryptococcus neoformans* var. *gattii* in Spain. *Journal of Clinical Microbiology*. 43: 3548-3550.
- 8 Criseo G., Bolignano M.S., De Leo F. & Staib F. 1995. Evidence of canary droppings as an important reservoir of *Cryptococcus neoformans*. *Zentralblatt Fuer Bakteriologie*. 282: 244-254.
- 9 Dora J.M., Kelbert S., Deutschendorf C., Cunha V.S., Aquino V.R., Santos R.P. & Goldani L.Z. 2006. Cutaneous cryptococcosis due to *Cryptococcus gattii* in immunocompetent hosts: case report and review. *Mycopathologia*. 161: 235-238.
- 10 Duncan C.G., Stephen C. & Campbell J. 2006. Evaluation of risk factors for *Cryptococcus gattii* infection in dogs and cats. *Journal of American Veterinary Medical Association*. 228: 377-382.
- 11 Duncan C., Schwantje H., Stephen C., Campbell J. & Bartlett K. 2006. *Cryptococcus gattii* in wildlife of Vancouver Island, British Columbia, Canada. *Journal of Wildlife Diseases*. 42:175-178.
- 12 Duncan C., Stephen C., Lester S. & Bartlett K. 2005. Sub-clinical infection and asymptomatic carriage of *Cryptococcus gattii* in dogs and cats during an outbreak of cryptococcosis. *Medical Mycology*. 43: 511-516.
- 13 Duncan C., Stephen C., Lester S. & Bartlett K. 2005. Follow-up study of dogs and cats with asymptomatic *Cryptococcus gattii* infection or nasal colonization. *Medical Mycology*. 43: 663-666.
- 14 Ellis D. & Pfeiffer T.J. 1990. Natural habitat of *Cryptococcus neoformans* var. *gattii*. *Journal of Clinical Microbiology*. 28: 1642-1644.
- 15 Escandon P., Quintero E., Granados D., Huérfano S., Ruíz A. & Castañeda E. 2005. Isolation of *Cryptococcus gattii*, serotype B from detritus of Eucalyptus trees in Colombia. *Biomedica*. 25: 390-397.
- 16 Ferreiro L., Loretta A.P., Appelt C.E. & Oliveira F.M. 2001. Disseminated cryptococcosis caused by *Cryptococcus neoformans* var. *gattii* in an immunocompromised siamese cat: a case report. [MV.029]. In: *Resumos do III Congresso Brasileiro de Micologia* (Águas de Lindóia, Brasil). p.129.
- 17 Gokulshankar S., Ranganathan S., Ranjith M.S. & Ranjithsingh A.J. 2004. Prevalence, serotypes and mating patterns of *Cryptococcus neoformans* in the pellets of different avifauna in Madras, India. *Mycoses*. 47: 310-314.
- 18 Gugnani H.C., Mitchell T.G., Litvintsera A.P., Lengeler K.B., Heitman J., Kumar A., Basu S. & Paliwal-Joshi. 2005. Isolation of *Cryptococcus gattii* and *Cryptococcus neoformans* var. *grubii* from the flowers and bark of Eucalyptus trees in India. *Medical Mycology*. 43: 565-569.
- 19 Hill F.I., Woodgyer A.J. & Lintott M.A. 1995. Cryptococcosis in a North Island brown kiwi (*Apteryx australis mantelli*) in New Zealand. *Journal of Medical & Veterinary Mycology*. 33: 305-309.
- 20 Kobayashi C., Souza L., Fernández O., Brito S., Silva A., Sousa E. & Silva M. 2005. Characterization of *Cryptococcus neoformans* isolated from urban environmental sources in Goiânia, Goiás, Brazil. *Revista do Instituto de Medicina Tropical de São Paulo*. 47: 203-207.
- 21 Krockenberger M.D., Candfield P.J. & Malik R. 2003. *Cryptococcus neoformans* var *gattii* in the koala (*Phascolarctus cinereus*): a review of 43 cases of cryptococcosis. *Medical Mycology*. 41: 225-234.
- 22 Kwon-Chung K.J., Boekhout T. & Fell J.W., 2002. Proposal to conserve the name *Cryptococcus gatti* against *C. honduricus* and *C. bacillisporus* (Basidiomycota, Hymenomycetes, Tremenomycetidae). *Taxon*. 51: 805-806.
- 23 Lagrou K., Van Eldere J., Keuleers S., Keuleers S., Hagen F., Merckx R., Verhaegen J., Peetermans W.E. & Boekhout T. 2005. Zoonotic transmission of *Cryptococcus neoformans* from a magpie to an immunocompetent patient. *Journal of Internal Medicine*. 257: 385-388.
- 24 Lazera M.S., Wanke B. & Nishikawa M.N. 1993. Isolation of both varieties of *Cryptococcus neoformans* from saprophytic source in the city of Rio de Janeiro. Brazil. *Journal of Medical & Veterinary Mycology*. 31: 449-454.
- 25 Lester S.J., Kowalewich N.J., Bartlett K.H., Krockenberger M.B., Fairfax T.M. & Malik R. 2004. Clinicopathologic features of unusual outbreak of Cryptococcosis in dogs, cats, ferrets and a bird: 38 cases (January to July 2003). *Journal of American Veterinary Medical Association*. 225: 1716-1722.
- 26 Mahmoud Yehia A.-G. 1999. First environmental isolation of *Cryptococcus neoformans* var. *neoformans* and var. *gatti* from the Gharbia Governorate, Egypt. *Mycopathologia*. 148: 83-86.
- 27 Meyer W., Castaneda A., Jackson S., Huynh M. & Castaneda E. 2003. Ibero American Cryptococcal Study Group. Molecular typing of Ibero American *Cryptococcus neoformans* isolates. *Emerging Infectious Diseases*. 9: 189-195.

- 28 Miller W.G., Padhye A.A., van Bonn W., Jensen E., Brandt M.E. & Ridgwa S.H. 2002. Cryptococcosis in a bottlenose dolphin (*Tursiops truncatus*) caused by *Cryptococcus neoformans* var *gattii*. *Journal of Clinical Microbiology*. 40: 721-724.
- 29 Morera-López Y., Torres-Rodríguez J.M., Jiménez-Cabello T., Baró-Tomás T., Alía-Aponte C. & Lázera M. 2005. DNA fingerprinting pattern and susceptibility to antifungal drugs in *Cryptococcus neoformans* variety *grubii* from Barcelona city and rural environmental samples. *Mycopathologia*. 169: 9-14.
- 30 Nosanchuk J.D., Shoham S., Fries B.C., Shapiro D.S., Levitz S.M. & Casadevall A. 2000. Evidence of zoonotic Transmission of *Cryptococcus neoformans* from a Pet Cockatoo to an immunocompromised. *Annals of Internal Medicine*. 132: 205-208.
- 31 O'Brien C.R., Krockenberger M.B., Wigney D.I., Martin P. & Malik R. 2004. Retrospective study of feline and canine cryptococcosis in Australia from 1981 to 2001: 195 cases. *Medical Mycology*. 42: 449-460.
- 32 Pollock C. 2003. Fungal diseases of Columbiformes and Anseriformes. *Veterinary Clinics of North America: Exotic Animal Practice*. 6: 351-361.
- 33 Raso T.F., Werter K., Miranda E.T. & Mendez-Giannini M.J. 2004. Cryptococcosis outbreak in psittacine birds in Brazil. *Medical Mycology*. 42: 355-362.
- 34 Riley C.B., Bolton J.R., Mills J.N. & Thomas J.B. 1992. Cryptococcosis in seven horses *Australian Veterinary Journal*. 69: 135-139.
- 35 Sorell T.C. 2001. *Cryptococcus neoformans* var. *gattii*. *Medical Mycology*. 39: 155-168.
- 36 Speed B. & Dunt D. 1995. Clinical and host differences between infections with the two varieties of *Cryptococcus neoformans*. *Clinical Infectious Diseases*. 21:28-34.
- 37 Swinne-Desgain D. 1974. The pigeon as reservoir of *Cryptococcus neoformans*. *Lancet*. 842-843.
- 38 Torres-Rodríguez J.M., Morera Y. & Baró-Tomás, T. 2004. Spontaneous infection of animals with *Cryptococcus neoformans* and *Cryptococcus gattii*. *Mikologia Lekarska*. 11: 303-307.
- 39 Weber A. & Schafer R. 1991. The occurrence of *Cryptococcus neoformans* in fecal samples from birds kept in human living areas. *Berliner und Munchener Tierärztliche Wochenschrift*. 104: 419-421.
- 40 Wegener H.H. & Staib F. 1983. Fatal cryptococcosis in a bird fancier. A clinical case report on pathology, diagnosis and epidemiology of cryptococcosis. *Zentralblatt Fuer Bakteriologie, Mikrobiologie und Hygiene*. 256: 231-238.