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***Leptospira* Seroprevalence in Capybaras from a Brazilian Urban Area**

Alexandre Alberto Tonin¹, Felipe da Silva Krawczak², Jéssica Carolina Gomes Noll³, Camila Tochetto⁴, Jorge Luiz Rodrigues Martins³, Manoel Renato Teles Badke³, Marcelo Bahia Labruna² & Aleksandro Schafer Da Silva⁵

ABSTRACT

Background: Leptospirosis remains the most widespread zoonotic disease in the world. As a clinical entity it is strongly associated with regional occupational and environmental exposures. While the exact global disease burden remains unknown, recent estimates by the leptospirosis Burden Epidemiology Reference Group (LERG) at the World Health Organization have set the number of human cases of severe leptospirosis to over 500,000 per year. This number almost certainly represents an under-representation due to poor surveillance and difficult diagnosis. *Leptospira* spp. belong to the order Spirochaetales, family Leptospiraceae, composed by 13 pathogenic *Leptospira* species with more than 260 serovars. Wildlife species are commonly considered to be important epidemiological carriers, mainly because of their frequent reactivity to *Leptospira* serovars native to their habitat. Capybara (*Hydrochaeris hydrochaeris*), a known leptospirosis host, is a widespread species in South America. However, reports regarding the importance of this animal in the epidemiology of leptospirosis are rare. Therefore, the objective of this study was to report the results of leptospirosis serological survey of capybaras from a residential park area in southeastern Brazil.

Materials, Methods & Results: A total of 172 capybaras were sampled at Itú Municipality, state of São Paulo, southeastern Brazil, from December 2012 to May 2013. Sera samples were examined for *Leptospira* antibodies by the microscopic agglutination test (MAT), using live antigens grown in liquid medium (EMJH). A complete panel of 7 serogroups (including 10 reference serovars) was used as antigens: serogroup Sejroe (serovars Hardjo and Wolffi), serogroup Grippotyphosa (serovar Grippotyphosa), serogroup Canicola (serovar Canicola), serogroup Icterohaemorrhagiae (serovars Icterohaemorrhagiae and Copenhageni), serogroup Australis (serovars Australis and Bratislava), serogroup Pomona (serovar Pomona), and serogroup Autumnalis (serovar Butembo). As a results, 46 (26.75%) were serologically positive: 29 (63.05%) for serogroup Sejroe, 7 (15.22%) for serogroup Icterohaemorrhagiae; 9 (19.56%) were seropositive for serogroups Sejroe and Icterohaemorrhagiae; and 1 (2.17%) was positive for serogroups Sejroe and Grippotyphosa.

Discussion: *L. interrogans* sensu stricto is responsible for the most frequent and severe cases of human and animal leptospirosis. Considering the results of our serological survey, it is important to reinforce that in tropical countries, mainly Brazil, India, Thailand, Vietnam, Australia and Barbados, serovars belonging to the Icterohaemorrhagiae serogroup (Icterohaemorrhagiae and Copenhageni) are generally the most prevalent. Therefore, the report of different serogroups in capybaras (as observed in our study) it is an important observation reported. Human leptospirosis is usually due to serovars that are maintained by the animal populations of a region, which spread the bacterium on the environment; thus, it may represent an important additional risk factor for human population. Our study identified a greater serum reactivity to strains that belongs to serogroup Sejroe. In Brazil this serogroup is highly predominant in livestock; thus, our main hypothesis is that some animals were contaminated with Sejroe serovars when they were at their natural habitat and, once in the park, they maintained these serovars through cross transmission. Therefore, based on our results, it was possible to observe a significant prevalence of serovars belonging to serogroup Icterohaemorrhagiae, and there was also a high prevalence of samples positive to serogroup Sejroe. Thus, our serologic survey showed that capybaras living in an urban area could represent a risk factor for leptospirosis for the human population eventually exposed.

Keywords: leptospirosis, wild host, *Hydrochaeris hydrochaeris*, urban environment.

INTRODUCTION

Leptospirosis has traditionally been described in the medical literature as a treatable zoonotic disease endemic to low-income countries in temperate and tropical regions [5]. As a clinical entity it is strongly associated with regional occupational and environmental exposures [27]. While the exact global disease burden remains unknown, recent estimates by the leptospirosis Burden Epidemiology Reference Group (LERG) at the World Health Organization have set the number of human cases of severe leptospirosis to over 500,000 per year [1]. This number almost certainly represents an under-representation due to poor surveillance and difficult diagnosis [20].

This agent belongs to the order Spirochaetales, family Leptospiraceae [14], composed by 13 pathogenic *Leptospira* species with more than 260 serovars [3]. Several species of water plants contaminated by leptospires have potential to be sources of most serogroups [24]; however, the primary reservoirs for infection are microtine rodents [2,9,30] and various game species [6,21]. Wildlife species are commonly considered to be important epidemiological carriers, mainly because of their frequent reactivity to *Leptospira* serovars native to their habitat [23].

Capybara (*Hydrochaeris hydrochaeris*) is the largest rodent in the world and a widespread species in South America [13]. However, reports regarding the importance of this animal in the epidemiology of leptospirosis are rare, especially in urban areas. Capybaras could be efficient reservoirs for *Leptospira*, because they live in areas with abundant water, an important environmental factor for disease transmission [19]. Based on these aspects, the goal of this study was to report the results of a leptospirosis serological survey of capybaras from a residential park area in southeastern Brazil.

MATERIALS AND METHODS

Animals and blood sampling

Capybaras were sampled in a residential park area at Itú Municipality, state of São Paulo, southeastern Brazil. The residential park had an area of 484 ha, which contained four lakes, and ≈400 homes interposed by conserved forest areas. Because of a Brazilian spotted fever case (caused by the bacterium *Rickettsia rickettsii* and transmitted by the capybara tick *Amblyomma cajennense*) in a child of the residential park in 2012, its entire

capybara population was legally authorized to be culled with the purpose to reduce environmental tick burdens [22]. For this purpose, from December 2012 to May 2013, a total of 172 capybaras were collected by corrals [26], and before been culled, individual blood samples were collected, the sera separated by centrifugation, and stored frozen at -20°C until further testing.

Microscopic agglutination test (MAT)

Samples were examined for *Leptospira* antibodies by the microscopic agglutination test (MAT) [8,15], using live antigens grown in liquid medium (EMJH)¹ free of contamination or self-agglutination., and as recommended by the World Health Organization [34]. A complete panel of 7 serogroups (including 10 reference serovars) was used as antigens: serogroup Sejroe (serovars Hardjo and Wolffi), serogroup Grippotyphosa (serovar Grippotyphosa), serogroup Canicola (serovar Canicola), serogroup Icterohaemorrhagiae (serovars Icterohaemorrhagiae and Copenhageni), serogroup Australis (serovars Australis and Bratislava), serogroup Pomona (serovar Pomona), and serogroup Autumnalis (serovar Butembo). Briefly, live suspensions of leptospires representing the 10 serovars were added to serially diluted serum specimens in 96-well microtiter plates and incubated at room temperature for 2 h. Agglutination was examined using dark-field microscopy at × 100 magnification. Titers were calculated as the highest serum dilution that agglutinated at least 50% of the leptospires for each serovar used [7].

RESULTS

Our results showed that from the 172 sera samples tested, 46 (26.75%) were serologically positive. The serogroups distribution showed that 29 (63.05%) samples were positive only for serogroup Sejroe, 7 (15.22%) only for serogroup Icterohaemorrhagiae, while 9 (19.56%) samples were seropositive for serogroups Sejroe and Icterohaemorrhagiae and 1 (2.17%) sample was positive for serogroups Sejroe and Grippotyphosa. Recent studies conducted in many tropical countries reinforce the complex epidemiological relationship between human/animal leptospirosis [1], reinforcing the importance of our results.

DISCUSSION

L. interrogans sensu stricto is responsible for the most frequent and severe cases of human and animal leptospirosis [34]. Synanthropic and wild rodents have

been considered important *Leptospira* reservoirs [33]. In Brazil, a serological *Leptospira* survey in capybaras culled in a slaughterhouse was performed [31], showing animals with high titers of agglutinating antibodies against a serovar Bratislava and serovar Australis, serovars that belong to serogroup Australis. Differently from this result, our findings serologically classified the samples in other different serogroups, highlighting the potential of infection of this wild species.

The report of different serogroups in capybaras living in an urban area it is an important observation, since human leptospirosis is usually due to serovars that are maintained by the animal populations of a region, which spread the bacterium on the environment [28,32]. Considering the results of our serological survey, it is important to reinforce that in tropical countries, mainly Brazil, India, Thailand, Vietnam, Australia and Barbados, serovars belonging to the Icterohaemorrhagiae serogroup, for example, Icterohaemorrhagiae and Copenhageni, are generally the most prevalent [4,17]. Thus, it may represent an important risk factor, especially considering that changes in human demography during the last 50 years have raised awareness of the emergence of leptospirosis as an urban health problem [20]. For example, in slum settings, endemic transmission of leptospirosis is largely due to circulation of a single serogroup, *L. interrogans* serogroup Icterohaemorrhagiae [10,16,20,29]. In this context, animals subjected to contaminated water sources living in urban areas may have potential to become hosts and diseases carriers for the general population or even to other domestic animals.

Our study identified a greater serum reactivity to strains that belongs to serogroup Sejroe. In Brazil this serogroup is highly predominant in livestock (especially cattle, sheep and goat) [11,12,18]. Considering our studied area as urban, the main hypothesis for this high Sejroe seroprevalence is the report of epizootiological

studies indicating that there is a high degree of adaptation between house mice (*Mus musculus*) and serogroup Sejroe [31]. The transition through ruminant hosts was ruled out because the park area is closed, and on that time, there were only capybaras into the area. Another explanation laid on the hypothesis that some animals were contaminated with Sejroe serovars when they were at their natural habitat and, once in the park, they maintained these serovars through cross transmission. Considering the absence of natural hosts for Sejroe serogroup, this serologic information confirms that capybaras may are good hosts for leptospirosis, setting them as important epidemiologic subjects.

CONCLUSION

Based on our results, it was possible to observe a significant prevalence of serovars belonging to serogroup Icterohaemorrhagiae, which use to cause public health problems, due to the severe presentation for some patient. There was also a high prevalence of samples positive to serogroup Sejroe, which considering the absence of natural host for these serovars, indicate that capybaras are able to maintain these serovars into the environment. Therefore, our serologic survey showed that capybaras living in an urban area could represent a risk factor for leptospirosis for the human population eventually exposed.

MANUFACTURER

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Ethical Approval. This work was authorized by the Environment State Secretary of the state of São Paulo (authorization no. 96/2012) and was approved by the Ethical Committee of Animal Use of the Faculty of Veterinary Medicine of the University of São Paulo (protocol No. 3104/2013).

Declaration of interest. The authors report no conflicts of interest. The authors alone are responsible for the content and writing of the paper

REFERENCES

- 1 Abela-Ridder B., Sikkema R. & Hartskeerl R.A. 2010. Estimating the burden of human leptospirosis. *International Journal of Antimicrobial Agents*. 36 (Suppl. 1): S5-S7.
- 2 Adler H., Vonstein S., Deplazes P., Steiger C. & Frei R. 2002. Prevalence of *Leptospira* spp. in various species of small mammals in an inner-city area Switzerland. *Epidemiology and Infection*. 128: 107-109.
- 3 Adler B. & de la Penã Moctezuma A. 2010. *Leptospira* and leptospirosis. *Veterinary Microbiology*. 140: 287-296.
- 4 Båverud V., Gunnarsson A., Engvall E.O., Franzén P. & Egenvall A. 2009. *Leptospira* seroprevalence and associations between seropositivity, clinical disease and host factors in horses. *Acta Veterinaria Scandinavica*. 51: 1-10.
- 5 Bharti A.R., Nally J.E., Ricaldi J.N., Matthias M.A., Diaz M.M., Lovett M.A., Levett P.N., Gilman R.H., Wiliig M.R., Gotuzzo E. & Vinetz J.M. 2003. Leptospirosis: a zoonotic disease of global importance. *Lancet Infectious*

- Diseases*. 3: 757-771.
- 6 **Bondarenko A.L., Utenkova E.O., Russkikh G.A. & Khmelevskaia N.S. 2002.** Epidemiology of leptospirosis in the Kirov region. *Zhurnal mikrobiologii, epidemiologii, i immunobiologii*. 3: 27-30.
 - 7 **Centers for Disease Control and Prevention (CDC). 1997.** Case definitions for infectious conditions under public health surveillance. *Morbidity and Mortality Weekly Report*. 46 (RR-10), 49.
 - 8 **Cole J.R., Sulzer C.R. & Pulsely P.R. 1973.** Improved microtechnique for the leptospiral microscopic agglutination. *Applied Microbiology*. 25: 976-980.
 - 9 **Collares-Pereira M., Mathias M.L., Santos-Reis M., Ramalhinho M.G. & Duarte-Rodrigues P. 2000.** Rodents and *Leptospira* transmission risk in Terceira Island (Azores). *European Journal of Epidemiology*. 16: 1151-1157.
 - 10 **De Faria M.T., Calderwood M.S., Athanazio D.A., McBride A.J., Hartskeerl R.A., Pereira M.M., Ko A.I. & Reis M.G. 2008.** Carriage of *Leptospira interrogans* among domestic rats from a high endemic urban setting for leptospirosis in Brazil. *Acta Tropica*. 108: 1-5.
 - 11 **De Nardi G., Genovez M.E., Ribeiro M.G., Castro V. & Jorge A.M. 2010.** An *in vitro* growth inhibition test for measuring the potency of *Leptospira* spp. Sejroe group vaccine in buffaloes. *Biologicals*. 38: 474-478.
 - 12 **Dos Santos J.P., Lima-Ribeiro A.M.C., Oliveira P.R., dos Santos M.P., Ferreira A., Medeiros A.A. & Tavares T.C.F. 2012.** Seroprevalence and risk factors for leptospirosis in goats in Uberlândia, Minas Gerais, Brazil. *Tropical Animal and Health Production*. 44: 101-106.
 - 13 **Emmons L.H. 1990.** *Neotropical Rainforest Mammals - a field guide*. 2nd edn. Chicago: University of Chicago Press, 396p.
 - 14 **Faine S., Adler B., Bolin C. & Perolat P. 1999.** *Leptospira and leptospirosis*. Mediscience, Melbourne.
 - 15 **Galton M.M., Sulzer C.R., Santa Rosa C.A. & Fields M.J. 1965.** Application of microtechnique to the agglutination test for leptospiral antibodies. *Applied Microbiology*. 13: 81-85.
 - 16 **Gouveia E.L., Metcalfe J., de Carvalho A.L., Aires T.S., Villasboas-Bisneto J.C., Queirroz A., Santos A.C., Salgado K., Reis M.G. & Ko A.I. 2008.** Leptospirosis-associated severe pulmonary hemorrhagic syndrome, Salvador, Brazil. *Emerging Infectious Diseases*. 14: 505-508.
 - 17 **Hamond C., Martins G., Reis J., Kraus E., Pinna A. & Lilenbaum W. 2011.** Pulmonary hemorrhage in horses seropositive to leptospirosis. *Pesquisa Veterinária Brasileira*. 31: 413-415.
 - 18 **Ishikawa M.M., Fonseca A.H., Soares C.O. & Yoshinari N.H. 1999.** Comparative serological study of Lyme Borreliosis, Brucellosis and Leptospirosis in cattle. *Revista de Patologia Tropical* 28: 195-291.
 - 19 **Ito F.H., Vasconcellos S.A., Bernardi F., Nascimento A.A., Labruna M.B. & Arantes I.G. 1998.** Evidência sorológica de brucelose e leptospirose e parasitismo por ixodídeos em animais silvestres do pantanal sul-mato-grossense. *Ars Veterinaria*. 14: 302-310.
 - 20 **Ko A.I., Galvao Reis M., Ribeiro Dourado C.M., Johnson Jr. W.D. & Riley L.W. 1999.** Urban epidemic of severe leptospirosis in Brazil. Salvador Leptospirosis Study Group. *Lancet*. 354: 820-825.
 - 21 **Koizumi N., Muto M., Yamamoto S., Baba Y., Kudo M., Tamae Y., Shimomura K., Takatori I., Iwakiri A., Ishikawa K., Soma H. & Watanabe H. 2008.** Investigation of reservoir animals of *Leptospira* in the northern part of Miyazaki Prefecture. *Japanese Journal of Infectious Diseases*. 61: 465-468.
 - 22 **Krawczak F.S., Nieri-Bastos F.A., Nunes F.P., Soares J.S., Moraes-Filho J. & Labruna M.B. 2014.** Rickettsial infection in *Amblyomma cajennense* ticks and capybaras (*Hydrochoerus hydrochaeris*) in a Brazilian spotted fever-endemic area. *Parasites & Vectors*. 7: 7.
 - 23 **Lins Z.C. & Lopes M.L. 1984.** Isolation of *Leptospira* from wild forest animals in Amazonian Brazil. *Transactions of the Royal Society of Tropical Medicine and Hygiene*. 78, 124-126.
 - 24 **Mailloux M. 1980.** Leptospiroses et environnement. *Revue d'Epidémiologie et de Santé Publique*. 28: 323-327.
 - 25 **Martins G. & Lilenbaum W. 2013.** The panorama of animal leptospirosis in Rio de Janeiro, Brazil, regarding the seroepidemiology of the infection in tropical regions. *BMC Veterinary Research*. 9: 237.
 - 26 **Moreira J.R., Ferraz K.M.P.M.B., Herrera E.A. & Macdonald D.W. 2013.** *Capybara: Biology, Use and Conservation of an Exceptional Neotropical Species*. New York: Springer Science, 422p.
 - 27 **Murhekar M.V., Sugunan A.P., Vijayachari P., Sharma S. & Sehgal S.C. 1998.** Risk factors in the transmission of leptospiral infection. *Indian Journal of Medicine Research*. 107: 218-223.
 - 28 **Oliveira T.V., Marinho D.P. & Costa Neto C. 2012.** Climate variables, living conditions and the health of the popula-

- tion: leptospirosis in the city of Rio de Janeiro from 1996 to 2009. *Ciência e Saúde Coletiva*. 17: 1569-1576.
- 29 Reis R.B., Ribeiro G.S., Felzemburgh R.D.M., Santana F.S., Mohr S., Melendez A.X.T.O., Queiroz A., Santos A.C., Ravines R.R., Tassinari W.S., Carvalho M.S., Reis M.G. & Ko A.I. 2008.** Impact of Environment and Social Gradient on *Leptospira* Infection in Urban Slums. *PLoS Neglected Tropical Diseases*. 2: e228.
- 30 Songer J.G., Chilelli C.J., Reed R.E. & Trauman R.J. 1983.** Leptospirosis in rodents from an arid environment. *American Journal of Veterinary Research*. 44: 1973-1976.
- 31 Silva E.F., Seyffert N., Jouglard S.D.D., Athanzio D.A., Dellagostin O.A. & Brod C.S. 2009.** Soroprevalência da infecção leptospiral em capivaras (*Hydrochoerus hydrochaeris*) abatidas em um frigorífico do Rio Grande do Sul. *Pesquisa Veterinária Brasileira*. 29: 174-176.
- 32 Tassinari W.S., Pellegrini D.C.P., Sá C.B.P., Reis R.B., Ko A.I. & Carvalho M.S. 2008.** Detection and modelling of case clusters for urban leptospirosis. *Tropical Medicine & International Health*. 13: 503-512.
- 33 Turk N., Milas Z., Margaletic J., Staresina V., Slavica A., Riquelme-Sertour N., Bellenger E., Baranton G. & Postic D. 2003.** Molecular characterization of *Leptospira* spp. strain isolated from small rodents in Croatia. *Epidemiology & Infection*. 130: 159-166.
- 34 WHO-World Health Organization, Leptospirosis 2012.** Available in http://www.wpro.who.int/mediacentre/factsheets/fs_13082012_leptospirosis/en/. [Accessed September 2015].

