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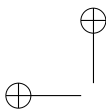
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## Associative diazotrophic bacteria in grass roots and soils from heavy metal contaminated sites

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### ABSTRACT

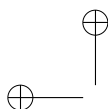
This work aimed to evaluate density of associative diazotrophic bacteria populations in soil and grass root samples from heavy metal contaminated sites, and to characterize isolates from these populations, both, phenotypically (Zinc, Cadmium and NaCl tolerance *in vitro*, and protein profiles) and genotypically (16S rDNA sequencing), as compared to type strains of known diazotrophic species. Densities were evaluated by using NFb, Fam and JNFb media, commonly used for enrichment cultures of diazotrophic bacteria. Bacterial densities found in soil and grass root samples from contaminated sites were similar to those reported for agricultural soils. *Azospirillum* spp. isolates from contaminated sites and type strains from non-contaminated sites varied substantially in their *in vitro* tolerance to Zn<sup>+2</sup> and Cd<sup>+2</sup>, being Cd<sup>+2</sup> more toxic than Zn<sup>+2</sup>. Among the most tolerant isolates (UFLA 1S, 1R, S181, S34 and S22), some (1S, S34 and S22) were more tolerant to heavy metals than rhizobia from tropical and temperate soils. The majority of the isolates tolerant to heavy metals were also tolerant to salt stress as indicated by their ability to grow in solid medium supplemented with 30 g L<sup>-1</sup> NaCl. Five isolates exhibited high dissimilarity in protein profiles, and the 16S rDNA sequence analysis of two of them revealed new sequences for *Azospirillum*.

**Key words:** *Azospirillum*, Zinc, Cadmium, soil contaminants, Nitrogen-fixing bacteria.

### INTRODUCTION

Mining and metallurgical industry play an important role in worldwide economy, particularly in Brazil. However, these industries can be responsible for environmental impacts caused mainly by heavy metal (HM) contamination of soil, vegetation and water bodies. Soil contamination with HM is a major constraint for plant growth (Marques et al. 2000) and for microbial density, diversity and processes, including biological Nitrogen fixation (Tyler 1981, Hirsch et al. 1993, Dias-Júnior et al. 1999, Gill et al. 1999, Klauber-Filho et al. 2002).

Biological Nitrogen fixation is mediated by several prokaryotic species occurring both in soil and rhizosphere, that can establish either symbiotic or associative relationships with a variety of plant species concerning to their N-nutrition. Among associative N-fixers, *Azospirillum* spp., *Burkholderia* and *Herbaspirillum* spp., are reported to occur in the ecto and rhizosphere of grasses and in other monocotyledonous species (Döbereiner 1978, Magalhães and Döbereiner 2001, Santos et al. 2001, Baldani and Baldani 2005). In view of the great volume of information about these



nitrogenase activity by heterotrophic soil bacterial populations in HM contaminated soil treated with sewage sludge. These authors also found high negative correlation between nitrogenase activity and Zn, Cd and Cu concentrations in the soil. Hayat et al. (2002) found that asymbiotic Nitrogen fixer populations, in soil receiving oil refinery wastewater, tolerate Ni, Cd, Pb, Cu and Zn tested individually, at concentrations up to  $400\mu\text{g mL}^{-1}$ . In the only study with associative diazotrophic isolates (Strzelczyk et al. 1997), it was found a similar behavior of isolates to different HM (Zn, Cd and Cu). However, results of these work are highly contradictory to what has been reported for other metal-bacteria interactions showing markedly differentiated response to distinct metals. The understanding of associative diazotroph behavior in HM contaminated soil may provide good insights for the ecology and application of these organisms in rehabilitation practices.

In this work we assessed associative diazotrophic bacterial population densities in heavy metals contaminated soils and grass roots. We also characterized, phenotypically (Zinc, Cadmium, and NaCl tolerance *in vitro* and protein profiles) and genotypically (16S rDNA sequencing) isolates from these sites in comparison to type strains.

## MATERIALS AND METHODS

### STUDIED SITES AND SAMPLING

Soil and grass root samples were collected in October (dry season) from five dumping sites near a Zinc processing industry (Companhia Mineira de Metais – CMM), located at Três Marias (MG). Zn and Cd were the main soil contaminants in all sites, as their contents were very high considering the allowed limits according to the European Communities (Table I). All these sites have the predominance of grass species such as: *Brachiaria decumbens*, *B. brizantha*, *B. mutica* and *Andropogon* sp. From each site, three samples of rhizosphere soil and three samples of roots were randomly collected at 0–20 cm depth. The samples were placed in heat-isolated protective cases, and transferred to the laboratory where they were stored at  $-4^{\circ}\text{C}$  until used for analysis.

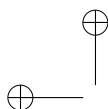
lation of successive decimal dilutions of samples (first dilution: 10 g root or soil/95 mL sterile NaCl solution  $5.5\text{ g L}^{-1}$ ) in semi-solid media, known to favor growth of *Azospirillum* spp. (*A. lipoferum*, *A. brasilense*, *A. amazonense*) (Magalhães and Döbereiner 1984, Döbereiner et al. 1995) and *Herbaspirillum* spp. (*H. seropedicae*, *H. rubrisubalbicans*) (Döbereiner et al. 1995) as well as other diazotrophs (Nóbrega et al. 2004) such as *Burkholderia* spp. The following media were used: Fam (Magalhães and Döbereiner 1984), NFB and JNFB (Döbereiner et al. 1995). Media were distributed into 10 mL vials (4 mL medium/vial). From each sample dilution, 0.1 mL was inoculated in each medium with three replicates (vials). Positive cultures for growth of diazotrophs were indicated by pellicle formation near medium surface. Isolates were obtained from single colonies and identified according to procedures described by Magalhães and Döbereiner (1984) and Döbereiner et al. (1995).

### TYPE STRAINS

Type and reference strains of *Azospirillum brasilense* (BR 11001<sup>T</sup>), *A. lipoferum* (BR 11080<sup>T</sup>), *A. amazonense* (BR 11140<sup>T</sup>), *A. irakense* (KCB1<sup>T</sup>), *Herbaspirillum seropedicae* (BR 11175<sup>T</sup>) and reference strain *Burkholderia* sp. (previously named *B. brasiliensis*) (BR 11340), as well as reference strain BR 9004 identified as *Burkholderia* sp. (F.M.S. Moreira, unpublished results) were also included in the phenotypic assays for comparison with isolates from contaminated sites.

### INOCULUM

Isolates and type strains were grown in liquid potato dextrose medium under 105 rpm at  $28^{\circ}\text{C}$ . After 10 days, one mL of each cell culture was submitted to centrifugation (Eppendorf centrifuge 5402, 14 rpm, 5 min,  $10^{\circ}\text{C}$ ), discarding the supernatant and re-suspending cells in 1 mL saline solution ( $\text{NaCl } 5.5\text{ g L}^{-1}$ ), respectively. This washing procedure was repeated three times to remove completely medium components from cell cultures that could cause a false positive growth in contaminated media. One mL of washed cells suspension in saline solution was transferred to modified solid media (potato dextrose)



## N<sub>2</sub>-FIXING-BACTERIA FROM HEAVY METAL CONTAMINATED SITES

**TABLE I**  
**Heavy metal concentrations (Mehlich1) in soil samples**  
**from the various sites.**

Site/grass species	Heavy metal (mg.kg <sup>-1</sup> ) <sup>(1)</sup> in soil				
	Zn	Cd	Mn	Cu	Pb
1 – <i>Brachiaria decumbens</i>	5739	67	267	4	30
2 – <i>Andropogon</i> sp.	5902	105	114	530	90
3 – <i>B. decumbens</i>	12873	46	34	548	144
4 – <i>B. mutica</i>	3960	223	76	215	134
5 – <i>B. mutica</i>	15279	76	78	722	198

(1) Permissible limits in European Communities: Zn = 300; Cd = 3; Cu = 140; Pb = 300 mg.kg<sup>-1</sup> (“aqua regia” extracted).

### TOLERANCE TO ZN, CD

Tolerance to Zn and Cd was assessed in potato dextrose agar and in semi-solid Nitrogen-free media (Fam, NFb) in which diazotrophs grow acquiring atmospheric Nitrogen through nitrogenase activity. In both experiments, solid media and N-free semi-solid media were modified by the addition of biological buffers HEPES (1.3 mg L<sup>-1</sup>) and MES (1.1 mg L<sup>-1</sup>) and were supplemented with metals at concentrations ranging from 0 to 1,000 mg Zn<sup>+2</sup> L<sup>-1</sup> and from 0 to 54.1 mg Cd<sup>+2</sup> L<sup>-1</sup>. Assayed concentrations were defined from results of the effects of these metals on tropical rhizobia isolates (Trannin et al. 2001a, Matsuda et al. 2002). Modified potato dextrose agar media was supplemented after autoclaving, with sterile solutions of ZnSO<sub>4</sub>.7H<sub>2</sub>O and/or CdSO<sub>4</sub>.8H<sub>2</sub>O to give final desirable concentrations of 0; 125; 250; 500 and 1,000 mg Zn<sup>2+</sup> L<sup>-1</sup> or 0; 1.7; 3.4; 6.8; 13.5; 27.1 and 54.1 mg Cd<sup>2+</sup> L<sup>-1</sup>. Zn<sup>+2</sup> and Cd<sup>+2</sup> were tested separately or in combinations of different concentrations in potato dextrose medium. Metals combined in potato dextrose agar were only tested for *A. amazonense*. Fam and NFb semi-solid N-free media were supplemented with sterile solutions of ZnSO<sub>4</sub>.7H<sub>2</sub>O or CdSO<sub>4</sub>.8H<sub>2</sub>O at the following concentrations of Zn<sup>2+</sup> and Cd<sup>2+</sup>: 0; 62.5; 125; 250; 500, 1,000 mg L<sup>-1</sup> and 0; 2.5; 5.0; 10.0; 20.0; 40.0, 60.0 mg L<sup>-1</sup>, respectively. Media receiving Zn sulfate (ZnSO<sub>4</sub>.7H<sub>2</sub>O) had the pH adjusted to 6.8 by addition of KOH 1N

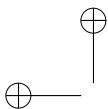
0 = no visual growth, 1 = very reduced growth, 2 = growth, 3 = intermediate growth, 4 = abundant growth, 5 = very abundant growth as compared to the of the tested isolate/strain in the same medium without heavy metal supplementation. Maximum tolerance concentration (MTC) was determined as being the one with a growth score equal or higher than 1. Formation of typical pellicles near the surface was considered as positive growth in semi-solid modified media (Magalhães and Döbereiner 1984, Döbereiner et al. 1995). Nitrogenase activity was measured by acetylene reduction technique (Dilworth 1966) in semi-solid media in four day old cultures by using gas chromatograph Varian 3400-CX.

### TOLERANCE TO NaCl

In order to verify relationships between tolerance to different stresses, strains were also tested for NaCl tolerance. Potato dextrose agar medium was modified to obtain the following final NaCl concentrations: 0, 30, 50 and 80 g L<sup>-1</sup>, based on ranges commonly found in the literature (Tortora et al. 2000).

### TOTAL PROTEIN PROFILES

Cultures were obtained as described previously for associative diazotrophs (Nóbrega et al. 2004). Cell protein extracts were prepared and sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE) performed by small modifications of



were compared for similarity degree estimated by Jacard coefficient (Sj), clustered by UPGMA method (*average linkage clustering*) (Everitt 1993) and graphically represent through a dendrogram, obtained by the program NTSYS-pc, version 2.0 t. (Rohlf 2002).

#### ANALYSES OF 16S rDNA SEQUENCING

Genomic DNA was obtained from potato medium log phase cultures of two isolates (UFLA1S and UFLA S181 – which were among the most tolerant to NaCl) and extracted with Ultra-clean Soil DNA isolation kit from MOBIO laboratories. For 16S rRNA sequencing, near full-length 16S rDNA genes were amplified with prime pair 8F (AGAGTTTGATCCTGGCTCAG) and 1492R (GGTTACCTTGTTACGACTT). PCR conditions were: 30 cycles of 94°C 40s; 55°C 40s; 72°C 90s. Directional partial sequencing of PCR-products was performed with 8F primer. Purification of PCR products was performed with centrifugal filter devices Microcon Millipore. The isolates were sequenced at an ABI 3700 sequencer (Michigan State University). Sequences obtained were compared through BLAST software and submitted to the “National Center for Biotechnology Information” data base (<http://www.ncbi.nlm.nih.gov/>).

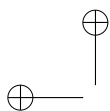
### RESULTS

#### BACTERIAL DENSITIES AND ISOLATES

Bacterial growth varied markedly amongst samples. Densities found were higher in roots than in soil samples (Table II). Densities found in soil samples (log most probable number  $g^{-1}$  dry soil) by using Fam, NFb and JNFB were respectively in the ranges: 0.57 to 2.34, 1.91 to 4.34 and 0 to 2.72, whereas in root samples, the range of densities (log most probable numbers  $g^{-1}$  fresh root) were: 2.32 to 3.68, 3.5 to 5.63 and 2.27 to 5.5. Seven isolates were obtained from soil (S) or root (R) samples inoculated in these media. Four of them had typical cultural characteristics of *A. brasilense* (UFLA 1R, S181, R214) or *A. lipoferum* (UFLA 1S) (Döbereiner et al. 1995) and three were similar to *A. amazonense* (UFLA R204, S34, S22) (Magalhães and Döbereiner 1984).

isolates (Figs. 1 and 2). Only *A. amazonense* isolates UFLA S34 and UFLA S22 and *A. brasilense* isolate UFLA 1R had MTC of  $1,000 \text{ mg Zn}^{+2} \text{ L}^{-1}$ . They had growth score equal to 2 (UFLA S34 and S22) or 4 (UFLA 1R) at this concentration (Fig. 1). Considering that  $1,000 \text{ mg L}^{-1} \text{ Zn}^{+2}$  was the highest concentration tested, perhaps these isolates could tolerate concentrations higher than that. No strain or isolate grew at  $54.1 \text{ mg Cd}^{+2} \text{ L}^{-1}$ . All isolates and strains tested in this study had MTC of  $\text{Cd}^{+2}$  equal to  $27.1 \text{ mg L}^{-1}$  (Fig. 2). However isolates UFLA S34, UFLA S22 and UFLA 1R, which were more tolerant to  $\text{Zn}^{+2}$ , had the highest growth score (4) at this concentration while the others had a growth score of 3 (UFLA 1S), 2 (UFLA R204, UFLA S181) or 1 (type strains and UFLA R214). *A. amazonense* did not grow at  $\text{Zn}^{+2}$  concentration of  $500 \text{ mg L}^{-1}$  in the presence of different concentrations of  $\text{Cd}^{+2}$  (Fig. 3). When  $\text{Zn}^{+2}$  concentrations were raised at fixed  $\text{Cd}^{+2}$  concentrations growth was depressed and the MTCs of  $\text{Zn}^{+2}$  were lower than when it was tested separately, indicating an additive toxic effect of these metals for *A. amazonense*. However, at fixed  $\text{Zn}^{+2}$  concentrations of 62.5 and  $125 \text{ mg L}^{-1}$  increasing the  $\text{Cd}^{+2}$  concentration did not affect or had little effect on growth score of UFLA S34 and UFLA S22 indicating antagonism of  $\text{Zn}^{+2}$  to  $\text{Cd}^{+2}$  at these combinations of concentrations. Time required for appearance of isolated colonies was delayed and colony size decreased with increasing concentrations of combined metals (Fig. 4). Also, it was observed that color of *A. amazonense* colonies shifted from white to yellowish under high  $\text{Zn}^{+2}$  and  $\text{Cd}^{+2}$  concentrations. However, the white color was recovered when colonies grown in high heavy metal concentration medium were replicated to metal free medium, indicating a metal induced response.

The most tolerant *A. amazonense* isolates (UFLA S34, S22) to  $\text{Zn}^{+2}$  and  $\text{Cd}^{+2}$  growing at combined N (potato dextrose media) were also the most tolerant in Nitrogen-fixing conditions (N-free semi-solid media), but in this medium, BR 11140<sup>T</sup> exhibited the same tolerance of isolates belonging to this species. Tolerance to  $\text{Zn}^{+2}$  of *A. amazonense* strains/isolates in  $\text{N}_2$  fixing con-



# N<sub>2</sub>-FIXING-BACTERIA FROM HEAVY METAL CONTAMINATED SITES

TABLE II

Density (log most probable number g<sup>-1</sup> dry soil or fresh roots) of diazotrophic bacteria grown in media: Fam, NFb and JNFB in soil and roots samples collected at different sites of a dumping site of zinc industry.

Site	Soil			Roots		
	Fam	NFb	JNFB	Fam	NFb	JNFB
1	2.34 ± 2.03 <sup>1</sup>	2.50 ± 2.28	2.72 ± 2.39	3.12 ± 2.72	3.5 ± 1.9	4.6 ± 0.7
2	1.03 ± 1.78	4.34 ± 1.57	2.17 ± 1.88	3.68 ± 1.34	5.63 ± 0.89	5.5 ± 2.3
3	1.31 ± 1.14	1.30 ± 1.22	0	2.32 ± 2.24	5.31 ± 0.73	4.86 ± 0.86
4	1.02 ± 0.1	2.59 ± 1.15	2.10 ± 1.87	3.04 ± 0.97	5.14 ± 0.53	2.27 ± 1.98
5	0.57 ± 0.98	1.91 ± 1.66	1.0 ± 1.73	2.81 ± 2.44	4.03 ± 0.82	4.44 ± 0.65

<sup>1</sup> mean standard deviation.

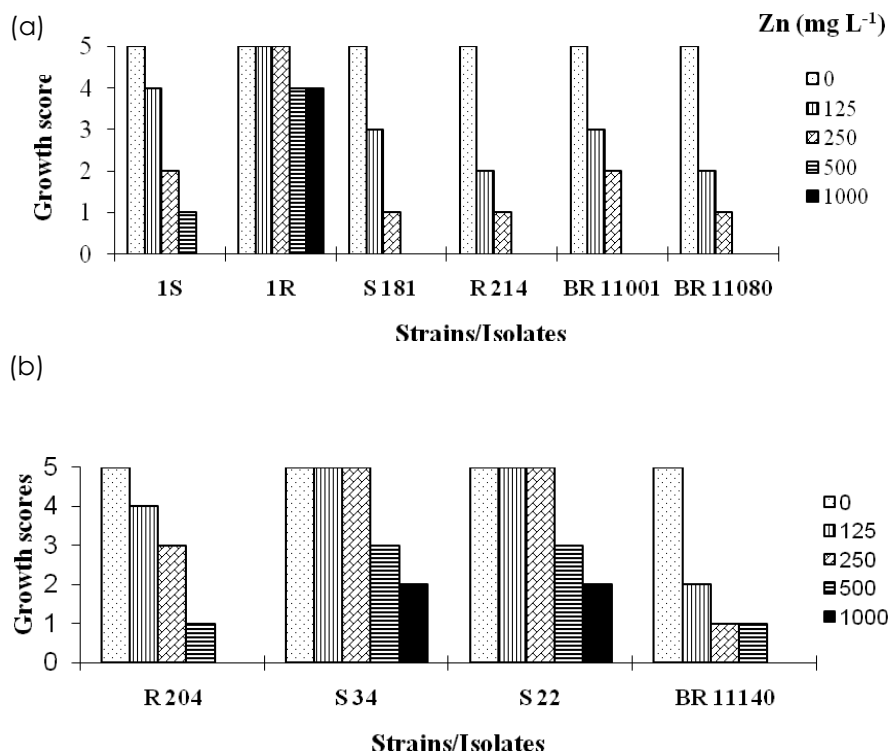


Fig. 1 – Growth scores of associative diazotrophic bacteria isolated from heavy metal contaminated area and type strains in potato dextrose medium supplemented with different concentrations of Zn<sup>2+</sup> (mg L<sup>-1</sup>) ten days after inoculation. a) *Azospirillum lipoferum* (1S and 1R) and *A. brasilense* (1R, S181, R214, BR 11001<sup>T</sup>); b) *A. amazonense* (R204, S34, S22, BR 11140<sup>T</sup>). 0 = no visual growth, 1 = very reduced growth, 5 = full growth.

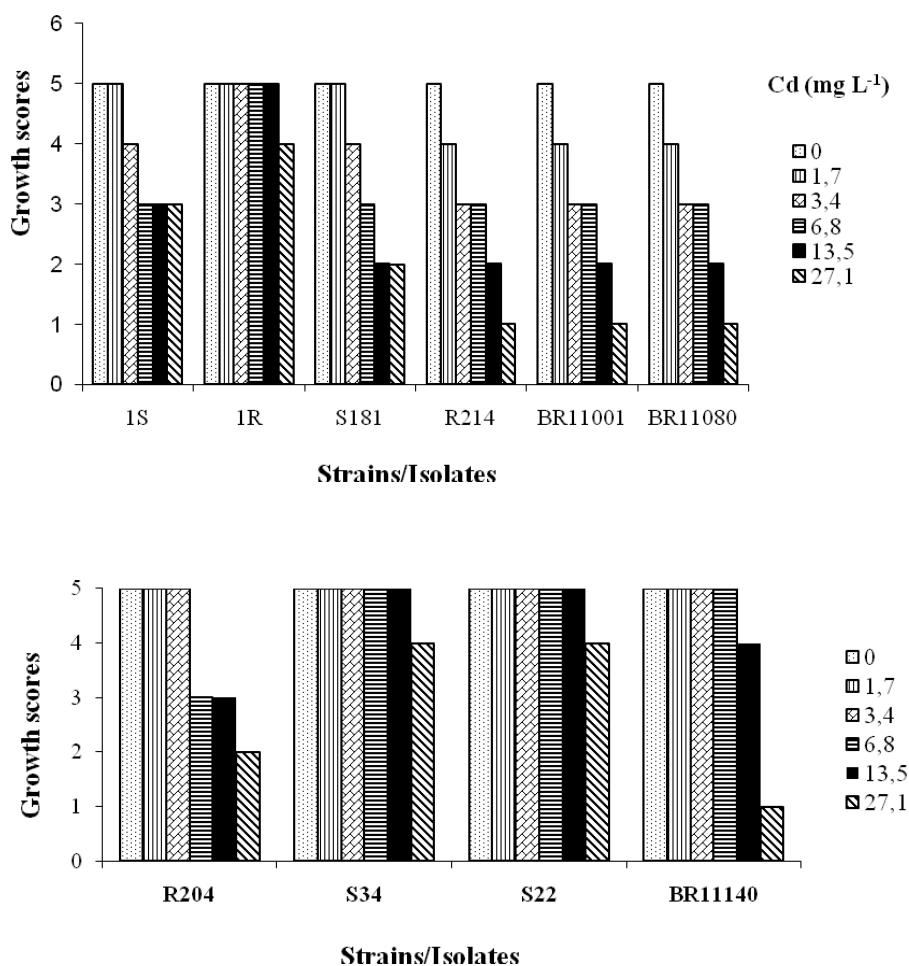
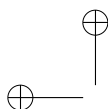


Fig. 2 – Growth scores of associative diazotrophic bacteria isolated from heavy metal contaminated area and type strains in potato dextrose agar medium supplemented with different concentrations of  $\text{Cd}^{2+}$  ( $\text{mg L}^{-1}$ ) ten days after inoculation. a) *Azospirillum lipoferum* (1S and BR 11080<sup>T</sup>) and *A. brasilense* (1R, S181, R214, BR 11001<sup>T</sup>); b) *A. amazonense* (R204, S34, S22, BR11140<sup>T</sup>). 0 = no visual growth, 1 = very reduced growth, 2 = low growth, 3 = intermediate growth, 4 = abundant growth, 5 = very abundant growth as compared to the growth of the strain/isolate in the same medium without heavy metal supplementation.

However, except for isolate UFLA 1R, *A. brasilense*/*A. lipoferum* behaved in a different way to *A. amazonense*, i.e., they tolerated the same or higher  $\text{Zn}^{+2}$  concentrations under  $\text{N}_2$  fixing conditions than with combined Nitrogen (Table III, Figs. 1a and 2a). Strain BR 11001<sup>T</sup> had the lowest MTC under  $\text{N}_2$  fixing conditions (Table III) as compared to media with combined Nitrogen (Figs. 1a and 2a). Nitrogenase activity decreased

was delayed as shown by no acetylene reduction and absence of pellicles after four days at concentrations where growth were observed only after 10 days (Table III). Isolate UFLA 1S was the only one found to tolerate the highest  $\text{Cd}^{+2}$  concentrations. Growth was delayed as indicated by acetylene reduction activity or pellicle presence after 4 days at the highest  $\text{Cd}^{+2}$  concentrations, i.e., the same behavior as in the media with  $\text{Zn}^{+2}$ . Although

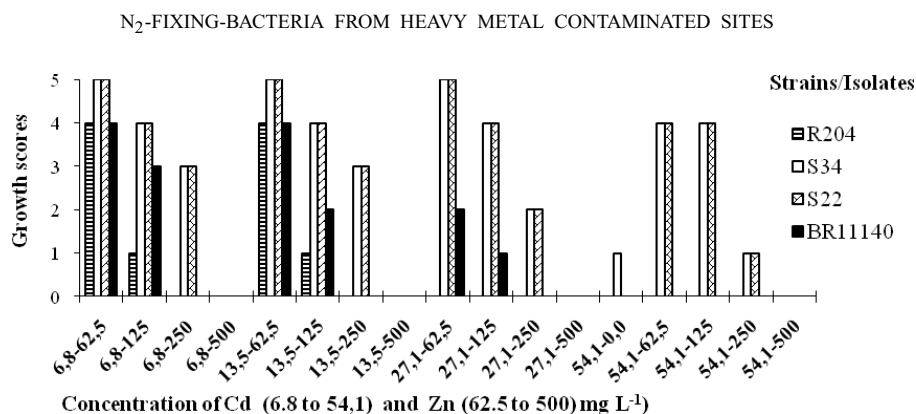
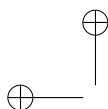


Fig. 3 – Growth scores of *Azospirillum amazonense* type strain (BR11140<sup>T</sup>) and isolates (R204, S34, S22) from heavy metal contaminated sites in potato dextrose agar medium supplemented with different concentrations of Cd<sup>2+</sup> + Zn<sup>2+</sup> (mg L<sup>-1</sup>) ten days after inoculation. 0 = no growth, 1 = very reduced growth, 2 = low growth, 3 = intermediate growth, 4 = abundant growth, 5 = very abundant growth as compared to growth of the strain/isolate in the same medium without heavy metal supplementation.

#### TOLERANCE TO NaCl

No type strain or isolate was able to grow in medium containing 80 g L<sup>-1</sup> NaCl. The type strain BR 11140<sup>T</sup> grew only in media supplemented with 10 g L<sup>-1</sup> NaCl or lower, therefore it was considered the most sensitive to saline stress. BR 11080<sup>T</sup>, BR 11340<sup>T</sup>, BR 9004 (*Burkholderia* sp.), UFLA R214, R204 and S34 tolerated up to 20 g L<sup>-1</sup> of NaCl, and they were classified as facultative halophytes. UFLA 1S, 1R, S181 and S22, BR 11175<sup>T</sup> and BR 11001<sup>T</sup> grew in up to 30 g L<sup>-1</sup> NaCl, and *Azospirillum irakense* (KBC1<sup>T</sup>) grew in medium with 50 g L<sup>-1</sup> NaCl, so they were considered tolerant to saline stress.

#### TOTAL PROTEIN PROFILES

The protein profiles of isolates UFLA 1R and R204 were not revealed in the polyacrylamide gel. Figure 5 shows that no isolate was clustered with any type or reference strain of described species at similarity levels equal or above 75%. Strains belonging to genus *Azospirillum* clustered together with type strains of these species at similarity levels of 43% (UFLA S181, 1S) and 50% (UFLAS22), except for UFLA R214 that clustered with *H. seropedicae* (56% and UFLA S34 (30%), clustered with all the other strains.

#### ANALYSES OF 16S rDNA SEQUENCING

(96%, corresponding to 251 of 259 bases) followed by *A. lipoferum* strain B2 (95%, corresponding to 259 bases) and UFLAS181 sequence was more related to *Azospirillum brasilense* strain MTC (98%, corresponding to 715 of 729 bases). Sequences were submitted to NCBI database and received accession numbers: DQ977745 (UFLA 1S) and DQ977746 (UFLA S181).

#### DISCUSSION

All the three semi-solid media tested were efficient to detect the presence of diazotrophs in the studied soil and roots samples from HM contaminated sites. Diazotrophic bacteria densities were, in general, higher in root than soil samples. Considering that JNFB medium is reported to favor growth of *Herbaspirillum* spp. (Döbereiner et al. 1995), it could be inferred that these organisms occurred in high numbers both in soil and root samples. However, these findings are not in accordance with the fact that *Herbaspirillum* spp. are endophytic organisms and therefore rarely occur in soil (Baldani and Baldani 2005). However, Nobre et al. (2004) pointed out that JNFB medium also detects species other than *Herbaspirillum* spp. such as *Azospirillum* spp., which was probably the case in the present study. In a previous study with soil samples from



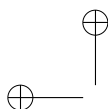
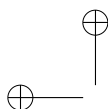


Fig. 4 – Growth scores (GS) of UFLA S34 at different combinations of  $\text{Zn}^{+2}$  and  $\text{Cd}^{+2}$  concentrations supplemented in dextrose agar medium. Figure shows colony color changing from white to yellow as well as growth decreasing with increasing concentrations of heavy metals. All plates received the same numbers of cells as inoculum. 2 = low growth, 3 = intermediate growth, 5 = very abundant growth as compared to the growth of the isolate in the same medium without heavy metal supplementation.

zosphere effect in soil samples might also have had a negative influence on diazotroph densities. The occurrence of diazotrophs in relatively higher densities in roots than in soil samples is probably due to a protective and stimulant microenvironment in roots, as suggested for acidity stress on *Azospirillum* spp. (Döbereiner 1978). Density values found in the present study were similar to the range found in several reports for non-contaminated agricultural soils (Magalhães et al. 1979, Baldani and

The *Azospirillum* spp. isolates from contaminated soil exhibited higher tolerance to  $\text{Zn}^{+2}$  and  $\text{Cd}^{+2}$  than type strains from non-contaminated soils. This finding corroborates results for rhizobia isolates from the same area (Trannin et al. 2001a), when these were compared to isolates from non-contaminated soils. Tolerance of these diazotrophic isolates as compared to rhizobia from other studies (Angle et al. 1993, Matsuda et al. 2002) suggest a better adaptation of associative bacte-



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**TABLE III**  
**Nitrogenase activity (acetylene reduction: nmoles ethylene/culture/hour) measured four days after inoculation by *Azospirillum* spp. cultures and Maximum Tolerated concentration (MTC) for growth measured by pellicle formation ten days after inoculation in media supplemented with increasing concentrations of Zn<sup>+2</sup> (Zn SO<sub>4</sub>.7H<sub>2</sub>O) or with Cd<sup>+2</sup> (Cd SO<sub>4</sub>.8H<sub>2</sub>O).**

Strains/Isolates	Nitrogenase activity Zn <sup>+2</sup> concentration (mg L <sup>-1</sup> )						MTC (mg L <sup>-1</sup> )	
	0	62.5	125	250	500	1000		
UFLA R204	63	25	3	0	0	0	125	
UFLA S34	36	50	6	0	0	0	250	
UFLA S22	83	65	27	0	0	0	250	
BR 11140 <sup>T(1)</sup>	57	70	50	0	0	0	250	
UFLA S181	168	8	23	11	0	0	500	
UFLA R214	2	0	0	0	0	0	500	
UFLA 1R	45	6	11	0	0	0	500	
UFLA 1S	18	11	0	0	0	0	500	
BR 11001 <sup>T(2)</sup>	13	0	0	0	0	0	250	
	Cd <sup>+2</sup> concentration (mg L <sup>-1</sup> )							
	0	2.5	5.0	10.0	20.0	40.0	60.0	
UFLA R204	25	20	4	1	0	0	0	40
UFLA S34	41	49	6	0	0	0	0	40
UFLA S22	41	49	6	0	0	0	0	20
BR 11140 <sup>T</sup>	67	3	1	0	0	0	0	20
UFLA S181	5	3	9	11	12	0	0	20
UFLA R214	66	0	0	0	0	0	0	40
UFLA 1R	8	9	12	10	12	0	0	40
UFLA 1S	8	9	10	9	10	0	0	60
BR 11001 <sup>T</sup>	18	20	32	28	24	0	0	20

Type strains of (1) *A. amazonense* and (2) *A. brasilense*.

800 mg L<sup>-1</sup> Zn<sup>+2</sup>. Angle et al. (1993) found tolerances of rhizobia isolates from temperate regions only up to 2.5 mg L<sup>-1</sup> Cd<sup>+2</sup> and 500 mg L<sup>-1</sup> Zn<sup>+2</sup>. Associative diazotrophic isolates in our study tolerated up to 1.000 mg L<sup>-1</sup> Zn<sup>+2</sup> and 60 mg L<sup>-1</sup> Cd<sup>+2</sup>.

Isolates UFLA S34 and UFLA S22 grew poorly or did not grow when 54.1 mg L<sup>-1</sup> Cd<sup>+2</sup> was tested alone, but grew when this concentration was tested in combination with 62.5 or 125 mg L<sup>-1</sup> Zn<sup>+2</sup>. This may be explained by antagonistic effects between these metals, as was re-

Cd<sup>+2</sup>. It was found that only isolates UFLA S22 and UFLA S34 tolerated the highest concentrations of combined Zn<sup>+2</sup> and Cd<sup>+2</sup>, which indicated that these isolates can be promising for plant inoculation in phytoremediation programs in multi-contaminated areas.

Yellow pigmentation associated with HM contamination occurred in a different way of previous studies. The dark pigment production by *A. brasilense* strain in minimum culture medium with 2 mM Zn<sup>+2</sup> (130.8 mg L<sup>-1</sup>) reported by Gowri and Srivastava

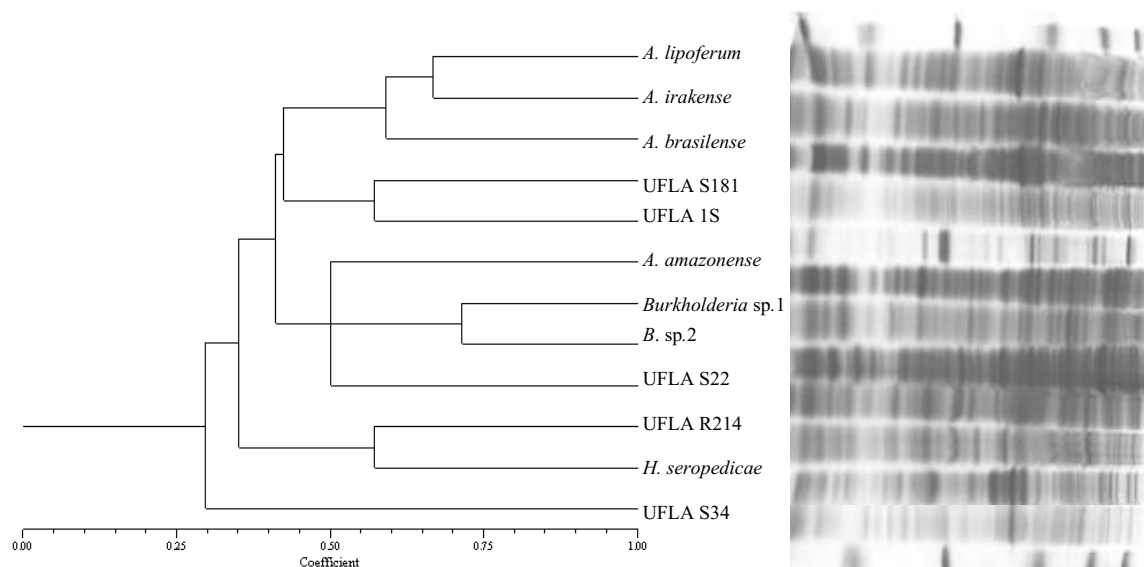
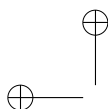


Fig. 5 – Dendrogram and total protein profiles of associative diazotrophic bacteria: MWM (molecular weight marker), *A. lipoferum* (BR11080<sup>T</sup>), *A. irakense* (KCB1<sup>T</sup>), *A. brasilense* (BR11001<sup>T</sup>), UFLA S181, UFLA 1S, *A. amazonense* (BR11040<sup>T</sup>), *Burkholderia* sp.1 (BR 9004), *B. sp.2* (BR11340), UFLA S22, UFLA R214, *H. seropedicae*, (BR11175<sup>T</sup>), UFLA S34 e MWM.

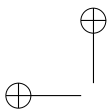
lished results) it was found that pellets from centrifuged cultures of *A. brasilense* type strain and isolates grown in dextrose potato liquid medium, supplemented with 125 and 250 mg Zn<sup>+2</sup> L<sup>-1</sup>, were darker than pellets from cultures grown in medium without metal amendment.

Studies regarding tolerance of associative diazotrophs to heavy metals have been widely neglected. Strzelczyk et al. (1997) found variation of three *Azospirillum* isolates concerning their tolerance to Zn<sup>+2</sup>, Cd<sup>+2</sup> and Cu<sup>+2</sup>. They also found similar tolerance to each one of the three metals at concentrations ranging from 1 to 100 mg L<sup>-1</sup>, and at 500 mg L<sup>-1</sup> growth was completely inhibited. Although we also found variation in tolerance of strains/isolates to Zn<sup>+2</sup> and Cd<sup>+2</sup>, we found growth of some isolates in up to 1.000 mg L<sup>-1</sup> Zn<sup>+2</sup> and 60 mg Cd<sup>+2</sup> L<sup>-1</sup>. In our study, Cd<sup>+2</sup> was more toxic than Zn<sup>+2</sup> to all strains/isolates tested, regardless culture conditions. This is in accordance with other results for rhizobia isolated from both temperate and tropical soils (Angle et al. 1993, Trannin et al. 2001a, Matsuda et al. 2002). Hayat et al. (2002) demonstrated soil populations of

Zn but not for Cd, in our study, as well as by Angle et al. (1993), Trannin et al. (2001a) and Matsuda et al. (2002) when studying rhizobia.

It is well known that plants using Nitrogen from symbiosis with rhizobia are less tolerant to various stress than when they utilize combined Nitrogen (Trannin et al. 2001b). This was the case of all *A. amazonense* strains and isolates, and *A. brasilense* strain 1R, however the other isolates/strains behaved in an opposite way. Further studies are necessary to explore this potential when these organisms are associated with plants.

The large variation observed for NaCl tolerance of isolates and type strains indicated that this analysis allowed discrimination of these bacteria against abiotic stress. Thirteen out of the fourteen isolates/strains grew at concentration higher than 2% NaCl, and could be considered facultative halophytes, a commonly found soil bacterial behavior. This observation also corroborated results obtained by Sanches (2002), who utilized different saline concentrations for characterizations of indigenous rhizobia strains, and those from Nóbrega et



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in general, the most tolerant to HM, showing a relationship between tolerance to HM and to NaCl.

Bacterial diversity is a source of valuable genetic resources for biotechnology and microbiologists have just begun to access the complexity and the potentialities of tropical diversity. The existence of associative diazotrophic bacteria in disturbed areas such as heavy metal contaminated sites, can indicate that even under adverse soil conditions these organisms can survive and establish associations with plants. Thus, tolerance of some associative diazotrophic isolates to heavy metals can be utilized for further studies, aiming rehabilitation of these sites, by improving functionality and sustainability of ecosystems. On the other hand, diazotrophic populations presenting low tolerance, can be used as quality indicators.

Species identification of UFLA 1S and S181 based on 16S rDNA sequencing confirmed previous identification based on cultural characteristics described by Magalhães and Döbereiner (1984) and Döbereiner et al. (1995), i.e, their 16S rDNA sequences revealed they are really closest related to *A. lipoferum* and *A. brasilense*. However, these two isolates are similar to these species at 95% and 98%, respectively. Similarity of 97% is usually considered the boundary for species differentiation (Rosseló-Mora and Amann 2001). However, these authors also reported that different species can have identical or nearly identical 16S rDNA sequences. Furthermore, protein profiles of isolates UFLA S181 and 1S have a dissimilarity equal or more than 75% with other *Azospirillum* type strains, including *A. lipoferum* and *A. brasilense*. Many studies have shown different species of different genera as having protein profiles at levels below 80% (Moreira et al. 1993, De Lajudie et al. 1994) and that, protein profiles similarity highly correlated with DNA homology, i.e, strains with highly similar protein profiles share high DNA hybridization values and therefore belong to the same species (Vandamme et al. 1996). Thus, further studies could indicate if these isolates do belong to these species, or if they are sub-species or new species of this genus.

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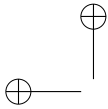
#### RESUMO

Objetivou-se avaliar a densidade de populações de bactérias diazotróficas associativas em amostras de solos e de raízes de gramíneas oriundas de sítios contaminados com metais pesados, e caracterizar isolados destas populações através de análises fenotípica (tolerância aos metais pesados zinco e cádmio, NaCl *in vitro*, perfis protéicos), e genotípica (sequenciamento de 16S rDNA), comparados às estirpes tipo das mesmas espécies. As densidades foram avaliadas nos meios NFB e LGI, comumente utilizados para culturas de enriquecimento de populações de bactérias diazotróficas associativas. As densidades encontradas em amostras de solo e raiz de sítios contaminados foram semelhantes àquelas relatadas na literatura em solos agrícolas. Isolados de *Azospirillum* spp. de solos contaminados e estirpes tipo oriundas de solos não contaminados variaram substancialmente com relação à tolerância a Cd<sup>+2</sup>, sendo que Cd<sup>+2</sup> mais tóxico que Zn<sup>+2</sup>. Dentre os isolados mais tolerantes (UFLA 1S, 1R, S181, S34, e S22) os isolados (1R, S34 e S22) foram mais tolerantes a metais pesados. Os rizóbios isolados de solos de áreas tropicais e temperadas variaram substancialmente com relação à tolerância a Cd<sup>+2</sup>. A maioria dos isolados mais tolerantes a metais pesados também foi tolerante ao estresse salino, o que foi indicado pelo seu crescimento em meio sólido suplementado com 3% de NaCl *in vitro*. Cinco isolados apresentaram alta dissimilidade em perfis protéicos e o sequenciamento do gene 16S rDNA em dois deles revelou que apresentam novas sequências de *Azospirillum*.

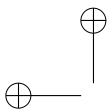
**Palavras-chave:** *Azospirillum*, zinco, cádmio, contaminação do solo, bactérias fixadoras de N<sub>2</sub>.

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