



Anais da Academia Brasileira de Ciências

ISSN: 0001-3765

[aabc@abc.org.br](mailto:aabc@abc.org.br)

Academia Brasileira de Ciências

Brasil

RODRIGUES, ELAINE G.; DOBROFF, ANDREY S.; TABORDA, CARLOS P.; TRAVASSOS, LUIZ R.

Antifungal and antitumor models of bioactive protective peptides

Anais da Academia Brasileira de Ciências, vol. 81, núm. 3, septiembre, 2009, pp. 503-520

Academia Brasileira de Ciências

Rio de Janeiro, Brasil

Available in: <http://www.redalyc.org/articulo.oa?id=32713479015>

- How to cite
- Complete issue
- More information about this article
- Journal's homepage in [redalyc.org](http://redalyc.org)

[redalyc.org](http://redalyc.org)

Scientific Information System

Network of Scientific Journals from Latin America, the Caribbean, Spain and Portugal

Non-profit academic project, developed under the open access initiative



Anais da Academia Brasileira de Ciências (2009) 81(3): 503-520  
(Annals of the Brazilian Academy of Sciences)  
ISSN 0001-3765  
www.scielo.br/aabc

## Antifungal and antitumor models of bioactive protective peptides

ELAINE G. RODRIGUES<sup>1</sup>, ANDREY S. DOBROFF<sup>1</sup>, CARLOS P. TABORDA<sup>2</sup> and LUIZ R. TRAVASSOS<sup>1</sup>

<sup>1</sup>Unidade de Oncologia Experimental, Departamento de Microbiologia, Imunologia e Parasitologia, Universidade Federal de São Paulo  
Rua Botucatu, 862, 8º andar, 04023-062 São Paulo, SP, Brasil

<sup>2</sup>Departamento de Microbiologia, Instituto de Ciências Biomédicas, Universidade de São Paulo  
Av. Prof. Lineu Prestes 1374, Cidade Universitária, 05508-900 São Paulo, SP, Brasil

Manuscript received on June 27, 2008; accepted for publication on March 31, 2009;  
contributed by LUIZ R. TRAVASSOS\*

### ABSTRACT

Peptides are remarkably reactive molecules produced by a great variety of species and able to display a number of functions in uni- and multicellular organisms as mediators, agonists and regulating substances. Some of them exert cytotoxic effects on cells other than those that produced them, and may have a role in controlling subpopulation and protecting certain species or cell types. Presently, we focus on antifungal and antitumor peptides and discuss a few models in which specific sequences and structures exerted direct inhibitory effects or stimulated a protective immune response. The killer peptide, deduced from an antiidiotypic antibody, with several antimicrobial activities and other Ig-derived peptides with cytotoxic activities including antitumor effects, are models studied *in vitro* and *in vivo*. Peptide 10 from gp43 of *P. brasiliensis* (P10) and the vaccine perspective against paracoccidioidomycosis is another topic illustrating the protective effect *in vivo* against a pathogenic fungus. The cationic antimicrobial peptides with antitumor activities are mostly reviewed here. Local treatment of murine melanoma by the peptide gomesin is another model studied at the Experimental Oncology Unit of UNIFESP.

**Key words:** bioactive peptides, *Paracoccidioides brasiliensis*, tumor cells, killer peptide, melanoma, apoptosis.

### INTRODUCTION

Bioactive peptides arise from proteins by the action of peptidases or are chemically synthesized based on certain templates of natural sequences that have been selected by a variety of screening methods. Peptides can be designed aiming at enhanced functional activity by using amino acid substitutions and chemical modification. Owing to their great diversity of binding properties, peptides can play roles of biochemical reagents, pharmacological drugs, hormones, antibiotics, vaccines and mediators of neural and immunological signaling. Peptides interact with membrane structures, are specifically

recognized by cell surface receptors or act as ligands interacting with intracellular compounds and subcellular structures. Peptides can include epitopes recognized by antibodies and TCRs, and those called protective peptides elicit a protective immune response. On the other hand, the actual fungal and tumor models, peptides that exert direct cytotoxicity on target cells or elicit a protective immune response in animals experimentally infected and challenged with tumor cells have been investigated.

### ANTIFUNGAL PEPTIDES

During the past decades, an increase in the incidence of fungal diseases has been recognized mainly caused by



the use of antimycotics is the only option for the treatment of fungal infections. Currently used antimycotics, however, frequently have a limited activity spectrum, are available only in intravenous formulations, favor resistance development, and cause serious side-effects (reviewed in François et al. 2005). Thus, the search for new antifungal therapies is strongly stimulated, and the use of antifungal peptides is a promising alternative.

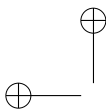
Antifungal properties of peptides have been reviewed by De Lucca and Walsh (2000). There are 405 peptides with antifungal properties described, comprising linear or cyclic, hydrophobic or amphipathic structures (<http://aps.unmc.edu/AP/main.php>).

Their cytotoxicity may involve binding to and disruption of the membrane (Shai 1995), membrane penetration and interaction with the mitochondria (Helmerhorst et al. 1999) or pore formation (Bechinger 1997). Antifungal peptides have been studied in bacteria, fungi, plants, insects, amphibians and mammals. Relevant examples are given next.

Syngomycins, syngostatins and syngotoxins from *Pseudomonas syringae* are lipodepsipeptides highly lethal to *Candida albicans*, *Aspergillus* and *Fusarium* species (De Lucca and Walsh 2000, De Lucca et al. 1999, Sorensen et al. 1996). Glycopeptide cepacidines from *Burkholderia cepacia* are active against *Candida sp.*, *Aspergillus niger*, *Fusarium oxysporum* and *Cryptococcus neoformans* (Lee et al. 1994, Lim et al. 1994). Antifungal peptidylnucleoside nikkomycins are produced by *Streptomyces tendae*, act by inhibiting chitin biosynthesis and were effective in murine infections by *Coccidioides immitis* and *Blastomyces dermatitidis* (Hector et al. 1990). Zeamatin, the 22 kilodalton (kDa) peptide produced by *Zea mays*, permeabilizes the fungal membrane and kills *C. albicans* with a minimal inhibitory concentration (MIC) of 0.5  $\mu$ g/ml (Roberts and Selitrennikoff 1991). Cecropins from the silk moth *Hyalopora cecropia* are linear, lytic peptides effective against germinating conidia of *F. oxysporum* and *A. fumigatus* (De Lucca et al. 1998). Both the L- and D-isomeric forms of cecropin B were fungicidal (De Lucca et al. 2000). Drosomycin is a 44 amino acids (aa)

do not coevolve with pathogens (Jiggins and Kim 2005). In contrast, antimicrobial peptides (AMPs) appear to undergo a rapid adaptive evolution in vertebrates. In frogs, each species produces 10-20 AMPs that differ in size, sequence and specificity, and this rapid diversification is driven by evolutionary selection (Duda et al. 2002). Dermaseptins, produced by *Phyllomedusa sauvagii*, a South American frog, are lysine-rich linear peptides fungicidal for *A. flavus*, *A. fumigatus* and *F. oxysporum* (Mor et al. 1994). Magainins are antifungal peptides produced by the African frog *Xenopus laevis* (De Lucca and Walsh 2000). They are not hemolytic and inhibit *Candida albicans* (Zasloff 1987). Plant [DmAMP1 from dahlia (*Dahlia merckii*), RsAFP2 from radish (*Raphanus sativus*), HsAFP1 from coral bells (*Heuchera sanguinea*), Psd1 from pea (*Pisum sativum*), MsDef1 from alfalfa (*Medicago sativa*) and MtDef2 from barrel medic (*Medicago truncatula*)], insects (Termicin from the termite *Pseudacanthotermes spiniger*, Drosomycin from the fruitfly *Drosophila melanogaster*, Heliomicin from the tobacco budworm *Heliothis virescens*) and human [ $\beta$ -defensin 1 (HBD1),  $\beta$ -defensin 2 (HBD2),  $\beta$ -defensin 3 (HBD3)] defensins showed antifungal properties (reviewed in Aerts et al. 2008). Although there are no clear similarities in the mode of action of these defensins, the presence of sphingolipid glucosyl ceramide (GlcCer) in fungal membranes seems to play a central role in the action of some defensins (Thevissen et al. 2004). Only Psd1 was internalized in the fungal cell, affecting the normal progression of the cell cycle (Lobo et al. 2007), and it is possible that the other defensins stay outside the cell inducing fungal cell death after interaction with their target (e.g. sphingolipids) and modulation of intracellular signaling cascades (Aerts et al. 2008). RsAFP2 was also effective in an *in vivo* prophylactic model of murine candidiasis (Tavares et al. 2008).

$\beta$ -Defensins include porcine cationic, cysteine-rich protegrins which inhibited *C. albicans* (Cho et al. 1998). Gomesin, a cationic AMP isolated from the hemocytes of the unchallenged Brazilian spider *Acanthoscurria gomesiana* (Silva et al. 2000), is structurally related to protegrins and exerts microbicidal activity against fila-



#### ANTI-FUNGAL AND ANTITUMOR PEPTIDES

the presence of the peptide, induced a decrease in capsule expression, rendering cells more susceptible to brain phagocytes and, in association with fluconazole, in concentrations with low antimicrobial activity ( $0.1\text{--}1\mu\text{M}$ ), inhibited fungal growth and enhanced the antimicrobial activity of brain phagocytes (Barbosa et al. 2007). One of the models described in the present review is that of gomesin cytotoxicity in murine and human tumor cells (Rodrigues et al. 2008).

Among the antifungal peptides produced by fungi, the echinocandins interfere with the cell wall biosynthesis (Denning 1997) and the pneumocandins, aculeacins, WF11899, and mulundocandins have a modified echinocandin B peptide core (Debono and Gordee 1994, Kurtz and Douglas 1997). Echinocandins are produced by *Aspergillus nidulans* and *A. rugulosus* and are effective against *Candida* (MIC =  $0.6\mu\text{g/ml}$  for echinocandin B and *C. albicans*) (reviewed in De Lucca and Walsh 2000). Clinical trials have started with molecules of the echinocandin group, VER-002, FK463 and caspofungin (MK-0991) modified for increased solubility and active against *Candida* spp. and *Aspergillus* spp. V-echinocandin and FK463 were effective in the treatment of esophageal candidiasis, the latter in AIDs patients (reviewed in De Lucca and Walsh 2000). Clinical trials with caspofungin (derived from pneumocandin), a drug that inhibits  $\beta$ -1,3 D-glucan synthase, have shown excellent results in the treatment of *Candida* infections and invasive aspergillosis refractory to other antifungal agents (i.e., conventional or lipid formulations of amphotericin B and/or itraconazole). Aureobasidins are produced by *Aureobasidium pullulans*, interfere with sphingolipid synthesis and are effective against murine candidiasis (Nageic et al. 1997, Takesako et al. 1993).

#### KILLER TOXINS AND KILLER PEPTIDES

Killer yeasts secrete killer toxins that target susceptible cells in a two-step receptor-mediated manner. They bind to cell wall receptors and translocate to the plasma membrane. They can then interact with secondary receptors or enter susceptible cells to exert a cytotoxic effect (Maoliani et al. 1997, Schmitt and Breinig

susceptible cells by various mechanisms, including induction of cation-selective ion channels in the membrane, interference in the cell cycle (G1, C arrest), chromosomal DNA synthesis and antitopoisomerase (Schmitt and Breinig 2006, Santos and Maoliani 2004, Jablonowski and Schaffrath 2007, Klasse 2004). Killer toxins can induce apoptosis mediated by yeast caspase Yca1p, characterized by DNA fragmentation, and phosphatidylserine external membrane exposure. This could be a general cell death mechanism under natural environmental conditions (Paluszynski 2007, Schmitt and Reiter 2008).

The direct use of killer toxins in antifungal therapy was discouraged owing to some of their properties. They are generally heat-labile, protease-sensitive and act within a narrow pH and temperature range. They are antigenic and toxic, as shown for *Pichia anomala* killer toxin (Pettoello-Mantovani et al. 1995). To overcome these pitfalls of a potential therapeutic agent, immunological derivatives were generated on the basis of the idiotype network that mimicked the toxic effect of *P. anomala* killer toxin (Polonelli et al. 1991). These antibodies with the internal image of the active killer toxin, which acted as antibiotics, were then maintained. They exerted significant therapeutic effect in experimental models of candidiasis, aspergillosis and pneumocystosis.

Toxic effects were also obtained with single-chain variable fragment (scFv) preparations and they were further examined by synthesizing overlapping decapetides which correspond to the light chain of antibodies and heavy chain of antibodies ( $V_H$ ) regions. These regions include the complementary determining regions (CDRs) that were tested *in vitro* against *C. albicans*. Several peptides were active and one of them, corresponding to the framework sequence with the final amino acids belonging to  $V_L$  CDR1, was selected. It was very cytotoxic and the substitution of the terminal glutamic acid by alanine generated a peptide with the AKVTMTCSAS sequence that was several times more active and was called killer peptide (KP). This peptide interacted with  $\beta$ -glucan and this binding was im-



dition to *C. albicans*, and was effective even in normal and immunocompromised animals against vaginal and systemic candidiasis (Polonelli et al. 2003), disseminated cryptococcosis (Cenci et al. 2004) and paracoccidioidomycosis (Travassos et al. 2004a). The KP is very stable forming dimers in non-reducing conditions without loss of activity (Magliani et al. 2004a, b).

The remarkable cytotoxicity of KP was also examined by electron microscopy. *C. albicans* cells treated with KP showed important internal alterations, including cell wall swelling with middle electron-dense region, collapse of the plasma membrane, condensation and fragmentation of nuclear material, and alteration of mitochondria structure (Fig. 1A). In a dividing cell with a big vacuole and chromatin condensation and fragmentation, cellular alterations were seen beyond the septum separating both cells, with the daughter cell already affected by the KP showing an altered cell wall (Fig. 1B).

#### A MODEL OF DIRECT ANTIFUNGAL EFFECT OF A PEPTIDE

Glucans, chitin and mannoproteins, in addition to plasma membrane sterols, are natural targets of antifungal drugs. Additional targets are ceramide monohexosides, ubiquitously present on the fungal cell wall and displaying several roles in fungal cells (Nimrichter et al. 2008). In *C. neoformans* (Rodrigues et al. 2000), *C. albicans* and *Pseudallescheria boydii*, these glycolipids were identified as targets of human antibodies that inhibited fungal growth. Other targets are melanin, adhesion factors, and cell wall enzymes. The killer decapeptide (KP) described above was synthesized and engineered demonstrating a strong candidacidal activity *in vitro* and curing rat vaginal infections caused by fluconazole-susceptible and -resistant *C. albicans* strains (Polonelli et al. 2003). The fungicidal activity of KP *in vitro* against *P. brasiliensis* and its therapeutic activity *in vivo* have been reported (Travassos et al. 2004a).

Paracoccidioidomycosis (PCM) is the prevalent systemic mycosis in South America with most reported cases in Brazil. It is a major cause of disability and death among young adult rural workers. Sequels are fre-

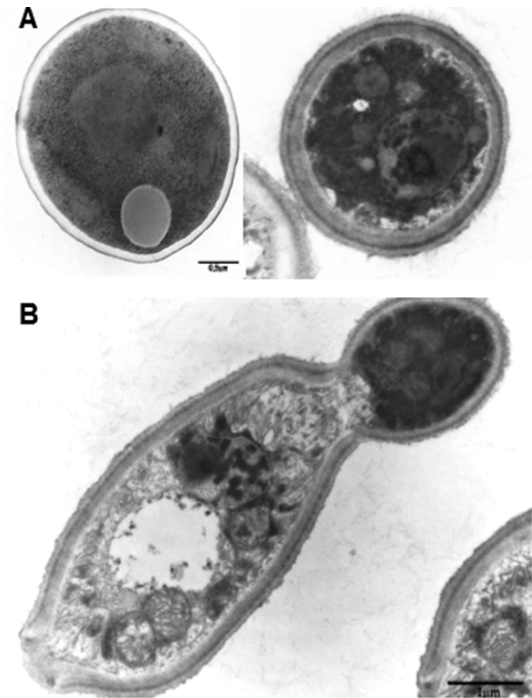


Fig. 1 – Electron micrographs showing the cytotoxic effects of the killer peptide (KP) on *Candida albicans*. (A) Normal untreated or treated with the inactive scrambled peptide *C. albicans* yeast cell (left) as compared with the KP-treated yeast cell (right). Major alterations can be seen as the swelling of the cell wall, plasma membrane collapse, chromatin condensation and nuclear fragmentation. (B) An elongated *C. albicans* cell with a budding cell, both affected by KP treatment. The same alterations as in (A) are seen with nuclear fragmentation and cytoplasmic blebs invading the daughter cells beyond the septum.

apy with itraconazole, amphotericin or sulfamethoxazole/trimethoprim are used in clinical practice, relapses are a significant unsolved problem (Travassos et al. 2008b). Vaccination against PCM is now a prospective goal after P10, and four other peptides derived from the major diagnostic antigen gp43 were found to be promiscuously presented by several human leukocyte antigens DR, MHC class II molecules (HLA-DR) (Iwai et al. 2003). Such a vaccine could function as an adjuvant to chemotherapy significantly reducing the time of treatment (Travassos et al. 2008a, b).

Wide-spectrum antimicrobial peptides, such as KP



#### ANTI-FUNGAL AND ANTITUMOR PEPTIDES

tion in cases of anergy and drug resistance. Multiply-budding yeast cells of *P. brasiliensis* had their viability hampered at 39 ng of KP/yeast in distilled water. The D-isomeric form of KP was also active. Further, the decapeptide was therapeutic in B10A mice infected intravenously with  $3 \times 10^6$  cells of *P. brasiliensis* Pb18 isolate administered intraperitoneally at 3.3  $\mu\text{g/g}$  of body weight, 1 h after infection and 1 and 2 days later. With this protocol, no colony forming units (CFUs) were obtained from lung, spleen and liver after 8 days of fungal challenge in the KP treated animals. In these animals compared to those injected with the scrambled peptide, the liver granulomas were smaller and fewer with no visible fungi. The lungs were less infiltrated with extensive areas of normal alveoli and no visible fungi. Spleens also were little affected, with no detectable fungi.

It was clear therefore that KP was an effective inhibitor of *P. brasiliensis* *in vitro* and *in vivo* (Travassos et al. 2004a).

It is still not clear whether  $\alpha$ -1,3 glucan, the predominant polysaccharide of yeast forms of *P. brasiliensis*, is a target of KP. There is, however, evidence that yeast forms may have  $\beta$ -glucans at the cell surface. Macrophages from pentraxin 3 transgenic (PTX3 Tg) mice showed improved opsonin-independent phagocytosis of zymosan particles and yeast forms of *P. brasiliensis*. In the case of *P. brasiliensis*, an enhanced microbicidal activity accompanied by high production of nitric oxide was observed in macrophages from transgenic mice. Blockade of dectin-1 receptor for  $\beta$ -1,3 glucan inhibited the phagocytosis of zymosan particles by PTX3 Tg macrophages, pointing out the relevant role of dectin-1 as the main receptor involved in zymosan and possibly also of *P. brasiliensis* uptake (Diniz et al. 2004).

#### BIOACTIVE PEPTIDES EXPRESSED AS IMMUNOGLOBULIN ISOLATED CDRs

The discovery by Polonelli et al. (Polonelli et al. 2003, Magliani et al. 2004a, b) that internal sequences of immunoglobulin variable regions may display antibiotic properties prompted us to investigate the activity of monoclonal antibody (mAb) CDRs tested as synthetic pen-

to the hypervariable domains called complement determining regions (CDRs). There are 6 CDRs in the variable regions of light ( $V_L$ ) and heavy chain ( $V_H$ ) with background variability on each side of the framework. The CDRs are named H1, H2, H3 and L1, L2, L3 in heavy and light chains, respectively. The framework sequences between CDRs can be similar or identical. Although all CDRs are expected to contribute to antigen binding with variable affinity, only the CDR3 of  $V_H$  when tested as an isolated linear or cyclic peptide was found to have the same specificity of the corresponding antibody, sharing some of its biological properties. CDR3 (H3) peptides with such properties have been called micro (mini) antibodies (Levi et al. 1993, Polonelli et al. 1998). They can even compete with a full antibody for binding to a certain antigen. The other CDRs generally do not show a similar reactivity when tested as isolated peptides.

Recently we showed, in collaboration with Polonelli's and Ponton's groups from Parma and Billingham, respectively, that, independently of the specificity of the native Ab, CDRs other than H3 may display, with high frequency, antimicrobial, antiviral and antitumor activities in a way reminiscent of molecules of early humoral immunity (Litman et al. 2005). The following peptides were studied as sources of the CDRs: Ab (mAb C7) raised against a *C. albicans* antigen; mouse mAb C7, sharing H1 and H2 with mAb C7; and human mAb C7, sharing no CDR either with mAb C7 or mAb C7, with specificity for difucosylated blood group antigens. mAbs generated CDRs that, represented by synthetic peptides, showed *in vitro*, *ex vivo* and/or *in vivo* direct antimicrobial (*C. albicans*), antiviral (HIV-1) and antitumor activities (Polonelli et al. 2008).

CDRs C7/pc42 H2 and HuA L1 were directly cytotoxic for melanoma and HL-60 (human leukemia) cells, causing caspase-dependent apoptosis. H2 peptide activity was receptor-mediated in melanoma cells. C7 H2 and HuA L1 peptides in the C-terminal amino acid form were active against lung colonization by melanoma cells by intravenous injection (i.v.). Peptides were also administered by intraperitoneal injection (i.p.). (2



ules in the lungs of peptide treated animals were very few. Presumably, even better results could have been obtained by optimization of the peptide administration protocol (Polonelli et al. 2008). C7 H3 but not C7/pc42 H2 competed with mAb C7 for binding to phosphatidylcholine, the probable ligand of polyreactive C7 (IgM) on melanoma cells. This CDR (C7 H3) together with the H3 CDRs of two anti-melanoma mAbs (A4 and A4M), that competed with the antibodies for binding to melanoma cells, were three examples of micro (mini) antibodies shown in our laboratory (unpublished results).

#### A PEPTIDE VACCINE AGAINST PARACOCIDIOIDOMYCOSIS

The main diagnostic antigen of *P. brasiliensis* was identified in our laboratory in 1986 (Puccia et al. 1986; reviewed Travassos et al. 2004b). Glycoprotein gp43 reacts with 100% sera of patients with paracoccidioidomycosis from a vast region of South America, with the possible exception of sera from certain Western areas. It elicits an immune response that protects against the intratracheal challenge by virulent *P. brasiliensis* yeast cells. This molecule has been cloned and sequenced (Cisalpino et al. 1996). Apart from B cell epitopes, which are beginning to be identified, the gp43 carries an immunodominant epitope that elicits a predominant IFN- $\gamma$ -mediated Th-1 response. It is responsible for delayed type sensitive (DTH) reactions in infected animals (Rodrigues and Travassos 1994). The T-CD4<sup>+</sup> cell epitope was mapped to a peptide called P10 with the QTLIAHTLAIRYAN sequence, the HTLAIR hexapeptide core being essential for priming the immune response (Taborda et al. 1998). P10 was as protective as the gp43 in intratracheal injection (i.t.) challenged mice, being administered i.p with complete Freund's adjuvant (CFA). The nucleotide sequence encoding P10 was conserved in a number of isolates (Travassos et al. 2004b).

The T cell epitope in peptide P10 is presented by major histocompatibility complex (MHC) class II molecules from three different mouse haplotypes (Taborda et al. 1998). Promiscuity of P10 was also observed with

Caucasian HLA-DR molecules (Iwai et al. 2003). Additional gp43 peptides were also identified using the TEPITOPE algorithm, which bound promiscuously to several HLA-DR molecules. As pointed out before (Travassos et al. 2008a, b) this is an essential property of a vaccine peptide candidate considering the genetic diversity of the target immunizable population.

In 29 patients with PCM and submitted to chemotherapy, 79% of them recognized one peptide selected by the TEPITOPE algorithm. By pooling peptides gp43<sup>45-59</sup>, gp43<sup>106-120</sup>, gp43<sup>181-195</sup> or P10, and gp43<sup>283-298</sup>, the recognition frequency increased to 86% (Iwai et al. 2007). Overall for 25 Caucasian HLA-DRs, P10 and neighboring peptides were predicted to bind (TEPITOPE) to 90% or more of these molecules. Very few healthy individuals had peripheral blood mononuclear cells (PBMC) proliferating with gp43 and even fewer with gp43 derived peptides. They may have been exposed to *P. brasiliensis* on a trip to reserve areas of the fungus or cross-reacted with related fungal antigens, possibly also exo- $\beta$ -1,3-D-glucanases. Site homologous but unidentical sequences, in comparison with P10, were found in  $\beta$ -1,3-glucanases from *Aspergillus nidulans*, *Histoplasma capsulatum*, *Blastomyces dermatitidis* and *Lacazia loboi* (a gp43-like protein).

The rationale for a peptide vaccine based on P10 has been discussed recently (Travassos et al. 2008a). Basically: “Stimulation of an effective IFN- $\gamma$ -producing T-helper response can simultaneously trigger the production of potentially protective antibodies and the activation of CD8<sup>+</sup> T cells in addition to activation of phagocytic cells. In the presence of several immunogenic molecules of the fungal agent, stimulation of one arm of the immune system may alter a state of early or installed immunosuppression”. Since treatment of fungal infections and particularly of PCM involves chemotherapeutic drugs, a peptide vaccine could work as an adjuvant to reduce the treatment period, which is usually long, avoid relapses and reverse the potentially lethal anergic cases. It also could help to treat those cases of fungal drug resistance.

To tackle the above issues while using experimental



#### ANTI-FUNGAL AND ANTITUMOR PEPTIDES

with P10 and/or a chemotherapeutic drug starting after 48h of infection. In the second protocol, P10 and/or drug treatment was started after 30 days of infection. It aimed at reproducing a condition of established infection as in patients with PCM. The treatment was held for 30 days, during which groups of mice received i.p. doses of itraconazole, fluconazole, ketoconazole, sulfamethoxazole or trimethoprim-sulfamethoxazole at every 24 h. Amphotericin B was given at every 48 h. P10 was administered weekly for 4 weeks, initially in CFA and three times in incomplete Freund's adjuvant (Marques et al. 2006).

In all cases, there was an additive protective effect with the combination of P10 immunization and chemotherapy. Animals treated with sulfamethoxazole showed early protection followed by relapse. Significantly, the association of sulfamethoxazole and P10 successfully controlled the infection. In the second protocol, the fungal burden was examined after 60 and 120 days of infection. An additive protective effect of P10 immunization and drug treatment was also observed, with 60 to 80% reduction in lung CFUs. Chemotherapy alone induced a predominant Th-2 response with increased production of IL-4 and IL-10 detected in lung homogenates, whereas P10 vaccination stimulated a Th1 response, rich in IFN- $\gamma$  and IL-12 without suppressing the Th-2 response (Marques et al. 2006). These are encouraging results in short term experiments. It is probable that an increased protective effect will be obtained in long term trials in which the animals will have time to completely recover of the fungal infection.

The condition of anergy was addressed as follows. Balb/c mice were treated with dexamethasone-21 phosphate added to drinking water. Negative DTH with *P. brasiliensis* antigen was obtained after 30 days. Immunosuppressed mice (n=10), infected with virulent *P. brasiliensis*, began to die 10 days after infection, and all animals were dead after 70 days. Chemotherapy and/or P10 immunization of immunosuppressed animals was started 15 days after i.t. infection and all treated animals survived thereafter. Chemotherapy and P10 immunization conferred additive protection. A significant increase

results suggest that P10 immunization can be protective in anergic patients.

Delivery of peptides for an efficient immunization has always been a concern of our group because previous experiments have always used CFA as an adjuvant. Following alternatives therefore have been investigated.

Early studies have shown that immunization of Balb/c mice with a mammalian expression vector (VR-gp43) carrying the full gene of gp43 with Cytomegalovirus (CMV) promoter induced B and T cell-mediated immune responses which were protective against challenge by virulent *P. brasiliensis* yeast forms (Marques et al. 2000). The cellular immune response in mice immunized with VR-gp43 was kept for at least 6 months after immunization. A similar construction with a P10 minigene in plasmid DNA alone or associated with a plasmid carrying mIL-12 insert was tested in Balb/c mice i.t. infected with a virulent isolate (Pb18) of *P. brasiliensis*. A significant reduction of fungal burden in lung, spleen and liver was obtained with production of IL-12 and IFN- $\gamma$  and reduction of IL-4 levels in homogenates (G. Rittner et al., unpublished results).

The construction of MAP (multiple antigenic peptide) was also tried to deliver a tetravalent antigen containing P10 sequence. MAP-10, or M10, had four LIAHTLAIRYAN (QT-less P10) chains synthesized on a branched lysine core. Lymph node cell proliferation from P10 or M10-sensitized mice was identical to *in vitro* stimulation with either P10 or M10. Immunization with single dose of M10 without adjuvant was protective with few lung, spleen and liver CFUs and no yeasts in lung histopathological sections (Taborda et al. 2006).

In Balb/c mice infected i.t. for 30 days, the protective effect of P10 was tested alone or mixed with adjuvants: alum, monophosphoryl lipid A or complete Freund's adjuvant (Travassos et al. 2008a, b). Unexpectedly, P10 administered in phosphate-buffered saline was most effective with a significant reduction in lung burden with no fungi detected in spleens and livers.

The protective effect of P10 has also been





therapy, both gp43 and gp70 are markers for monitoring successive treatment and cure through their decreased antigenemia and specific antibody response (Marques da Silva et al. 2004, Silva et al. 2004). In the experimental Balb/c model of PCM infection, anti-gp43 mAb 3E effectively reduced the fungal burden and promoted phagocytosis *in vitro* (Buisa-Filho et al. 2008). The recognized epitope in the gp43 was mapped to the sequence NHVRIPIGYWAV shared with *Aspergillus fumigatus*, *A. oryzae* and *B. graminis* internal sequences of  $\beta$ -1,3-glucanases. This peptide could increase the protective effect of P10 in a possible peptide vaccine against PCM.

Again, as stressed, we quote our own thought expressed before (Travassos et al. 2008a): “Short term protocols (30 to 45 days) have the advantage of allowing repeated experiments to define a certain response. However, longer periods of treatment and observation may lead to even more effective results, aiming at sterilization in experimental models with massive infection loads”.

#### ANTITUMOR PEPTIDES

Cancer remains as a major source of mortality and morbidity around the world, despite numerous recent advances in treatment alternatives. Chemotherapy and, more recently, biochemotherapy, is still the choice treatment for advanced and metastatic disease (Espinosa et al. 2003). It is, though, often associated with deleterious side effects caused by drug-induced damage to healthy cells and tissues (Buzaid and Atkins 2001). Quiescent or slowly proliferating cancer cells are refractory to the cytotoxic effect of drugs interfering with DNA synthesis (Naumov et al. 2003) and, frequently, cellular changes affected sensitivity to chemotherapeutic drugs by increased expression of drug-detoxifying enzymes and/or drug transporters, altered interactions between the drug and its target, increased ability to repair DNA damage and defects in the apoptotic pathway (Gatti and Zunino 2005). Development of a new class of anticancer drugs that lack toxicity to healthy cells and are

tides (CAPs), are promising candidates for antitumor treatment.

CAPs have been found in all species that have been tested so far, including bacteria, fungi, plants and animals, and they probably represent one of the first evolved forms of defense of eukaryotic cells against pathogens (Zasloff 2002). An updated list of CAPs can be found in <http://aps.unmc.edu/AP/main.php>, with 1,393 entries. Most CAPs have a broad spectrum of antimicrobial activities; only 82 of the listed CAPs were active, however, against tumor cells.

Despite their diverse origins, antimicrobial peptides have common biophysical parameters, including small size, positive charge, and amphipathicity, that are likely important for peptide activity. These molecules are grouped according to structural characteristics, and are usually separated in three classes: (1) linear, often forming alpha-helical structures; (2) cysteine stabilized, beta-sheet structures; and (3) peptides with one or more predominant amino acid residues, but variable in structure (Yount et al. 2006). As stated before, not all CAPs are able to kill cancer cells, and to date, it has not been possible to predict an antitumor activity based on the peptide structure.

The short length and cationic/amphipathic properties of these molecules enable CAPs to interact and disrupt lipid membranes. Positively charged amino acid residues, such as lysine and arginine, and hydrophobic residues are frequently found in large numbers in CAPs (Hoskin and Ramamoorthy 2008). The high expression of anionic molecules, such as phosphatidylserine in the outer membrane leaflet of human tumor cells (Utsugi et al. 1991, Dobrzynska et al. 2005), as well as O-glycosylated mucins (Yoon et al. 1996) on cancer cell membranes, account for the net negative charge of these cells and their electrostatic interactions with cationic CAPs. In the case of magainin peptides, the cytotoxic activity for tumor cells was abolished by eliminating the electrical gradient across the plasma membrane. Apparently, the cellular potential is critical for peptide channel formation in tumor cell membranes and could determine the selective killing of tumor cells by



#### ANTI-FUNGAL AND ANTITUMOR PEPTIDES

major membrane components, such as sphingomyelin, phosphatidylethanolamine and phosphatidylcholine (Zachowski 1993).

CAPs interaction with cancer cell membranes is not mediated by receptors, since D-amino acid peptide analogues displayed an activity similar to the all-L-amino acid peptide (Rodrigues et al. 2008, Hetru et al. 2000).

Another mechanism for cancer cell killing by CAPs is the induction of apoptosis by permeation of mitochondrial membrane after internalization, release of cytochrome c, leading to caspase 9 and 3 activation (Pardo et al. 2001). Both cationic and hydrophobic amino acids play a role in the peptide permeation of mitochondrial membranes (Horton et al. 2008). Alternatively, apoptosis may be induced by CAPs interaction with cell death receptors, such as Fas ligand, leading to caspase 8 activation. Interestingly, arginine, glycine and asparagine, integrin homing domain (RGD)-conjugated tachyplesin induced both pathways, suggesting that some CAPs may have more than one effect on cancer cells (Chen et al. 2001).

Protein glycosylation may alter the secondary structure of a membrane-associated protein or peptide, and altered glycosylation of membrane proteins is frequently found in malignant cells. Moreover, differential branching and sialic acid content of N-linked glycans are associated with an increase in the net negative charge in the membrane of many cancer cells. Interestingly, peptide-glycosylation was associated with increased potency of drosocin *in vitro* (McManus et al. 1999). It is therefore likely that glycosylation of CAPs and/or cancer cell membrane proteins may influence the binding affinity of some CAPs for the cancer cell.

CAPs may be used in combination with conventional chemotherapeutic antitumor drugs in order to reduce effective doses, and thereby reduce harmful side-effects frequently observed in treated patients. Cecropin A, in combination with 5-fluorouracil and cytarabine, showed a synergistic cytotoxic effect on human leukemia cells (Hui et al. 2002).

Representative naturally occurring CAPs with antitumor activities are depicted on Table I.

tides can exhibit direct tumor cell cytotoxicity, immunomodulators or as antiangiogenic factors. For a review on these peptides, see Daffre et al. (2008).

#### A MODEL OF ANTITUMOR EFFECT OF A PEPTIDE

Gomesin is a CAP isolated from hemocytes of the challenged Brazilian spider *Acanthoscurria gomesi*. It is a hairpin-like two-stranded antiparallel  $\beta$ -sheet structure formed by 18 amino acid residues and two translational modifications, the N-terminal pyroglutamic acid (Z) and the C-terminal amidated arginine (R) (Silva et al. 2000, Mandard et al. 2002; Table I). The rigid conformation is maintained by two intramolecular disulfide bridges formed by four cysteine residues, Cys<sup>6-11</sup>, together with six hydrogen bonds in the central part of the molecule, as well as at each end of the  $\beta$ -sheet (Mandard et al. 2002). The peptide is amphipathic, with a hydrophobic face (residues Leu<sup>1</sup>, Val<sup>12</sup> and Tyr<sup>14</sup>) and three hydrophilic regions containing positively charged and polar amino acids at the N-terminus (Arg<sup>3</sup> and Arg<sup>4</sup>), at the C-terminus (Arg<sup>18</sup>) and within the canonical  $\beta$ -turn (Lys<sup>8</sup>, Glu<sup>9</sup>, Arg<sup>10</sup>) (Fazio et al. 2006). A representation of gomesin is depicted on Figure 2.

As stated before, gomesin has a broad and potent antimicrobial activity. The peptide is active against Gram-positive and Gram-negative bacteria, filamentous fungi, yeast (Silva et al. 2000), *Cryptococcus neoformans* (Silva et al. 2007) and parasites, such as *Plasmodium falciparum* and *Plasmodium berghei* (Moreira et al. 2007).

The antitumor activity of gomesin was tested *in vitro* and *in vivo* (Rodrigues et al. 2008). Gomesin exerted direct cytotoxic effects on murine and human tumor cells *in vitro*. The estimated IC<sub>50</sub> for the melanoma cell line B16F10-Nex2 was 3.58  $\mu$ M. The IC<sub>50</sub> was below 10  $\mu$ M for human tumor cell lines (Table I). Human endothelial cells were also sensitive to gomesin *in vitro*, with an IC<sub>50</sub> of 5.30  $\mu$ M. The cytotoxicity was time- and dose-dependent, and was not reversed after peptide removal. The  $\beta$ -hairpin structure and amphipathicity of the peptide are important for antitumor activity, since substitution of cysteine residues



**TABLE I**  
**Naturally occurring CAPs with antitumor activity.**

Peptides	AA sequence*	Source	Antitumor Activity	Refs.
<b><math>\alpha</math>-helical</b>				
BMP27, BMP28	GRFKRFRKKFKKLFKKLSPVIPLLHL, GGLRSLGRKILRAWKKYGPIIVPIIRI	Bovine Cathelicidin- derived	<i>In vitro</i>	Risso et al. 1998, Risso et al. 2002
Cecropin A, Cecropin B	KWKLFKKIEKVGQNIRDGIIKAG- PAVAVVGQATQIAKY KWKVFKKIEKMGRNIRNGIVKAG- PAIAVLGEAKAL	Insects and mammals	<i>In vitro</i> , Xenogeneic model <i>in vivo</i>	Moore et al. 1994, Chan et al. 1998, Winder et al. 1998, Hui et al. 2002, Ye et al. 2004, Suttman et al. 2008
LL-37/hCAP-18	LLGDFFRKSKEKIGKEFKRIVQRIK- DFLRNLVPRTES	Human	<i>In vitro</i>	Okumura et al. 2004, Li et al. 2006
Magainins and analogues	GIGKFLHSAKKFGKAFVGEIMNS (magainin 2)	Frog skin	<i>In vitro</i> , Xenogeneic model <i>in vivo</i> (local therapy)	Cruciani et al. 1991, Soballe et al. 1995, Takeshima et al. 2003, Cruz-Chamoro et al. 2006, Lehman et al. 2006
Gaegurin 5, Gaegurin 6	FLGALFKVASKVLPSVKCAITKKC FLPLLAGLAANFLPTIICFISYKC	Frog skin	<i>In vitro</i>	Kim et al. 2003, Won et al. 2006
Aurein 1.2	GLFDIIKKIAESF	Frog skin	<i>In vitro</i>	Rozek et al. 2000
Citropin 1.1	GLFDVIKKVASVIGGL	Frog skin	<i>In vitro</i>	Doyle et al. 2003
Melittin	GIGAVLKVLTTGLPALISWIKRKRQQ	Insect venom	<i>In vitro</i> , <i>In vivo</i> (melittin- avidin conjugate)	Tosteson and Tosteson 1981, Killion and Dunn 1986, Saini et al. 1999, Holle et al. 2003
Epinecidin-1	GFIFHIIKGLFHAGKMIHGLV	Fish	<i>In vitro</i>	Lin et al. 2009
Polybia-MP1	I D W K K L L D A A K Q I L	Wasp venom	<i>In vitro</i>	Wang et al. 2008
<b><math>\beta</math>-sheet</b>				
Defensins HNP-1 HNP-2 HNP-3	ACYCRIPACIAGERRYGTCTIYQGRWAFCC CYCRIPACIAGERRYGTCTIYQGRWAFCC DCYCRIPACIAGERRYGTCTIYQGRWAFCC	Human	<i>In vitro</i> , <i>In vivo</i> xenogeneic model (HNP-1)	Lichtenstein et al. 1986, Müller et al. 2002, McKeown et al. 2006, Xu et al. 2008
Bovine Lactoferricin	FKCRRWQWRMKKLGAPSITCVRRAF	milk	<i>In vitro</i> , <i>In vivo</i> Xenogeneic model, antiangiogenic	Yoo et al. 1997a, b, Eliassen et al. 2002, Mader et al. 2005, Eliassen et al. 2006
Tachyplesin I	KWCFRVCYRGICYRRCR	Crustacean hemocytes	<i>In vitro</i> , <i>In vivo</i> (RGD- tachyplesin)	Li et al. 2000, Chen et al. 2001, Ouyang et al. 2002, Chen et al. 2005, Shi et al. 2006
Gomesin	ZCRRLCYKQRCVTYCRGR	Insect	<i>In vitro</i> , <i>In vivo</i> (local therapy)	Rodrigues et al. 2008
<b>Linear, with predominant AA</b>				
PR-39, Proline arginine-rich porcine	RRRPRPPYLPRPRPPFPRLPRIPP- GFPPRFPPRF	Porcine cathelicidin- derived	<i>In vitro</i>	Ohtake et al. 1999



# ANTI-FUNGAL AND ANTITUMOR PEPTIDES

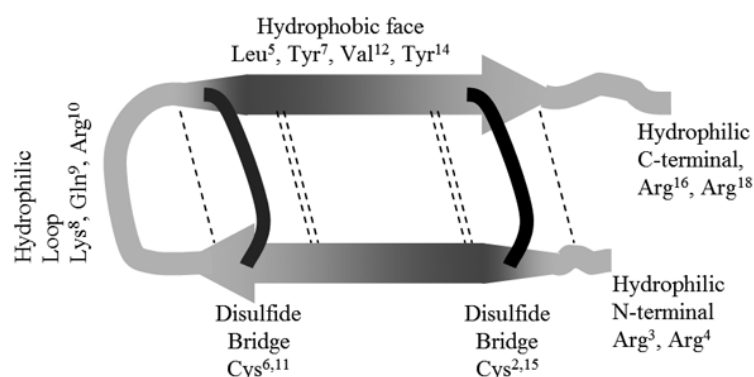


Fig. 2 – Schematic representation of gomesin. The molecule is formed by two antiparallel  $\beta$ -strands stabilized by 2 disulphide bridges (black lines) and 6 hydrogen bonds (hatched lines). Gomesin contains a hydrophobic face and three hydrophilic regions.

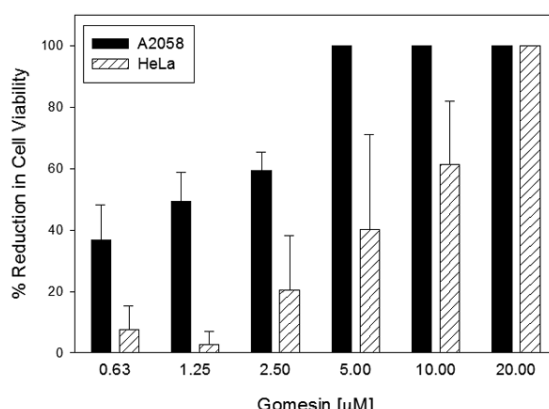


Fig. 3 – Gomesin cytotoxicity *in vitro* against human tumor cells. Human melanoma (A2058) and cervical cancer (HeLa) cells were treated *in vitro* with different concentrations of gomesin for 12 hours, and viable cells were counted in presence of Trypan Blue. The percentage of reduction of cell viability in relation to untreated cells is shown. The melanoma A2058 was the most sensitive and HeLa the most resistant cell line amongst all lineages studied (Rodrigues et al. 2008).

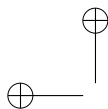
the cytotoxic effect. The enantiomer D-gomesin, synthesized employing D-amino acids and containing both disulfide bridges, was equally cytotoxic for tumor cells, suggesting that chiral recognition is not required for the antitumor effect (Rodrigues et al. 2008).

The peptide concentrates at the tumor cell mem-

tion, caused (1) early morphological alteration, (2) increased granularity and loss of cytoplasmic content, (3) release of lactate dehydrogenase (LDH) in a dose-dependent way; (4) partial inhibition of the membrane-dependent proton gradient; (5) internalization of immunoglobulins that reacted with tubulin filaments and with nuclear histone H1 (monoclonal A4M). Gomesin peptide did not induce apoptosis of tumor cells (Rodrigues et al. 2008).

Interestingly, the monoclonal antibody (mAb) A4M is an IgM that recognizes nuclear histone H1 in B16F10 and Nex2 murine melanoma cells, but is not cytotoxic to an intact tumor cell (A.S. Dobroff et al., unpublished results). After treatment with low doses of gomesin, however, the mAb A4M was internalized in B16F10 cells and showed additive cytotoxic activity *in vitro*. Therefore, gomesin at low concentrations could facilitate the penetration of drugs inside tumor cells, partially reducing toxic doses and allowing penetration of molecules that are not directly cytotoxic to cells with intact membranes.

More importantly, topical *in vivo* treatment with gomesin significantly delayed subcutaneous melanoma development and significantly increased the survival of animals with tumors below the allowed maximal size limit. Male mice with established



oil-in-water cream. This effect can be explained by the direct effect of gomesin on tumor cells, but also by an effect on tumor neoangiogenesis, since endothelial cells were sensitive to low concentrations of the peptide. Repeated topical applications of gomesin did not affect the peripheral healthy skin of treated mice (Rodrigues et al. 2008).

Some patients may develop extensive, confluent regional metastases near the primary nodular melanoma. In these cases, surgical excision or radiotherapy are unsuitable, and topical treatment is a preferred alternative. Some topical treatments have indeed been used tentatively, but only partial responses were obtained with 5-aminolevulinic acid photodynamic therapy (Wolf et al. 1993), imiquimod (Steinmann et al. 2000, Hesling et al. 2004), dinitrochlorobenzene (Malek-Mansour 1973, Illig et al. 1984, von Nida and Quirk 2003), and diphenacyprone (Damian and Thompson 2007). Gomesin could be an alternative for treatment of these patients and eventually also patients with other skin cancers.

#### PERSPECTIVES

Peptides used in protective protocols against pro- and eukaryotic cells, including fungi and tumor cells, can act directly on target cells or will elicit an immune response that may be effective to control infections and tumor development. Peptides allow structural changes to incorporate protective substitutions, chiral derivatives, non-natural amino acids and other modifications aiming at increased stability, efficiency and resistance to proteolysis. In this sense, they are much more drug-like than recombinant proteins. A great number of peptide sequences with biological activity is now recognized, and the finding that fragments of immunoglobulin variable chains have increased frequency of bioactivity opens a broad field of investigation. Peptide-based vaccines are now in development for various pathologies including cancer (Purcell et al. 2007). The possibility of chemical synthesis of a limitless variety of peptide sequences and derivatives poses the question of how many more reagents can be produced compared to our capacity to test them in different biological systems. The use of

vaccine candidate being presented by most Caucasian HLA-DR molecules, and being able to protect against massive *P. brasiliensis* infection in normal and immunosuppressed mice. The combination of chemotherapy and P10 vaccination is therefore a very promising strategy to treat human PCM. Antitumor peptides for systemic and topical treatment are additional tools that can be largely developed as adjuvants of conventional treatment.

#### ACKNOWLEDGMENTS

The authors thank Dr. Edna Haapalainen for the technical supervision with the electron microscopy. The present review was supported by grants from Fundação de Amparo à Pesquisa do Estado de São Paulo (FAPESP) and research fellowships from Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq) and Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES).

#### RESUMO

Peptídeos são moléculas particularmente reativas produzidas por uma grande variedade de espécies, aptos a exercer um número de funções em organismos uni- e multicelulares como mediadores, agonistas e substâncias regulatórias. Alguns deles exercem efeitos citotóxicos em células outras das que os produzem, e podem ter um papel controlando subpopulações e protegendo certas espécies ou tipos celulares. No presente, focalizamos peptídeos antifúngicos e antitumorais e discutimos alguns modelos nos quais seqüências específicas e estruturas exercem efeitos inibitórios diretos ou estimulam uma resposta imune protetora. O peptídeo letal (“killer”), deduzido de um anticorpo anti-idiotípico, com várias atividades antimicrobianas bem como outros peptídeos derivados de imunoglobulinas com atividades citotóxicas incluindo efeitos antitumorais são modelos estudados *in vitro* e *in vivo*. O peptídeo P10 da gp43 de *P. brasiliensis* e a perspectiva de vacina contra a paracoccidiodomicose é outro tópico ilustrando o efeito protetor *in vivo* contra um fungo patogênico. Peptídeos antimicrobianos catiônicos com atividades antitumorais são os principais revisados aqui. O tratamento local do melanoma murino com o peptídeo gomesina é outro modelo estudado na Unidade de



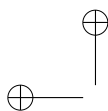
## ANTI-FUNGAL AND ANTITUMOR PEPTIDES

### REFERENCES

- AERTS A, FRANÇOIS IEJA, CAMMUE BPA AND THEVISEN K. 2008. The mode of antifungal action of plant, insect and human defensins. *Cell Mol Life Sci* 65: 2069–2079.
- BARBOSA FM, DAFFRE S, MALDONADO RA, MIRANDA A, NIMRICHTER L AND RODRIGUES ML. 2007. Gomesin, a peptide produced by the spider *Acanthoscurria gomesiana*, is a potent anticryptococcal agent that acts in synergism with fluconazole. *FEMS Microbiol Lett* 274: 279–286.
- BECHINGER B. 1997. Structure and functions of channel-forming peptides: magainins, cecropins, mellitin and alamethicin. *J Membrane Biol* 156: 197–211.
- BOURGEOIS C, BOUR JB, AHO LS AND POTHIER P. 1998. Prophylactic administration of a complementarity-determining region derived from a neutralizing monoclonal antibody is effective against respiratory syncytial virus infection in BALB/c mice. *J Virol* 72: 807–810.
- BRAKHAGE AA. 2005. Systemic fungal infections caused by *Aspergillus* species: epidemiology, infection process and virulence determinants. *Curr Drug Targets* 6: 875–886.
- BUISSA-FILHO R, PUCCIA R, MARQUES AF, PINTO FA, MUÑOZ JE, NOSANCHUK JD, TRAVASSOS LR AND TABORDA CP. 2008. Monoclonal antibody against the major diagnostic antigen of *Paracoccidioides brasiliensis* mediates immune protection in infected Balb/c mice challenged intratracheally with the fungus. *Infect Immun* 76: 3321–3328.
- BUZAID AC AND ATKINS M. 2001. Practical guidelines for the management of biochemotherapy-related toxicity in melanoma. *Clin Cancer Res* 7: 2611–2619.
- CENCI E, BISTONI F, MENCACCI A, PERITO S, MAGLIANI W, CONTI S, POLONELLI L AND VECCHIARELLI A. 2004. A synthetic peptide as a novel anticryptococcal agent. *Cell Microbiol* 6: 953–961.
- CHAN SC, YAU WL, WANG W, SMITH DK, SHEU F-S AND CHEN HM. 1998. Microscopic observations of the different morphological changes caused by anti-bacterial peptides on *Klebsiella pneumoniae* and HL-60 leukemia cells. *J Pept Sci* 4: 413–425.
- CHEN J, XU X-M, UNDERHILL CB, YANG S, WANG L, CHEN Y, HONG S, CRESWELL K AND ZHANG L. 2005. Tachyplesin activates the classic complement pathway to kill tumor cells. *Cancer Res* 65: 4614–4622.
- CHO Y, TURNER J, DIHN N-G AND LEHRER R. 1999. Activity of protegrins against yeast-phase *Candida albicans*. *Infect Immun* 66: 2486–2493.
- CISALPINO PS, PUCCIA R, YAMAUCHI LM, CANO JF, SILVEIRA JF AND TRAVASSOS LR. 1996. Cloning, characterization, and epitope expression of the major cell wall antigen of *Paracoccidioides brasiliensis*. *J Biol Chem* 271: 4553–4560.
- CRUCIANI RA, BARKER JL, ZASLOFF M, CHEN H, COLAMONICI O. 1991. Antibiotic magainins exhibit cytolytic activity against transformed cell lines and inhibit channel formation. *Proc Natl Acad Sci USA* 88: 3796.
- CRUZ-CHAMORO L, PUERTOLLANO MA, PUERTO J, E, CIENFUEGOS GA DE AND DE PABLO MA. 2000. *In vitro* biological activities of magainin 1 alone or in combination with nisin. *Peptides* 27: 1201–1209.
- DAFFRE S, BULET P, SPISNI A, EHRET-SABATIER E, RODRIGUES EG AND TRAVASSOS LR. 2008. Bacterial peptides. In: ATTA-UR-RAHMAN (Ed), *Antibiotics in Natural Product Chemistry*, vol 35, Chapter 1, p. 597–691.
- DAMIAN DL AND THOMPSON JF. 2007. Treatment of cutaneous metastatic melanoma with topical cecropin. *J Am Acad Dermatol* 56: 869–871.
- DE LUCCA A, BLAND J, JACKS T, GRIMM C AND WALSH T. 1998. Fungicidal and binding properties of the peptides cecropin B and dermaseptin. *Med Mycol* 36: 291–298.
- DE LUCCA A, BLAND J, VIGO C, JACKS T, PETER J AND WALSH T. 2000. D-cecropin: proteolytic activity, lethality for pathogenic fungi, and binding properties. *Med Mycol* 38: 301–308.
- DE LUCCA AJ AND WALSH TJ. 2000. Antifungal peptides: Origin, activity, and therapeutic potential. *Rev Microbiol* 17: 116–120.
- DE LUCCA AJ, JACKS TJ, TAKEMOTO J, VINYARD J, PETER J, NAVARRO E AND WALSH TJ. 1999. Antifungal activity, lethality, binding, and cytotoxicity of syringomycin A. *Antimicrob Agents Chemother* 43: 371–373.
- DEBONO M AND GORDEE R. 1994. Antibiotics that inhibit fungal cell wall development. *Annu Rev Microbiol* 48: 471–497.
- DENNING D. 1997. Echinocandins and pneumocandins: a new antifungal class with a novel mode of action. *Antimicrob Chemother* 40: 611–614.



- for the dectin-1-dependent internalization of zymosan by macrophages. *J Leukoc Biol* 75: 649–656.
- DOBRYNSKA J, SZACHOWICZ-PETELSKA B, SULKOWSKY S AND FIGASZEWSKI Z. 2005. Changes in electric charge and phospholipids composition in human colorectal cancer cells. *Mol Cell Biochem* 276: 113–119.
- DOYLE J, BRINKWORTH CS, WEGENER KL, CARVER JA, LLEWELLYN IN, OLVER JH, BOWIE PA AND WABNITZ MJ. 2003. nNOS inhibition, antimicrobial and anticancer activity of the amphibian skin peptide citropin 1.1, and synthetic modifications. The solution structure of a modified citropin 1.1. *Eur J Biochem* 270: 1141–1153.
- DUDA TFJR, VANHOYE D AND NICOLAS P. 2002. Roles of diversifying selection and coordinated evolution in the evolution of amphibian antimicrobial peptides. *Mol Biol Evol* 19: 858–864.
- ELIASSEN LT, BERGE G, SVEINBJORNSSON B, SVENDSEN JS, VORLAND LH AND REKDAL O. 2002. Evidence for a direct antitumor mechanism of action of bovine lactoferricin. *Anticancer Res* 22: 2703–2710.
- ELIASSEN LT ET AL. 2006. The antimicrobial peptide, Lactoferricin B, is cytotoxic to neuroblastoma cells in vitro and inhibits xenograft growth *in vivo*. *Int J Cancer* 119: 493–500.
- ESPINOSA E, ZAMORA P, FELIU J AND GONZALEZ BARON M. 2003. Classification of anticancer drugs – A new system based on therapeutic targets. *Cancer Treat Rev* 29: 515–523.
- FAZIO MA, OLIVEIRA VX, BULET P, MIRANDA MTM, DAFFRE S AND MIRANDA A. 2006. Structure-activity relationship studies of gomesin: importance of the disulfide bridges for conformation, bioactivities and serum stability. *Biopolymers* 84: 205–218.
- FRANÇOIS IE, AERTS AM, CAMMUE BP AND THEVISSSEN K. 2005. Currently used antimycotics: spectrum, mode of action and resistance occurrence. *Curr Drug Targets* 6: 895–907.
- GATTI L AND ZUNINO F. 2005. Overview of tumor cell chemoresistance mechanisms. *Methods Mol Med* 111: 127–148.
- HECTOR R, ZIMMER B AND PAPPAGIANIS D. 1990. Evaluation of nikkomycins X and Z in murine models of coccidioidomycosis, histoplasmosis and blastomycosis. *Antimicrob Agents Chemother* 34: 587–593.
- HELMERHORST EJ, BREEUWER P AND VANIT HOF W. HESLING C ET AL. 2004. *In vivo* and *in situ* modulation of the expression of genes involved in metastasis and angiogenesis in a patient treated with topical imiquimod for melanoma skin metastases. *Br J Dermatol* 150: 761–767.
- HETRU C, LETELLIER L, OREN Z, HOFFMAN JA AND SHAI Y. 2000. Androctonin, a hydrophilic disulphide-bridged non-haemolytic anti-microbial peptide: a plausible mode of action. *Biochem J* 345: 653–664.
- HOLLE L, SONG W, HOLLE F, WEI Y, WAGNER T AND YU X. 2003. A matrix metalloproteinase 2 cleavable melittin/avidin conjugate specifically targets tumor cells *in vitro* and *in vivo*. *Int J Oncol* 22: 93–98.
- HORTON KL, STEWART KM, FONSECA SB, GUO Q AND KELLEY SO. 2008. Mitochondria-penetrating peptides. *Chem Biol* 15: 375–382.
- HOSKIN DW AND RAMAMOORTHY A. 2008. Studies on anticancer activities of antimicrobial peptides. *Bioch Biophys Acta* 1778: 357–375.
- HUI L, LEUNG K AND CHEN HM. 2002. The combined effects of antibacterial peptide cecropin A and anti-cancer agents on leukemia cells. *Anticancer Res* 22: 2811–2816.
- ILLIG L, PAUL E AND BODEKER RH. 1984. Epifocal dinitrochlorobenzene therapy in malignant melanoma (experience during the last eight years). *Anticancer Res* 4: 293–298.
- IWAI LK ET AL. 2003. In silico prediction of peptides binding to multiple HLA-DR molecules accurately identifies immunodominant epitopes from gp43 of *Paracoccidioides brasiliensis* frequently recognized in primary peripheral blood mononuclear cell responses from sensitized individuals. *Mol Med* 9: 209–219.
- IWAI LK ET AL. 2007. T-cell recognition of *Paracoccidioides brasiliensis* gp43-derived peptides in patients with paracoccidioidomycosis and healthy individuals. *Clin Vaccine Immunol* 14: 474–476.
- JABLONOWSKI D AND SCHAFFRATH R. 2007. Zymocin, a composite chitinase and tRNase killer toxin from yeast. *Biochem Soc Trans* 35: 1533–1537.
- JIGGINS FM AND KIM K-W. 2005. The evolution of antifungal peptides in *Drosophila*. *Genetics* 171: 1847–1859.
- KILLION JJ AND DUNN JD. 1986. Differential cytolysis of murine spleen, bone marrow and leukemia cells by melittin reveals differences in membrane topography. *Biochem Biophys Res Commun* 139: 222–227.
- KIM S, KIM SS, BANG Y-J, KIM S-J AND LEE B-J. 2003.



#### ANTI-FUNGAL AND ANTITUMOR PEPTIDES

- KLASSEN R, TEICHERT S AND MEINHARDT F. 2004. Novel yeast killer toxins provoke S-phase arrest and DNA damage checkpoint activation. *Mol Microbiol* 53: 263–273.
- KURTZ M AND DOUGLAS C. 1997. Lipopeptide inhibitors of fungal glucan synthesis. *Antimicrob Agents Chemother* 35: 79–86.
- LEE CH, KIM S, HYUN B, SUH JW, YON C, KIM C, LIM Y AND KIM C. 1994. Cepacidine A, a novel antifungal antibiotic produced by *Pseudomonas cepacia* I. Taxonomy, production, isolation, and biological activity. *J Antibiot* 47: 1402–1405.
- LEHMAN J, RETZ M, SIDHU SS, SUTTMANN H, SELL M, PAULSEN F, HARDER J, UNTEREGGER G AND STÖCKLE M. 2006. Antitumor activity of the antimicrobial peptide magainin II against bladder cancer cell lines. *Eur Urol* 50: 141–147.
- LEMAITRE B, REICHHART J AND HOFFMAN J. 1997. *Drosophila* host defense: differential induction of antimicrobial peptide genes after infection by various classes of microorganisms. *Proc Natl Acad Sci USA* 94: 14614–14619.
- LEVI M, SÄLLBERG M, RUDÉN U, HERLYN D, MARUYAMA H, WIGZELL H, MARKS J AND WAHREN B. 1993. A complementarity-determining region synthetic peptide acts as a miniantibody and neutralizes human-immunodeficiency-virus type-1 *in vitro*. *Proc Nat Acad Sci USA* 90: 4374–4378.
- LI QF, OUYANG GL, LI CY AND HONG SG. 2000. Effects of tachyplesin on the morphology and ultrastructure of human gastric carcinoma cell line BGC-823. *World J Gastroenterol* 6: 676–680.
- LI X, LI Y, HAN H, MILLER DW AND WANG G. 2006. Solution structures of human LL-37 fragments and NMR-based identification of a minimal membrane-targeting antimicrobial and anticancer region. *J Am Chem Soc* 128: 5776–5785.
- LICHTENSTEIN A, GANZ T, SELSTED ME AND LEHRER RI. 1986. *In vitro* tumor cell cytolysis mediated by peptide defensins of human and rabbit granulocytes. *Blood* 68: 1407–1410.
- LIM Y, SUH J-W, KIM S, HYUN B, KIM C AND LEE C. 1994. Cepacidine A, a novel antifungal antibiotic produced by *Pseudomonas cepacia*. II. Physicochemical properties and structure elucidation. *J Antibiot* 47: 1406–1416.
- LIN WJ, CHIEN YL, PAN CY, LIN TL, CHEN JY, CHIU SJ AND HU SC. 2003. Fungicidal activity of a novel peptide derived from the antifungal protein of *Trichoderma reesei*. *J Antibiot* 56: 140–145.
- LITMAN GW, CANNON JP AND DISHAW LJ. 2005. Structuring immune phylogeny: New perspectives. *Immunol* 5: 866–879.
- LOBO DS, PEREIRA IB, FRAGEL-MADEIRA L, MELO LN, CABRAL LM, FARIA J, BELLIO M, COSTA RC, LINDEN R AND KURTENBACH E. 2007. A fungal *Pisum sativum* defensin 1 interacts with *Neurospora crassa* cyclin F related to the cell cycle. *Biochem J* 405: 987–996.
- MADER JS, SALSMAN J, CONRAD DM AND HOSKINS J. 2005. Bovine lactoferricin selectively induces apoptosis in human leukemia and carcinoma cell lines. *Mol Ther* 4: 612–624.
- MAGLIANI W, CONTI S, GERLONI M, BERTOLOTTO L AND POLONELLI L. 1997. Yeast killer systems. *Clin Microbiol Rev* 10: 369–400.
- MAGLIANI W, CONTI S, SALATI A, ARSENI S, RATTI L, FRAZZI R AND POLONELLI L. 2004a. Engineered killer mimotopes: new synthetic peptides for antimicrobial therapy. *Curr Med Chem* 11: 1793–1800.
- MAGLIANI W, CONTI S, SALATI A, VACCARI S, RATTI L, MAFFEI DL AND POLONELLI L. 2004b. Therapeutic potential of yeast killer toxin-like antimicrobial mimotopes. *FEMS Yeast Res* 5: 11–18.
- MALEK-MANSOUR S. 1973. Remission of melanoma after D.N.C.B. treatment. *Lancet* 2: 503–504.
- MANDARD N, BULET P, CAILLE A, DAFFREY C AND VOVELLE F. 2002. The solution structure of gongylosporin, an antimicrobial cysteine-rich peptide from the spider *Gongylus*. *J Biochem* 269: 1190–1198.
- MARQUES AF, DA SILVA MB, JULIANO MA, TRAVASSOS LR AND TABORDA CP. 2006. Peptide immunization as an adjuvant to chemotherapy in mice challenged intratracheally with virulent yeast cells of *Paracoccidioides brasiliensis*. *Antimicrob Agents Chemother* 50: 2819.
- MARQUES AF, DA SILVA MB, JULIANO MA, MANTOVANI JE, TRAVASSOS LR AND TABORDA CP. 2008. Protective effect of P10 immunization and chemotherapy in immunocompetent mice challenged intratracheally with virulent cells of *Paracoccidioides brasiliensis*. *Microbes Infect* 10: 1251–1258.
- MARQUES DA SILVA SH, QUEIROZ-TELLES F, COSTA AL, BLOTTA MH, LOPES JD AND CAMARGO Z. 2008. Monitoring gp43 antigenemia in paracoccidioidomycosis patients during therapy. *J Clin Microbiol* 42: 2411–2415.





- MAVOR AL, THEWES S AND HUBE B. 2005. Systemic fungal infections caused by *Candida* species: epidemiology, infection process and virulence attributes. *Curr Drug Targets* 6: 863–874.
- MCKEOWN STW, LUNDY FT, NELSON J, LOCKHART D, IRWIN CR, COWAN CG AND MARLEY JJ. 2006. The cytotoxic effects of human neutrophil peptide-1 (HNP-1) and lactoferrin on oral squamous cell carcinoma (OSCC) *in vitro*. *Oral Oncol* 42: 685–690.
- MCMANUS AM, OTVOS L, HOFFMAN R AND CRAIK DJ. 1999. Conformational studies by NMR of the antimicrobial peptide, drosocin, and its non-glycosylated derivatives: effects of glycosylation on solution conformation. *Biochemistry* 38: 705–714.
- MOORE AJ, DEVINE DA AND BIBBY MC. 1994. Preliminary experimental anticancer activity of cecropins. *Pept Res* 7: 265–269.
- MOR A, HANI K AND NICOLAS P. 1994. The vertebrate peptide antibiotics dermaseptins have overlapping structural features but target specific organisms. *J Biol Chem* 269: 31635–31641.
- MOREIRA CK, RODRIGUES FG, GHOSH A, VAROTTI FD, MIRANDA A, DAFFRE S, JACOBS-LORENA M AND MOREIRA LA. 2007. Effect of the antimicrobial peptide gomesin against different life stages of *Plasmodium spp.* *Exp Parasitol* 116: 346–353.
- MÜLLER CA ET AL. 2002. Human  $\alpha$ -defensins HNP-1, -2, and -3 in renal cell carcinoma. Influences on tumor cell proliferation. *Am J Pathol* 160: 1311–1324.
- NAGEIC M, NAGEIC E, BALTISBURGER J, WELL G, LESTER R AND DICKSON R. 1997. Sphingolipid synthesis as a target for antifungal drugs. *J Biol Chem* 272: 9807–9817.
- NAUMOV GN, TOWSON JL, MACDONALD IC, WILSON SM, BRAMWELL VH, GROOM AC AND CHMBERS AF. 2003. Ineffectiveness of doxorubicin treatment on solitary dormant mammary carcinoma cells or late-developing metastasis. *Breast Cancer Res Treat* 82: 199–206.
- NIMRICHTER L, RODRIGUES ML, BARRETO-BERGTER E AND TRAVASSOS LR. 2008. Sophisticated functions for a single molecule: The role of glucosylceramides in fungal cells. *Lipid Insights* 2: 61–73.
- OHTAKE T, FUJIMOTO Y, IKUTA K, SAITO H, OHHIRA M, ONO M AND KOHGO Y. 1999. Proline-rich antimicrobial peptide, PR-39 gene transduction altered invasive activity, ABIKO Y, SHIBATA T, HIRATA M AND ISOGAI H. 2004. C-terminal domain of human CAP18 antimicrobial peptide induces apoptosis in oral squamous cell carcinoma SAS-H1 cells. *Cancer Lett* 212: 185–194.
- OUYANG GL, LI QF, PENG XX, LIU QR AND HONG SG. 2002. Effects of tachyplesin on proliferation and differentiation of human hepatocellular carcinoma SMMC-7721 cells. *World J Gastroenterol* 8: 1053–1058.
- PALUSZYNSKI JP, KLASSEN R AND MEINHARDT F. 2007. *Pichia acaciae* killer system: genetic analysis of toxin immunity. *Appl Environ Microbiol* 73: 4373–4378.
- PARDO J ET AL. 2001. A role of the mitochondrial apoptosis-inducing factor in granulysin-induced apoptosis. *J Immunol* 167: 1222–1229.
- PETTOELLO-MANTOVANI M, NOCERINO A, POLONELLI L, MORACE G, CONTI S, DI MARTINO L, DE RITIS G, IAFUSCO M AND GUANDALINI S. 1995. *Hansenula anomala* killer toxin induces secretion and severe acute injury in the rat intestine. *Gastroenterology* 109: 1900–1906.
- PINTO AR, PUCCIA R, DINIZ SN, FRANCO MF AND TRAVASSOS LR. 2000. DNA-based vaccination against murine paracoccidioidomycosis using the gp43 gene from *Paracoccidioides brasiliensis*. *Vaccine* 18: 3050–3058.
- POLONELLI L, CONTI S, GERLONI M, MAGLIANI W, CASTAGNOLA M, MORACE G AND CHEZZI C. 1991. ‘Antibodies’: antibiotic-like anti-idiotypic antibodies. *J Med Vet Mycol* 29: 235–242.
- POLONELLI L, MAGLIANI W, CONTI S, BRACCI L, LOZZI L, NERI P, ADRIANI D, DE BERNARDIS F AND CASSONE A. 2003. Therapeutic activity of an engineered synthetic killer anti-idiotypic antibody fragment against experimental mucosal and systemic candidiasis. *Infect Immun* 71: 6205–6212.
- POLONELLI L ET AL. 2008. Antibody complementarity-determining regions (CDRs) can display differential antimicrobial, antiviral and antitumor activities. *PLoS One* 3(6): e2371. doi: 10.1371/journal.pone.0002371.
- PUCCIA R, SCHENKMAN S, GORIN PA AND TRAVASSOS LR. 1986. Exocellular components of *Paracoccidioides brasiliensis*: identification of a specific antigen. *Infect Immun* 53: 199–206.
- PURCELL AW, MCCLUSKEY J AND ROSSJOHN J. 2007. More than one reason to rethink the use of peptides in vaccine design. *Nat Rev Drug Discov* 6: 404–414.



#### ANTI-FUNGAL AND ANTITUMOR PEPTIDES

- RISSE A, BRAIDOT E, SORDANO MC, VIANELLO A, MACRI F, SKERLAVAJ B, ZANETTI M, GENNARO R AND BERNARDI P. 2002. BMAP-28, an antibiotic peptide of innate immunity, induces cell death through opening of the mitochondrial permeability transition pore. *Mol Cell Biol* 22: 1926–1935.
- ROBERTS W AND SELITRENNIKOFF C. 1991. Zeamatin, an antifungal protein made from maize with membrane-permeabilizing activity. *J Gen Microbiol* 40: 1771–1778.
- RODRIGUES EG AND TRAVASSOS LR. 1994. Nature of the reactive epitopes in *Paracoccidioides brasiliensis* polysaccharide antigen. *J Med Vet Mycol* 32: 77–81.
- RODRIGUES EG ET AL. 2008. Effective topical treatment of subcutaneous murine B16F10-Nex2 melanoma by the antimicrobial peptide gomesin. *Neoplasia* 10: 61–68.
- RODRIGUES ML, TRAVASSOS LR, MIRANDA KR, FRANZEN AJ, ROZENTAL S, DE SOUZA W, ALVIANO CS AND BARRETO-BERGTER E. 2000. Human antibodies against a purified glucosylceramide from *Cryptococcus neoformans* inhibit cell budding and fungal growth. *Infect Immun* 68: 7049–7060.
- ROZEK T, WEGENER KL, BOWIE JH, OLVER IN, CARVER JA, WALLACE JC AND TYLER MJ. 2000. The antibiotic and anticancer active aurein peptides from the Australian Bell Frogs *Litoria aurea* and *Litoria raniformis*. The solution structure of aurein 1.2. *Eur J Biochem* 267: 5330–5341.
- SAINI SS, CHOPRA AK AND PETERSON JW. 1999. Melittin activates endogenous phospholipase D during cytolysis of human monocytic leukemia cells. *Toxicon* 37: 1605–1619.
- SANTOS A AND MARQUINA D. 2004. Ion channel activity by *Pichia membranifaciens* killer toxin. *Yeast* 21: 151–162.
- SCHMITT MJ AND BREINIG F. 2006. Yeast viral killer toxins: lethality and self-protection. *Nat Rev Microbiol* 4: 212–221.
- SCHMITT MJ AND REITER J. 2008. Viral induced yeast apoptosis. *Biochim Biophys Acta* 1783: 1413–1417.
- SHAI Y. 1995. Molecular recognition between membrane-spanning polypeptides. *TIBS* 20: 460–464.
- SHI SL, WANG YY, LIANG Y AND LI QF. 2006. Effects of tachyplesin and n-sodium butyrate on proliferation and gene expression of human gastric adenocarcinoma cell line BGC-823. *World J Gastroenterol* 12: 1694–1698.
- SILVA PI, DAFFRE S AND BULET P. 2000. Isolation and characterization of gomesin, a 18-residue cysteine-rich defensin-like peptide from *Paracoccidioides brasiliensis*. *Antonie van Leeuwenhoek* 77: 101–108.
- SILVA SH, MATTOS GROSSO D, LOPES JD, COLOMBO R, BLOTTA MH, QUEIROZ-TELLES F AND CAMARAO JR. 2004. Detection of *Paracoccidioides brasiliensis* circulating antigen and follow-up of patients under antimycotic therapy. *J Clin Microbiol* 42: 4480–4484.
- SOBALLE PW, MALOY WL, MYRGRA ML, JACOBSON KA AND HERLYN M. 1995. Experimental local therapy with magainin peptides. *Int J Cancer* 60: 280–284.
- SORENSEN K, KIM K-H AND TAKEMOTO JY. 1996. Antifungal and fungicidal activities and erythrocytic hemolysis of *Pseudomonas syringae* pv. *syringae*. *Antonie van Leeuwenhoek* 70: 271–273.
- STEINMANN A, FUNK JO, SCHULER G AND VOIGT M. 2000. Topical imiquimod treatment of cutaneous melanoma metastasis. *J Am Acad Dermatol* 42: 555–556.
- SUTTMANN H, RETZ M, PAULSEN F, HARTMANN J, ZWARGEL U, KAMRADT J, WULFICH B, UNTERWIESINGER G, STÖCKLE M AND LEHMANN J. 2008. Antimicrobial peptides of the Cecropin-family show potent cytotoxic activity against bladder cancer cells. *BMJ Open* 8: 5.
- TABORDA CP, JULIANO MA, PUCCIA R, FRANCO RF AND TRAVASSOS LR. 1998. Mapping of the T-cell epitopes in the major 43-kilodalton glycoprotein of *Paracoccidioides brasiliensis* which induces a Th-1 response protective against fungal infection in BALB/c mice. *Infect Immun* 66: 786–793.
- TABORDA CP, NAKAIE CR, CILLI EM, RODRIGUES ML, SILVA LS, FRANCO MF AND TRAVASSOS LR. 2000. Synthesis and immunological activity of a synthetic peptide carrying the T-cell epitope of gp43, the major cell surface antigen of *Paracoccidioides brasiliensis*. *J Immunol* 164: 58–65.
- TAKESAKO K, KURODA H, INOUE T, HARUNA F, KAWA Y AND KATO I. 1993. Biological properties of aureobasidin A, a cyclic depsipeptide antifungal antibiotic. *J Antibiot* 46: 1414–1420.
- TAKESHIMA K, CHIKUSHI A, LEE K-K, YONEHARA A AND MATSUZAKI K. 2003. Translocation of analogs of the antimicrobial peptides magainin and buforin II across human cell membranes. *J Biol Chem* 278: 1310–1315.
- TAVARES PM, THEVISSSEN K, CAMMUE BP, FRANCO RF, BARRETO-BERGTER E, TABORDA CP, MARQUES M, RODRIGUES ML AND NIMRICHTER L. 2008. Antifungal activity of the antifungal peptide defensin BnAFP1 against *Candida albicans*. *Antonie van Leeuwenhoek* 94: 101–108.



- THEVISSSEN K, WARNECKE DC, FRANÇOIS IE, LEIPELT M, HEINZ E, OTT C, ZHRINGER U, THOMMA BP, FERKET KK AND CAMMUE BP. 2004. Defensins from insects and plants interact with fungal glucosylceramides. *J Biol Chem* 279: 3900–3905.
- TOSTESON MT AND TOSTESON DC. 1981. The sting melittin forms channels in lipid bilayers. *Biophys J* 36: 109–116.
- TRAVASSOS LR, SILVA LS, RODRIGUES EG, CONTI S, SALATI A, MAGLIANI W AND POLONELLI L. 2004a. Therapeutic activity of a killer peptide against experimental paracoccidioidomycosis. *J Antimicrob Chemother* 54: 956–958.
- TRAVASSOS LR, TABORDA CP, IWAI LK, CUNHA-NETO E AND PUCCIA R. 2004b. The gp43 from *Paracoccidioides brasiliensis*: A major diagnostic antigen and vaccine candidate. In: DOMER JE, KOBAYASHI GS (Eds), *The Mycota XII, Human Fungal Pathogens*, Springer-Verlag, Berlin-Heidelberg, p. 279–296.
- TRAVASSOS LR, RODRIGUES EG, IWAI LK AND TABORDA CP. 2008a. Attempts at a peptide vaccine against paracoccidioidomycosis, adjuvant to chemotherapy. *Mycopathologia* 165: 341–352.
- TRAVASSOS LR, TABORDA CP AND COLOMBO AL. 2008b. Treatment options for paracoccidioidomycosis and new strategies investigated. *Expert Rev Anti Infect Ther* 6: 251–262.
- UTSUGI T, SCHROIT AJ, CONNOR J, BUCCANA CD AND FIDLER IJ. 1991. Elevated expression of phosphatidylserine in the outer leaflet of human tumor cells and recognition by activated human blood monocytes. *Cancer Res* 51: 3062–3066.
- VON NIDA J AND QUIRK C. 2003. Successful treatment of in-transit melanoma metastases using topical 2,4-dinitrochlorobenzene. *Australas J Dermatol* 44: 277–280.
- XU N ET AL. 2008. Human alpha-defensin-1 inhibits growth of human lung adenocarcinoma xenograft in nude mice. *Mol Cancer Ther* 7: 1588–1597.
- WANG KR, ZHANG BZ, ZHANG W, YAN JX, LI J AND WANG R. 2008. Antitumor effects, cell selectivity and structure-activity relationship of a novel antimicrobial peptide polybia-MPI. *Peptides* 29: 963–968.
- WINDER D, GUNZBURG WH, ERFLE V AND SALMONS B. 1998. Expression of antimicrobial peptides has an antitumor effect in human cells. *Biochem Biophys Res Commun*
- WOLF P, RIEGER E AND KERL H. 1993. Topical photodynamic therapy with endogenous porphyrins after application of 5-aminolevulinic acid. An alternative treatment modality for solar keratoses, superficial squamous cell carcinomas, and basal cell carcinomas? *J Am Acad Dermatol* 28: 17–21.
- WON H-S, SEO M-D, JUNG S-J, LEE S-J, KANG S-J, SON W-S, KIM H-J, PARK T-K, PARK S-J AND LEE B-J. 2006. Structural determinants for the membrane interaction of novel bioactive undecapeptides derived from gaegurin 5. *J Med Chem* 49: 4886–4895.
- YE J-S, ZHENG J-X, LEUNG KW, CHEN HM AND SHEU F-S. 2004. Induction of transient ion channel-like pores in a cancer cell by antibiotic peptide. *J Biochem* 136: 255–259.
- YOO Y-C, WATANABE R, KOIKE Y, MITOBE M, SHIMAZAKI K, WATANABE S AND AZUMA I. 1997a. Apoptosis in human leukemic cells induced by lactoferricin, a bovine milk-derived peptide: involvement of reactive oxygen species. *Biochem Biophys Res Commun* 237: 624–628.
- YOO Y-C, WATANABE R, WATANABE K, HATA K, SHIMAZAKI K AND AZUMA I. 1997b. Bovine lactoferrin and lactoferricin, a peptide derived from bovine lactoferrin, inhibit tumor metastasis in mice. *Jpn J Cancer Res* 88: 184–190.
- YOON WH, PARK HD, LIM K AND HWANG BD. 1996. Effect of O-glycosylated mucin on invasion and metastasis of HM7 human colon cancer cells. *Biochem Biophys. Res Commun* 222: 694–699.
- YOUNT NY, BAYER AS, XIONG YQ AND YEAMAN MR. 2006. Advances in antimicrobial peptide immunobiology. *Biopolymers* 84: 435–458.
- ZACHOWSKI A. 1993. Phospholipids in animal eukaryotic membranes: transverse asymmetry and movement. *Biochem J* 294: 1–14.
- ZASLOFF M. 1987. Magainins, a class of antimicrobial peptides from *Xenopus* skin: isolation, characterization of two active forms and partial cDNA sequence of a precursor. *Proc Natl Acad Sci USA* 84: 5449–5453.
- ZASLOFF M. 2002. Antimicrobial peptides of multicellular organisms. *Nature* 415: 389–395.