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## Diet influence on egg production of the copepod *Acartia tonsa* (Dana, 1896)

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### ABSTRACT

Egg production in the copepod *Acartia tonsa* was evaluated using different densities of the microalgae *Thalassiosira weissflogii*, *Chaetoceros muelleri* and *Isochrysis galbana*. Male and female were kept under controlled conditions (salinity 30, 20°C, photoperiod 12L:12D), acclimated to the experimental conditions and left over a period of 24 h to allow copulation. Algal densities tested were equivalent in biovolume and corresponded to 0, 2.5, 5, 10, 20, 40 and 60.10<sup>3</sup> cells.mL<sup>-1</sup> of *T. weissflogii*. Ten acclimated female were separated, transferred to glass bottles and exposed for further 24 h to the corresponding experimental medium. After this period, the eggs were fixed and counted. Copepod egg production reached a threshold value when *T. weissflogii*, *C. muelleri* and *I. galbana* were supplied at 10.10<sup>3</sup>, 140.10<sup>3</sup> and 640.10<sup>3</sup> cells.mL<sup>-1</sup>, respectively. Mean egg production corresponded to 28.0 ± 0.5, 20.1 ± 1.0 and 22.0 ± 3.5 eggs.female<sup>-1</sup>.day<sup>-1</sup>, respectively. Copepods fed *T. weissflogii* showed the highest mean egg production while those fed *I. galbana* reached a maximum egg production when the algae were supplied at a density two- to four-fold higher, considering the biovolume of *T. weissflogii* and *C. muelleri*. These differences are explained considering the different sizes of the microalgae used to feed the copepods.

**Key words:** egg production, *Acartia tonsa*, microalgae.

### INTRODUCTION

Copepods from the genus *Acartia* play an important role in the food webs of estuaries in both tropical and subtropical areas (Björnberg 1981, Mauchline 1998). Generally, they show the higher biomass values in most shallow enclosed bays and estuaries (Azaiteiro et al. 2005, Leandro et al. 2007). This fact may be related to their omnivorous feeding behavior, being able to survive and

2007). Such characteristic makes *Acartia* species relatively easy to cultivate at small- and large-scale laboratory conditions, providing enough biomass as an alternative live food in marine aquaculture (Santos and Nosker 1997, McKinnon et al. 2003) and a classical model in toxicological tests (Bielmyer et al. 2007, Pedroso et al. 2007, Pinho et al. 2007).

The increased use of copepods as live food



of different sizes. The high nutritional value of copepods, which is characterized by a rich content in phospholipids, highly unsaturated fatty acids (HUFA), and natural antioxidants (Watanabe et al. 1983, Kraul et al. 1992, Sargent et al. 1997, Støttrup and Nosker 1997, Støttrup 2000, Helland et al. 2003), can also explain their actual broad use in aquaculture.

The success of a copepod culture depends on survival and fecundity rates of the cultivated animals, which are limited by food availability (Checkley 1980), temperature (Gaudy et al. 2000), and water salinity (Cardozo 2004). Egg production estimation is commonly used as an indicator of the nutritional quality of the food employed to feed the cultivated copepods (Butler and Dam 1994, Ceballos and Ianora 2003). Also, some studies have been performed to characterize the influence of different natural diets on copepod growth and egg production (Jakobsen et al. 2005, Kaminski and Montú 2005). All the information generated from these studies is important to select feeding regimes in order to improve copepod culture in laboratory (Støttrup and Jensen 1990, Castro-Longoria 2003, Cardozo 2004).

In light of the above, the aim of the present study was to evaluate the influence of the source and amount of food available on egg production in the copepod *A. tonsa*, using three different species of marine microalgae at different densities.

#### MATERIALS AND METHODS

The marine microalgae *Thalassiosira weissflogii* (Ø 13.2 µm), *Chaetoceros muelleri* (Ø 7.3 µm) and *Isochrysis galbana* (Ø 4.7 µm) were used in the present study as food source for copepods. They were grown in 5-L bottles, using the Guillard F/2 adapted method (Guillard and Ryther 1962).

Copepods used in the tests were isolated from field samples collected in the Patos Lagoon estuary (Rio Grande, RS, Southern Brazil) and kept under laboratory conditions for approximately 5 generations. The culture was carried out in non-toxic plastic tanks (70 L) containing saltwater (salinity 30). Tanks were kept in a experimental room with fixed temperature (20 ± 1°C) and photo-

For each feeding regime (food source and density) adult copepods (50 males and 50 females) were acclimated for 24 h in glass bottles (1 L) containing filtered (1 µm) saltwater (salinity 30) and the microalgae to be tested at the desired density. For each feeding regime, 6 different microalgae densities were tested, including one control condition where no food was added to the experimental medium. Therefore, 19 glass bottles were used, each one containing 100 copepods (50 males and 50 females). Bottles containing copepods and the experimental medium were kept under fixed temperature (20 ± 1°C) and photoperiod (12L:12D) and gently aerated to prevent algae sedimentation.

Algae densities were selected based on the saturation curve of egg production for *A. tonsa* fed on *Rhodomonas baltica* (Kjørboe et al. 1985). These authors have shown that *A. tonsa* achieved a stabilization on egg production at ~ 500 µg C.L<sup>-1</sup>. Considering the amount of carbon and the volume of *R. baltica*, the equivalent in biovolume required to achieve ~ 500 µg C.L<sup>-1</sup> would be 20.10<sup>3</sup>, 280.10<sup>3</sup> and 320.10<sup>3</sup> cells.mL<sup>-1</sup> for *T. weissflogii*, *C. muelleri* and *I. galbana*, respectively. The other 5 algae densities were selected to bracket that giving ~ 500 µg C.L<sup>-1</sup> and were calculated based on the *T. weissflogii* densities (Table I), using the equivalent biovolume approach (Hillebrand et al. 1999).

TABLE I  
Algal densities used in the experiments to measure egg production in the copepod *Acartia tonsa*.  
Algae densities are equivalent in biovolume.

Densities (10 <sup>3</sup> cells.mL <sup>-1</sup> )		
<i>Thalassiosira weissflogii</i>	<i>Chaetoceros muelleri</i>	<i>Isochrysis galbana</i>
0	0	0
2.5	35	40
5	70	80
10	140	160
20	280	320
40	560	640
60	840	960

The first 24-h period of test was used to acclimate



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domly collected and transferred to 300-mL glass bottles (10 female copepods per bottle) containing the corresponding experimental medium. Therefore, each experimental condition (food source and density) was tested in triplicate, totalizing 63 experimental units. Female copepods were then kept for a further 24 h under the same experimental conditions (feeding regime, salinity 30, temperature  $20 \pm 1^\circ\text{C}$ , and photoperiod 12L:12D). Experimental medium were gently aerated to prevent algae sedimentation. Every 4 h, bottles were carefully agitated to keep algae cells in suspension.

After the second 24-h period of test, the experimental medium from each bottle was filtered (20- $\mu\text{m}$  mesh filter). Eggs retained in the filter were transferred to 20-mL bottles, fixed in 4% formaldehyde solution, and counted in squared Petri dishes under a stereomicroscope. Egg production was expressed as  $\text{eggs.female}^{-1}.\text{day}^{-1}$ , considering the number of living females after the 48-h period of test.

Data from each experimental condition were expressed as mean  $\pm$  standard error ( $n = 3$ ). Considering the facts that food availability is a natural factor limiting *A. tonsa* egg production and that the relationship between egg production and food quantity measured as algal density follows a saturation curve (e.g. Berggreen et al. 1988), the maximum egg production for each microalgae was determined through non-linear regression analysis (exponential with saturation), using SigmaPlot 2001 for Windows version 7.0 (SPSS Inc., USA).

#### RESULTS

For the three microalgae species used, mean egg production rate was dependent on the density of food supplied to copepods (Fig. 1). Mean egg production in copepods fed on *T. weissflogii*, *C. muelleri* and *I. galbana* increased as the algae cell density increased, until reaching  $10.10^3$ ,  $140.10^3$  and  $640.10^3$   $\text{cells.mL}^{-1}$ , respectively. In all cases, egg production did not change at higher cell densities. In fact, data obtained fitted well to a saturation-type kinetic model, especially for *T. weissflogii* and *C. muelleri*, where very high regression coefficients were observed ( $R^2 = 0.99$  and  $0.95$ , respec-

This lower  $R^2$  value would be associated with an outlier mean value observed at the density of 1  $\text{cells.mL}^{-1}$  (Fig. 1).

Copepod egg production was also dependent on the food source provided to animals tested (Fig. 1). On the data regression model employed, the maximum mean egg production obtained was  $28.0 \pm 0.5$ ,  $3.5$ , and  $20.1 \pm 1.0$   $\text{eggs.female}^{-1}.\text{day}^{-1}$  with *T. weissflogii*, *I. galbana* and *C. muelleri*.

#### DISCUSSION

Data from the present study shows an influence of microalgae species and density on the egg production of the copepod *Acartia tonsa*. At low microalgae density, a low egg production was observed for the three microalgae tested. This fact could be explained by a decrease in the efficiency of food capture at low microalgae density, as observed in *A. tonsa* fed with *T. weissflogii* (Richardson and Stearns 1988). At high microalgae density, egg production became stable was obtained for the three microalgae tested. Considering the equivalent volume approach (Hillebrand et al. 1999), density of *T. weissflogii* and *C. muelleri* showing stabilization of egg production corresponded to half of that using *Monas baltica* ( $\sim 500 \mu\text{g C.L}^{-1}$ ). On the other hand, it corresponded to double for *I. galbana* (Kjørboe et al. 1985). These findings suggest that different microalgae concentrations are required to obtain stabilization of egg production in *A. tonsa* when different food sources are employed.

For the three microalgae species, mean values at saturation in egg production ( $20$  to  $28$   $\text{eggs.female}^{-1}.\text{day}^{-1}$ ) were always higher than those measured in the field ( $1$  to  $16$   $\text{eggs.female}^{-1}.\text{day}^{-1}$ ) for *A. tonsa* (Kleppel et al. 1998, Kleppel and Hazzard 2000). For copepods fed on the same algae (*T. weissflogii*) under controlled conditions, the mean value observed for *A. tonsa* in the present study ( $28$   $\text{eggs.female}^{-1}.\text{day}^{-1}$ ) was only higher than that reported for *A. clausi* ( $21$  to  $22$   $\text{eggs.female}^{-1}.\text{day}^{-1}$ ) by Richardson and Verheye (1999).

*A. tonsa* showed a higher egg production when fed on *T. weissflogii* than on *C. muelleri* or *I. galbana*.

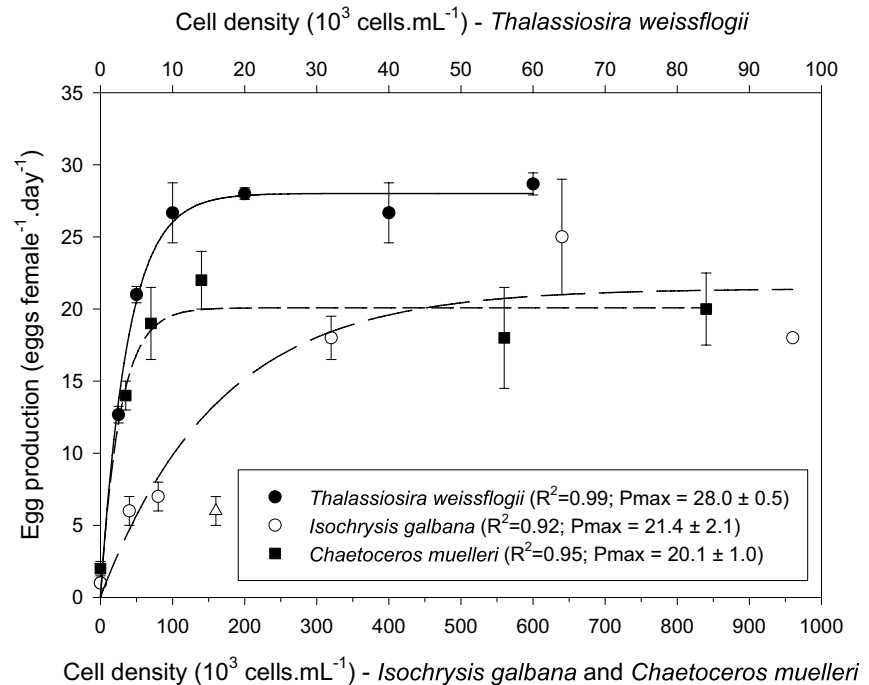


Fig. 1 – Egg production in the copepod *Acartia tonsa* using three different food sources: the microalgae *Thalassiosira weissflogii* (closed circles; solid line; top x axis), *Chaetoceros muelleri* (closed squares; short dashed line; bottom x axis) and *Isochrysis galbana* (open circles; long dashed line; bottom x axis). Different cell densities were tested for each microalgae species to give an equivalent biovolume value at each experimental condition. Copepods were acclimated to fixed salinity (30), temperature (20°C) and photoperiod (12L:12D) before tests. Tests were performed under the same acclimation conditions. Results are expressed as mean  $\pm$  standard error. Data were analyzed by non-linear regression analysis (exponential with saturation). Pm = maximum egg production expressed as eggs.female<sup>-1</sup>.day<sup>-1</sup>. R<sup>2</sup> = regression coefficient. Egg production data with *I. galbana* at 160.10<sup>3</sup> cells.mL<sup>-1</sup> (open triangle) was considered as an outlier.

was needed. These findings could be explained by the reduced food capture efficiency in *A. tonsa* when particles are between 2 and 4  $\mu$ m (Irigoien et al. 2003, Katechakis et al. 2004). In this context, it is important to emphasize that *I. galbana* has a smaller size ( $\sim 5 \mu$ m) than *T. weissflogii* ( $\sim 13 \mu$ m) and *C. muelleri* (5-10  $\mu$ m) (Berggreen et al. 1988). According to the size preference for food capture, *T. weissflogii* shows a more ideal size ( $\sim 13 \mu$ m) for the adult stage of the copepod *A. tonsa* (Berggreen et al. 1988). Furthermore, *A. tonsa* can have low sensitive chemoreception mechanisms (Paffenhofers and Stearns 1988), requiring a higher biomass of smaller algae (e.g. *I. galbana*) than that of larger ones (e.g. *T.*

*weissflogii*). It is important to note that the sloppy feeding was not considered when comparing feeding efficiency among the different microalgae tested. According to Møller and Nielsen (2001), *A. tonsa* does not show sloppy feeding when capturing algae of sizes similar to those employed in the present study ( $\sim 10 \mu$ m or less).

Ismar et al. (2008) reported that both *T. weissflogii* and *C. muelleri* allow the development of all life stages of *A. tonsa*. In the present study, *A. tonsa* fed on *T. weissflogii* and *C. muelleri* reached a saturated egg production at concentrations equivalent in biovolume. However, copepods fed on *T. weissflogii* produced in average 40% more eggs (28 eggs.female<sup>-1</sup>.day<sup>-1</sup>) than



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higher fecundity observed with *T. weissflogii* suggests a preference for use of this microalgae under laboratory conditions. The fact that higher survival rates and better physiological responses and development of various stages of copepods are observed with *T. weissflogii* than with *I. galbana* under controlled conditions (Tirelli and Mayzaud 2005, Ismar et al. 2008, Koski et al. 2008) also supports this choice.

Despite the fact that a maximum egg production was obtained in the present study with a single food source, especially for *T. weissflogii*, a combination of at least two microalgae species could enhance *A. tonsa* egg production. For example, a higher egg production (32 eggs.female<sup>-1</sup>.day<sup>-1</sup>) was obtained in *A. tonsa* fed with a mixture of *I. galbana* and *Rhinomonas reticulata* (Medina and Barata 2004) than in those fed only with *I. galbana* (22 eggs.female<sup>-1</sup>.day<sup>-1</sup>) at an equivalent biovolume concentration. In other copepod species, better results were also obtained with pluralalgal diets (Buttino et al. 2009).

In summary, data reported in the present study indicate that the feeding regime (microalgae species and density) influences egg production in the copepod *Acartia tonsa*. Differences in egg production with the different feeding regimes tested in the present study were explained considering the different sizes of the microalgae employed to feed the copepods.

#### RESUMO

A produção de ovos do copépode *Acartia tonsa* foi avaliada utilizando diferentes densidades das microalgas *Thalassiosira weissflogii*, *Chaetoceros muelleri* e *Isochrysis galbana*. Machos e fêmeas foram colocados sob condições controladas (salinidade 30, 20°C, fotoperíodo 12L:12D), aclimatados às condições experimentais e mantidos juntos por 24 h para permitir a copula. As densidades de algas foram equivalentes em biovolume e corresponderam a 0, 2,5, 5, 10, 20, 40 e 60,10<sup>3</sup> células.mL<sup>-1</sup> de *T. weissflogii*. Dez fêmeas aclimatadas foram separadas, transferidas para frascos de vidro e expostas por mais 24 h ao meio experimental correspondente. Após este período, os ovos foram fixados e contados. A produção de ovos alcançou um valor limiar quando *T. weissflogii*, *C. muelleri* e *I.*

ovos.fêmea<sup>-1</sup>.dia<sup>-1</sup>, respectivamente. Copépodes alimentados com *T. weissflogii* mostraram a maior produção média enquanto os alimentados com *I. galbana* alcançaram uma produção de ovo máxima quando as algas foram presentes a uma densidade de duas a quatro vezes maior, considerando o biovolume de *T. weissflogii* e *C. muelleri*. Estas diferenças podem ser explicadas considerando os diferentes tamanhos das microalgas utilizadas para alimentar os copépodes.

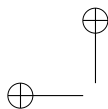
**Palavras-chave:** produção de ovos, *Acartia tonsa*, microalgas.

#### REFERENCES

- AZAITEIRO UM, MARQUES SC, VIEIRA LMR, RINHO MRD, PEREIRA PAB, RÉ MJ AND MOREIRA FMR. 2005. Dynamics of the *Acartia* genus (Copepoda) in a temperate shallow estuary (the Mondego estuary) on the west coast of Portugal. *Acta Oecologica* 26: 7–20.
- BERGGREEN U, HANSEN B AND KIØRBOE T. 1983. Food size spectra, ingestion and growth of the copepod *Acartia tonsa* during development: implications for determination of copepod production. *Mar Biol* 99: 341–352.
- BIELMYER GK, GROSEL M AND BRIX KV. 2006. Effects of silver, zinc, copper, and nickel to the copepod *Acartia tonsa* exposed via a phytoplankton diet. *Environmental Technology* 40: 2063–2068.
- BJÖRNBERG TSK. 1981. Copepoda. In: BOLTOVSKOY D (Ed), *Atlas del zooplankton del Atlantico Sudoccidental y Metodos de Trabajos con el Zooplankton Marino*. CEP, Mar del Plata, Argentina, p. 587–677.
- BUTLER M AND DAM HG. 1994. Production rates and characteristics of fecal pellets of the copepod *Acartia tonsa* under simulated phytoplankton bloom conditions: implications for vertical fluxes. *Mar Ecol Prog Ser* 114: 1–10.
- BUTTINO I, IANORA A, BUONO S, VITELLO V, SACCONE G AND MIRALTO A. 2009. Are monoalgal diets sufficient to pluralalgal diets to maximize cultivation of the copepod *Temora stylifera*? *Mar Biol* 156: 1171–1180.
- CARDOZO AP. 2004. Influência de diferentes salinidades na reprodução e crescimento de *Acartia tonsa* (Copepoda, Calanoida). Disponível em: <http://www.pluridoc.org>
- CASTRO-LONGORIA E. 2003. Egg Production and hatching success of four *Acartia* species under different



- the copepod *Temora stylifera*. J Exp Mar Biol Ecol 294: 189–202.
- CHECKLEY DM. 1980. The egg production of a marine planktonic copepod in relation to its food supply: laboratory studies. Limnol Oceanogr 25: 430–446.
- DELBARE D, DHERT P AND LAVENS P. 1996. Zooplankton. In: MANUAL ON THE PRODUCTION AND USE OF LIVE FOOD FOR AQUACULTURE. FAO Fisheries Technical Paper, p. 252–281.
- GAUDY R, CERVETTO G AND PAGANO M. 2000. Comparison of the metabolism of *Acartia clausi* and *Acartia tonsa*: influence of temperature and salinity. J Exp Mar Biol Ecol 247: 51–65.
- GUILLARD RRL AND RYTHER JH. 1962. Studies of marine planktonic diatoms *Cyclotella nanna* (Hustedt) and *Detonula convolvacea* (Cleve). Gran Can J Microbiol 8: 229–239.
- HELLAND S, TERJESEN BF AND BERG L. 2003. Free amino acid and protein content in the planktonic copepod *Temora longicornis* compared to *Artemia franciscana*. Aquaculture 215: 213–228.
- HILLEBRAND H, DÜRSELEN CD, POLINGHER U AND ZOHARY T. 1999. Biovolume calculation for pelagic and benthic microalgae. J Phycol 35: 403–424.
- IRIGOIEN X, TITELMAN J AND HARRIS RP. 2003. Feeding of *Calanus finmarchicus* nauplii in the Irminger Sea. Mar Ecol Prog Ser 262: 193–200.
- ISMAR MH, HANSEN T AND SOMMER U. 2008. Effect of food concentration and type of diet on *Acartia* survival and naupliar development. Mar Biol 154: 335–343.
- JAKOBSEN HH, HALVORSEN E, HANSEN BW AND VISSER AW. 2005. Effects of prey motility and concentration on feeding in *Acartia tonsa* and *Temora longicornis*: the importance of feeding modes. J Plankton Res 27: 775–785.
- KAMINSKI SM AND MONTÚ MA. 2005. Produção de ovos dos copépodes costeiros *Acartia tonsa*, *Temora stylifera* e *Temora turbinata* da Praia do Cassino, Rio Grande, RS. Atlântica 27: 103–111.
- KATECHAKIS A, STIBOR H, SOMMER U AND HANSEN T. 2004. Feeding selectivities and food-niche separation of *Acartia clausi*, *Penilia avirostris* (Crustacea) and *Dolium denticulatum* (Thaliacea) in Blanes Bay (Catalan Sea, NW Mediterranean). J Plankton Res 26: 589–603.
- and composition of specific dynamic action. Mar Ecol Prog Ser 26: 85–97.
- KLEPPEL GS. 1992. Environmental regulation of feeding and egg production by *Acartia tonsa* off southern California. Mar Biol 112: 57–65.
- KLEPPEL GS AND HAZZARD SE. 2000. Diet and egg production of the copepod *Acartia tonsa* in Florida Bay. II. Role of the nutritional environment. Mar Biol 137: 111–121.
- KLEPPEL GS, BURKART CA, HOUCHIN L AND TOMAS C. 1998. Egg production of the copepod *Acartia tonsa* in Florida Bay during Summer. 1. The roles of food environment and diet. Estuaries 21: 328–339.
- KOSKI M, WICHARD T AND JÓNASDÓTTIR SH. 2008. “Good” and “bad” diatoms: development, growth and juvenile mortality of the copepod *Temora longicornis* on diatom diets. Mar Biol 154: 719–734.
- KRAUL S, NELSON A, BRITTAI K, AKO H AND OGASAWARA A. 1992. Evaluation of live feeds for larval and postlarval mahimahi *Coryphaena hippurus*. J World Aquacult Soc 23: 299–307.
- LEANDRO SM, MORGADO F, PEREIRA F AND QUEIROGA H. 2007. Temporal changes of abundance, biomass and production of copepod community in a shallow temperate estuary (Ria de Aveiro, Portugal). Est Coast Shelf Sci 74: 215–222.
- MAUCHLINE J. 1998. The Biology of Calanoid Copepods. Advances in Marine Biology. London, Academic Press, 710 p.
- MCKINNON AD, DUGGANA S, NICHOLS PD, RIMMER MA, SEMMENS G AND ROBINO B. 2003. The potential of tropical paracalanid copepods as live feeds in aquaculture. Aquaculture 223: 89–106.
- MEDINA M AND BARATA C. 2004. Static-renewal culture of *Acartia tonsa* (Copepoda: Calanoida) for ecotoxicological testing. Aquaculture 229: 203–213.
- MØLLER EF AND NIELSEN TG. 2001. Production of bacterial substrate by marine copepods: effect of phytoplankton biomass and cell size. J Plankton Res 23: 527–536.
- PAFFENHOFER GA AND STEARNS DE. 1988. Why is *Acartia tonsa* (Copepoda: Calanoida) restricted to nearshore environments? Mar Ecol Prog Ser 42: 33–38.
- PEDROSO MS, BERSANO JG AND BIANCHINI A. 2007.



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- PINHO GLL, PEDROSO MS, RODRIGUES SC, SOUZA SS AND BIANCHINI A. 2007. Physiological effects of copper in the euryhaline copepod *Acartia tonsa*: waterborne versus waterborne plus dietborne exposure. *Aquat Toxicol* 84: 62–70.
- RICHARDSON AJ AND VERHEYE HM. 1998. The relative importance of food and temperature to copepod egg production and somatic growth in the southern Benguela upwelling system. *J Plankton Res* 20: 2379–2399.
- SAIZ E, CALBET A, ATIENZA D AND ALCARAZ M. 2007. Feeding and production of zooplankton in the Catalan Sea (NW Mediterranean). *Progr Oceanog* 74: 313–328.
- SARGENT JR, MCEVOY LA AND BELL JG. 1997. Requirements, presentation and sources of polyunsaturated fatty acids in marine larval feeds. *Aquaculture* 155: 117–127.
- SCHIPP GR, BOSMANS JMP AND MARSHALL AJ. 1999. A method for hatchery culture of tropical calanoid copepods, *Acartia* spp. *Aquaculture* 174: 81–88.
- STØTTRUP JG. 2000. The elusive copepods: their production and suitability in marine aquaculture. *Aquacult Res* 31: 703–711.
- STØTTRUP JG AND JENSEN J. 1990. Influence of food quality on feeding and egg-production of the calanoid copepod *Acartia tonsa* Dana. *J Exp Mar Biol Ecol* 141: 87–100.
- STØTTRUP JG AND NOSKER NH. 1997. Production and survival of copepods in marine fish larviculture. *Aquaculture* 155: 231–247.
- TIRELLI V AND MAYZAUD P. 2005. Relationship between feeding rate, functional response and gut transit time in the copepod *Acartia clausi*: role of food quantity and quality. *J Plankton Res* 27: 557–568.
- UCHIMA M. 1988. Gut content analysis of neritic copepods *Acartia omorii* and *Oithona davisae* by a new method. *Mar Ecol Prog Ser* 48: 93–97.
- WATANABE T, KATAJIMA C AND FUJITA S. 1983. The nutritional value of live organisms used in Japan for marine fish propagation of fish: a review. *Aquaculture* 34: 115–127.
- WU CH, HWANG JS AND YANG JS. 2004. Diets of marine fish larvae (Poecilostomatoida) in the Southern Ocean. *Strait Zool Stud* 43: 388–392.