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## Bacterioplankton abundance, biomass and production in a Brazilian coastal lagoon and in two German lakes

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### ABSTRACT

The bacterioplanktonic abundance, biomass, and production within a tropical lagoon (Cabiúnas, Brazil) and two temperate lakes (Stechlin and Dagow, Germany) were compared. Bacterial abundance and production were significantly different among the three water bodies. The lowest bacterial production ( $0.8 \mu\text{g C l}^{-1} \text{ d}^{-1}$ ) was observed in the tropical Cabiúnas Lagoon despite its higher mean temperature and dissolved organic carbon concentration. Highest bacterioplankton abundance ( $2.6 \times 10^9 \text{ cells l}^{-1}$ ) and production ( $68.5 \mu\text{g C l}^{-1} \text{ d}^{-1}$ ) were measured in eutrophic Lake Dagow. In oligotrophic Lake Stechlin, the lowest bacterial biomass ( $48.05 \mu\text{g C l}^{-1}$ ) was observed because of lower bacterial biovolume ( $0.248 \mu\text{m}^3$ ) and lower bacterial abundance. Bacterial populations in the temperate lakes show higher activity (production/biomass ratio) than in the tropical lagoon. The meaning of isotopic dilution and leucine incorporation by non-bacterial micro-organisms were evaluated in the oligotrophic temperate system. Leucine uptake by non-bacterial micro-organisms did not have significant influence on bacterial production.

**Key words:** bacterial production, leucine incorporation, tropical lagoon, temperate lake.

### INTRODUCTION

Tropical aquatic environments are very different from temperate environments, influencing the type of organisms and their metabolic processes in these different environments. For instance, tropical freshwater communities are mainly regulated by variations in the hydrological conditions rather than by temperature (Junk & Furch 1993, Anésio et al. 1997, Gomes & Agostinho 1997). As the temperature varies seasonally in temperate regions, primary and secondary production reaches a maximum during the warm months (Lampert & Sommer 1993).

Furthermore, Hecky et al. (1993) suggested that particulate matter in temperate freshwater ecosystems frequently exhibits low P concentration, while in tropical aquatic systems either N or P may be limiting or even non-limiting. Many tropical water bodies are warm and shallow, and thus remain polymictic. These latter water bodies often have large littoral zones colonized by macrophytes. In these water ecosystems, eutrophication enhances the growth of macrophytes in contrast to the over-growth of phytoplankton in temperate waters (Oskam & van Genderen 1996), where the principal source of DOM and POM comes from phytoplankton or allochthonic inputs.

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Bacterial abundance, chlorophyll *a* (which reflects the trophic state) and temperature have been described as the most important factors influencing bacterial production in marine and freshwater aquatic habitats (White et al. 1991). But in tropical ecosystems, since the temperature is relatively constant, it may be conjectured that the natural dynamics of bacterial production and density is controlled by others factors such as nutrient availability or interaction between micro-organisms.

The principal objective of this study was directly to compare the influence of temperature and trophic state on abundance, biomass and production of heterotrophic bacteria in the water column and their spatial variability within a tropical lagoon and two temperate lakes. The tropical Cabiúnas Lagoon (Brazil) and the temperate Lakes Stechlin (oligotrophic) and Dagow (eutrophic), both in Germany, were used for comparison. Pronounced differences in temperature, trophic state, and morphological characteristics were observed in these environments and described in this study.

In addition, earlier research describing problems using labeled markers (as isotopic dilution; Kirchman 1993) and uptake by micro-organisms other than bacteria (van Looij & Riemann 1993, Kamjunke & Jähnichen 2000), were proofed with samples from Lake Stechlin in this report.

#### MATERIAL AND METHODS

Located on the coastal zone of Rio de Janeiro State (Brazil), Cabiúnas Lagoon was formed during the last Holocene transgression that occurred in the Pleistocene (Perrin 1984). It is separated from the Atlantic Ocean by a 100m sand bar. Its water is colored by humic substances and has an average salinity of about 0.3. The littoral zone is principally colonized by the macrophytes *Typha domingensis* and *Potamogeton stenostachys*. The climate is tropical wet and the long-term average minimum temperature is 18.7°C.

The lagoon is situated in a sand-dune habitat (Restinga) and surrounded by dense vegetation

formed mainly by representatives of the Theopharastaceae, Anarcadiaceae, Myrtaceae, Clusiaceae, Ericaceae, Melastomataceae and Bignoniaceae (Araújo & Henriques 1984). Panosso et al. (1998) described the morphology in detail and Petrucio (1998) reported the limnological characteristics. The oligotrophic Lake Stechlin and the eutrophic Lake Dagow are both situated in the Baltic Lake district in Northeast Germany. Both lakes are of glacial origin and the shores are forested mainly by *Pinus sylvestris* and *Fagus sylvatica*. Both lakes were described by Casper (1985). In contrast to Cabiúnas Lagoon, the lakes present typical stratification during summer. Some morphological and limnological characteristics of all three sampling sites are summarized in Table I.

Water samples were collected in Lakes Stechlin and Dagow in February 1998 and in Cabiúnas Lagoon in September 1998. Samples were taken in each aquatic environment at three stations along a transect. In all cases, station L1 was located in a littoral zone without macrophytes, station L2 in a littoral zone colonized by macrophytes, and station P in the pelagic zone. At the pelagic zone, samples were collected at three depths: near the water surface (Pa), at half water depth (Pb) and near the bottom (Pc). In Cabiúnas Lagoon, the near bottom samples were taken at 3.5m. In Lakes Stechlin and Dagow the near bottom subsamples were taken at 30m and 9m, respectively.

In the German lakes, temperature and oxygen profiles were measured using a Microprocessor Oximeter Oxi 196 (WTW, Germany) and N and P-concentrations by a FIA-5010 Analyser (Tecator, Sweden). In the Brazilian lagoon, temperature was measured using a Cole Parmer thermistor Model 8402-10 and oxygen concentration by the Winkler method. N and P analysis were done using the Kjeldahl method (Golterman et al. 1978). Dissolved organic and inorganic carbon concentrations were determined for all sites using a TOC-5050 Analyser (Shimadzu, Japan).

Bacterial abundances were determined after fixing with formalin (4% v/v). Subsamples (n=3)

TABLE I

**Morphological and limnological characteristics of Cabiúnas Lagoon, Lake Stechlin and Lake Dagow.**

	area km <sup>2</sup>	depth meter		water temperature °C		pH		conductivity μS cm <sup>-1</sup>	
		max.	mean	min.	max.	min.	max.	min.	max.
Cabiúnas Lagoon	0.34 <sup>1</sup>	4.0	2.4 <sup>1</sup>	21.0	28.5 <sup>3</sup>	6.0	7.4 <sup>3</sup>	300	14.300
L. Stechlin	4.3 <sup>2</sup>	68.0	22.8 <sup>2</sup>	3.7	19.9 <sup>5</sup>	7.5	8.6 <sup>5</sup>	246	336 <sup>5</sup>
L. Dagow	0.3 <sup>2</sup>	9.5	4.9 <sup>2</sup>	1.7	21.5 <sup>6</sup>	7.1	9.1 <sup>6</sup>	360	500 <sup>6</sup>
	volume × 10 <sup>6</sup> m <sup>3</sup>	transparency Secchi disk meter		oxygen concentration mg l <sup>-1</sup>		N-total mg l <sup>-1</sup>		P-total μg l <sup>-1</sup>	
		min.	max.	min.	max.	min.	max.	min.	max.
Cabiúnas Lagoon	1.4 <sup>1</sup>	0.8	2.8 <sup>3</sup>	2.6	7.7 <sup>4</sup>	0.27	1.07 <sup>4</sup>	4.4	24.0 <sup>4</sup>
L. Stechlin	96.9 <sup>2</sup>	7.2	12.3 <sup>5</sup>	5.3	12.5 <sup>5</sup>	0.34	1.2 <sup>5</sup>	6.0	15.0 <sup>5</sup>
L. Dagow	1.2 <sup>2</sup>	1.8	3.2 <sup>6</sup>	0.0	11.8 <sup>6</sup>	0.77	2.37 <sup>6</sup>	23.0	199.0 <sup>6</sup>

<sup>1</sup>Panosso et al. (1998). <sup>2</sup>Casper (1985). <sup>3</sup>data from 1993 to 1995, Petrucio (1998). <sup>4</sup>data from 1993 to 1995, Marinho (pers. comm.). <sup>5</sup>data from 1998 from 0-10 m, Koschel (pers. comm.). <sup>6</sup>data from 1996 from 0-8 m, Koschel (pers. comm.).

were stained with the fluorochrome 4',6-diamidino-2-phenylindole (DAPI, 5 mg l<sup>-1</sup>) for 10 minutes in the dark and filtered onto black polycarbonate membrane filters (0.2 μm; Nuclepore Corp.) under low vacuum pressure < 30 mm Hg (Kepner & Pratt 1994). Bacterial cells were counted using a fluorescence microscope (Leica DMBR, HBO 50 W, BP 355-425, RKP 455 and LP 470) with a magnification of 2000 × under immersion. Ten randomly selected microscopic fields or a minimum of 200 bacteria per filter were counted.

The bacterioplankton production (BP) was measured according to Smith and Azam (1992). Samples (n=5) were incubated for 1 hour in the dark with L-[4,5-<sup>3</sup>H] leucine (specific activity 136 Ci mmol<sup>-1</sup>, 50 nM final concentration). Formalin (4% v/v) was added to controls prior to the isotope addition and to the samples directly after incubation to terminate reactions. Samples were shaken and then centrifuged at 14,000 × g for 10 minutes. The supernatant was aspirated and the remaining pellet was resuspended and washed with 5% TCA and 80% ethanol. To

evaluate the saturation level of leucine uptake, samples from Lake Stechlin were incubated as described with 20, 50, 100, 150 and 200 nM <sup>3</sup>H-leucine. In all experiments, labeled leucine was diluted three times with nonlabeled leucine. The samples were counted in a Packard 1600 TR liquid scintillation counter after adding a liquid scintillation cocktail (Ultima Gold, Packard Co.). The BP was calculated according to Simon and Azam (1989):

$$\text{BP(g)} = (\text{mol leucine incorporated}) \times (0.073) \times (131.2) \times (0.86) \times (\text{isotopic dilution}) \quad (1)$$

where 0.073 is the leucine/protein ratio, 131.2 is the molecular weight of leucine and 0.86 is the ratio of cellular carbon to protein. The isotope dilution was considered as 2.

To assess the effect of non-bacterial micro-organisms on the bacterial production, unfiltered and filtered water samples from Lake Stechlin (polycarbonate filters, 0.8 μm, Nuclepore Corp.) were incubated with leucine as described earlier.

To determine bacterial size, microphotographs (Kodak Elite II 100) of DAPI stained samples were

projected onto a screen, and a total of 50 bacteria were randomly measured per sample.

Bacterial biovolume was calculated from sized bacteria (Bratbak 1985):

$$\text{Bacterial biovolume} = (\pi/4) \times W^2 \times (L-W/3) \quad (2)$$

where L is length and W is width of the cells, applying this formula to cocci ( $L=W$ ).

Carbon content was estimated using the factor  $105 \text{ fg C } \mu\text{m}^{-3}$  (Theil-Nielsen & S ndergaard 1998).

#### RESULTS AND DISCUSSION

Some limnological parameters measured in the three water bodies at the time of sampling are summarized in Table II. Temperature and dissolved organic carbon were higher in Cabi nas Lagoon, in contrast to both German lakes. The lagoon is a typical coastal tropical lagoon that does not stratify and remains warm during the austral winter. Both German lakes were sampled at the end of winter. Each had turned over and exhibited higher concentrations of inorganic carbon than the lagoon. Oxygen was found to be saturated in all lakes; and nutrient concentrations were similar at all sites. The P- and N-concentrations indicate the humic Cabi nas Lagoon as an oligotrophic system.

The highest bacterial abundance (BA) was estimated in the eutrophic Lake Dagow, which varied from  $1.6 \times 10^9$  to  $2.6 \times 10^9$  cells  $\text{l}^{-1}$  (Fig. 1C). In Cabi nas Lagoon, bacterial abundance (BA) was lower than in Lake Dagow. BA varied from  $1.0 \times 10^9$  to  $1.5 \times 10^9$  cells  $\text{l}^{-1}$  (Fig. 1A) which is in the range of the most oligotrophic lakes (Chrzanowski et al. 1996, Lindell & Edling 1996, Reche et al. 1997). The lowest BA of the three systems examined was observed in the oligotrophic Lake Stechlin (Fig. 1B), varying from  $7.8 \times 10^8$  cells  $\text{l}^{-1}$  at station L1 to  $1.2 \times 10^9$  cells  $\text{l}^{-1}$  at station L2. Lake Stechlin was the only site where the bacterial abundance was significantly different among the stations (Kruskal-Wallis,  $p < 0.05$ ).

Bacterial production (BP) measured in the different systems ranged from 0.8 to  $68.5 \mu\text{g C l}^{-1} \text{ d}^{-1}$ .

The lowest bacterial production was detected in stations of Cabi nas Lagoon ( $0.8$  to  $4.3 \mu\text{g C l}^{-1} \text{ d}^{-1}$ , Fig. 1A) and Lake Stechlin ( $3.6$  to  $6.4 \mu\text{g C l}^{-1} \text{ d}^{-1}$ ). The highest BP was calculated for the littoral zone colonized by macrophytes (L2) in Lake Dagow ( $68.5 \mu\text{g C l}^{-1} \text{ d}^{-1}$ ).

Bacterial production was not significantly different among the stations (L1, P and L2) for all studied ecosystems (Kruskal-Wallis,  $p < 0.05$ ), but comparing the three water bodies, both BA and BP were significantly different (Kruskal-Wallis,  $p < 0.05$ ).

Values of BP measured in February in the temperate lakes were higher than those previously measured by Babenzien and Babenzien (1985) based on  $^{14}\text{C}$ -glucose uptake. In this study bacterial production in Lake Stechlin ranged from  $0.024$  to  $1.4 \mu\text{g C l}^{-1} \text{ d}^{-1}$  and in Lake Dagow  $0.24$  to  $13.2 \mu\text{g C l}^{-1} \text{ d}^{-1}$  over the course of a year.

Even though Cabi nas Lagoon exhibited higher mean temperature and DOC (Table II), the lowest bacterial carbon production was measured in this lagoon. Bacterial production observed in this lagoon was comparable to those of temperate lakes during autumn (Tulonen 1993) and lower than observed in other Brazilian aquatic ecosystems such as the Patos Lagoon (Abreu et al. 1995) or the Amazonian Lake Batata (An sio et al. 1997). The higher content of humic substances in this brown-water tropical lagoon may inhibit decomposition (Qualls & Haines 1990). The concentration of labile DOC (considered here as soluble carbohydrates) in Cabi nas Lagoon represented less than 7% of total DOC (Marinho, pers. comm.). Similar dissolved labile carbon and DOC concentrations were observed in temperate eutrophic lakes (S ndergaard et al. 1995). However, the BP observed by these latter authors were higher than in Cabi nas Lagoon, indicating that only the concentration of labile DOC alone cannot explain the low BP. Apparently, the inter-relationship between temperature and nutrient availability may better explain bacterial production than either of these factors alone (Felip et al. 1996). Nutrient availability did not seem to limit BP in

TABLE II

Measurements of water temperature, dissolved oxygen, dissolved organic carbon (DOC) and inorganic (DIC) and nutrients in Cabiúnas Lagoon on September 1998 and in Lake Stechlin and Lake Dagow on February 1997. The values were recorded on the day of sampling.

	Water temperature °C	Oxygen mg l <sup>-1</sup>	DOC mg l <sup>-1</sup>	DIC mg l <sup>-1</sup>	P-total μg l <sup>-1</sup>	SRP μg l <sup>-1</sup>	N-total μg l <sup>-1</sup>	NO <sub>3</sub> μg l <sup>-1</sup>	NH <sub>4</sub> μg l <sup>-1</sup>
Cabiúnas Lagoon									
L1	26.1	5.9	11.9	1.6	9	4	500	5	2
P <sup>1</sup>	26.3	6.7	12.4	1.8	18	2	590	14	14
L2	26.8	3.5	8.6	1.5	18	3	590	3	3
Lake Stechlin									
L1	4.0	12.1	3.9	21.7	32	3	298	14	29
P <sup>1</sup>	3.8	12.8	3.2	21.6	15	11	333	22	16
L2	4.0	12.0	7.6	21.7	6	9	305	12	6
Lake Dagow									
L1	4.8	12.9	8.62	22.0	14	4	782	98	23
P <sup>1</sup>	4.5	13.7	8.42	22.2	15	4	850	78	30
L2	4.9	13.6	8.02	23.5	16	4	770	84	24

<sup>1</sup> average of pelagic samples a, b and c. <sup>2</sup> data from samples collected in August 1998.

Cabiúnas Lagoon (see Table I and II). In this lagoon, the production attributed to picoplankton ( $< 1\mu\text{m}$ ) represents about 35% of the total planktonic primary production. Picoplankton also exhibit a high rate of carbon excretion, which can support a higher rate of bacterial production (Roland 1998). In addition, the concentration of total phosphorus in the lagoon was similar to that found in Lake Stechlin. Thus, the lagoon has reduced bacterial activity despite high temperature and nutrient availability, suggesting that the microbial community is regulated by other mechanisms.

In the small bay of the tropical lagoon, which is colonized by both emergent and submersed macrophytes, BP was statistically higher and varied from 1 to  $33\mu\text{g C l}^{-1} \text{ d}^{-1}$  (Mann-Whitney,  $p < 0.05$ ; Furtado et al. 2000). This result indicates that the littoral zones are sites of greater microbial activity than the principal water body.

In Table III, bacterial biovolume, carbon content and production-biomass ratio (production/biomass) of water samples from all water bodies are summarized. The lowest bacterial biovolume

( $0.25\mu\text{m}^3$ ) was measured in Lake Stechlin, therefore the carbon content ( $26.04 \text{ fg C cell}^{-1}$ ) and bacterial biomass ( $26.04\mu\text{g C l}^{-1}$ ) were low. The bacterial biovolume was generally higher than observed in most temperate (Tulonen 1993, Tulonen et al. 1996, Hwang & Heath 1997) and tropical water bodies (Kroer 1994). Bacterial biovolume and corresponding carbon content in Cabiúnas Lagoon and Lake Dagow were similar (Mann-Whitney,  $p < 0.05$ ), but the bacterial biovolume and carbon content were significantly different among the stations (Kruskal-Wallis,  $p < 0.05$ ). The low bacterial biovolume of cells from Lake Stechlin was responsible for this result suggesting that biovolume was more related to trophic state than activity. Also, the production/biomass ratio has the same tendency as production, with the lowest P/B ratio in Cabiúnas Lagoon ( $0.03 \text{ d}^{-1}$ ) and the highest in Lake Dagow ( $0.78 \text{ d}^{-1}$ ). Bacterial metabolism was higher in both temperate lakes than in the tropical lagoon.

To ensure that our measurements of BP were not effected by leucine uptake by other micro-organisms and to reduce the effect of isotopic dilu-

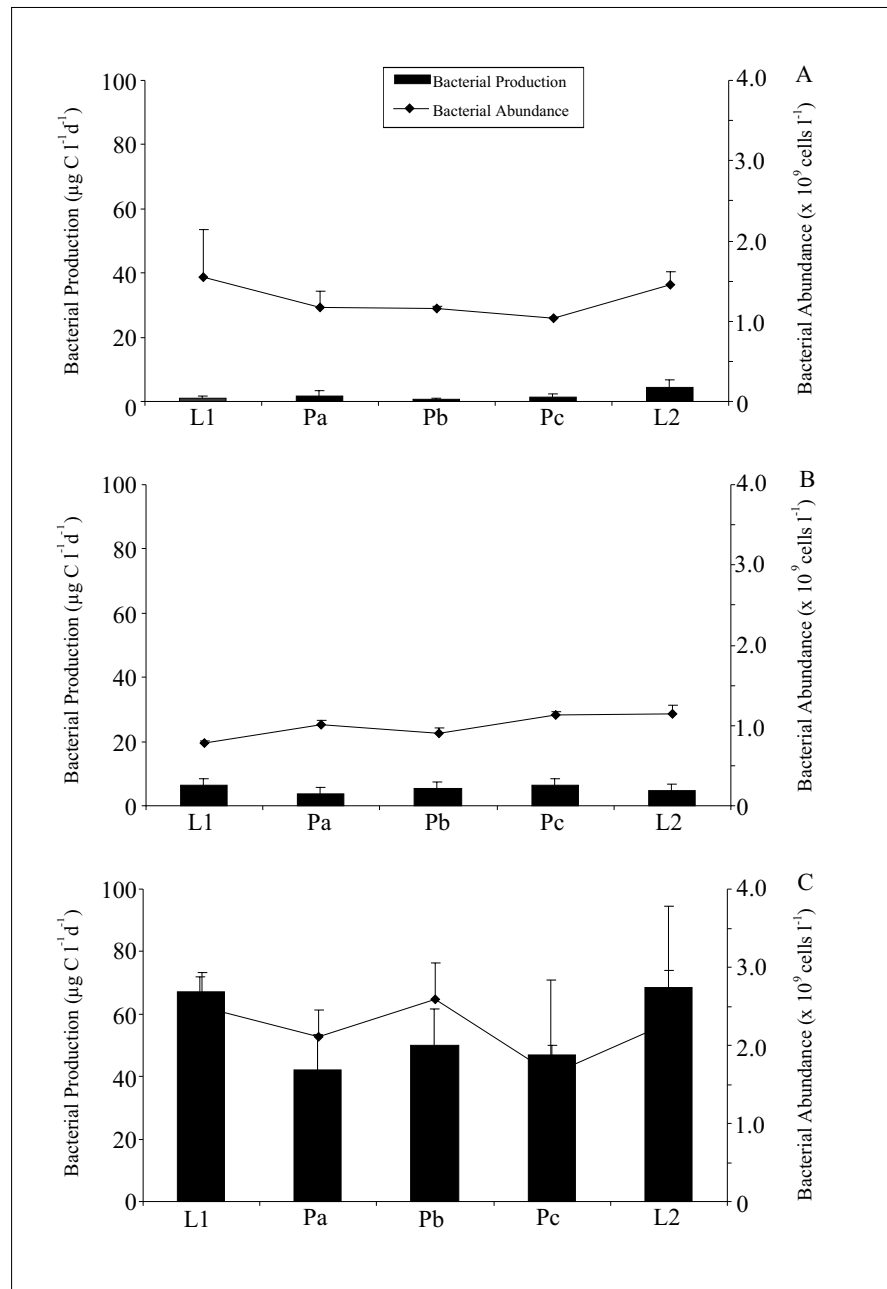


Fig. 1 – Bacterial production and abundance in samples from Cabiúnas Lagoon (A), Lake Stechlin (B) and Lake Dagow (C). L1 – littoral without macrophytes, Pa – pelagial surface, Pb – pelagial intermediate, Pc – pelagial bottom and L2 – littoral with macrophytes. Vertical bars represent the average + standard deviation.

TABLE III

**Bacterial biovolume, carbon content, abundance, biomass, production and P/B ratio in water samples from Cabiúnas Lagoon, Lake Stechlin and Lake Dagow. The values represent the average of the three stations (L1, L2 and P)  $\pm$  standard deviation.**

	Volumen <sup>1</sup> $\mu\text{m}^3$	C content <sup>2</sup> fg C cell <sup>-1</sup>	Bacterial abundance cell l <sup>-1</sup> $\times 10^9$	Bacterial biomass $\mu\text{g C l}^{-1}$	Bacterial production $\mu\text{g C l}^{-1} \text{d}^{-1}$	P/B ratio <sup>3</sup> d <sup>-1</sup>
Cabiúnas Lagoon	0.352 $\pm$ 0.209	36.96	1.3	48.05	1.22	0.03
Lake Stechlin	0.248 $\pm$ 0.268	26.04	1.0	26.04	5.30	0.2
Lake Dagow	0.304 $\pm$ 0.171	31.92	2.2	70.22	54.88	0.78

<sup>1</sup>calculated according to Bratbak (1985). <sup>2</sup>according to Theil-Nielsen and S ndergaard (1998). <sup>3</sup>Production/Biomass ratio.

tion, we refined our leucine uptake experiments. We determined the saturation concentration for leucine in water samples from Lake Stechlin. Leucine uptake increased linearly up to 200 nM leucine ( $r^2 = 0.9062$ ), following the expression (Fig. 1):

$$Y = 0.0393 X \quad (3)$$

where Y is the leucine rate uptake ( $\text{nmol l}^{-1} \text{h}^{-1}$ ) and X is the concentration of leucine (nM).

In these experiments, isotopic saturation was not reached. Usually, the leucine saturation level is found at a concentration of 10 nM in waters of oligotrophic and marine systems (Kirchman 1993). At this concentration, variation does not occur in the bacterial growth rate and minimizes intracellular isotopic dilution (Simon & Azam 1989). In contrast, higher leucine concentration ( $> 200\text{nM}$ ) has been recommended for eutrophic environments (van Looij & Riemann 1993) to reduce both the effect of the intracellular *de novo* synthesis, and external dilution (J rgensen 1992). Also, according to Kirchman et al. (1986) the addition of low concentrations of extracellular leucine are enough to inhibit intracellular synthesis. Riemann and Azam (1992), using leucine concentrations up to 100 nM, could not find saturation levels in 6 of 7 investigated water samples for several coastal marine and freshwater ecosystems in Denmark.

A concentration of 50 nM leucine was used throughout our experiments. A leucine concentration of  $> 40\text{nM}$  is noted to reduce the effect of intracellular dilution by about 90% in eutrophic lakes (J rgensen 1992). Additionally, leucine uptake by non-bacterial micro-organisms was minimized in Lake Stechlin samples, when incubated with 50 nM. The bacterial production varied from  $5.0\mu\text{g C l}^{-1} \text{d}^{-1}$  in the filtered samples ( $< 0.8\mu\text{m}$ ) to  $6.4\mu\text{g C l}^{-1} \text{d}^{-1}$  in unfiltered samples. The difference was not statistically significant (Mann-Whitney,  $p < 0.05$ ; Fig. 2). The concentration of 50 nM leucine was enough to saturate samples from Cabi nas Lagoon minimizing isotopic dilution. Farjalla (1998), using a concentration of 10 nM leucine in the same lagoon, observed higher values for bacterial production than measured in this investigation.

## CONCLUSIONS

Despite the lower water temperatures measured on the winter sampling dates, both temperate lakes exhibited higher bacterial production and consequently a higher P/B ratio (production/bacterial biomass). Bacterial production was not coupled with abundance, because average bacterial production in Lake Stechlin was about four times higher than in Cabi nas Lagoon, but bacterial abundance in Lake Stechlin was significantly lower. Highest



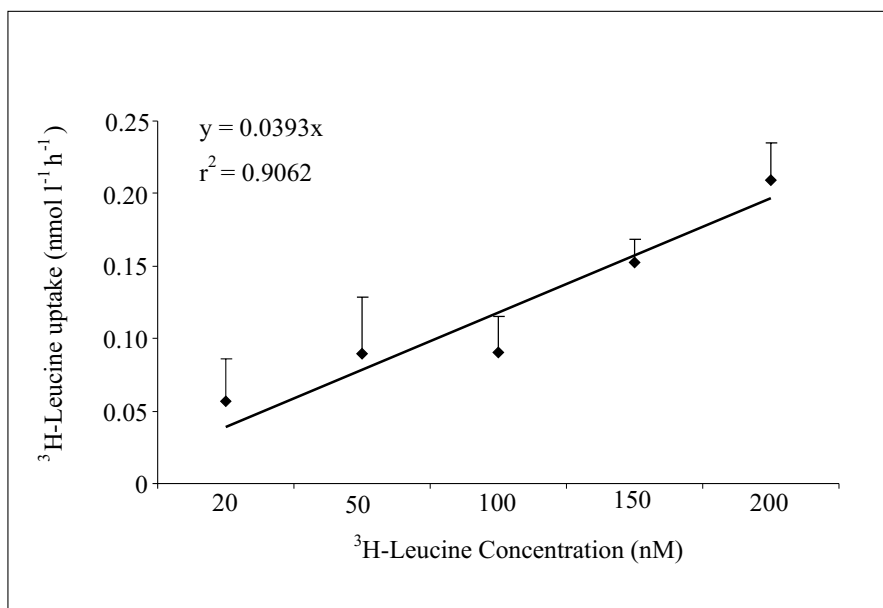


Fig. 2 – Leucine uptake versus leucine concentration in water samples from Lake Stechlin. Vertical bars represent the average ( $n=5$ ) + standard deviation.

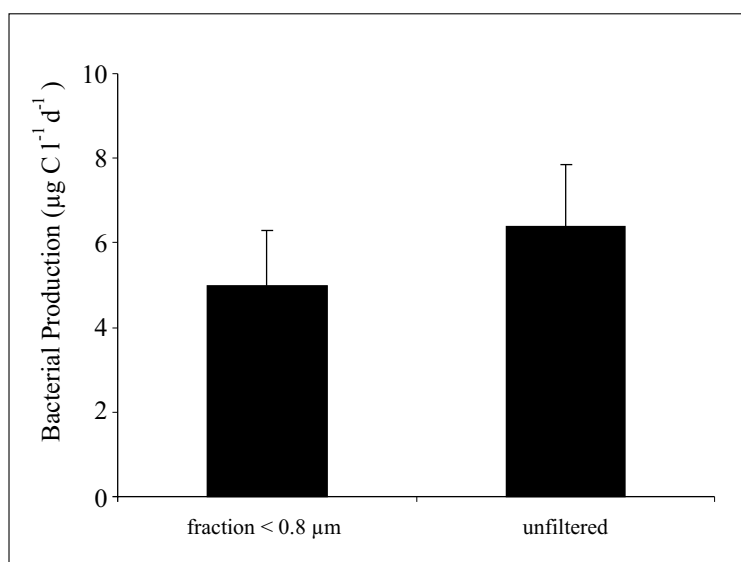


Fig. 3 – Bacterial production in water samples from Lake Stechlin at  $< 0.8 \mu\text{m}$  and unfiltered sample. Vertical bars represent the average ( $n=5$ ) + standard deviation.

bacterial abundance and production were detected in eutrophic Lake Dagow. In this study, trophic state was determined to be most important factor for increasing bacterial production instead of temperature.

The microbial activity in tropical Cabiúnas Lagoon was not regulated by temperature and N- and P-concentrations, indicating that bacterial community is regulated by other more complex interactions, which limited microbial activity in the water column.

Leucine uptake did increase linearly up to 200 nM indicating that oligotrophic water bodies may show high values of bacterial production and measurements using low concentrations of leucine (10 nM) to measure BP may underestimate BP in some systems.

Finally, the leucine uptake by micro-organisms greater than  $> 0.8\mu\text{m}$  in size did not influence estimates of BP for oligotrophic Lake Stechlin.

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#### RESUMO

A abundância, biomassa e produção bacterioplânctônica em uma lagoa tropical (lagoa Cabiúnas, Brasil) e em dois lagos temperados (lago Stechlin e lago Dagow, Alemanha) foram comparadas. A abundância e a produção bacteriana foram significativamente diferente entre os três ecossistemas aquáticos. A menor produção bacteriana ( $0.8\mu\text{g C l}^{-1} \text{ d}^{-1}$ ) foi observada na lagoa Cabiúnas, apesar da alta

temperatura da água e concentração de carbono orgânico dissolvido. A maior abundância ( $2.6 \times 10^9$  células  $\text{l}^{-1}$ ) e produção bacterioplânctônica ( $68.5\mu\text{g C l}^{-1} \text{ d}^{-1}$ ) foram medidas no eutrófico lago Dagow. No oligotrófico lago Stechlin, foi observada a menor biomassa bacteriana ( $48.05\mu\text{g C l}^{-1}$ ), refletindo o menor volume ( $0.248\mu\text{m}^3$ ) e abundância bacteriana. Populações bacterianas nos lagos temperados mostraram maior atividade (razão produção/biomassa) que na lagoa tropical. O efeito da diluição isotópica e a incorporação de leucina por microorganismos não bacterianos foram avaliadas no ecossistema oligotrófico temperado. A absorção de leucina por microorganismos não bacterianos não influenciou significativamente a produção bacteriana.

**Palavras-chave:** produção bacteriana, incorporação de leucina, lagoa tropical, lagos temperados.

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