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New enamine derivatives of lapachol and biological activity

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ABSTRACT

A convenient synthesis of the new enamine derivatives 2-(4-morpholinyl)-3-(3-methyl-2-butenyl)-1,4-naphthalenedione, 2-(1-piperidinyl)-3-(3-methyl-2-butenyl)-1,4-naphthalenedione and 2-(1-pyrrolidinyl)-3-(3-methyl-2-butenyl)-1,4-naphthalenedione was carried out from natural 2-hydroxy-3-(3-methyl-2-butenyl)-1,4-naphthalenedione (lapachol) and morpholine, piperidine and pyrrolidine. The structures of the products were established mainly by NMR analysis, including 2D experiments. Biological activities of these products were evaluated against *Artemia salina*, *Aedes aegypti* and cytotoxicity using A549 human breast cells.

Key words: lapachol, enamine derivatives, biological activities.

INTRODUCTION

Lapachol (**1**) is a natural quinone (see Figure 1) which has been isolated in very good yield from several species of Bignoniaceae found in Brazil, including Ceará State, where the species *Tabebuia serratifolia* is popularly known as “ipê-amarelo” (Correia 1984). From an ethanolic bark extract of a specimen of *Tabebuia serratifolia* lapachol (**1**) was obtained in 2.9% yield (Fernandes-de-Oliveira 2000).

Several biological activities are reported for this natural quinone, including anti-cancer, antiviral, antimicrobial, analgesic, antiinflammatory, antimalarial, cercaricidal, schistosomicidal and also a potential activity against *Trypanosoma cruzi*, the causal agent of Chagas' disease (Carvalho et al.

1988, Driscoll et al. 1974, Grazziotin et al. 1992, Lagrota 1983, Lopes et al. 1978, Pinto et al. 1977, 1987).

Nitrogen containing indoloquinone derivatives have been recently evaluated as novel anticancer agents, and the amine moiety was identified as an important feature for oxic and hipoxic potency and for the ability to act as a substract for reductase enzymes (Naylor et al. 1997).

Aedes aegypti is the vector for the transmission of “yellow fever” or “dengue” and it is responsible for the contamination of the urban population in developing countries, specifically in Brazil, causing serious public health hazards (Pan American Health Organization 1994).

This prompted us to synthesize new nitrogen lapachol derivatives in order to examine their larvicidal activities against *Aedes aegypti* and *Artemia*

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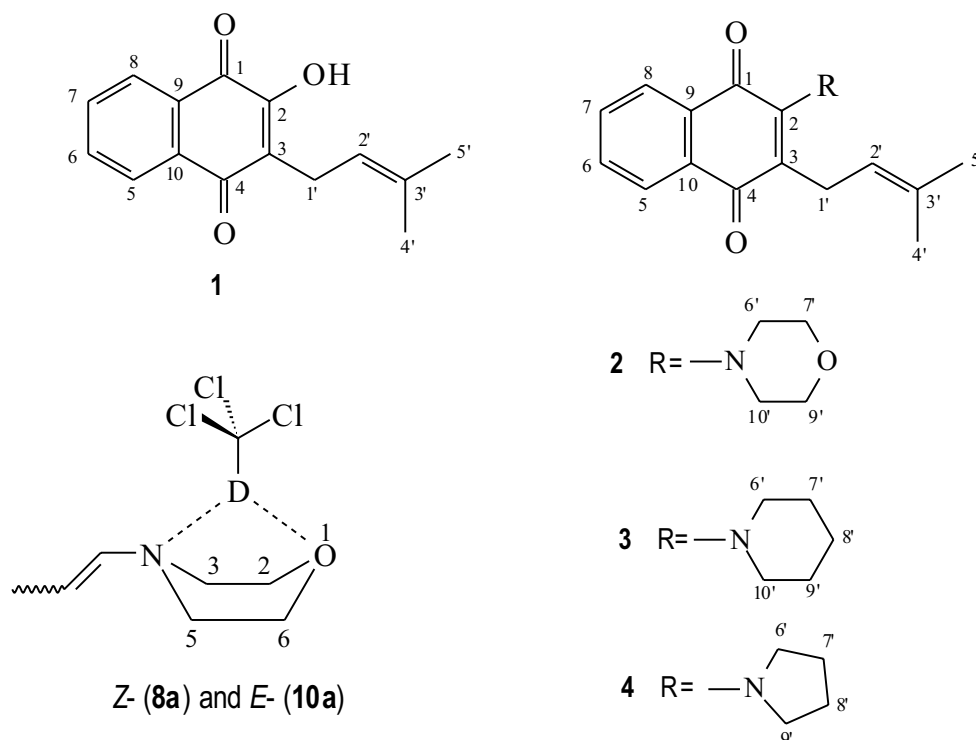


Fig. 1 – Structures lapachol (**1**), lapachol derivatives (**2,3,4**) and model compounds **8** and **10** involved in intermolecular hydrogen bond with CDCl_3 (**8a** and **10a**).

salina as well as cytotoxicity against A549 human breast tumor cells.

MATERIALS AND METHODS

GENERAL EXPERIMENTAL PROCEDURE

Melting points (mp) were determined on a Mettler FP5 apparatus and are uncorrected. ^1H - and ^{13}C -NMR spectra were obtained on a Bruker Avance DRX 500 MHz (500 MHz for ^1H and 125 MHz for ^{13}C) spectrometer. IR spectra were run on a Perkin-Elmer 1000 FT-IR spectrometer using KBr pellets. Lapachol (**1**) was obtained from an ethanolic extract of the bark of a specimen of *Tabebuia serratifolia* Bertol, Bignoniaceae, collected in Mulungu, Ceará, Brazil.

GENERAL PROCEDURE FOR DERIVATIVES

In a typical experiment, a solution of lapachol (**1**, 100mg, 0.41 mmol) in freshly redistilled amine (35 mmol) was stirred for 6h at room temperature and the exceeding amine was evaporated under vacuum. The residue was recrystallized from a mixture of EtOAc/hexane to yield **2** (94%), **3** (79%) and **4** (77%).

2-HYDROXY-3-(3-METHYL-2-BUTENYL)-1, 4-NAPHTHALENEDIONE (**1**)

Yellow solid, mp 139-140°C. IR (cm^{-1} , KBr): ν_{max} 3353 (O-H), 1657 (C=O), 1642 (C=O), 1592 (aromatic ring), 1273 (C-O). ^1H -NMR (500 MHz, CDCl_3): see Table I; ^{13}C -NMR (125 MHz, CDCl_3): see Table II.

TABLE I

¹H NMR spectral data for lapachol (1) and the enamine derivatives 2-4. Chemical shifts in δ_H (ppm) and coupling constants (J , in parenthesis) in Hz, in CDCl₃ and TMS as internal standard.*

H	1	2	3	4
5	8.07 (d, $J=7.5$)	8.07 (d, $J=7.4$)	8.03 (d, $J=7.5$)	8.02 (d, $J=7.5$)
6	7.70 (t, $J=7.5$)	7.68 (t, $J=7.4$)	7.59 (t, $J=7.5$)	7.58 (t, $J=7.5$)
7	7.62 (t, $J=7.5$)	7.60 (t, $J=7.4$)	7.47 (t, $J=7.5$)	7.45 (t, $J=7.5$)
8	8.02 (d, $J=7.5$)	7.98 (d, $J=7.4$)	7.85 (d, $J=7.5$)	7.84 (d, $J=7.5$)
1'	3.27 (d, $J=7.3$)	3.26 (d, $J=7.2$)	3.23 (d, $J=6.1$)	3.17 (d, $J=5.9$)
2'	5.17 (t, $J=7.3$)	5.18 (t, $J=7.2$)	5.18 (m)	5.13 (m)
4'	1.75 (s)	1.73 (s)	1.76 (s)	1.76 (s)
5'	1.64 (s)	1.65 (s)	1.63 (s)	1.63 (s)
6'	—	2.97 (t, $J=4.2$)	3.08 (br s)	3.25 (br s)
7'	—	3.30 (t, $J=4.2$)	1.61 (br s)	1.94 (br s)
8'	—	—	1.53 (br s)	1.94 (br s)
9'	—	3.30 (t, $J=4.2$)	1.61 (br s)	3.25 (br s)
10'	—	2.97 (t, $J=4.2$)	2.97 (t, $J=4.2$)	—

*Homonuclear 2D ¹H-¹H-COSY and heteronuclear 2D ¹H-¹³C-COSY- nJ_{CH} ($n=1$, HMQC; $n=2$ and 3, HMBC) were also used in these assignments (Table II).

2-(4-MORPHOLINYL)-3-(3-METHYL-2-BUTENYL)-
1, 4-NAPHTHALENEDIONE (2)

Dark red solid, mp 81.7-83.2°C. IR (cm⁻¹, KBr): ν_{\max} 1659 (C=O), 1590, 1540 (aromatic ring), 1340 (C-N), 1271 (C-O). ¹H-NMR (500 MHz, CDCl₃): see Table I. ¹³C-NMR (125 MHz, CDCl₃): see Table II.

2-(1-PIPERIDINYL)-3-(3-METHYL-2-BUTENYL)-
1, 4-NAPHTHALENEDIONE (3)

Dark red solid, mp 87.7-90.1°C. IR (cm⁻¹, KBr): ν_{\max} 1663 (C=O), 1589, 1539 (aromatic ring), 1359 (C-N). ¹H-NMR (500 MHz, CDCl₃): see Table I. ¹³C-NMR (125 MHz, CDCl₃): see Table II.

2-(1-PYRROLIDINYL)-3-(3-METHYL-2-BUTENYL)-
1, 4-NAPHTHALENEDIONE (4)

Dark red syrup, IR (cm⁻¹, KBr): ν_{\max} 1668 (C=O), 1589 (aromatic ring), 1361 (C-N). ¹H-NMR (500Mz, CDCl₃): see Table I. ¹³C-NMR (125Mz, CDCl₃): see Table II.

LARVICIDAL ACTIVITY

Larvicidal activity against *Artemia salina* was determined using a procedure described in the literature (McLaughlin 1991). Compounds **1-4** were tested in a concentration ranging from 1 to 500 ppm and LD₅₀ values were obtained using the Probit Program. Umbelliferone was included as a control (Table III).

Compounds **1-4** were tested against *Aedes aegypti* being placed in a beaker and dissolved

TABLE II

¹³C NMR and heteronuclear long-range couplings (²J_{CH} and ³J_{CH}) spectral data for lapachol (1) and the enamine derivatives 2-4. Chemical shifts in δ_C (ppm), in CDCl₃ and TMS as internal standard.*

	1	2		3		4	
	δ_{C}	δ_{C}	$^2J_{\text{CH}}$ and $^3J_{\text{CH}}$	δ_{C}	$^2J_{\text{CH}}$ and $^3J_{\text{CH}}$	δ_{C}	$^2J_{\text{CH}}$ and $^3J_{\text{CH}}$
C							
1	181.71	182.60	H-8	186.88	H-8	187.17	H-8
2	152.75	154.74	2H-1'	164.47	2H-1'	165.85	2H-1'
3	123.53	123.58	2H-1'	122.26	2H-1'	121.43	2H-1'
4	184.54	184.58	H-5, 2H-1'	183.97	H-5, 2H-1'	183.79	H-5, 2H-1'
9	129.48	129.85	H-5, H-7	131.12	H-7	131.25	H-5, H-7
10	132.94	133.11	H-6, H-8	134.55	H-6, H-8	134.63	H-6, H-8
3'	133.80	133.40	2H-1', 3H-4', 3H-5'	131.42	2H-1', 3H-4', 3H-5'	131.67	2H-1', 3H-4', 3H-5'
CH							
5	126.78	126.62	H-7	126.01	H-8	125.92	H-7
6	134.83	134.61	H-8	133.86		133.60	H-6
7	132.85	132.55	H-5	131.12		130.92	
8	126.05	125.90	H-6	125.32	125.24		
2'	119.71	120.35	2H-1', 3H-4', 3H-5'	123.18		123.24	2H-1', 3H-4', 3H-5'
CH ₂							
1'	22.64	22.70		22.94	2H-8'	22.93	2H-7' and/or 2H-8'
6'	–	45.45		44.85		45.54	
7',	–	66.80		22.94		24.69	
8'	–	–		22.56		24.69	
9'	–	66.80		22.94		45.54	
10'	–	45.45		44.85		–	
CH ₃							
4'	17.90	17.94	3H-5'	18.04	H-2'	18.09	3H-5'
5'	25.76	25.77	3H-4'	25.74	H-2'	25.76	3H-4'

*The number of bound hydrogens for each carbon signal was deduced by comparative analysis of HBBD- and DEPT-¹³C NMR spectra. Homonuclear 2D ¹H-¹H-COSY spectra of **1-4** and heteronuclear 2D ¹H-¹³C-COSY-ⁿJ_{CH} (n=1, HMQC) of **1** were also used in these assignments.

in DMSO (0.3 mL) and water (19.7 mL) at concentrations ranging from 1 to 500 ppm followed by addition of 50 larvae at the third stage. Mortality counts were made after 24 hours of treatment. A control solution was prepared using DMSO and water. Tests were done in triplicate and the results are presented in Table III.

CITOTOXICITY ASSAY AGAINST A549 HUMAN TUMOR CELLS

Selective toxicity to hypoxic A549 human breast tumor cells was determined for all compounds using the MTT [3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-2H-tetrazolium bromide] assay. Cells were

TABLE III

Activity against *Aedes aegypti* and *Artemia salina* of lapachol (**1**) and the enamine derivatives (**2-4**).

Compounds	LD ₅₀ (<i>Aedes aegypti</i>)	LD ₅₀ (<i>Artemia salina</i>)
1	20.79	12.75
2	242.61	10.00
3	899.38	55.26
4	397.05	31.62
Umbelliferone	—	100.00

TABLE IV

Cytotoxicity against A549 human breast tumor cells.

Compounds	IC ₅₀ (air) (mM)	IC (N ₂) (mM)
1	0.783 ± 0.058	1.0 ± 3
2	.054 ± 0.001	0.091 ± 0.032
3	.05 ± 0.21	0.84 ± 0.11
4	.68 ± 0.0059	0.99 ± 0.08

treated with drug for 3 hours at 37°C under aerobic or hypoxic (N₂) conditions. The drug was then removed and the cells allowed to proliferate for 3 days prior to MTT assay. The IC₅₀ (air) and IC₅₀ (N₂) values are the concentrations required to kill 50% of the cells under aerobic and under hypoxic conditions, respectively. The results are derived from at least three independent experiments and are presented in Table IV.

RESULTS AND DISCUSSION

Reaction of lapachol (**1**) with the amines morpholine, piperidine and pyrrolidine gave the 2-aminonaphthalenedione derivatives 2-(4-morpholinyl)-3-(3-methyl-2-butenyl)-1,4-naphthalenedione (**2**), 2-(1-piperidinyl)-3-(3-methyl-2-butenyl)-1,4-naphthalenedione (**3**) and 2-(1-pyrrolidinyl)-3-(3-methyl-2-butenyl)-1,4-naphthalenedione (**4**) in relatively good yields. A solution of lapachol (**1**) in freshly re-

distilled amine (morpholine or piperidine or pyrrolidine) was stirred for 6h at room temperature. The excess amine was evaporated under vacuum and the residue recrystallized from a mixture of EtOAc/hexane to yield the corresponding enamine derivatives **2** (94%), **3** (79%) and **4** (77%). The reaction, is initiated by intermolecular attack of the nucleophile nitrogen on the 2-enol carbon atom (path a) or 2-keto-carbonyl group (path b, much probable by the presence of a basic amine reagent) to produce the intermediates **I** and **II**, respectively, which dehydrate to give the corresponding enamine derivatives (**2-4**), as postulated and summarized in Figure 2.

The ¹H spectral data comparison (Table I) between lapachol (**1**) and the derivatives **2-4** indicate that the signals of the aromatic ring and side-chain attached at C-3 are practically unchanged and reveal the absence of the hydroxyl group at C-2 in the obtained products. Changes are mainly observed in the ¹³C NMR spectral data (Table II), which show a deshielding effect at C-2 [δ_C 154.74 (**2**), 164.47 (**3**), 165.85 (**4**)] when compared with lapachol (**1**: δ_C 152.75). The ¹³C signals related to the amine moieties were observed at δ_C 66.80 (CH₂-7' and CH₂-9') and 45.45 (CH₂-6' and CH₂-10') for **2**; δ_C 44.85 (CH₂-6' and CH₂-10'), 22.94 (CH₂-7' and CH₂-9') and 22.56 (CH₂-8') for **3**; δ_C 45.54 (CH₂-6' and CH₂-9') and 24.69 (CH₂-7' and CH₂-8') for **4** (Table II). The ¹H (1D and 2D ¹H-¹H-COSY), ¹³C (HBBD and DEPT) and heteronuclear 2D ¹H-¹³COSY-ⁿJ_{CH} (n=1, HMQC; n=2 and 3, HMBC) NMR spectra were used to establish the structures and to assign unambiguously the chemical shifts for all the hydrogen and carbon atoms of **2-4** (Tables I and II). The presence of the amine moieties at C-2 was confirmed by the heteronuclear long-range couplings of the hydrogen (Table I) and carbon atoms (Table II) revealed by cross-peaks observed in the HMBC spectra corresponding to the spin-spin interactions of H-8 [δ_H 7.98 (**2**), 7.85 (**3**) and 7.84 (**4**)] and C-1 [δ_C 182.60 (**2**), 186.88 (**3**) and 187.17 (**4**), ³J_{CH}], 2H-1' [δ_H 3.26 (**2**), 3.23 (**3**) and 3.17 (**4**)] and C-2 [δ_C 154.74 (**2**), 164.47 (**3**) and 165.85 (**4**), ³J_{CH}], C-3 [δ_C 123.58 (**2**), 122.26 (**3**) and 121.43 (**4**), ²J_{CH}]

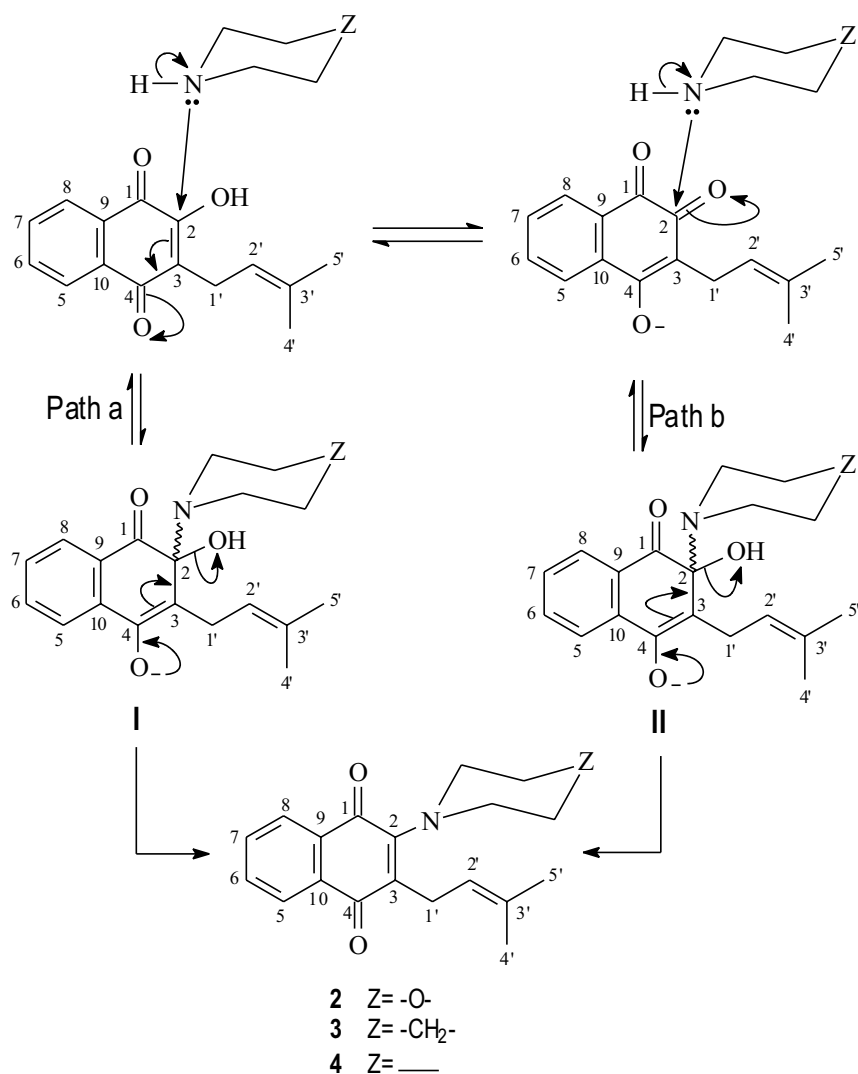
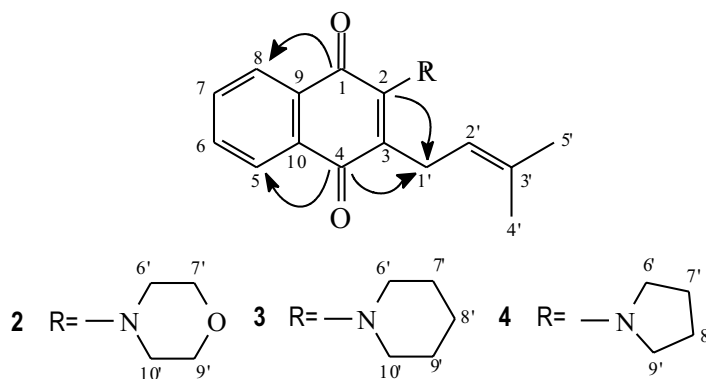


Fig. 2 – Proposed mechanism for the reaction of the amines morpholine, piperidine and pyrrolidine with lapachol (**1**) leading to the formation of the corresponding enamine derivatives **2-4**.

and C-4 [δ_C 184.58 (**2**), 183.97 (**3**) and 183.79 (**4**), $^3J_{CH}$] and H-5 [δ_H 8.07 (**2**), 8.03 (**3**) and 8.02 (**4**)] and C-4 [δ_C 184.58 (**2**), 183.97 (**3**) and 183.79 (**4**), $^3J_{CH}$], as summarized in Figure 3. The differences observed in the chemical shifts of the carbon atoms C-1 [δ_C 186.88 (**3**) and 187.17 (**4**)] and mainly C-2 [δ_C 164.47 (**3**) and 165.85 (**4**)] of **3** and **4** when compared with those of **1** [δ_C 181.71 (C-1) and 152.75 (C-2)] and **2** [δ_C 182.60 (C-1) and 154.74 (C-2)]

cannot be only accounted for by intramolecular electronic (inductive and mesomeric effects) and stereochemical arguments. However, the heteronuclear long-range couplings observed in the HMBC spectra (Table II and Figure 3) are clearly consistent with the proposed structures.

The ^{13}C chemical shifts of the enamine derivatives **2-4** were compared with data reported in the literature (Giasuddin et al. 1978, Breitmaier and

Fig. 3 – Selected HMBC correlations for compounds **1-4**.

Voelter 1987) for compounds **5-10** (Figure 4). As shown in Figure 4, the electronic effect of a nitrogen atom [reduced electron density at the α carbon (C-3) by inductive withdrawal effect and increased electron density at the β carbon by mesomeric effect] in **5** and **6** is practically identical, revealing $\Delta\delta_C=1.1$ ppm. Major differences ($\Delta\delta_C=2.4$ to 10.4 ppm) were observed in the comparison of the ^{13}C chemical shifts of (*Z*)-1-pyrrolidinyl-1- (**7**), (*Z*)-morpholinyl-1- (**8**), (*E*)-1-pyrrolidinyl-1- (**9**) and (*E*)-morpholinyl-1-propene (**10**), showing that, in fact, the electronic effects of the nitrogen atom in morpholinyl and pyrrolidinyl groups are significantly different, along with the anticipated shielding by a γ -effect which clearly allows to define the geometric isomer (*E* or *Z*). Comparative analysis of the chemical shifts of the non-hydrogenated carbon atoms C-1 [δ_C 182.60 (**2**), 186.88 (**3**) and 187.17 (**4**)] and C-2 [δ_C 154.74 (**2**), 164.47 (**3**) and 165.85 (**4**)] of the enamine derivatives **2-4** revealed major electron density reduction at C-2 of **3** (δ_C 164.47) and **4** (δ_C 165.85), which are in accord with values δ_C 163.4 and 164.6 reported in the literature for the carbon atom C-3 of the model compounds **5** and **6**, respectively (Figure 4). The chemical shifts of C-1 at δ_C 186.88 (**3**) and 187.17 (**4**) compared with those corresponding to C-4 [δ_C 183.97 (**3**) and 183.79 (**4**)] show major electron density at C-4 and may be justified by the mesomeric effect involving the conjugated unpaired electrons of

the nitrogen atom. However, the chemical shifts of these carbon atoms C-1 and C-4 in the compounds **1** and **2** cannot be justified on the basis of electronic effects (mainly mesomeric in this case), since they reveal a lower electron density at C-4 [δ_C 184.54 (**1**) and 184.58 (**2**)] than at C-1 [δ_C 181.71 (**1**) and 182.60 (**2**)]. Thus, these data show significant differences when compared with the values observed for the compounds **3** and **4** above considered (Figure 4). Consequently, the electronic participation of the nitrogen atom of the morpholinyl group of **2** is different from that conferred by the pyrrolidinyl and piperidinyl groups. Comparative analysis of the ^{13}C NMR spectral data reported in the literature (Giasuddin et al. 1978, Breitmaier and Voelter 1987) for the model compounds **7-10** allowed again to observe different participation of the nitrogen atom of the morpholinyl group (**8**: δ_C 140.0; **10**: δ_C 141.0), indicating a major inductive effect at the α carbon and a minor mesomeric effect at the β carbon (**8**: δ_C 107.5; **10**: δ_C 95.8) than those revealed by compounds **7** [δ_C 137.6 (C- α) and 97.1 (C- β)] and **9** [δ_C 136.6 (C- α) and 92.3 (C- β)], as shown in Figures 4 and 5.

All these data may be used to postulate the influence of other effects in the chemical shifts of C-1 and C-4 of lapachol (**1**) and its derivative **2**, the same occurring with α and β carbon atoms of the compounds **8** and **10** also containing the morpholinyl

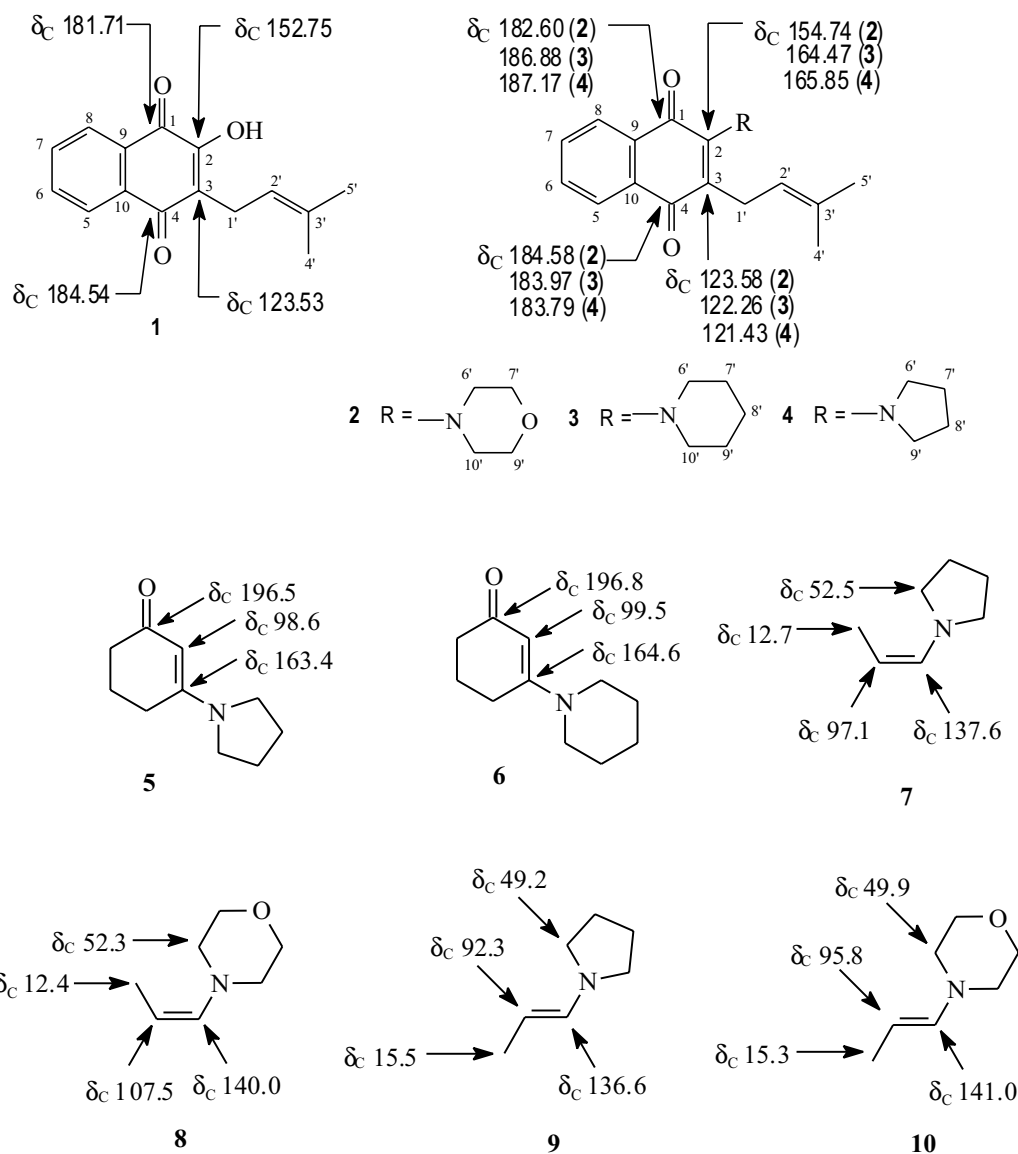


Fig. 4 – ^{13}C NMR spectral data of enamine derivatives **2-4** and model compounds **5-10** described in the literature (Giasuddin et al. 1978, Breitmaier and Voelter 1987).

group as **2**. Among other possible contributing effects can be included the participation of intramolecular hydrogen bonding in **1** and intermolecular deuterium bonding involving the deuterium of the solvent CDCl_3 , the carbonyl at C-1 and the nitrogen atom of **3** and **4** in the NMR experiments. However,

in the case of **2**, a repulsion between the non bonding electrons of chlorine (solvent) and the oxygen atom of the morpholinyl group prevents the formation of the deuterium bond (Figure 5) thus accounting for the minor changes of the chemical shifts of C-1 and C-2 of **2** when compared with **3** and **4**.

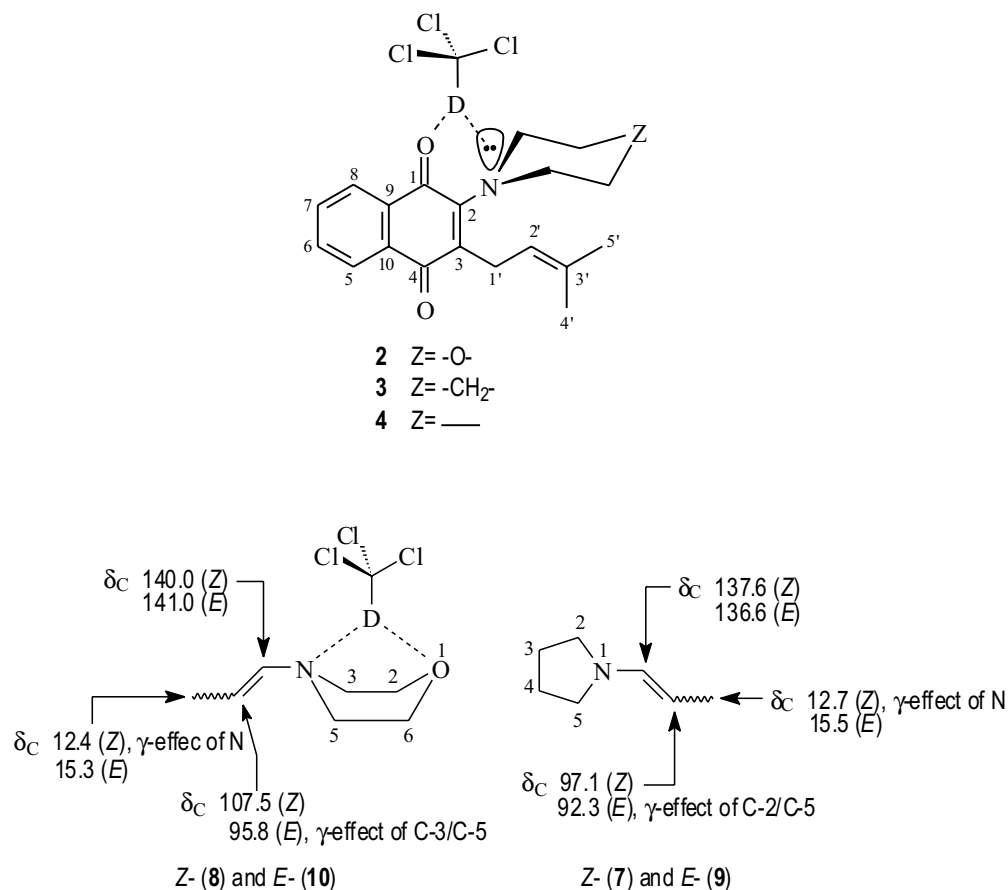


Fig. 5 – Intermolecular deuterium bonding of solvent $CDCl_3$ with derivatives **2-4** and model compounds **7-10** described in the literature (Giasuddin et al. 1978, Breitmaier and Voelter 1987).

In the compounds (Z)-(**8**) and (E)-1-morpholinyl-1-propene (**10**) the postulated speculative intermolecular deuterium bonding (Figure 5) may involve the nitrogen and oxygen atoms of the morpholinyl group in a bark conformation (**8a** and **10a**). Thus, the mesomeric effect involving the conjugated unpaired electrons of the nitrogen is attenuated and the inductive withdrawal effect encouraged. In this structural situation, the influence of these effects induces an electron density reduction and, consequently, major chemical shifts for the α and β carbon atoms of **8** and **10** when compared with those of **7** and **9**. This postulation requires additional investigations involving solvent, concentration and temper-

ature variations, including methyl ether and acetyl derivatives of lapachol (**1**) and other analogous compounds, using dried samples to assure the absence of H_2O .

The larvicidal activity of compounds **1-4** using *Artemia salina* was evaluated (Table III). All compounds were found to be quite active. However, no significant difference was observed with substitution of the hydroxyl group by an amine at C-2 position. A slight decrease in activity was observed for compounds **3** and **4** compared to lapachol (**1**). LD_{50} data are summarized in Table III.

In a bioassay against *Aedes aegypti*, lapachol (**1**), with a LD_{50} of 20.79 ppm, was found to be

more active than the amine derivatives with LD₅₀ values of 242.6, 899.4 and 397.0 ppm, respectively, showing the importance of the hydroxyl group at the C-2 position. The values of LD₅₀ of these bioassays are also summarized in Table III.

The cytotoxicity against A549 human tumor cells was evaluated using compounds **1-4** (Table IV). On the basis of our experimental observations, lapachol (**1**) and their analogues **2-4** showed mM activity against cells kill, under aerobic as well as hypoxic conditions. Compound **2** is 10 fold more potent than its analogues, indicating the importance of the morpholine moiety in the toxicity of these compounds.

CONCLUSION

We have shown that lapachol (**1**) can be converted easily into enamine derivatives (**2-4**) using morpholine, piperidine and pyrrolidine. In a bioassay against *Aedes aegypti*, lapachol (**1**) was found to be more active than the amine derivatives **2-4**, showing the importance of the hydroxyl group at the C-2 position. The morpholine moiety in compound **2** may be important in the toxicity against A549 human tumor cells.

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RESUMO

Uma síntese conveniente dos novos derivados enamínicos 2-(4-morfolinil)-3-(3-metil-2-butenil)-1,4-naftalenodiona, 2-(1-piperidinil)-3-(3-metil-2-butenil)-1,4-naftalenodiona e 2-(1-pirrolidinil)-3-(3-metil-2-butenil)-1,4-naftalenodiona foi conseguida de 2-hidroxi-3-(3-metil-2-butenil)-1,4-naftalenodiona (lapachol) natural e morfolina, piperidina e pirrolidina. As estruturas dos produtos foram estabelecidas principalmente pela análise de RMN, inclusive experimentos 2D. Atividades biológicas destes produtos foram avaliadas contra *Artemia salina*, *Aedes aegypti*

e citotoxicidade usando células humanas A549.

Palavras-chave: lapachol, derivados enamínicos, atividades biológicas.

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