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**A second note on the geographical differentiation of
Amphisbaena fuliginosa L., 1758 (Squamata, Amphisbaenidae),
with a consideration of the forest refuge
model of speciation**

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ABSTRACT

The geographical differentiation of *Amphisbaena fuliginosa* L. is studied with basis on 220 specimens from 99 definite and 2 generic localities, ranging from Panamá to Puno in Peru and Goiás in Brasil. The pattern found is considered to support Vanzolini's 1951 scheme of five subspecies, defined by color pattern and by four meristic characters, and differentiated in consequence of Quaternary paleoecological events. Recent developments relating to models of differentiation in Quaternary tropical South America are briefly considered.

Key words: subspecies, paleoclimates, *Amphisbaena*, refuge model.

INTRODUCTION

I published in 1951 a study of the geographic variation of *Amphisbaena fuliginosa* Linnaeus, 1758, concluding that five subspecies could be recognized, based mainly on color pattern, but with substantial support from scale counts. The paper was written in the flush and heat of the Mayrian paradigm (Mayr 1942): if two taxa are similar and allopatric, they should be considered as geographical races or subspecies. I have since come to question the unqualified application of the paradigm. The concept of subspecies involves more than likeness and allopatry; it entails too the presence of broad areas of morphological stability connected by relatively narrow belts of integration (Vanzolini and Williams 1970). Additionally, I came to worry about the statistical tools: the methods then available in textbooks within my reach were not ideally suited to the

study of geographical differentiation.

This Museum has recently received a fine sample of *A. fuliginosa* from an entirely unexpected area, not forested, as were all represented before, but sited in the core of the domain of the cerrados, eminently open formations (Ab'Saber 1977, Projeto Radambrasil 1981). The need to place in context this sample, so distant ecologically and geographically from those hitherto known, led me to re-examine the problem, including materials gathered since 1951, as well as better, if still elementary, statistical methods.

MATERIALS AND METHODS

SPECIMENS

I used in the present analysis in part the data of the 1951 paper, in part specimens collected since. The former were 135 specimens from 63 definite localities and 5 from generic ones ("Surinam" and

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TABLE I

Amphisbaena fuliginosa, body annuli, statistics of the distributions of frequencies.

Sample	N	R	m	s	V
Oriximiná	8	191–199	194.8 ± 1.10	3.1	1.6
Roraima	10	183–201	194.9 ± 2.22	7.0	3.6
Serra da Mesa	13	189–202	193.5 ± 1.51	5.5	2.8
Rondonia	9	184–205	195.4 ± 2.64	7.9	4.1
Colombia	10	190–205	197.7 ± 1.32	4.2	2.1
Panamá	10	195–202	198.4 ± 0.81	2.5	1.3
Ecuador West	14	190–205	198.5 ± 1.12	4.2	2.1
Aripuanã	6	195–206	200.3 ± 1.69	4.1	1.7
Ecuador East	11	196–209	202.1 ± 1.25	4.2	2.1
Surinam	10	193–216	203.5 ± 2.06	6.5	3.2
Pará	9	196–214	204.8 ± 1.89	5.7	2.8
Trinidad	17	202–212	206.2 ± 0.68	2.8	1.4
Guyana	10	199–218	206.8 ± 1.71	5.4	2.6
Iquitos	18	204–213	207.8 ± 0.59	2.5	0.6
Ucayali	11	204–215	210.5 ± 0.95	3.1	1.5
Manaus	18	206–220	212.8 ± 0.94	4.0	1.9

TABLE II

Amphisbaena fuliginosa, body annuli, Tukey's test.

	m	N
1. Serra da Mesa	193.5	13
2. Oriximiná	194.8	8
3. Roraima	194.9	10
4. Rondonia	195.4	9
5. Colombia	197.7	10
6. Panamá	198.4	10
7. Ecuador West	198.5	14
8. Aripuanã	200.3	6
9. Ecuador East	202.1	11
10. Suriname	203.5	10
11. Pará	204.8	9
12. Guyana	206.8	10
13. Iquitos	207.8	18
14. Ucayali	210.5	11
15. Manaus	212.8	18

“Trinidad”). The latter were 80 specimens from 37 definite localities. In all, 220 specimens, only 2 localities present in both samples. The materials are listed in the Appendix.

SCALE COUNTS AND MEASUREMENTS

I have at hand not many, but enough of the 1951 specimens to be sure that measurements and scale counts, taken so far apart in time, are fully compatible.

Four scale counts and two body proportions are here studied:

Body annuli, counted from the first complete post-occipital annulus to the one in front of the row of preanal pores. The annuli of *A. fuliginosa* usually show many irregularities, with partial splits and fusions. Differences due to these causes between successive counts seldom reached more than 2-3 annuli, i.e., 1% of the count. This, combined with broad range and small variability, makes the character, to me, very important in taxonomy. I adopted the first count of each specimen, checked the extremes of each distribution, and did some additional spot-checking.

Tail annuli, counted ventrally, from the first complete post-anal annulus to the tip of the tail, the terminal conical segment included.

Segments to a midbody annulus were, due to

TABLE III

Amphisbaena fuliginosa, body annuli, southern transect.

Annuli	Ucayali	Acre	Rondonia	Aripuanã	Sa. da Mesa	Sta. Maria
184			1			
185			1			
186			-			
187			-			
188			-			
189			1		1	
190			-		1	
191			-			
192			-		2	
193			-		1	
194			-		1	
195			1	1	1	
196			1	-	2	
197			-	-	-	1
198		1	-	2	1	-
199		-	1	-	1	-
200		-	-	-	1	-
201		-	-	1	-	-
202		-	1	-	1	1
203		-	-	-		-
204	1	-	1	1		-
205	1	-	1	-		-
206	-	-		1		1
207	-	-				
208	2	-				
209	-	1				
210	2					
211	3					
212	1					
213	-					
214	-					
215	2					
Sum	12	2	9	6	13	3

TABLE IV

Amphisbaena fuliginosa, body annuli, West-East transect.

Annuli	W Ecuador	E Ecuador	Iquitos	Leticia	Taba- tinga	Berurí	Manaus	Itapi- ranga	Orix- miná	Pará
190	1									
191	–								2	
192	–								1	
193	1								–	
194	–								–	
195	–								1	
196	2	1							2	1
197	2	1							–	1
198	1	1							1	–
199	2	–							1	–
200	1	2								–
201	–	–								–
202	1	3								1
203	1	–								–
204	1	–	2							–
205	1	1	2							1
206		–	3				1			1
207		1	1	1			2			1
208		–	3	–	1		–			2
209		1	2	–			–			–
210			3	–			1			–
211			1	–			1			–
212			–	–		1	4			–
213			1	–			2			–
214				1			2			1
215				–			2			
216				–			–			
217				–			–			
218				–			–			
219				–			2			
220				–			1			
221				1						
222										
223										
224								1		
Sum	14	11	18	3	1	1	18	1	8	9

TABLE V

Amphisbaena fuliginosa, body annuli, Manaus-Colombia transect.

Annuli	Colombia	Villavicencio	Cucuí	Vista Alegre	Balbina	Manaus
190	1					
191	–					
192	–					
193	–					
194	–					
195	1					
196	3					
197	1					
198	–					
199	–	1				
200	1	–				
201	2	1				
202	–	–		1		
203	–	–				
204	–	–				
205	1	–				
206		1				1
207	1					2
208		1	1			–
209			–			–
210			–		1	1
211			–		–	1
212					–	4
213			–		–	2
214			1		2	2
215						2
216						–
217						–
218						–
219						2
220						2
Sum	10	5	2	1	3	19

TABLE VI

Amphisbaena fuliginosa, tail annuli, statistics of the distributions of frequencies.

Sample	N	R	m	s	V
Pará	8	19–23	21.3 ± 0.53	1.5	7.0
Oriximiná	6	20–23	21.5 ± 0.43	1.1	4.9
Rondonia	7	23–24	23.6 ± 0.20	0.5	2.3
Roraima	9	23–26	24.3 ± 0.33	1.0	4.1
Aripuanã	5	24–25	24.4 ± 0.24	0.5	2.2
Panamá	5	23–27	24.4 ± 0.68	1.5	6.2
Manaus	14	22–26	24.6 ± 0.33	1.2	4.9
Colombia	9	23–27	24.7 ± 0.41	1.2	5.0
Ecuador West	12	23–29	24.9 ± 0.68	2.4	9.4
Serra da Mesa	8	24–27	25.0 ± 0.46	1.3	5.2
Guyana	9	24–29	25.4 ± 0.60	1.7	6.6
Trinidad	15	24–27	25.6 ± 0.29	1.1	4.4
Suriname	7	25–27	25.9 ± 0.34	0.9	3.5
Ecuador East	5	26–27	26.6 ± 0.24	0.5	2.1
Ucayali	11	26–29	27.9 ± 0.34	1.1	4.1
Iquitos	16	26–30	28.4 ± 0.33	1.3	4.6

TABLE VII

Amphisbaena fuliginosa, tail annuli, Tukey's test.

Sample	m	N
1. Pará	21.3	8
2. Oriximiná	21.5	6
3. Rondonia	23.6	7
4. Roraima	24.3	9
5. Aripuanã	24.4	5
6. Panamá	24.5	5
7. Manaus	24.6	14
8. Colombia	24.7	9
9. Ecuador West	24.9	12
10. Serra da Mesa	25.0	8
11. Guyana	25.4	9
12. Suriname	25.9	7
13. Ecuador East	26.6	5
14. Ucayali	27.9	11
15. Iquitos	28.4	11

irregularities in the annuli, counted on several adjacent annuli, sited roughly at midbody, and a mode adopted.

Preanal pores.

Two measurements can be meaningfully taken in amphisbaenids, *tail length* and *head width*, both proxies for attenuation/elongation of the body, essential information in the case of subterranean limbless animals (Vanzolini 1991).

Tail length was measured ventrally, from the edge of the post-anal semicircle to the tip of the tail.

Head width was measured at the widest point, with calipers, to the nearest millimeter. I see no point in looking for more precision; nobody can go back to a measuring point in a soft-bodied animal within one tenth of a millimeter. Ideally, given the cylindrical shape of amphisbaenids, head width should be regressed on total (body plus tail) length, but there were simply not enough intact tails, and I have had to use body length as the independent variable. Anyway, this turned out to be an uninformative character (Table XXI), and was abandoned.

STATISTICS

I used, for the scale counts, the usual descriptive statistics: number of specimens, range, mean and its standard deviation, sample standard deviation and

TABLE VIII

Amphisbaena fuliginosa, tail annuli, southern transect.

Annuli	Ucayali	Acre	Rondonia	Aripuanã	Serra da Mesa	Santa Maria
23		1	3			
24			4	3	2	1
25				2	2	–
26	2				3	–
27	1				1	–
28	3					1
29	1	1	7	5		2
Sum	7	1	14	10	8	3

TABLE IX

Amphisbaena fuliginosa, tail annuli, West-East transect.

Annuli	W Ecuador	E Ecuador	Iquitos	Leticia	Berurí	Manaus	Itapiranga	Oriximiná	Pará
19									1
20									2
21								2	
22						1		2	2
23	2					1		1	2
24	–				1	3	1		
25	2		3	1		5			
26	2	2	–	1		3			
27	1	3	3			–			
28	1	1	7			1			
29	2		3						
30									
Sum	10	6	16	2	1	14	1	6	8

coefficient of variation.

For multiple comparisons I performed analyses of variance, followed by Tukey's test for means. I checked Tukey's test against Kramer's (1956, as modified by Duncan 1957); there were no disagreements. I cite here only Tukey's test, because it is the more widely known.

For the body proportions I first decided on the anamorphosis to be used (all regressions turned out to be linear, with excellent fits) and compared the

regressions by analysis of covariance and Tukey's tests for the slope and intercept.

In all cases I tested initially for the presence of sexual differences, based on the 1951 materials. I did not sex the new materials, since the preliminary analysis showed no dimorphism and also because sexing amphisbaenids demands dissection of the base of the tail, which greatly increases the chances of breakage at the autotomy level.

All statistical methods used can be found in

TABLE X

Amphisbaena fuliginosa, segments to a midbody annulus, statistics of the distributions of frequencies.

Sample	N	R	m	s	V
Ucayali	12	38–44	41.8 ± 0.63	2.2	5.2
Ecuador East	10	40–44	42.8 ± 0.44	1.4	3.3
Iquitos	18	42–46	43.9 ± 0.20	0.8	1.9
Ecuador West	14	42–46	44.1 ± 0.33	1.2	2.8
Trinidad	17	40–50	44.2 ± 0.57	2.3	5.3
Colombia	10	40–50	44.6 ± 1.08	3.4	7.6
Rondonia	9	40–46	44.7 ± 0.67	2.0	4.5
Oriximiná	8	42–48	44.8 ± 0.65	1.8	4.1
Guyana	10	40–50	45.6 ± 1.02	3.2	7.1
Panamá	10	44–48	45.8 ± 0.55	1.8	3.8
Aripuanã	6	44–52	46.7 ± 1.12	2.7	5.9
Manaus	19	44–50	47.1 ± 0.44	1.9	4.1
Pará	10	46–52	48.4 ± 0.58	1.8	3.8
Suriname	9	46–54	49.1 ± 1.01	3.0	6.2
Roraima	9	46–52	49.8 ± 0.40	1.2	2.4
Serra da Mesa	13	48–52	50.9 ± 0.43	1.6	3.1

elementary texts, such as Dixon and Massey (1983), Vanzolini (1993a) or Zar (1999).

Conventions used in the tables of frequency distributions are: N, individuals in sample; R, range of the variable; m, mean ± its standard deviation; s, sample standard deviation; V, coefficient of variation.

In the regression tables: N, individuals in sample; R(x), R(y), ranges of the variables; b, coefficient of regression (slope) ± its standard deviation; a, regression constant (intercept) ± its standard deviation; F, Fisher's variance ratio, for the evaluation of the significance of the regression; r^2 , coefficient of determination.

In all tables and in the text:
 ns, not significant at the 5% level;
 *, significant at the 5% level;
 **, significant at the 1% level;
 ***, significant at the 0.1% level.

TACTICS

The ideal basic sample in the study of geographic differentiation is a good series of specimens from a single well-defined locality. The concept of a "good series" depends on the variability of the character and on the number of specimens. It should be kept in mind that, at the 5% level of significance, t for 5 degrees of freedom is 2.571; for 10 degrees, 2.228; for 15, 2.131; for 30, 2.042; and, for infinity, 1.960. Not much gain after 15. In the case of characters with little variability and sharp differences (the only ones of real practical interest), samples with 6 or 8 specimens usually turn out quite useful.

For the present study I had the following samples from single localities: Manaus (20 specimens), Iquitos (18), Oriximiná (8), Serra da Mesa (12) and Aripuanã (6).

The next choice is to group localities reasonably close together geographically and ecologically homogeneous. This is obviously a subjective pro-

TABLE XI

Amphisbaena fuliginosa, segments to a midbody annuli, Tukey's test.

Sample	m	N
1. Ucayali	41.8	12
2. Ecuador East	42.8	10
3. Iquitos	43.9	18
4. Ecuador West	44.1	14
5. Colombia	44.6	10
6. Rondonia	44.7	9
7. Oriximiná	44.8	8
8. Guyana	45.6	10
9. Panamá	45.8	10
10. Aripuanã	46.7	6
11. Manaus	47.1	19
12. Pará	48.4	10
13. Suriname	49.1	9
14. Roraima	49.8	9
15. Serra da Mesa	50.9	13

TABLE XII

Amphisbaena fuliginosa, segments to a midbody annulus, Manaus-Colombia transect.

Segments	Colombia	Villavicencio	Cucuí	Vista Alegre	Balbina	Manaus
40	1					
41	–					
42	4		1			
43	–		–			
44	1		1			3
45	–					–
46	2			1		6
47	–					–
48	1	1			2	8
49	–	–			–	–
50	1	3			–	3
51		–			–	
52		1			1	
Sum	10	5	2	1	3	20

cedure. Its adequacy can be tested by comparing the coefficients of variation of the grouped samples with those of the single-locality ones e.g, Tables I, VI, X and XIV). My grouped samples are Colom-

bia, Ecuador West and East, Guyana, Panamá, Pará, Roraima, Surinam, Trinidad and Ucayali.

Localities represented by one or very few specimens may be used in two ways, depending on the ge-

TABLE XIII

Amphisbaena fuliginosa, segments to a midbody annuli, West-East transect.

Segments	W Ecuador	E Ecuador	Iquitos	Leticia	Berurí	Manaus	Itapi- ranga	Oriximiná	Pará
40		1							
41		–							
42	2	5	2	1				1	
43	–	–	–	–				–	
44	7	5	15	–		3		4	
45	–		–	–		–		–	
46	2		1	–		6		2	2
47				–		–		–	–
48				1	1	8	1	1	5
49				–		–		–	–
50				1		3		–	2
51									–
52									–
53									–
54									–
55									–
56									1
Sum	11	11	18	3	1	20	1	8	10

TABLE XIV

Amphisbaena fuliginosa, preanal pores, distributions of frequencies.

Sample	Pores	6	7	8	9	10	Sum
Colombia		4	2	3			9
Trinidad		7	3				13
Rondonia		1	2	6			9
Ecuador West			5	8	1		14
Ucayali			3	9			12
Pará			1	8			9
Iquitos		1	–	17			18
Ecuador East				2	6	2	10
Roraima			2	4	2		8
Panamá				10			10
Oriximiná				7	1		8
Serra da Mesa		1	–	9	3		13
Manaus				15	4		19
Suriname				7	2	1	10
Guyana				5	3	2	10
Aripuanã				2	1	1	4

ography. Ideally, they may constitute steps in transects between basic samples, indispensable in the search for intergradation. Alternatively, they may be compared with the geographically close basic samples, to refine the areal definitions of the taxa involved.

The 16 basic samples used are (coordinates in the Gazetteer):

Aripuanã (state of Mato Grosso, Brasil), 6 specimens.

Colombia, 10 specimens from 6 localities in Andean central Colombia (all in my 1951 paper): Barrancabermeja (Departamento de Santander); Medellín (Antioquia), Muzo (Boyacá), Paime (Cundinamarca); Honda (Tolima); San Pedro (Valle del Cauca).

Ecuador East, 11 specimens from Upper Amazonian Ecuador (Provincias) and Peru (Departamentos): Loreto (Prov. Napo); Avila (Prov. Napo); Baños to Canelos (Prov. Pastaza); Turula, Rio

TABLE XV

Amphisbaena fuliginosa, preanal pores, statistics of the distributions of frequencies.

Sample	N	R	m	s	V
Colombia	9	6-8	6.9 ± 0.31	0.9	13.5
Trinidad	16	6-8	7.1 ± 0.23	0.0	13.2
Rondonia	9	6-8	7.6 ± 0.24	0.7	9.6
Ecuador West	14	7-9	7.7 ± 0.16	0.6	7.9
Ucayali	12	7-8	7.8 ± 0.13	0.5	5.8
Pará	9	7-8	7.9 ± 0.11	0.3	4.2
Iquitos	18	6-8	7.9 ± 0.11	0.5	6.0
Ecuador East	10	7-9	8.0 ± 0.21	0.7	8.3
Roraima	8	7-9	8.0 ± 0.27	0.8	9.5
Panamá	10	8	8.0		
Oriximiná	8	8-9	8.1 ± 0.13	0.4	4.4
Serra da Mesa	13	6-9	8.1 ± 0.21	0.8	9.4
Manaus	19	8-9	8.2 ± 0.10	0.4	5.1
Suriname	10	8-10	8.4 ± 0.22	0.7	8.3
Guyana	10	8-10	8.7 ± 0.26	0.8	9.5
Aripuanã	4	8-10	8.8 ± 0.48	1.0	10.9

TABLE XVI

Amphisbaena fuliginosa, preanal pores, Tukey's test.

Sample	m	N
1. Colombia	6.9	9
2. Rondonia	7.6	9
3. Ecuador West	7.7	14
4. Ucayali	7.8	12
5. Pará	7.9	9
6. Iquitos	7.9	18
7. Ecuador East	8.0	10
8. Roraima	8.0	8
9. Panamá	8.0	10
10. Oriximiná	8.1	8
11. Serra da Mesa	8.1	13
12. Manaus	8.2	19
13. Suriname	8.4	10
14. Guyana	8.7	10
15. Aripuanã	8.8	4

Upano (frequently misspelled "Luoula", e.g. in my 1951 paper) (Prov. Morona - Santiago); Canelos (Prov. Pastaza); Rio Pastaza (Prov. Pastaza); Boca Santiago (Dept. Amazonas); Rio Cenipa (frequently misspelled "Cenepa") (Dept. Amazonas).

Ecuador West, 14 specimens from 7 localities in Ecuador west of the Andes: Santo Domingo de los Colorados (Prov. Pichincha); Riobamba (Chimborazo); 50 km E Loja (Zamora-Chinchipec); Bucay (Guayas).

Guyana, 10 specimens from Maccasseema, Georgetown, Kaieteur, Kalacoon, Kamakusa, Marudi River, Nappi (Kanuku Mts.).

Iquitos (Dept. Loreto, Peru), 18 specimens, all in the 1951 paper.

Manaus (state of Amazonas, Brasil), 18 specimens.

Oriximiná (state of Pará, Brasil), 8 specimens.

Panamá, 10 specimens from the 1951 paper.

Pará, 9 specimens from the eastern part of the

TABLE XVII

Amphisbaena fuliginosa, preanal pores, West-East transect.

Pores	W Ecuador	E Ecuador	Iquitos	Leticia	Taba- tinga	Berurí	Manaus	Itapi- ranga	Orixí- miná	Pará
6			1							
7	5		–							
8	8	2	17	1		1	15	1	7	1
9	1	6			1		4		1	8
10		2								
Sum	14	10	18	1	1	1	19	1	8	9

TABLE XVIII

Amphisbaena fuliginosa, preanal pores, Manaus-Colombia transect.

Pores	Colombia	Villavicencio	Cucuí	Vista Alegre	Balbina	Manaus
6	4			1		
7	2					
8	3	4	2		3	16
9		1				4
Sum	9	5	2	1	3	20

state of Pará, Brasil, and adjacent Maranhão: Belém; Highway BR-010 (“Belém–Brasília”) km 93; Canindé, Rio Gurupí; Tomé-Açu; Açailândia.

Rondonia, 9 specimens from Santa Barbara; Alto Paraíso; Cachoeira de Nazaré, Rio Machado (or Ji-Paraná); Highway RO-399 km 1; Highway RO-399 km 5.

Roraima, 10 specimens from Vila Pacaraima; Tepequém Mt.; Island of Maracá, Rio Uraricoera; Maloca Mangueira; Cachoeira do Cujubim, Rio Catrimani; Rio Barauana at Highway BR-210; Apiaú.

Serra da Mesa (state of Goiás, Brasil), 12 specimens, plus one from nearby Piracanjuba (13 in all).

Suriname, 10 specimens from Paramaribo, Christiaankondre and unspecified localities.

Trinidad, 17 specimens from several localities in the island. This sample is included only to make the data available. It cannot at present be included in the analysis for lack of Venezuelan materials.

Ucayali, 12 specimens from the valley of the Rio Ucayali, Peru (Depts. Loreto and Ucayali): Cashiboya; Contamana; Rio Cushabatay; Pampa Hermosa; Pucallpa; Reforma; Requena; Roaboya.

The localities with one or a few specimens are listed in the transect tables (Tables III, IV, V, VIII, IX, XII, XIII, XVII, XVIII). The sample “Acre” comprises two specimens, one from Porto Walter, the other from Tarauacá.

All localities except those in the state of Goiás (Serra da Mesa and Piracanjuba), which are in cerrado, are in areas primarily covered by rain forest.

The Gazetteer at the end contains the coordinates of all localities.

ANALYSIS

COLOR PATTERN (FIGURE 1)

The color pattern of *Amphisbaena fuliginosa* is an intriguing subject. On one side, it is unique in



Fig. 1 – 1, *Amphisbaena fuliginosa varia*, MZUSP 2101, Panamá. 2, *A. f. amazonica*, MZUSP 57863, Manaus, Amazonas. 3, *A. f. bassleri*, MZUSP 6430, Loreto, Napo, Ecuador. 4, *A. f. wiedi*, MZUSP 87695, Serra da Mesa, Goiás.

the genus, unequivocally linking together a large number of populations over a vast area. It is, in fact, so characteristic that, until I (on general principles) started counting scales in 1950, there had been no preoccupation with geographical differentiation. On the other hand, once one is aware, it is easy to see that there are marked variations of the basic type, with definite spatial arrangements.

It is a brindled pattern, dark spots on a lighter background, with variations on back, belly and top of the head. The ground color varies from creamy yellow to dusky pink to purplish. The spotting varies

from definitely black to just a little darker than the ground color, either coinciding with the borders of individual segments or being more or less diffuse.

In Panamá (Figure 1: 1) and Colombia the ground color is a dirty pinkish; the spotting is smudgy, a little darker than the background.

In Iquitos, the Ucayali and eastern Ecuador (Figure 1: 3), the design is strikingly different: the ground color is very light, clear yellow or white; the dorsal pattern is reduced, tending to form narrow transverse well-marked black bands that reach down the flanks but not across the belly.

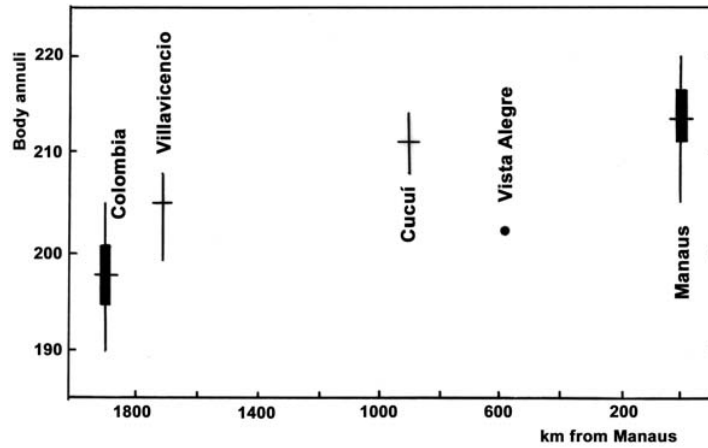


Fig. 2 – *Amphisbaena fuliginosa*, body annuli, Colombia–Manaus transect.

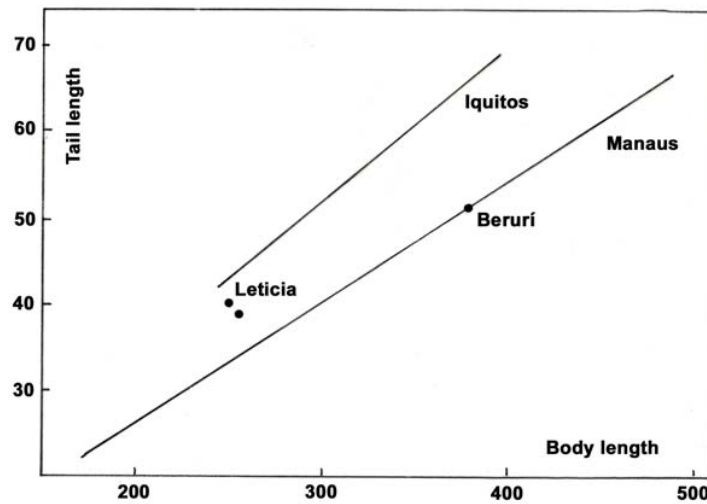


Fig. 3 – *Amphisbaena fuliginosa*, regression of tail length on body length, Manaus–Iquitos transect.

In Manaus (Figure 1: 2) the ground color is light, yellowish; the spotting is black, denser in front and sparser behind and especially on the belly. Twenty of the 21 specimens seen have immaculate heads; one shows a minuscule black spot. All Amazonian specimens east of Manaus, show this color pattern; only one example from Belém has a small spot on the head.

Guianan and Roraiman specimens have a color pattern similar to that of Manaus, but the heads are consistently spotted.

Finally, the Goiás (Figure 1: 4) and Santa Maria specimens have dark purplish backs, at times distinctly mottled. The head is smudged; there is frequently on each side of the nape a lighter, but still smudged marking, almost an interrupted collar. The

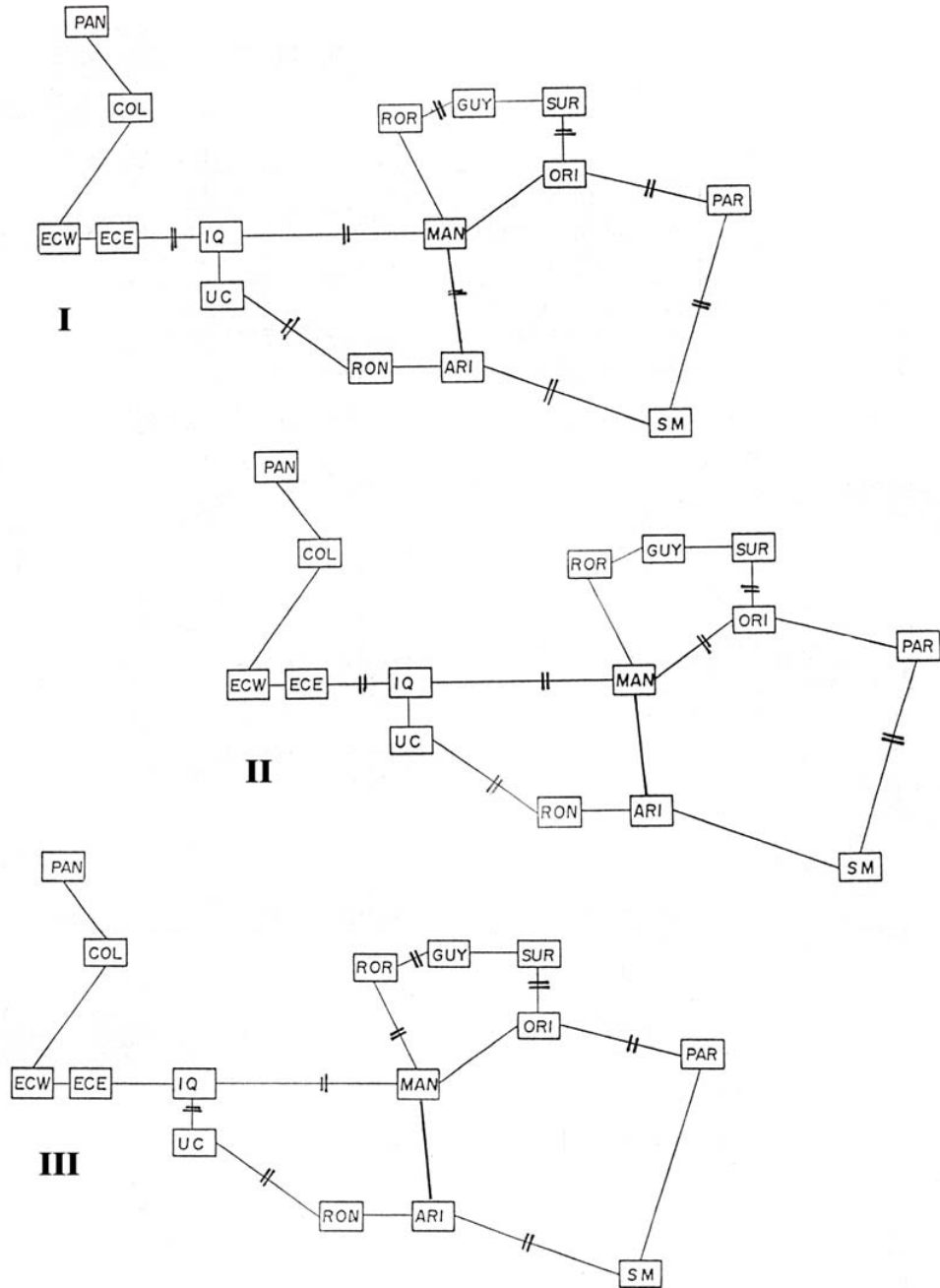


Fig. 4 1-3 – Spatial relationships of the distributions of meristic characters. 1. Body annuli. 2. Tail annuli. 3. Segments to a midbody annulus. Samples: ARI, Aripuanã. COL, Colombia. ECE, ECW, East and West Ecuador. GUY, Guyana. IQ, Iquitos. MAN, Manaus. ORI, Oriximiná. PAN, Panama. PAR, Pará. RON, Rondônia. ROR, Roraima. SM, Serra da Mesa. SUR, Suriname. UC, Ucayali. Cross lines indicate statistically significant differences.

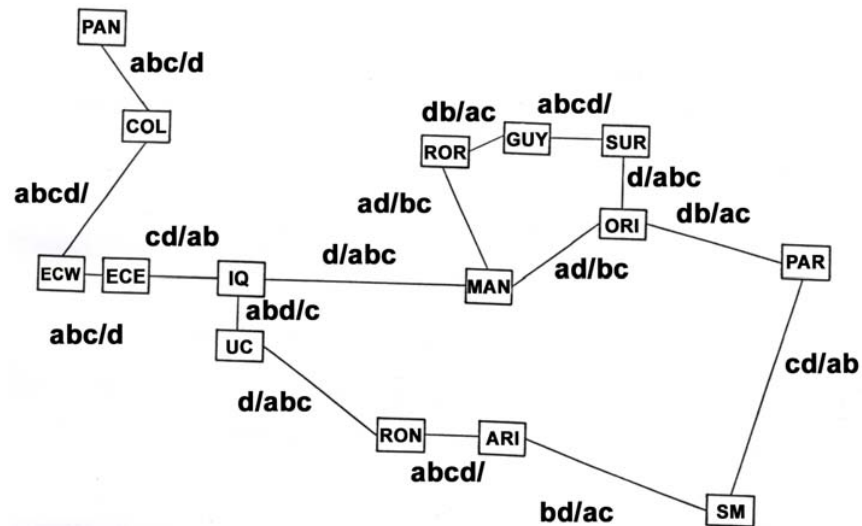


Fig. 5 – Summary of Figure 4 1-3. Conventions for samples as in Figure 4 1-3. Small letters to the left of the dash mean characters in which the two samples do not differ significantly; to the right of the dash, characters with significant differences. *a*, body annuli. *b*, tail annuli. *c*, segments to a midbody annulus.

belly is usually straw-colored, with incomplete dark crossbands, usually one annulus wide. This color pattern is also shown by the Aripuanã, Maranhão and Gurupí specimens.

Clear intergradation is found in several areas. In western Ecuador the Colombian and Iquitos patterns intergrade; for instance, of three Riobamba specimens, one has a definitely Colombian cast, the other two are intermediate. The Cucuí and Villavencio specimens are intermediate between Manaus and Colombia. Leticia agrees better with Iquitos than with Manaus, the head being distinctly spotted.

SCALE COUNTS

Body annuli (Tables I-V, Figure 2, and especially Figure 4-I). This, as said, is a very important character, showing considerable geographic variation. The major areas of homogeneity are Panamá + Colombia, Guyana + Surinam and Rondonia + Aripuanã + Serra da Mesa. Otherwise there is not a general pattern: it is apparently a case of marked inter-deme variation (Carson 1987).

The transects are highly interesting. The Ma-

naus–Colombia transect (Table V, Figure 4-I) shows clear clinal transition. The West-East transect (Table IV) shows a short western cline, broad stability in central Amazonia and a sharp break in Pará. Finally, a southern transect (Table III), including the southern Brazilian localities ordered from West to East, shows that the Acre, Rondonia and Aripuanã samples are intermediate between the Ucayali + Iquitos and the northeastern samples.

Tail annuli (Table VI-IX, Figure 4-II). Again the northwestern samples are in full agreement among themselves. From Iquitos to Manaus and from Manaus to Pará the variation is roughly clinal. The southern samples are homogeneous, as are the Guianas.

Segments (Tables X-XIII, Figure 4-III). From Panamá to Iquitos there is uniformity; breaks occur to the east and south of Acre. Other areas of stability are the Guianas, Manaus + Oriximiná + Rondonia + Aripuanã, and Serra da Mesa + Pará.

Preanal pores (Tables XIV-XVIII). The geography of preanal pores is thoroughly fragmentary.

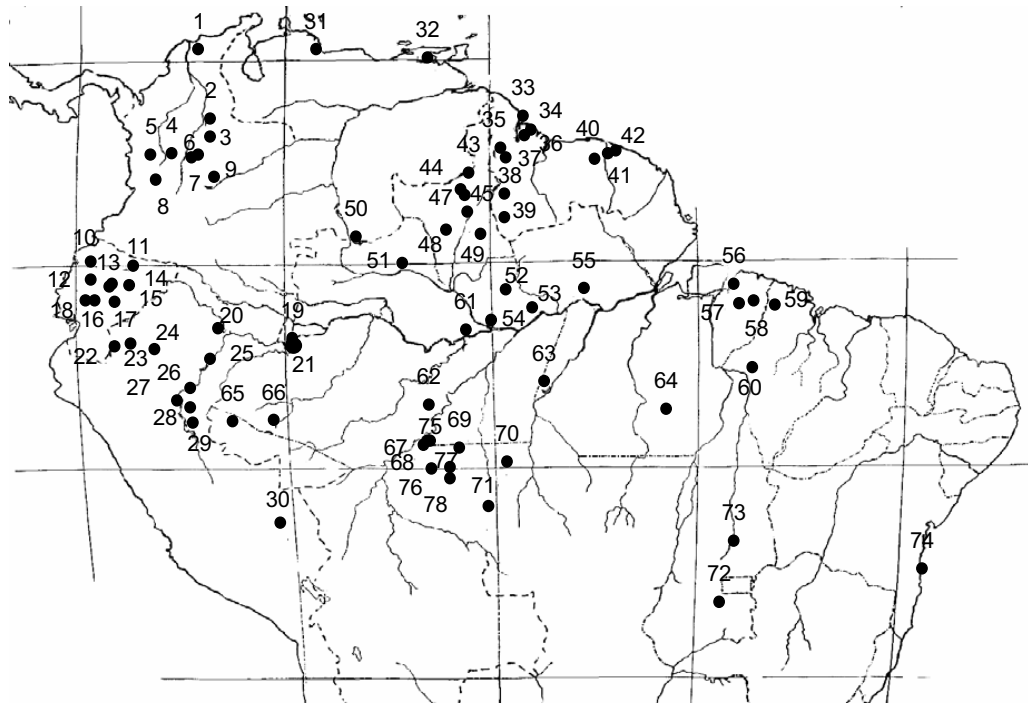


Fig. 6 – South American localities from which specimens were available. 1, Rio Frio. 2, Barrancabermeja. 3, Medellin. 4, Muzo. 5, Caño Las Animas ; Andagoya; Peña Lisa. 6, Honda. 7, Paime. 8, San Pedro. 9, Villavicencio. 10, Santo Domingo de los Colorados. 11, Avila; Loreto. 12, Pichilingue. 13, Baños. 14, Riobamba. 15, Canelos. 16, Bucay. 17, Turula. 18, Rio Daule; Guayaquil. 19, Loja. 20, Iquitos. 21, Leticia. 22, Rio Cenipa. 23, Boca Santiago. 24, Rio Pastaza. 25, Requena. 26, Cashiboya; Reforma. 27, Contamana; Rio Cushabatay; Pampa Hermosa. 28, Roaboya. 29, Pucallpa. 30, Rio San Gaban. 31, Riecito. 32, Caripito. 33, Maccasseema. 34, Georgetown. 35, Kamakusa. 36, Kalacoon. 37, Kaieteur. 38, Nappi. 39, Marudi River. 40, Paramaribo. 41, Chistiaankondre. 42, Saint Laurent du Maroni. 43, Vila Pacaraima. 44, Tepequém. 45, Maloca Mangueira. 46, Ilha de Maracá. 47, Apiaú. 48, Cachoeira do Cujubim. 49, Rio Barauana at Highway BR-210. 50, Cucuí, 51, Vista Alegre. 52, Balbina. 53, Itapiranga. 54, Manaus. 55, Oriximiná. 56, Belém. 57, Tomé Açu. 58, BR-010 km 93. 59, Canindé. 60, Açailandia. 61, Berurí. 62, Humaitá. 63, Jacareacanga. 64, Serra de Kukoinhokren. 65, Porto Walter. 66, Tarauacá. 67, Alto Paraíso. 68, Santa Barbara. 69, Cachoeira de Nazaré. 70, Aripuanã. 71, km 1; km 5 Highway RO-399. 72, Piracanjuba. 73, Serra da Mesa. 74, Santa Maria Eterna. 75, Charobamba. 76, Sorata. 77, Tipuani. 78, Las Yungas.

TAIL LENGTH

I feel incapable of analyzing the ensemble of the geographic variation of relative tail length (Tables XIX-XX). I find it worthwhile, though, to examine the relationships between some pairs of geographically adjacent samples.

The difference between Iquitos and Manaus is striking; the intervening samples (Leticia and Berurí) are clearly intermediate (Figure 3). East of

Manaus there is uniformity. Roraima differs significantly from Guyana.

CONCLUSION

The main features of this geographic scenario seem to be:

1. There is only one species, *A. fuliginosa*, involved.

TABLE XIX

Amphisbaena fuliginosa, statistics of the regression of tail length on body length.

Samples	N	R(x)	R(y)	b	a	F	r ²
Panamá	5	240–350	36–52	0.145 ± 0.0062	1.56 ± 2.640 ns	524.319***	0.9943
Colombia	8	115–310	19–45	0.135 ± 0.024	2.76 ± 3.162 ns	44.146***	0.8803
Ecuador West	14	220–360	34–62	0.148 ± 0.0228	2.00 ± 2.057 ns	41.922***	0.7921
Ecuador East	5	130–380	20–64	0.158 ± 0.0327	5, 360 ± 7.836 ns	23.402*	0.8864
Iquitos	15	245–450	39–75	0.170 ± 0.0183	0.62 ± 2.825 ns	80.584***	0.8695
Ucayali	11	105–365	16–62	0.170 ± 0.0163	–1.68 ± 3.740 ns	109.350**	0.9240
Manaus	11	170–330	21–40	0.135 ± 0.0158	–0.77 ± 1.902 ns	72.996***	0.8902
Roraima	8	250–360	38–56	0.147 ± 0.0386	2.78 ± 2.456 ns	14.463**	0.7068
Guyana	8	195–420	29–55	0.113 ± 0.0132	7.51 ± 2.899*	73.279***	0.9243
Surinam	7	130–350	19–51	0.142 ± 0.0107	1.86 ± 4.505 ns	176.409***	0.9861
Oriximiná	5	174–252	21–29	0.080 ± 0.0460		3.061 ns	
Pará	6	225–375	30–50	0.180 ± 0.0397	–13.44 ± 3.177 * **	25.503*	0.8368
Rondonia	5	150–275	20–40	0.148 ± 0.0151	–1.85 ± 3.318 ns	96.007***	0.9697
Serra da Mesa	8	284–353	39–55	0.205 ± 0.0292	–20.26 ± 1.985 * **	49.457***	0.8918

TABLE XX

Amphisbaena fuliginosa, regression of tail length on body length, Tukey's test for the intercept.

Sample
Guyana
Roraima
Surinam
Manaus
Colombia
Panamá
Rondonia
Ecuador West
Ecuador East
Ucayali
Iquitos
Serra da Mesa

- This species occupies continually one very large area, subdivided into two equally very large adjacent territories, in which occur two contrasting and well-characterized plant formations, rainforest and cerrado. There is one locality in the Atlantic forest.

- In the forested territory the species is obviously in a stage of advanced but still incomplete differentiation. There are broad areas of well-defined and characteristic color patterns, separated by relatively narrow transition belts. The data on quantitative characters do not quite permit to say that they are exactly coextensive with that of the color pattern, but the location of the transitional belts is highly suggestive.

- There are not yet adequate data on cerrado populations. A first impression is, as said, one of strong inter-deme differentiation (Carson 1987).

I have no doubt that this situation is the result of a process of differentiation in forest refugia, as proposed by Haffer (1969) and by Vanzolini and Williams (1970). The differentiation, if incomplete, is well-marked, and the geographic units can be easily and unambiguously identified. This why I decided in 1951, and reiterate it now, to treat them as subspecies.

The general subspecific pattern is superimposed on a scheme of pronounced inter-deme differentiation, as indicated by the relatively high number

TABLE XXI

Amphisbaena fuliginosa, statistics of the regression of head width on body length.

Sample	N	R(x)	R(y)	b	a	F	r ²
Aripuanã	6	288–385	11–16	0.0329 ± 0.01740		3.554 ns	
Manaus	8	193–490	7–17	0.0358 ± 0.00357	0.25 ± 1.321 ns	100.133***	0.9435
Oriximiná	7	174–300	7–11	0.0341 ± 0.00382	0.90 ± 0.580 ns	79.762***	0.9410
Pará	8	225–325	10–12	0.0158 ± 0.00359	6.84 ± 0.274 * **	19.416***	0.7639
Rondonia	9	150–320	7–12	0.0278 ± 0.0826	2.17 ± 0.632*	11.311*	0.7860
Roraima	11	250–365	9–15	0.0504 ± 0.00757	–3.69 ± 0.397 * **	44.328***	0.8312
Serra da Mesa	13	264–353	10–15	0.0552 ± 0.001230	–4.76 ± 0.510 * **	20.113***	0.6465

TABLE XXII

Amphisbaena fuliginosa, statistics of the distributions of frequencies of the meristic characters of the subspecies.

Character	Subspecies	N	R	m	s	V
Body annuli	<i>varia</i>	24	190–205	197.8 ± 0.72	3.5	1.8
	<i>fuliginosa</i>	21	192–218	205.4 ± 1.33	6.1	3.0
	<i>bassleri</i>	30	204–215	208.3 ± 0.53	2.9	1.4
	<i>amazonica</i>	22	206–220	213.1 ± 0.84	4.01	1.9
	<i>wiedi</i>	31	180–210	196.5 ± 1.23	6.9	3.5
Tail annuli	<i>varia</i>	19	23–27	24.6 ± 0.27	1.2	4.7
	<i>fuliginosa</i>	15	24–29	25.6 ± 0.35	1.4	5.3
	<i>bassleri</i>	27	26–30	28.2 ± 0.24	1.3	4.4
	<i>amazonica</i>	16	20–28	24.9 ± 0.35	1.4	5.7
	<i>wiedi</i>	22	23–28	24.5 ± 0.28	1.3	5.3
Segments	<i>varia</i>	24	40–50	45.2 ± 0.56	2.8	6.1
	<i>fuliginosa</i>	20	40–56	47.7 ± 0.89	4.0	8.3
	<i>bassleri</i>	30	38–46	43.1 ± 0.33	1.8	4.2
	<i>amazonica</i>	23	44–52	47.4 ± 0.43	2.0	4.3
	<i>wiedi</i>	31	40–52	48.3 ± 0.62	3.4	7.1
Pores	<i>varia</i>	23	6–8	7.4 ± 0.18	0.8	11.4
	<i>fuliginosa</i>	21	8–10	8.6 ± 0.18	0.8	9.3
	<i>bassleri</i>	30	6–8	7.8 ± 0.08	0.5	5.9
	<i>amazonica</i>	23	8–9	8.2 ± 0.08	0.4	4.7
	<i>wiedi</i>	29	6–10	8.1 ± 0.17	0.9	11.1

of statistically significant differences between adjacent areas of comparatively similar ecology. This fractured pattern can well be expected from a subterranean animal in early (or interrupted) stages of speciation. On the other hand, there is abundant evidence of gene flow, especially the presence of areas of character stability and the smoothness of

the transitions. It is to me quite obvious that there is no way of making sense of such a pattern without recourse to a causal explanation – a model of geographical differentiation. The fragmented character of the pattern indicates the presence of isolated demes at the time of differentiation.

TABLE XXIII

Amphisbaena fuliginosa, statistics of the regression of tail length on body length, subspecies.

Subspecies	N	R(x)	R(y)	b	a	F	r ²
<i>A.f. varia</i>	26	115–360	19–62	0.150 ± 0.0120	0.26 ± 1.656 ns	156.860***	0.8613
<i>A.f. fuliginosa</i>	15	130–420	19–55	0.125 ± 0.0086	4.74 ± 2.604 ns	212.134***	0.9423
<i>A.f. amazonica</i>	25	170–342	21–50	0.143 ± 0.0125	–2.85 ± 1.436 ns	130.540***	0.8502
<i>A.f. bassleri</i>	26	105–450	16–75	0.177 ± 0.0155	–2.27 ± 2.458 ns	235.468***	0.9075
<i>A.f. wiedi</i>	16	150–385	20–55	0.147 ± 0.0084	–1.82 ± 2.223 ns	304.939***	0.9561

How *A. fuliginosa*, which clearly originated in Amazonia, came to invade the cerrado is a very interesting problem, to which at present there are no decisive clues. Apparently there were two cycles in the general shaping of the territory: first differentiation in Amazonia and next occupation of the cerrado. Examining the obvious possibilities, the species may simply have expanded its area across the ecological frontier, protected by its subterranean habitus. Alternatively, it may have passed through an intermediate stage, involving a “vanishing refuge” (Vanzolini and Williams 1981): (i) a forest population gets locally adapted to ecotonal conditions; (ii) disappearance of the forest and persistence of the animal in the site lead to further in loco adaptation to open conditions; (iii) at the same time the retreat of the forest isolates the population in its new environment, with all attendant chances for differentiation.

Otherwise, *A. fuliginosa* may have taken part in the last wave of invasion of Central Brasil by Amazonian forests (Ledru 1993, Vanzolini 1997), being left behind in the succeeding withdrawal.

In trying to choose an alternative, one consideration is that it would at first sight seem easy for a fossorial animal to cross an ecological barrier occurring on the surface of the landscape. However the differences between rain forest and cerrado soils are extreme, in all physical and chemical properties. Contrariwise, it should be taken into account that the contacts between morphoclimatic domains are not gradual, but broad, complex, sectorial (Ab'Saber 1977) – which has always been an encumbrance to clinal theories of geographical speciation, but would

offer way-stations for animals crossing the barrier. Finally sharp differences in small spaces would tend to favor a vanishing refuge model.

Any of these alternatives may have happened as a single event at a single place, or as repeated crossings, at the same or different moments, at different places, but always at times of minimal climatic contrast. Any further consideration of these matters must await further data – more collections adequately covering the cerrados, or convincing negative evidence. At present it is only possible to say that the event (or events), seems to have been relatively recent, as most cerrado events seem to be (Vanzolini 1997).

A special problem is raised by the sample from Santa Maria – incidentally the hypodigm of *A. f. wiedi*. It dates from the early nineteenth century and is the only sample known from the Atlantic forest, herpetologically the best explored region of Brasil. Amazonian animals with distribution extending into the Atlantic forest are no novelty (Vanzolini 1994). The actual problems are the failure to collect the animal again after almost two hundred years, and the fact that it does not visibly differ from specimens from the cerrados. Prince Max's locality may conceivably be in error, but I have no reason at present to doubt his data. It is one more case for further collecting, if only to ascertain negative information.

ON IDENTIFICATION

A frequent criticism of subspecies is that they are rather identifiable by provenance than by morphology. This is not the case here. Identification is, as

said, ready and unambiguous. In fact, the practicality of the scheme has already been fully tested. I wrote the 1951 paper while a graduate student at the Museum of Comparative Zoology, Harvard University. When I submitted the paper, Mr. Arthur Loveridge, the curator of Herpetology, imposed, as a condition for acceptance, that I correctly identify at least 75% of a series of specimens of provenance unknown to me. This for two reasons: (i) that I, a Brazilian, was a "native" (Mr. Loveridge was an Englishman with a colonial African background), and (ii) that a renowned contemporary herpetologist was not able to identify 50% of his own subspecies. Mr. Loveridge gave me 14 specimens, and I provided correct provenances for the 14, which convinced him of the reality of the subspecies. (This immensely amused my then colleague W.L. Brown Jr, the myrmecologist, who aptly remarked that the feat had nothing to do with the reality of subspecies, only with the robustness of the statistics).

TAXONOMY

CHARACTERIZATION OF THE SPECIES

Head relatively massive, snout prominent, swollen. Body stocky, total length of adults usually 350 to 500 mm, one specimen 555 mm. Tail clavate; autotomy annulus, fifth to seventh.

Rostral moderate, low. Nasals large, in contact with the second supralabial. Prefrontal moderate. Postfrontals forming together a rough hexagon, wider than long. Three upper labials, the first smallest, the second and third variously divided and fused, at times suggesting preoculars or anterior suboculars. One true subocular, above the third labial, which may be horizontally split. Two irregular temporal series. Symphyseal roughly anvil-shaped, sometimes fused with one of the infralabials, which are 3 or 4, at times split or fused. Post-symphyseal elongate, sometimes longitudinally split. Remaining scales of the mental region variable.

Amphisbaena f. fuliginosa
Linnaeus, 1758

Amphisbaena fuliginosa Linnaeus, 1758: 29, Type locality "America".

Amphisbaena vulgaris Laurenti, 1768: 66. Type locality not mentioned.

Amphisbaena magnifica Laurenti, 1768: 67. Type locality "America".

Amphisbaena flava Laurenti, 1768: 67. Type locality "America".

Amphisbaena americana Gray, 1844: 70. *Nomen novum pro Amphisbaena fuliginosa* Linnaeus.

Amphisbaena fuliginosa fuliginosa, Vanzolini, 1951: 60. Revision.

Type locality: Linnaeus (1758: 229) had two specimens, which have survived (Vanzolini 1951: 56-59). A description of the specimen still preserved at Uppsala is contained in Linnaeus's *Amoenitates Academicae* (Linnaeus 1749, *Surinamensia Grilliana*; Lönnberg 1896). It seems clear, judging by the specimen's characters (Vanzolini 1951: 59), and is compatible with historical and geographical information, that the type locality must have been Suriname, very probably the region around Paramaribo. In agreement with Myers and Böhme (1996), I see no virtue in the designation of type localities by fiat, just to fill a void. However, there is always purpose in clearing moot points, using circumstantially relevant information, should the need ever arise (Vanzolini 1999: 12).

Meristic characters: Body annuli 192-218; tail annuli 24-29; segments to a midbody annulus 40-56; preanal pores 8-10.

Color pattern: Ground color whitish; spotting black, clear-cut, denser on the front part of the dorsum, sparsest on the belly. Top of head spotted.

Range: The Guianas.

Amphisbaena fuliginosa varia

Laurenti, 1768

Amphisbaena varia Laurenti, 1768: 66. Type locality, Barro Colorado Island, by designation of neotype (Vanzolini 1951).

Amphisbaena fuliginosa varia, Vanzolini, 1951: 61. Revision.

Meristic characters: Body annuli 192-205; tail annuli 23-27; segments to a midbody annulus 40-50; preanal pores 6-8.

Color pattern: Ground color a dirty pinkish; body markings smudgy, very dense, almost obliterating the ground color, equally dense on back, belly and top of the head.

Range: Panamá and Andean Colombia, extending into adjacent Ecuador and probably Venezuela.

Amphisbaena fuliginosa bassleri

Vanzolini, 1951

Amphisbaena fuliginosa bassleri Vanzolini, 1951: 61. Type locality Peru: Ucayali: Roaboya.

Meristic characters: Body annuli 204-215; tail annuli 26-30; segments to a midbody annulus 38-46; preanal pores 6-8.

Color pattern: Ground color white to yellowish. Dorsal markings black, transverse, not crossing the belly.

Range: Upper Amazonas in Ecuador and Peru, valley of the Ucayali into Peru. Rhodes (1963) discusses six specimens from La Paz (Charobamba, Sorata, Tipuani, Yungas) that can possibly be assigned to this subspecies. Using the data of my 1951 paper, she concludes that there is reasonable agreement in color pattern, and, principally, differences in the number of body and tail annuli, and relative tail length. I compared Rhodes's data with the present ones for the Ucayali sample: the differences in number of annuli persist, but there is excellent agreement in the regression of tail length on body length.

Amphisbaena fuliginosa amazonica

Vanzolini, 1951

Amphisbaena fuliginosa amazonica Vanzolini, 1951: 62. Type locality Brasil: Amazonas: Manaus.

Meristic characters: Body annuli 206-220; tail

annuli 20-28; segments to a midbody annulus 44-52; preanal pores 8-9.

Color pattern: Ground color yellowish. Body spotting clear-cut, denser on the back. Head practically always immaculate.

Range: "Baixo Amazonas", i.e., the course of the Rio Amazonas east of the mouth of the Rio Negro.

Amphisbaena fuliginosa wiedi

Vanzolini, 1951

Amphisbaena fuliginosa wiedi Vanzolini, 1951: 62. Type locality Brasil: Bahia, Santa Maria Eterna.

Note on the type locality: In 1951 I was not able to identify precisely the type locality of this subspecies. The hypodigm (AMNH 1091-1093) having been collected by Prince Maximilian zu Wied-Neuwied, it was extremely probable that the locality would be in the Atlantic forest of southern Bahia. Now I have found it. It is still called Santa Maria, in fact Santa Maria Eterna. It lies about 55 km due West of Belmonte, where the Prince spent some month and a half in 1816, actively exploring the valley of the river then called Belmonte, now Jequitinhonha (Wied 1820-1821: chapter 11; Bokermann 1957: 227-229).

Meristic characters: Body annuli 180-210; tail annuli 23-28; segments to a midbody annulus 46-52; preanal pores 6-10.

Color pattern: Ground color purplish; back densely smudged; belly with few spots, usually incomplete cross bands one annulus wide.

Range: Brasil south of the Rio Amazonas, to the latitude of the upper Rio Tocantins.

COMMENTS

Amphisbaena fuliginosa varia is a very coherent subspecies. It extends from Panamá to Ecuador, very uniform in color pattern, meristic characters and tail length, the only exception being a pocket of low preanal pores in Colombia. Its range coincides with that of many other species of amphibians and reptiles (Vanzolini and Heyer 1985). Its two neigh-

bors are *bassleri* to the south and *amazonica* to the east. From *bassleri* it differs in color pattern, an extreme difference, and in number of body and tail annuli. In the case of color pattern, intergradation is clear, the locality of Riobamba being especially informative in this respect. In the meristic characters no intergradation has been perceived. From *amazonica*, *varia* markedly differs in color pattern; of the geographically intermediate samples, Villavencio and Cucuf are also phenotypically intermediate and Vista Alegre is typical *amazonica*. There are also strong differences in the number of body annuli (Table V, intergradation clear) and in the number of tail annuli (no intergradation found).

A. f. fuliginosa, the subspecies that occurs in the Guianas, is also very coherent: character variation is concordant throughout. The color pattern is highly distinctive, very different from those of neighboring Roraima and Oriximiná. It is regrettable that no specimens are available from Amapá.

Roraima, although represented by a reasonably large sample, well prepared and with firm data, is hard to place at this time. It does not show much affinity with its next neighbor, *amazonica*. The distributions of frequencies of body annuli and of segments to a midbody annulus are peculiar. In color pattern it tends rather toward *varia*, geographically distant, than toward spatially closer *fuliginosa* and *amazonica*. On purely statistical grounds, Roraima might be made the site of still another subspecies, but clarification of its status must wait for good samples from Venezuela.

A. f. amazonica, most visibly identified by its immaculate white head and by the high number of body annuli, occupies the middle valley of the Rio Amazonas. It differs markedly from *bassleri*, to the west at the same latitude, in every character but pre-anal pores. In the case of the quantitative characters there is clear intergradation (Tables IV, IX, XIII, Fig. 3). To the east, toward Belém of Pará, the situation is less clear (Tables IV, IX, XIII). Unspotted heads prevail, but there is much irregularity in the quantitative characters.

When I first described this subspecies I duly stressed the immaculate head. Subsequently I had doubts and even regrets. In hindsight the character seemed somewhat unsubstantial in the context of such a complex color pattern: I expected numerous exceptions would soon start appearing. Quite the contrary: time showed it to be very reliable. Why this should be so in a fossorial animal is beyond me.

A. f. bassleri inhabits a stretch of the Upper Amazonas and goes deep south into Peru, by present evidence along the Ucayali and farther south into Puno, possibly, as said, into Bolivia. The striking color pattern, that is found in pure form also in Acre and Rondonia, is geographically quite stable; the scale counts are not: only the counts of tail annuli are characteristically high. To the east, on the southeastern edge of the range, there is intergradation with *wiedi*. The clinal belt is unmistakable, but extremely broad – some nineteen hundred kilometers (Tables III, VIII). This is certainly an area from which good collections are much wanted.

A. f. wiedi is, not surprisingly, the least coherent subspecies. The color pattern is uniform, but the quantitative characters show little regularity. I think this is to be expected: *wiedi* is the product of a late event or series of events. It is quite possible that it is still undergoing adaptation to local ecological conditions, under some regime of inter-deme competition and gene flow that can only be guessed at. One must wait for further materials from the domain of the cerrados, from which scarcely more than a dozen specimens are known.

A NOTE ON THE REFUGE MODEL

I have adopted, to explain the pattern of differentiation of *A. fuliginosa* on the northern, forested, part of its range, the model of Quaternary forest refuges, which to me continues to be the most parsimonious (besides orthodox) mechanism proposed to understand geographic differentiation in the American tropics – both for forest and for open formation species.

To my knowledge, a model of speciation based

on paleoclimates was first proposed by Lönnberg (1926), who maintained that some African birds had undergone speciation in isolated patches of forest during former dry climatic episodes. This was a time when subspecies, especially avian, were the focus of much interest, to culminate in the late thirties and early forties with the seminal books by Dobzhansky (1937), Huxley (1940) and Mayr (1942). In spite of this interest, there was no research on speciation in South America, and especially no cogitation of past climates, until a relatively late date (Vanzolini 1993).

The model of forest refugia was independently and almost simultaneously proposed by Haffer (1969) and by Vanzolini and Williams (1970). The animal groups studied, respectively birds and lizards, were different, but the approach was essentially the same: a search for geoscientific historical facts that would explain the distribution patterns observed. Both papers concluded that differentiation was principally geographic, allopatric and driven by the changing coarse mosaics of forest and open formations consequent upon climatic cycles. The present continuous Amazonian forest would have been preceded, in drier times, by a vast extension of open formations containing "islands" of forest. In these patches, isolated remains of forest fauna would differentiate, to come together again (with varying outcomes of interaction) in the ensuing humid leg of the climatic cycle.

The model was readily adopted by systematists (see, e.g., the papers in Prance 1982, Whitmore and Prance 1987), but, in time, aroused opposition from paleoecologists – systematists in general stayed faithful. There has been considerable controversy.

Recently (December, 2001), the issue (16, 3/4) of the journal *Amazoniana* honoring the limnologist Harald Sioli on his ninetieth birthday included four papers on the debate, by Haffer and Prance, by Colinvaux, Irion, Räsänen, Bush and Nunes de Mello, by van der Hammen and by Hooghiemstra. These papers very adequately summarize and bring to date the argument.

Haffer and Prance (2001: 579) review the history of the model, discuss pro and con arguments and, very important, analyze some misconceptions mistakenly erected as criticisms. Finally, they examine the applicability of rival models of allopatric speciation.

The discussion of the misconceptions (or misrepresentations), based essentially on Haffer (1993), includes, among others, the following points that I deem indispensable.

1. The model would have been based only on present biotic patterns. Actually, as said, the initial work combined distributional patterns with paleoecological (palynological and geomorphological) data. I know: I participated, and it has been a landmark in my life that this was the beginning of a long and rewarding association with Aziz Nacib Ab'Saber, better than whom nobody could have offered ingress into paleoclimatic matters, with ready and continuous application to the Brazilian reality around us. Haffer, a professional geologist, availed himself of his unmatched command both of the literature, ecologic and geoscientific, and of the field evidence, in Colombia and in Amazonia in general.
2. The model would have been designed for application to the Pleistocene only, and would thus imply that all patterns currently observable are Quaternary in age. This, as Haffer and Prance (2001: 590) express it, is a mere straw man, erected only to be knocked down. The claim was never made; stress was instead explicitly placed, both by Haffer (1969) and by Vanzolini and Williams (1970), on the recurrent nature of the process. Vanzolini and Williams (1970: 107) even mention the probability of phenomena of resonance, given the duration of topographic features, responsible for many relevant rainfall features, compared to the relative shortness of climatic cycles. It is matter of undisputed common knowledge among systematists

that some speciation events date back to the Tertiary (Frailey 1986, Maxson and Heyer 1988).

3. Another misrepresentation, and one frequently alleged by opponents of the model (e.g. Colinvaux (1996) is that it implies arid climatic phases. This is another straw man: the assertion was never made by the originators of the model. There were arid phases in the Pleistocene (Kronberg and Benchimol 1993, Latrubesse and Franzinelli 1995, Latrubesse and Rancy 1998), but, for the model, all that is required is a climate (such as that of many parts of present Brasil, that has vast cerrados and caatingas, but not one single arid area) that will favor open vegetation over forests. Modern analogy points out mainly (but not exclusively) to cerrados, of which sizable enclaves nowadays exist in Amazonia. In a case study of the symmetrical problem of differentiation in islands of cerrado dispersed in a general forest, I have (Vanzolini 1997) calculated that, broadly speaking, to substitute cerrado for forest, a decrease will suffice of about 30 percent in the rainfall of an area with a suitably cool and dry winter. This conjuncture is found in many areas of present Amazonia (Salati et al. 1978), and the required decrease is modest indeed in view of estimated climatic parameters for the Quaternary (van der Hammen 2001, Markgraf 1993, Iriondo and Latrubesse 1994).

Haffer and Prance deal in sequence with some alternative models of allopatric speciation proposed for Amazonia. They examine in detail the shortcomings of the proposals of Bush (1994), Colinvaux (1998), Fjelds  et al. (1999) and Patton et al. (2000). I think their analysis is conclusive and see no need to return to the issue.

The next paper in the volume, by Colinvaux et al. (2001: 609-646) is entitled "A paradigm to be discarded: geological and paleoecological data falsify the Haffer and Prance refuge hypothesis of Amazonian speciation". This title gives a good in-

dications of the emotional tone of the article. (Colinvaux, in another paper (1993: 475), says: "Thus the concept of Pleistocene forest refuges in the tropics is as intellectually barren as its predecessor concept of forest refuges in North America" – a revealing pearl of contumely and irrelevance).

Colinvaux et al. (2001) take arms again against the straw man of Amazonian aridity: "all available Amazonian pollen data, without exception and including new data, imply biome stability..."; "the 'aridity with refuges paradigm' now impedes Amazonian research, and should be discarded".

In the next two papers of the same issue of *Amazoniana*, van der Hammen and Hooghiemstra (2001) thoroughly demolish Colinvaux's (1998) argument, especially the palynological part, with which van der Hammen deals masterly (partly based on Hooghiemstra and van der Hammen 1998).

I think this limelight-seeking line of thought needs no further attention.

Another area of research that has been very fruitful in the context is paleomammalogy, since it is easy to ascertain the ecological preferences of faunal assemblages. Since early days (Paula Couto 1944), southwestern Amazonia has provided several fossil mammal faunas clearly adapted to open formations – I may add to cerrado. In the context of the Upper Tertiary "Great American Interchange", Webb and Rancy (1996; see also Rancy 1993) review the evidence and analyze the Late Pleistocene faunas of Acre. They sum up (1993: 350): "The Late Pleistocene fauna from western Amazonia lends support to this savanna hypothesis." The support, however, is restricted to the presence of "savannas" (cerrados), not extended to the refuge model as a whole: "A fundamental weakness of this rainforest theory is that the supporting evidence is derived from Recent biotic patterns" (*l.c.*: 352). This reveals only that Webb and Rancy did not read Haffer (1969) and Vanzolini and Williams (1970) in the original. It also, and very curiously, goes frontally against the uniformitarianist principle ("the present is the key to the past"), which, since enunciated by Hutton and

Lyell in the late eighteenth century, has been central to paleontological thought (e.g., Gould 1965). Since, however, two distinguished paleontologists disagree, we have to examine their reasons.

They argue (and this, incidentally, does not go against uniformitarianism, quite the contrary) that the locations of three of these fossil mammal faunas lie in areas designated as Late Pleistocene forest refuges. This is an obvious *non sequitur*. The existence of refuges and their location based on recent distributions are entirely different issues. “Patterns of distribution and differentiation do not prove the existence of refuges; they are simply compatible with the model”... “as to the location of refuges, it is a matter for geomorphologists and paleopalynologists.” (Vanzolini 1993: 51). This was written before Colinvaux et al. (1996) for the first time located a refuge through pollen analysis, ironically enough within an area (the “Imeri Refuge”) predicted by Haffer (1969).

Webb and Rancy (1996: 352) proceed: “Thus the western Amazon did not offer its mammal fauna a pure savanna setting, but rather a mobile habitat mosaic that included meandering belts of rain forest and perhaps seasonal flooding. Geomorphologists studying the same fluvial systems have proposed similar conclusions (Räsänen et al. 1987)”.

There are several points here in need of remark. I think that, inventing the expression “pure savanna setting”, Webb and Rancy may have had in mind an expanse of cerrado without gallery forest or forest enclaves. If so, this does not exist and was never included in the model. Webb and Rancy could have read in Ab’Saber’s papers (e.g., 1971, 1977) that gallery forests are an essential, integral part of the domain of the cerrados – Ab’Saber even calls the latter “domain of the highlands under cerrado, penetrated by gallery forests” (my translation).

Gallery forests, in the strict usage of the term an exclusive feature of the cerrados, begin at headwaters as marginal rows (“veredas”) of burití palms (*Mauritia*). Soon downstream these veredas become thicker and more heterogeneous, but still limited

to the levees (“restingas”) of the streams. Next, they widen laterally on the backswamp (“varzea”), and later climb the moderate interfluves. When the “eyelashes” of two convergent rivers meet, we acknowledge to be in Amazonia (Ab’Saber 1996 a,b). Today, a large and well-known enclave is the “Mato Grosso de Goiás” (Waibel 1948, Vanzolini 1994: 176).

As to “meandering belts of rainforest”, they are a figment of the imagination. There are neither rainforests nor meanders in the cerrados (Ab’Saber 1971: 2). On the other hand, enclaves of forest, of different types but (and this is the really significant point) with closed canopy, are another characteristic feature of cerrados (Labouriau 1966, Ab’Saber 1977).

Finally, “seasonal floodings” are also a constant trait of the physiology of cerrado rivers. They happen at the beginning of summer (wet season) and are responsible for two universally acknowledged key geomorphic processes, the shaping of the levees and the deposition of sediment in the backswamp.

Webb and Rancy (*l.c.*) consider their conclusion similar to that of Räsänen et al. (1987). However, the proposal of these Finnish authors has nothing to do with savannas or with meandering forest belts. They literally state in the Abstract of their article (*l. c.*: 1398): “...in the basinal forelands of the tectonically active Andes, the geological setting causes long-term fluvial perturbation. This leads to a temporally structured highly complex mosaic of fossil and present floodplains. These dynamics have been present with varying activity and geographic range during the Tertiary and Quaternary, providing site-turnover that has not been recognized by the biogeographic tradition of the Amazon basin”.

One must first consider how a model fitted to the basinal forelands of the Andes can be applied to the whole of Amazonia. Second, how could a fluid mosaic of “fossil and present floodplains” (of unspecified ecology, probably varzea forest) result in geographic, or any kind of isolation.

In what concerns me as a practicing systema-

tist and biogeographer, this model of Räsänen et al. (1987) is just a game of words. (It has also been criticized, and found wanting, from other viewpoints – Dumont et al. 1990, Latrubesse 1996).

One paleomammalogical paper that escaped Webb and Rancy's (1996) review is that by Campbell and Frailey (1985), who found a land mammal fauna with the same ecological implications in the nearby Rio Acre, in Peru.

Three additional paleomammalogical papers confirm the presence of cerrado-adapted faunas in the Late Pleistocene of southwestern Amazonia: Frailey (1986); Sant'Anna-Filho et al. (1996) and Latrubesse and Rancy (1998). The latter convincingly review the geology of this area so fertile in Quaternary fossils.

Some papers that I find important were omitted in the *Amazoniana* (2001) debate. Latrubesse and Ramonell (1994) review the paleontological literature and, with basis on recent analogs, propose a most interesting model of wind circulation and general climate for southwestern Amazonia during the Last Glacial. They conclude that the area, then under cerrado, had a longer and more pronounced dry season than at present, and that the short spells of cold weather ("friagens") consequent upon the penetration of polar fronts were more frequent and ecologically important (see also Iriondo and Latrubesse 1994).

Dumont et al. (1990) deal with the neotectonics of the Upper Amazon, and conclude that "the great age and stability of the upland ecosystems, and the rather constant ecological conditions (including gap-phase regeneration) in the under-story are still the most convincing explanation of the high species diversity within upland forest." This belated revival, simply stated, without any support, of the gratuitous "Urwald" concept (Schwabe 1969, critique in Vanzolini and Williams 1970: 108), apparently shared by Colinvaux (1996), needs hardly any comment in the light of current research on speciation. But, for the simple sake of argument, it could again be asked, how to extend a strictly juxta-Andean process to the

whole of Amazonia?

Finally, there is a most interesting detailed scenario of Amazonian ecology in the Late Pleistocene (Iriondo and Latrubesse 1994: 127): the general vegetation would have been cerrado, with a strongly seasonal climate, rainfall 850-1000 mm/yr, the dry atmosphere resulting in limited local recycling of water as precipitation. This seems to me a very sensible scenario, except for the rainfall, that I find too low.

The llanos of the Orinoco, that Iriondo and Latrubesse cite as a modern analog, receive 1300-1400 mm/yr (Tamayo 1972). Turning to Brasil, I took data (Brasil, Escritório de Meteorologia, 1968) from twelve localities I am sure are in the core of the cerrados; the range was 1072-1665 mm/yr, the mean 1428 ± 60.6 mm. For eight equally undoubted caatinga localities I got a range of 497-922 mm/yr, mean 712 ± 49.5 mm. Beyond doubt, the conjecture of Iriondo and Latrubesse fits caatingas better than cerrados. Caatingas certainly have existed in Amazonia: they left behind white sands and stone-lines (Ab'Saber 1979). But it would be rash to generalize their climatic features to all of Late Pleistocene Amazonia; Ab'Saber himself (*l.c.*) makes the point very clearly.

So I conclude that, after all these years and controversies, the refuge model stands virtually as originally proposed. The only change I would suggest would be to substitute a model of morphoclimatic domains for the simple consideration of types of vegetation.

A morphoclimatic domain (Ab'Saber 1967, 1977) is an area of sub-continental extent with characteristic features of topography, drainage, climate, soil and vegetation. In the case of the cerrados the topography is one of low plateaus, gentle, rolling at most. The drainage is dendritic, highly hierarchical. The climate is mesic, with a dry, cool winter and a wet warm summer. The soils are very profound (up to 30 meters), and guarantee water supply to the deep roots of the vegetation, during the dry season. Cerrados are not summer-deciduous,

APPENDIX

Specimens used.

Specimen	Locality	Body + tail length	Head width	Body + tail annuli	Segments	Pores
	Panamá					
MZUSP 2101	Panamá City	295 + x	10	200 + x	46	8
MCZ 18924	Gatun	280 + 41	–	195 + 27	44	8
MCZ 18925	Monte Lirio	315 + x	–	200 + x	44	8
MCZ 22070	Barro Colorado	315 + 46	–	197 + 24	44	8
MCZ 24003	Frijoles	240 + x	–	200 + x	46	8
MCZ 24004	Monte Lirio	285 + 42	–	196 + 23	48	8
MCZ 37106	Chagres River	330 + x	–	202 + x	46	8
MCZ 37874	Panamá City	240 + 36	–	195 + 24	44	8
USNM 37859	Canal Zone	305 + x	–	201 + x	48	8
USNM 38399	Bohio	350 + 52	–	198 + 24	48	8
	Colombia					
MZUSP 57566	Villavicencio	328 + x	12	201 + x	50	8
AMNH 18237	Caño Las Animas	315 + 47	–	192 + 25	48	8
AMNH 27495	Honda	285 + 45	–	201 + 25	46	8
AMNH 27496	Muzo	310 + 45	–	196 + 24	48	7
AMNH 37439	San Pedro	285 + 40	–	201 + 24	46	6
AMNH 37440	Medellin	280 + x	–	196 + x	40	6
AMNH 37441	''	115 + 19	–	195 + 25	48	7
AMNH 37455	Honda	315 + 50	–	205 + 26	50	8
AMNH 62137	Villavicencio	440 + x	–	206 + x	50	8
BM 714	Peña Lisa	325 + 45	–	202 + 24	46	7
CM 30583	Villavicencio	235 + 38	–	208 + 26	48	8
MCZ 17647	Paime	290 + 45	–	196 + 24	42	x
MCZ 25052	Rio Frio	320 + 50	–	199 + 25	42	7
CM 27575	Barrancabermeja	225 + 35	–	200 + 27	42	8
MCZ 29683	Andagoya	405 + 58	–	194 + 24	42	6
MCZ 48959	Leticia	340 + x	–	214 + x	42	8
MCZ 48960	''	250 + 40	–	221 + 26	50	10
MCZ 42189	Muzo	265 + 33	–	190 + 23	42	6
MCZ 46437	''	250 + 34	–	197 + 24	42	6
MCZ 48961	Leticia	255 + 39	–	207 + 25	48	8
SU 8272	Villavicencio	400 + 55	–	207 + 25	52	9
SU 8273	''	190 + 30	–	199 + 25	50	8
	Venezuela					
MNH 69728	Caripito	175 + 25	–	206 + 28	46	6
MNH 69729	''	260 + 40	–	209 + 27	48	6
MCZ 49049	Riecito	260 + 39	–	200 + 26	50	8

APPENDIX (continuation)

Specimen	Locality	Body + tail length	Head width	Body + tail annuli	Segments	Pores
	Trinidad					
AMNH 6968	Caparu	260 + 42	–	206 + 26	46	8
AMNH 64440	Tucker Valley	265 + 40	–	211 + 26	46	8
AMNH 64462	''	215 + 30	–	205 + 24	42	8
AMNH 64462 A	''	250 + 38	–	210 + 25	44	7
AMNH 64462 B	''	210 + 29	–	212 + 25	42	6
AMNH 64462 C	''	195 + 25	–	203 + 24	46	6
AMNH 64523	''	250 + x	–	204 + x	42	7
AMNH 64523 A	''	200 + 26	–	209 + 27	40	6
AMNH 64523 B	''	235 + 36	–	204 + 27	42	6
BM 746	–	270 + 36	–	204 + 27	44	6
BM 749	–	293 + 45	–	207 – 25	44	7
CM 49910	Brackfield	240 + 37	–	207 + 27	44	8
CM 49912	San Rafael	245 + 39	–	206 + 24	44	8
CM 49912 A	''	280 + 42	–	206 + 26	46	8
MCZ 10137	Toco	310 + x	–	205 + x	44	6
USNM 5751 A	–	262 + 41	–	202 + 25	46	6
USNM 5751 B	–	210 + 26	–	205 + 26	50	8
	Guyana					
MZUSP 2000	Kalacoon	250 + 39	8	205 + 26	44	9
AMNH 1085	Kaieteur Falls	195 + 29	–	199 + 25	44	9
AMNH 7295	Georgetown	305 + 39	–	207 + 24	46	9
AMNH 17688	''	230 + 31	–	208 + 25	46	8
AMNH 17689	''	420 + 55	–	213 + 24	42	8
AMNH 25088	Kamakusa	205 + x	–	204 + x	40	8
AMNH 60907	Marudi River	230 + 32	–	218 + 29	46	8
BM 743	Nappi, Kanuku Mts.	265 + 40	–	202 + 26	48	10
BM 747	Maccasseema	180 + x	–	205 + x	50	10
BM 748	''	235 + 35	–	207 + 24	50	8
	Suriname					
MZUSP 2001	Paramaribo	145 + 22	6	205 + 25	48	8
MZUSP 9678	Christiaankondre	130 + 19	6	192 + 26	x	8
AMNH 8134	Paramaribo	240 + 36	–	207 + 26	46	9
AMNH 8135	''	285 + x	–	206 + x	52	8
AMNH 8649	''	215 + x	–	196 + x	48	10
AMNH 8650	''	275 + 39	–	206 + 25	50	8
AMNH 8678	''	135 + 21	–	199 + 27	46	8
MCZ 15719	–	350 + 51	–	216 + 25	46	8
USNM 6118	–	370 + x	–	205 + x	52	8
USNM 58739	–	225 + 38	–	203 + 27	54	9
	French Guiana					
CAS 14546	St. Laurent du Maroni	270 + x	–	211 + x	56	10

APPENDIX (continuation)

Specimen	Locality	Body + tail length	Head width	Body + tail annuli	Segments	Pores
Brasil: Roraima						
MZUSP 49351	Cachoeira Cujubim	305 + 41	11	198 + 23	48	x
MZUSP 62841	Vila Pacaraima	315 + 50	10	201 + 25	50	9
MZUSP 62842	"	320 + 56	13	199 + 26	52	9
MZUSP 64961	"	360 + 54	15	197 + 24	50	8
MZUSP 69629	Apiáú	264 + 41	10	198 + 24	50	x
MZUSP 69630	Maloca Mangueira	280 + x	12	191 + 25	50	8
MZUSP 70266	Rio Barauana/BR-210	250 + 38	9	187 + 23	x	7
MZUSP 73760	Ilha de Maracá	255 + 42	9	185 + 25	50	7
MZUSP 78112	Tepequém	275 + 44	10	183 + 24	50	8
MZUSP 79301	Ilha de Maracá	257 + x	9	201 + x	48	8
Brasil: Amazonas						
IB 673	Manaus	260 + 33	–	212 + 25	48	8
IB 674	"	240 + 30	–	214 + 26	48	8
MZUSP 1927	"	290 + 40	–	219 + 26	50	9
MZUSP 7664	Tabatinga	210 + x	8	208 + x	48	9
MZUSP 10914	Manaus	227 + 32	9	213 + 23	46	8
MZUSP 16928	Itapiranga	247 + 33	9	224 + 24	48	8
MZUSP 26012	Manaus	445 + 67	8	219 + 25	46	9
MZUSP 26013	Manaus	490 + 65	17	206 + 22	46	9
MZUSP 26014	"	251 + x	10	212 + x	46	9
MZUSP 26015	"	393 + x	16	212 + x	50	8
MZUSP 28238	Humaitá	285 + 43	11	195 + 22	48	8
MZUSP 38371	Berurí	178 + 51	14	212 + 24	48	8
MZUSP 51617	Manaus	301 + 41	10	215 + 25	48	8
MZUSP 52876	"	193 + x	7	213 + x	46	8
MZUSP 57559	"	275 + x	12	207 + x	44	8
MZUSP 57863	"	285 + 39	10	214 + 24	48	8
MZUSP 57864	"	210 + x	8	–	44	8
MZUSP 63643	Balbina	325 + x	13	210 + x	48	8
MZUSP 68641	"	342 + 43	11	214 + 24	48	8
MZUSP 68642	"	270 + 42	10	214 + 26	52	8
AMNH 37442	Cucuí	390 + x	–	208 + x	44	8
AMNH 37443	"	205 + 32	–	214 + 29	42	6
AMNH 37444	Vista Alegre	285 + 41	–	202 + 25	46	8
AMNH 64917	Manaus	152 + x	–	210 + x	46	8
AMNH 64918	"	170 + 21	–	207 + 25	44	8
AMNH 64919	"	280 + 37	–	211 + 24	48	8
MCZ 19540	"	230 + 29	–	220 + 24	48	8
MCZ 20663	"	270 + 39	–	215 + 26	50	8
MCZ 20664	"	330 + 40	–	212 + 25	48	8
MCZ 20665	"	170 + 25	–	220 + 28	48	8

APPENDIX (continuation)

Specimen	Locality	Body + tail length	Head width	Body + tail annuli	Segments	Pores
	Brasil: Pará					
MZUSP 3364	Jacareacanga	295 + 38	13	198 + 22	48	9
MZUSP 4714	Tomé Açu	225 + 30	10	197 + 19	48	8
MZUSP 4715	Tomé Açu	275 + 34	12	202 + 20	48	8
MZUSP 4716	"	280 + 32	11	196 + 20	50	8
MZUSP 6822	Belém	-	-	206 + x	48	x
MZUSP 6823	"	-	-	214 + 22	46	8
MZUSP 6824	"	322 + x	12	208 + x	46	8
MZUSP 7653	BR-010 km 93	342 + x	12	208 + 21	48	8
MZUSP 8518	Belém	322 + 44	12	207 + 22	48	8
MZUSP 12337	Canindé	275 + 37	11	208 + 23	50	8
MZUSP 12356	Oriximiná	230 + 23	9	198 + 22	44	8
MZUSP 12357	"	206 + 27	8	191 + 22	42	8
MZUSP 13855	"	174 + 21	7	196 + 23	44	8
MZUSP 17436	"	280 + x	11	192 + 21	46	9
MZUSP 36497	"	220 + 27	8	191 + 20	44	8
MZUSP 36498	"	252 + 29	9	195 + 21	48	8
MZUSP 77476	"	300 + x	11	196 + x	44	8
MZUSP 77823	"	252 + x	9	199 + x	46	8
MZUSP 78409	Serra Kukoinhokren	410 + x	12	208 + x	46	8
	Brasil: Maranhão					
MZUSP 14439	Açailândia	335 + 50	12	205 + 23	56	7
	Brasil: Acre					
MZUSP 5384	Tarauacá	292 + 47	10	202 + 23	40	8
MZUSP 53456	Porto Walter	385 + x	17	198 + x	40	8
	Brasil: Rondonia					
MZUSP 64736	Santa Barbara	207 + 30	8	189 + 23	44	8
MZUSP 64738	RO-399 km 5	320 + x	12	205 + 24	46	6
MZUSP 64763	Alto Paraíso	317 + x	11	199 + x	46	8
MZUSP 64908	RO-399 km 1	255 + 34	9	204 + 24	40	8
MZUSP 64909	RO-399 km 1	248 + x	10	202 + x	44	7
MZUSP 64910	RO-399 km 1	275 + 40	11	196 + 23	46	8
MZUSP 66219	Cachoeira Nazaré	150 + 20	6	185 + 24	46	7
MZUSP 64923	Santa Barbara	265 + x	9	195 + 23	46	8
MZUSP 66348	Cachoeira Nazaré	245 + 34	10	184 + 24	44	8
	Brasil: Mato Grosso					
MZUSP 81603	Aripuanã	370 + x	16	198 + 25	46	-
MZUSP 82547	"	288 + 40	12	204 + 24	46	8
MZUSP 82798	"	341 + 48	11	198 + 24	44	9
MZUSP 82799	"	385 + 52	14	195 + 24	46	10
MZUSP 82800	"	310 + x	12	206 + x	52	-
MZUSP 82805	"	370 + x	14	301 + 25	46	8

APPENDIX (continuation)

Specimen	Locality	Body + tail length	Head width	Body + tail annuli	Segments	Pores
	Brasil: Goiás					
MZUSP 80223	Piracanjuba	294 + 39	10	180 + 23	52	8
MZUSP 83397	Serra da Mesa	345 + 50	13	192 + 24	52	8
MZUSP 83398	"	300 + 41	11	199 + 25	50	9
MZUSP 83399	"	324 + x	12	193 + x	52	8
MZUSP 83400	"	332 + 45	14	202 + 26	52	9
MZUSP 83401	"	284 + 40	11	192 + 26	52	8
MZUSP 83402	"	315 = 45	13	191 + 25	52	9
MZUSP 83403	"	338 + x	14	196 + x	48	8
MZUSP 83404	"	333 + x	13	196 + x	48	6
MZUSP 83405	"	335 + x	14	198 + x	52	8
MZUSP 83406	"	339 + x	15	194 + x	50	8
MZUSP 87694	"	353 + 55	15	195 + 24	50	8
MZUSP 87695	"	330 + 47	15	189 + 27	52	8
	Brasil: Bahia					
AMNH 1091	Santa Maria Eterna	335 + 51	–	206 + 24	52	9
AMNH 1092	"	280 + 49	–	197 + 28	52	10
AMNH 1093	"	250 + x	–	202 + x	50	8
	Ecuador					
MZUSP 3404	Canelos	130 + 20	6	197 + 27	40	7
MZUSP 6425	Santo Domingo de los Colorados	280 + 41	10	196 + 22	44	7
MZUSP 6426	"	235 + 42	12	197 + 22	46	7
MZUSP 6427	"	322 + 52	12	193 + 23	46	6
MZUSP 6428	Canelos	312 + x	11	198 + x	42	8
MZUSP 6429	Loreto	228 + 51	12	202 + 26	44	8
MZUSP 6430	"	310 + 55	9	202 + 27	44	8
MZUSP 6431	Pichilingue	339 + 50	12	190 + 23	44	7
AMNH 2285	Riobamba	340 + x	–	197 + x	44	8
AMNH 22090	Bucay	350 + 55	–	198 + 25	44	7
AMNH 23324	Riobamba	275 + 44	–	204 + 26	44	8
AMNH 23325	"	360 + 62	–	203 + 29	44	8
AMNH 23326	"	295 + 45	–	202 + 26	42	8
AMNH 28784	Turula	380 + 64	–	202 + 27	44	9
AMNH 28785	"	350 + x	–	200 + x	42	8
AMNH 37862	Baños to Canelos	250 + x	–	205 + x	44	9
AMNH 56593	Rio Cenipa	315 + x	–	196 + x	42	8
AMNH 56595	"	290 + x	–	209 + x	44	7
BM 741	Guayaquil	315 + 51	–	200 + 28	44	9
BM 742	Santo Domingo de los Colorados	295 + 42	–	193 + 23	46	8
BM 745	50 km E Loja	275 + 41	–	199 + 29	42	8

APPENDIX (continuation)

Specimen	Locality	Body + tail length	Head width	Body + tail annuli	Segments	Pores
Ecuador (cont.)						
MCZ 3572	Rio Daule	220 + 34	–	196 + 27	44	8
MCZ 7799 A	Riobamba	345 + 50	–	199 + 23	44	8
MCZ 7799 B	''	320 + 46	–	205 + 25	46	7
MCZ 37271	Rio Pastaza	370 + x	–	207 + x	–	–
SU 8259	Avila	365 + 60	–	200 + 26	42	8
Peru						
AMNH 56224	Iquitos	250 + x	–	208 + 26	44	8
AMNH 56310	Pampa Hermosa	365 + 62	–	208 + 28	44	8
AMNH 56588	Iquitos	450 + 75	–	206 + 27	44	8
AMNH 86589	''	280 + 47	–	209 + 28	44	8
AMNH 56590	''	270 + 46	–	205 + 29	44	8
AMNH 56592	''	330 + x	–	204 + x	42	8
AMNH 56594	Contamana	105 + 16	–	210 + 26	38	8
AMNH 56596	Iquitos	365 + 64	–	206 + 29	44	8
AMNH 56597	''	360 + 61	–	211 + 30	44	8
AMNH 56598	''	305 + 55	–	209 + 29	44	8
AMNH 56599	''	255 + 52	–	210 + 29	44	8
AMNH 56600	''	275 + 51	–	210 + 30	44	8
AMNH 56601	Reforma	315 + 56	–	204 + 29	44	8
AMNH 56602	Iquitos	300 + 50	–	204 + 28	42	8
AMNH 56603	''	245 + 39	–	207 + 27	44	8
AMNH 56605	''	340 + 57	–	205 + 29	44	8
AMNH 56606	Roaboya	275 + 42	–	211 + 28	42	7
AMNH 56607	''	275 + 45	–	211 + 28	42	7
AMNH 56608	Requena	205 + 35	–	215 + 29	42	8
AMNH 56609	Contamana	295 + 50	–	208 + 29	44	8
AMNH 56610	Iquitos	395 + 75	–	208 + 29	44	8
AMNH 56611	Rio Cushabatay	305 + 46	–	211 + 27	44	8
AMNH 56612	Boca Santiago	300 + 55	–	205 + 28	42	8
AMNH 56613	Roaboya	245 + 39	–	212 + 26	42	7
AMNH 56617	Iquitos	370 + 60	–	206 + 29	44	8
AMNH 56618	''	360 + 61	–	210 + 30	44	8
BM 703	Rio San Gaban	275 + 40	–	207 + 27	40	8
BM 704	''	335 + 73	–	205 + 27	42	7
BM 705	Rio Cushabatay	270 + x	–	210 + x	40	8
BM 740	Rio San Gaban	275 + 41	–	207 + 27	40	7
CM 45471	Iquitos	341 + x	–	213 + x	44	8
CM 45472	''	270 + 40	–	208 + 26	46	6
MCZ 45784	Pucallpa	325 + 48	–	215 + 29	38	8

GAZETTEER

South American localities of *Amphisbaena fuliginosa*.

Latitudes South and longitudes West taken for granted.

The last number of each entry corresponds to Map 1.

The coordinates of rivers are, unless otherwise specified, those of the mouth.

- Açailândia (Brasil, Maranhão) 0547, 4741, 49
Alto Paraíso (Brasil, Rondonia) 0937, 6327, 54
Andagoya (Colombia, Chocó) 0506 N, 7640, 5
Apiáú (Brasil, Roraima) 0240 N, 6115, 19
Aripuanã (Brasil, Mato Grosso) 1020, 5923, 5
Avila (Ecuador, Napo) 0038, 7725, 26
- Balbina, UHE (Usina Hidroelétrica) (Brasil, Amazonas) 0153, 5928, 32
Baños (Ecuador, Pastaza) 0124, 7825, 27
Barauana, Rio, at Highway BR-210 (Brasil, Roraima) 0128 N, 6052, 22
Barrancabermeja (Colombia, Santander) 0703 N, 7352, 2
Belém (Brasil, Pará) 0126, 4829, 28
Berurí (Brasil, Amazonas) 0354, 6122, 41
Boca Santiago (Peru, Amazonas) 0427, 7738, 45
Bucay (Ecuador, Guayas) 0210, 7907, 33
- Canelos (Ecuador, Pastaza) 0135, 7744, 29
Canindé, Rio Gurupí (Brasil, Pará) 0233, 4631, 37
Caripito (Venezuela, Monagas) 1007 N, 6305, 32
Cashiboya (Peru, Loreto) 0651, 7440, 50
Cenipa, Rio (Peru, Amazonas) 0439, 7810, 46
Charobamba (Bolivia, La Paz) 1437, 6844, 65
Christiaankondre (Suriname) 0543 N, 5401, 7
Contamana (Peru, Loreto) 0720, 7502, 51
Cucuí (Brasil, Amazonas) 0113 N, 6650, 23
Cujubim, Cachoeira do (falls), Rio Catrimani (Brasil, Roraima) 0145 N, 6217, 21
Cushabatay, Rio (Peru, Loreto) 0709, 7508, 51
- Daule, Rio (Ecuador, Guayas), 0210, 7952, 18
- Georgetown (Guyana) 0647 N, 5811, 3
Guayaquil (Ecuador, Guayas) 0210, 7952, 18
- Honda (Colombia, Tolima) 0512 N, 7445, 10
Humaitá (Brasil, Amazonas) 0731, 6302, 62
- Iquitos (Peru, Loreto) 0347, 7313, 40
Itapiranga (Brasil, Amazonas) 0244, 5801, 38
- Jacareacanga (Brasil, Pará) 0615, 744, 63
-

GAZETTEER (continuation)

Latitudes South and longitudes West taken for granted.

The last number of each entry corresponds to Map 1.

The coordinates of rivers are, unless otherwise specified, those of the mouth.

Kaieteur Falls (Guyana) 0510 N, 5928, 11

Kalacoon (Guyana) 0623 N, 5838, 4

Kamakusa (Guyana) 0556 N, 5954, 6

km 1 Highway RO-399 (Brasil, Rondonia) 1244, 6016, 57

km 5 Highway RO-399 (Brasil, Rondonia) 1242, 6011, 57

km 93 Highway BR-010 (Brasil, Pará) 0212, 4734, 34

Kukoinhokren, Serra de (Brasil, Pará) 0750, 5155, 64

Las Animas, Caño (Colombia, Chocó) 0520 N, 7643, 5

Leticia (Colombia, Amazonas) 0409, 6957, 43

Loja (Ecuador, Loja) 0400, 7910, 42

Loreto (Ecuador, Napo) 0038, 7719, 11

Maccasseema (Guyana) 0715 N, 5843, 1

Manaus (Brasil, Amazonas) 0307, 6000, 39

Mangueira, Maloca (Indian village) (Brasil, Roraima) 0318, N, 6127, 18

Maracá, Ilha de, Rio Uraricoera (Brasil, Roraima) 0325 N, 6140, 16

Marudí River (Guyana) 0207 N, 5934, 20

Medellin (Colombia, Antioquia) 0615 N, 7536, 5

Mesa, Serra da (Brasil, Goiás) 1415, 4835, 56

Muzo (Colombia, Boyacá) 0532 N, 7406, 8

Nappi, Kanuku Mts. (Guyana) 0325 N, 5934, 17

Nazaré, Cachoeira de (falls), Rio Machado or Ji-Paraná (Brasil, Rondonia) 0945, 6155, 55

Oriximiná (Brasil, Pará) 0146, 5551, 31

Pacaraima, Vila (border-marker BV-8) (Brasil, Roraima) 0430 N, 6109, 12

Paima (Colombia, Cundinamarca) 0522 N, 7410, 9

Pampa Hermosa (Peru, Loreto) 0712, 7518, 51

Paramaribo (Suriname) 0505 N, 5510, 40

Pastaza, Rio (Ecuador, Pastaza) 0455, 7624, 47

Peña Lisa (Colombia, Chocó) 0506 N, 7637, 5

Pichilingue (Ecuador, Los Rios) 0106, 7928, 12

Piracanjuba (Brasil, Goiás) 1718, 4901, 62

Porto Walter (Brasil, Acre) 0816, 7244, 52

Pucallpa (Peru, Ucayali) 0823, 7433, 53

Reforma (Peru, Loreto) 0644, 7458, 50

Requena (Peru, Loreto) 0504, 7353, 48

Riecito (Venezuela, Falcón) 1055 N, 6844, 31

Riobamba (Ecuador, Chimborazo) 0140, 7839, 30

GAZETTEER (continuation)

Latitudes South and longitudes West taken for granted.

The last number of each entry corresponds to Map 1.

The coordinates of rivers are, unless otherwise specified, those of the mouth.

Rio Frio (locality) (Colombia, Magdalena) 1054 N, 7411, 1

Roabaya (Peru, Loreto) 0748, 7446, 51

Saint Laurent du Maroni (French Guiana) 0530 N, 5402, 42

San Gaban, Rio (Peru, Puno) 1325, 7018, 30

San Pedro (Colombia, Valle del Cauca) 0400 N, 7613, 14

Santa Barbara (Brasil, Rondonia) 0910, 6304, 54

Santa Maria Eterna (Brasil, Bahia) 1551, 3925, 59

Santo Domingo de los Colorados (Ecuador, Pichincha) 0015, 7929, 24

Sorata (Bolivia, La Paz) 1547, 6840, 66

Tabatinga (Brasil, Amazonas) 0414, 6955, 44

Tarauacá (Brasil, Acre) 0810, 7046, 60

Tepequém (Brasil, Roraima) 0345 N, 6145, 15

Tipuani (Bolivia, La Paz) 1533, 6800, 67

Tomé Açu (Brasil, Pará) 0224, 4809, 36

Turula, Rio Upano (Ecuador, Morona – Santiago) 0213, 7811, 35

Villavicencio (Colombia, Meta) 0409 N, 7337, 13

Vista Alegre, Rio Negro (Brasil, Amazonas) 0021, 6427, 25

Yungas (Bolivia, La Paz) 1610, 6730, 68

contrary to other formations (e.g. African) also called ‘savannas’, and this makes untenable the adoption of a same name for both formations. Cerrado vegetation is highly characteristic, both physiognomically and taxonomically (Heringer et al. 1976); it includes several sub-types of cerrados, from open fields to dry forests, as well as gallery forests and forest enclaves of diverse types and extent. This is a very different, and much richer, scenario than the simplistic ones usually envisaged.

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RESUMO

A diferenciação geográfica de *Amphisbaena fuliginosa* (Reptilia, Amphisbaenia, Amphisbaenidae) é estudada com base em 220 exemplares de 99 localidades definidas e 2 localidades genéricas. O padrão encontrado apoia a conclusão de Vanzolini (1951) de que são reconhecíveis sub-espécies, definidas pelo padrão de colorido e por 4 caracteres merísticos. Esse padrão de diferenciação resultaria de eventos paleoclimáticos Quaternários. Contribuições recentes sobre modelos de diferenciação no Quaternário da América tropical são brevemente discutidos.

Palavras-chave: sub-espécie, paleoclima, *Amphisbaena*, modelo e refúgios.

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