



Ciência Rural

ISSN: 0103-8478

cienciarural@mail.ufsm.br

Universidade Federal de Santa Maria
Brasil

Hideki Sumida, Ciro; Canteri, Marcelo Giovanetti; Casaroto Peitl, Douglas; Tibolla, Fabiana; Orsini, Idenize Pedrina; Araújo, Felipe André; Fonseca Chagas, Débora; Sanches Calvos, Natália

Chemical and biological control of Sclerotinia stem rot in the soybean crop

Ciência Rural, vol. 45, núm. 5, mayo, 2015, pp. 760-766

Universidade Federal de Santa Maria

Santa Maria, Brasil

Available in: <http://www.redalyc.org/articulo.oa?id=33138346002>

- How to cite
- Complete issue
- More information about this article
- Journal's homepage in redalyc.org

redalyc.org

Scientific Information System

Network of Scientific Journals from Latin America, the Caribbean, Spain and Portugal

Non-profit academic project, developed under the open access initiative

Chemical and biological control of *Sclerotinia* stem rot in the soybean crop

Controle químico e biológico da podridão branca da haste na cultura da soja

Ciro Hideki Sumida^{1*} Marcelo Giovanetti Canteri¹ Douglas Casaroto Peitl¹
Fabiana Tibolla¹ Idenize Pedrina Orsini¹ Felipe André Araújo¹
Débora Fonseca Chagas¹ Natália Sanches Calvos¹

ABSTRACT

It was evaluated the effect of fungicides and the microbial control agent *Trichoderma harzianum* on the inhibition of the carpogenic and ascospore germination of *Sclerotinia sclerotiorum*. This study also evaluated the chemical, fungicidal and microbial control of white mold or *Sclerotinia* stem rot of soybean in the field. Three experiments were conducted, as follows: 1) inhibition of carpogenic germination of sclerotia, 2) inhibition of ascospore germination, and 3) control of *Sclerotinia* stem rot in a soybean crop under field conditions. The treatments evaluated were fluazinam, procymidone, iprodione, thiophanate-methyl, carbendazim, benzalkonium chloride + fluazinam, and *T. harzianum*. Procymidone resulted in an inhibition of 13.5% and benzalkonium chloride in an inhibition of 13.9% in an ascospore germination test. Fluazinam and procymidone were the most effective in reducing the production of ascospores/apothecium, representing 65.6% and 82.4% of inhibition. Procymidone and fluazinam if combined or not with benzalkonium chloride were the most effective in controlling *sclerotinia* stem rot under field conditions when applied at the onset of flowering and 15 days later. In the 2009-10 harvest, these two fungicides reduced the incidence of *Sclerotinia* stem rot by 73.1 and 71.6% and in the 2010-11 harvest by 75.7 and 77.6%, respectively.

Key words: *Glycine max*, *Trichoderma harzianum*, apothecium, ascospore, sclerotium, soybean.

RESUMO

Avaliaram-se o efeito de fungicidas e agente de controle biológico *Trichoderma harzianum* na inibição da germinação carpogênica e de ascósporos de *Sclerotinia sclerotiorum*. Também foram avaliados os controles químico e biológico do mofo branco ou podridão branca da haste na cultura da soja em condições de campo. Foram desenvolvidos três ensaios, sendo: 1) inibição da germinação carpogênica; 2) inibição da germinação de ascósporos; e 3) controle da podridão branca da haste na cultura da soja em condições de campo. Os tratamentos

avaliados foram fluazinam, procimidona, iprodione, tiofanato metílico, carbendazim, cloreto de benzalcônio + fluazinam e *T. harzianum*. O procimidona resultou em inibição de 13,5% e o cloreto de benzalcônio + fluazinam em 13,9% de inibição no teste de inibição da germinação dos ascósporos. Fluazinam e procimidona foram os mais eficientes na redução da produção de ascósporos/apotécio representando 65,6% e 82,4% de inibição. O procimidona e fluazinam se combinados com cloreto de benzalcônio ou não foram os mais eficientes no controle da podridão branca da haste em condições de campo, quando aplicados no início do florescimento e 15 dias após. Na safra 2009-10, esses dois fungicidas reduziram a incidência da podridão branca da haste, em 73,1 e 71,6% e, na safra 2010-11, reduziram em 75,7 e 77,6%, respectivamente.

Palavras-chave: *Glycine max*, *Trichoderma harzianum*, apotécio, ascósporos, escleródio, soja.

INTRODUCTION

White mold or *Sclerotinia* stem rot caused by *Sclerotinia sclerotiorum* (Lib.) de Bary is a growing threat to the soybean crop (DHINGRA et al., 2009) and is of great concern to producers and researchers in countries such as Brazil (LOBO JUNIOR & ABREU, 2000) and the United States, causing losses of some 1.6 million tons in 2004 alone (WRATHER & KOENNING, 2011).

Conventional methods to control the disease include crop management (FERRAZ et al., 2003), use of resistant cultivars, and chemical control (BARDIN & HUANG, 2001). With regard to chemical control, there are reports of inconsistent and ineffective results owing to application problems

¹Departamento de Agronomia, Universidade Estadual de Londrina (UEL), Rodovia Celso Garcia Cid, PR 445, Km 380, CP 6001, 86051-990, Londrina, PR, Brasil. E-mail: cirosumida@hotmail.com. *Corresponding author.

(COSTA & COSTA, 2004); however, according to TU (1989), several fungicides are considered effective in controlling *Sclerotinia* stem rot and are recommended for application during flowering. MUELLER et al. (2004) have also reported the use of fungicides for controlling the disease as an effective control strategy because of the flexibility of application timing in comparison with other crop control methods. Alternative methods such as biological control have also been attempted (HJELJORD & TROSMO, 2005) using fungi such as *Trichoderma* sp., which is characterized as a mycoparasite of *S. sclerotiorum*, capable of colonizing mycelia (ABDULLAH et al., 2008) and sclerotia (HUANG & ERICKSON, 2008).

Therefore, the aim of this study was to evaluate the effects of fungicides and a biological product based on *Trichoderma harzianum* on the inhibition of the carpogenic and ascospore germination of *S. sclerotiorum* and the control of *Sclerotinia* stem rot in the soybean crop under field conditions.

MATERIAL AND METHODS

Isolation of *S. Sclerotiorum*

An isolate of *S. sclerotiorum* was obtained by collecting diseased plants from commercial soybean crops in the municipality of Mauá da Serra, Paraná, Brazil. Sclerotia were produced, separated, and conditioned at $7.0 \pm 3^\circ\text{C}$ for 8 months before use in the experiments.

Fungicide inhibition of carpogenic germination of *S. Sclerotiorum*

The experiment was performed in plastic germination boxes ("gerboxes" of $0.11 \times 0.11 \times 0.35\text{m}$) containing 0.24L of soil [dark red latosol (Oxisol)] according to the method described by COSTA & COSTA (2004) but with different numbers of sclerotia/box. The experimental design was fully randomized with eight treatments and four replications. The soil was prepared by sieving, moistening and autoclaving for 2h at 120°C and 1atm pressure. After the soil was dried in an oven at 56°C for 24h, 0.2L of the soil and 0.06L of sterilized distilled water were placed in each box. Then, 100 sclerotia/gerbox were placed in the soil and covered with 40mL of the soil, forming a layer of approximately 2.0mm thickness over sclerotia.

Treatments were prepared at the recommended doses in 0.06L spray containers as follows: T1 (control), T2 (fluazinam 0.5L of the active ingredient. ha^{-1} or 0.06g a.i.gerbox $^{-1}$), T3 (procymidone 0.5kg a.i. ha^{-1} or 0.06g a.i.gerbox $^{-1}$), T4 (iprodione 0.5kg a.i. ha^{-1} or 0.06g a.i.gerbox $^{-1}$),

T5 (thiophanate-methyl 0.5kg a.i. ha^{-1} or 0.06g a.i.gerbox $^{-1}$), T6 (carbendazim 0.5kg a.i. ha^{-1} or 0.06g a.i.gerbox $^{-1}$), T7 (benzalkonium chloride 0.1kg a.i. ha^{-1} or 0.012g a.i.gerbox $^{-1}$ + fluazinam 0.5kg a.i. ha^{-1} or 0.06g a.i.gerbox $^{-1}$), and T8 (commercial product based on *T. harzianum* 5×10^{10} conidia. ha^{-1} or 6×10^6 conidia.gerbox $^{-1}$). Approximately 240 μL (200L ha^{-1}) of solution of fungicide and suspension of *Trichoderma* were sprayed into each gerbox, followed by incubation at $20 \pm 2^\circ\text{C}$ for 45 days with a photoperiod of 12h. First, it was evaluated the number of germinated sclerotia, stipes, and apothecia, and then, it was calculated inhibition by comparing treatments with the control. The variables evaluated were the inhibition of sclerotia germination (ISG), inhibition of carpogenic germination (ICG), number of apothecia (NA), and number of stipes (NS). For the evaluation of ISG, a sclerotium was considered to be germinated if stipes were present, and/or apothecia had germinated, regardless of whether one or more stipes or one or more apothecia were present.

In this same experiment, it was also evaluated the ascospores germination. Five apothecia were removed from each replication and placed in microtubes containing 1.0mL of sterilized distilled water. These apothecia were then crushed with tweezers to obtain a suspension of ascospores to determine the number of ascospores/apothecium using a hemacytometer. It was also placed 0.1mL of the suspension in Petri dishes containing water - agar (1L - 0.02kg) and incubated them in a BOD chamber at $20 \pm 2^\circ\text{C}$ in darkness for 6h. After incubation, it was evaluated the number of germinated ascospores under a stereoscopic microscope and calculated the inhibition percentage in comparison with the control.

Fungicide inhibition of ascospore germination of *S. Sclerotiorum*

It was evaluated the efficacy of the fungicides in direct contact with ascospores in Petri dishes containing water agar. The experimental design was fully randomized with eight treatments (see previous test) and five replications. Apothecia were obtained using the method described for the previous test, with the modification of using only five gerboxes. After day 45, the boxes were opened to identify mature apothecia based on the fact that ascospores were released from these apothecia. It was selected 10 mature apothecia from the five boxes and the stipes were sliced and crushed and, placed in 10-mL crucibles with sterilized distilled water. Next, 0.1mL of the ascospore suspension were placed in Petri dishes containing a water-agar culture medium

and sprayed with approximately 160 μ L (200Lha⁻¹) of fungicide solutions obtained as described for the previous test. These Petri dishes were incubated in a BOD chamber at 20 \pm 2°C in darkness for 6h to determine ascospore germination using a stereoscopic microscope, and the inhibition percentage inhibition was calculated in comparison with the control.

Chemical control of *Sclerotinia* stem rot in a soybean crop

Trials were conducted under field conditions in the municipality of Mauá da Serra, Paraná, Brazil, latitude 23°51'06", longitude 51°10'40", elevation 970 m. BRS283 soybean was sown under no tillage following oats in the 2009-10 harvest and wheat in 2010-11. The experimental area was subdivided into 32 plots of 4.0 \times 6.0m (24m²); thus, the total area was of 768m². The experimental design was of randomized blocks with eight treatments (see previous test) and four replications.

Treatments were applied with a backpack sprayer pressurized with CO₂ with four flat-fan nozzles of type AXI 110-02. Spray pressure was 206.8kPa, and spray volume 200L of water/ha. *Trichoderma* treatment was applied at stages V3 and R1 and the fungicides at stage R1 and 15 days later (R3). Temperature during these treatments was approximately 20-25°C and relative humidity 90-99%.

At 95 DAS (days after sowing), it was evaluated the incidence (percentage of plants with symptoms) and severity of *Sclerotinia* stem rot using the scale of GRAU et al. (1982): 0, no symptoms; 1, lesion on lateral branches; 2, lesion on the main stem; 3, lesions on the main stem resulting in plant death. Scores were

converted into angular values ω and then into infection intensity indices $I' = \sin^2 \omega$ as proposed by AMARAL (1969). Four points/plot and 40 plants/point were evaluated. At approximately 120 DAS, were harvested by hand 4.0m of the two central rows, totaling 8.0 linear meters of each plot. Then, it was evaluated the weight of grain/plot and moisture content. The weight of grain/plot was transformed to kilograms/hectare (kg ha⁻¹).

Statistical analysis

Data obtained were subjected to the Shapiro-Wilk normality test. Homogeneity of variances was tested using Hartley's Fmax test. Analysis of variance was performed using the Scott-Knott test at 5% error probability. The Pearson correlation coefficient (r) was calculated to verify data correlation.

RESULTS

Fungicide inhibition of carpogenic germination of *S. Sclerotiorum*

ISG was evaluated in terms of the percentage of sclerotia that produced apothecia or stipes, irrespective of the numbers of apothecia or stipes. Fluazinam, benzalkonium chloride + fluazinam, iprodione, and carbendazim exhibited the highest percentages of sclerotia inhibition at 63.9, 61.8, 46.5, and 44.4%, respectively (Table 1).

ICG was evaluated by counting the total number of apothecia. The best results were obtained for fluazinam (100% inhibition), benzalkonium chloride + fluazinam (82.8%), and iprodione (63.1%). These results differed statistically (P<0.05) from the

Table 1 - Percentage inhibition of sclerotia germination (ISG), inhibition of carpogenic germination (ICG), inhibition of ascospore germination (IAG), and number of ascospores produced/apothecium (ASC/APO \times 1000) after fungicide application to the soil in gerboxes (Londrina-PR, 2011).

Treatment	----- ISG -----		----- ICG -----		----- IAG -----		----- ASC/APO ^c -----	
Control	-	-	-	-	-	-	98.0 ^d	a
Fluazinam	63.9 ^d	a	100.0 ^d	a	-	-	0.0	-
Procymidone	22.2	b	34.0	b	13.5 ^d	a	33.6	b
Iprodione	46.5	a	63.1	a	1.1	c	23.4	b
Thiophanate-methyl	0.7	c	18.5	b	0.4	c	32.8	b
Carbendazim	44.4	a	39.4	b	9.8	b	28.2	b
BC ^a + Fluazinam	61.8	a	82.8	a	13.9	a	17.1	b
<i>T. harzianum</i> ^b	0.0	-	6.9	b	0.6	c	58.0	a
CV (%)	31.8		38.9		5.6		5.8	

^aBenzalkonium chloride.

^bCommercial product based on *T. harzianum*.

^cData transformed into log x.

^dMeans followed by the same letter in the column did not differ significantly in the Scott-Knott test at 5% error probability.

results for carbendazim, procymidone, thiophanate-methyl, and *T. harzianum* (Table 1).

Applying the fungicides did not influence the development of NS, which ranged from 3.3 to 16.5%, and there was no significant difference ($P < 0.05$) between treatments (data not shown).

In this experiment, apothecia grown with the application of each treatment were crushed, and the number of germinated ascospores was evaluated. The percentage inhibition of germination was calculated in comparison with the control. Benzalkonium chloride + fluazinam and procymidone were the most effective treatments. They inhibited 13.9 and 13.5% of germination; this finding differed statistically ($P < 0.05$) from that for other treatments, which produced a low level of inhibition that ranged from 9.8% (carbendazim) to 0.6% (*T. harzianum*). No ascospores were produced after treatment with fluazinam (because no apothecia developed in the gerbox, there were no ascospores to evaluate).

With regard to numbers of ascospores/apothecium, benzalkonium chloride + fluazinam and procymidone produced 17,100 and 33,600 ascospores.apothecium⁻¹, respectively, whereas the control produced 98,000 ascospores.apothecium⁻¹. These treatments resulted in reductions of 82.4% and 65.6%. However, none of the fungicides produced statistically different results ($P < 0.05$), but all differed from results for the control (98,000 ascospores.apothecium⁻¹) and treatment with *T. harzianum* (58,000 ascospores.apothecium⁻¹).

Fungicide inhibition of ascospore germination of *S. Sclerotiorum* in Petri dishes

Treatments with fluazinam and benzalkonium chloride + fluazinam completely inhibited ascospore germination and were statistically identical ($P < 0.05$) to treatment with carbendazim (95.9% inhibition). The results obtained for thiophanate-methyl (70.1%), iprodione (35.7%), procymidone (7.5%), and *T. harzianum* (0.2%) differed statistically ($P < 0.05$). Treatment with the commercial product based on *T. harzianum* was least successful in inhibiting ascospore germination and did not differ from the control (data not shown).

Chemical control of Sclerotinia stem rot in the soybean crop

In the 2009-10 harvest, all treatments were effective in controlling Sclerotinia stem rot (Table 2) in comparison ($P < 0.05$) with the control (incidence of 79.4%). The most effective treatments were procymidone (incidence of 21.3%), benzalkonium

chloride + fluazinam (21.3%), and fluazinam (22.5%), which achieved reductions in the incidence of Sclerotinia stem rot of 73.1%, 73.1%, and 71.6%, respectively, in comparison with the control. With regard to severity of Sclerotinia stem rot, the control exhibited 73.9% severity; this differed statistically ($P < 0.05$) from treatments with procymidone (7.8% severity), benzalkonium chloride + fluazinam (11.5%), and fluazinam (13.6%). It also exhibited cutting severity by 89.4%, 84.4%, and 81.5%, respectively. The best yields were obtained using benzalkonium chloride + fluazinam, fluazinam, procymidone and carbendazim, with higher yields ($P < 0.05$) than those of other treatments. There was a high negative correlation between the incidence and the yield and between severity and the yield, with correlation coefficients of -0.60 and -0.76, respectively. This finding indicated that the yield increased as treatments reduced the incidence and severity of the disease.

In the 2010-11 harvest, the initial inoculum concentration observed was 11 sclerotia.m⁻² with 83% viability, and the results were similar for the 2009-10 harvest with regard to the incidence and severity, in which treatment with fluazinam, procymidone, and benzalkonium chloride + fluazinam surpassed other treatments, with respective figures of 77.6, 75.7, and 77.6% control. However, there were no significant differences in the yield between treatments, probably because the incidence and severity of the disease had not reached levels that could cause losses and influence yield. As in experiment 1, a strong negative correlation was observed between the incidence and the yield, and between the severity and the yield, with correlation coefficients of -0.69 and -0.69, respectively.

The commercial product based on *T. harzianum* was not effective in reducing the damage caused by Sclerotinia stem rot in the soybean crop under field conditions, and the results did not differ statistically ($P < 0.05$) from those for the control for the majority of parameters evaluated, with the exception of the incidence in the 2009-10 harvest.

DISCUSSION

The fungicides inhibited *S. sclerotiorum* resistance structures, fruiting body, and spores and pathogen development under field conditions. The biological product based on *Trichoderma* also showed efficient inhibition of carpogenic and ascospore germination. However, although there are many reports of the success of biological control against Sclerotinia stem rot, the biological control

Table 2 - Incidence and severity of *Sclerotinia stem* rot treated with fungicides, soybean yields for the 2009-2010 and 2010-2011 harvests (Mauá da Serra, Paraná, Brazil).

Treatment	Incidence %	Severity I ^c	Yield kg ha ⁻¹
Control	79.4 ^d a	73.9 ^d a	2594.0 ^d b
Fluazinam	22.5 d	13.6 c	3131.0 a
Procymidone	21.3 d	7.8 c	3046.0 a
Iprodione	40.0 c	43.5 b	2346.0 b
Thiophanate-methyl	60.0 b	63.0 a	2751.0 b
Carbendazim	55.0 b	34.0 b	3087.0 a
BC ^a + Fluazinam	21.3 d	11.5 c	3250.0 a
<i>T. harzianum</i>^b	61.9 b	61.3 a	2548.0 b
CV (%)	24.0	29.2	15.4
Treatment	Incidence %	Severity I ^c	Yield kg ha ⁻¹
Control	36.3 ^d a	57.5 ^d a	3042.8 ^d a
Fluazinam	8.1 c	3.1 d	3860.8 a
Procymidone	8.8 c	1.5 d	4132.8 a
Iprodione	18.8 b	8.0 c	3783.0 a
Thiophanate-methyl	29.4 a	36.3 b	3517.6 a
Carbendazim	25.0 b	9.7 c	3677.1 a
BC ^a + Fluazinam	8.1 c	1.2 d	3562.7 a
<i>T. harzianum</i>^b	33.8 a	56.4 a	3650.6 a
CV (%)	13.7	26.7	16.4

^aBenzalkonium chloride.^bCommercial product based on *T. harzianum*.^cInfection intensity index as a percentage.^dMeans followed by the same letter in the column did not differ significantly in the ScottKnott test (P<0.05).

of *S. sclerotiorum* under field conditions was not effective. The use of biological control under field conditions is highly sensitive to the success of the establishment of biocontrol agents.

Similar reports of the chemical fungicide control of *S. sclerotiorum* are available COSTA & COSTA (2004) tested the efficacy of several fungicides applied to the soil to inhibit myceliogenic and carpogenic germination of sclerotia. Of the fungicides evaluated, fluazinam resulted in 100% inhibition of apothecia, procymidone in 95% inhibition, iprodione 85% inhibition, and thiophanate-methylin less than 85% inhibition. The authors reported that fluazinam was completely effective in inhibiting the formation of apothecia but not the formation of stipes. This is in agreement with the results obtained in our study, in which carpogenic germination was completely inhibited but stipe production remained unaffected. However, efficacy in controlling the disease is based on the inhibition of the apothecium, as even if stipes are formed, if apothecia do not develop and release ascospores, the disease is controlled.

Studies of the action of fungicides on the ascospores germination of *S. sclerotiorum* are few. In this study, in addition to observing the positive effect of chemical treatments on inhibiting apothecia production by sclerotia, it was observed the effect on the number of germinated or viable ascospores produced. The reduction observed in the number of ascospores and the decrease in ascospore germination can cause a decrease in the number of plants infected under field conditions due to the lower quantity of inoculum released, in addition to the possibility of nonviable ascospore production.

Many in vivo studies have been conducted to evaluate the efficacy of fluazinam and procymidone in the control of diseases caused by *S. sclerotiorum* in common bean crops (PAULA JÚNIOR et al., 2009).

Fluazinam affects infection processes such as spore germination, formation of appressoria, hyphae penetration and growth, and pathogen sporulation in plants by uncoupling mitochondrial oxidative phosphorylation. Procymidone paralyzes the growth of hyphae, causing swell, bursting the cell

wall, ejecting the cytoplasmic contents, and inhibiting protein, nucleic acid, and lipid synthesis and spore germination (PICININI & GOULART, 2002). According to REIS et al. (2010), dicarboximide fungicides (procymidone) probably also use another mechanism involving peroxidized lipids interacting with cytochrome-c flavin reductase. The transport of NADPH electrons to cytochrome-c is blocked by the fungicide's action, and as a result, NADPH and essential phospholipids that surround the center of the flavin enzyme are oxidized, probably by a flavin enzyme intermediate or free radicals. In this study, it was observed significant differences in the effectiveness of fluazinam and procymidone in the in vitro tests of the inhibition of mycelial growth, carpogenic germination, and ascospore germination (by direct application of the fungicide to ascospores). The control achieved by these fungicides was statistically the same ($P < 0.05$) only in the field trials and both cropping years. PICININI & GOULART (2002) reported on the action of both fungicides in inhibiting spore germination but obtained better results using fluazinam. The difference observed in the in vitro and field tests is probably due to the mode of action of the fungicides. Fluazinam is a non penetrating (contact) residual protectant fungicide that acts on the pathogen on the plant's surface, inhibiting structural development and preventing penetration. Procymidone is a systemic fungicide that acts inside the plant and takes effect after the fungus has penetrated, when symptoms of the disease are already present.

In summary, fluazinam showed the greatest efficacy in inhibiting mycelial growth, carpogenic germination, and ascospore germination in vitro and in controlling *Sclerotinia* stem rot in commercial soybean crops. Procymidone showed the same potential as fluazinam in both agricultural years in controlling *Sclerotinia* stem rot under field conditions.

Regarding conventional recommendations for the use of fungicides to control *Sclerotinia* stem rot, MUELLER et al. (2004) commented that application should be at the onset of flowering and possible reapplication after blooming, reducing the incidence of the disease and boosting yields. However, it is very difficult to control diseases caused by sclerotia-forming fungi by applying specific fungicides to inhibit sclerotia in the field (COSTA & COSTA, 2004). Inconsistent results regarding the control of *Sclerotinia* stem rot using fungicides are probably due to inadequate coverage on the plant canopy, flowers, stems, and leaves where ascospores are usually deposited. According to TU (1989), the fungicide must be applied in sufficient volume to

cover flowers, stems, and leaves completely and particularly the structures closest to the soil surface.

CONCLUSION

Fluazinam and procymidone showed the highest efficiency in controlling *Sclerotinia* stem rot in a soybean crop under field conditions. Carpogenic and ascospore germination in vitro was efficiently inhibited with fluazinam alone.

ACKNOWLEDGEMENTS

The financial support of the Coordination for the Improvement of Higher Education Personnel.

REFERENCES

- ABDULLAH, M.T. et al. Biological control of *Sclerotinia sclerotiorum* (Lib.) de Bary with *Trichoderma harzianum* and *Bacillus amyloliquefaciens*. **Crop Protection**, v.27, p.1354-1359, 2008. Available from: <<http://www.sciencedirect.com/science/article/pii/S0261219408000926>>. Accessed: 18 jan. 2013. doi: 10.1016/j.cropro.2008.05.007.
- AMARAL, E. Novo índice de intensidade de infecção. **Pesquisa Agropecuária Brasileira**, v.4, p.1-2, 1969.
- BARDIN, S.D.; HUANG, H.C. Research on biology and control of *Sclerotinia* diseases in Canada. **Canadian Journal of Plant Pathology**, v.23, p.88-98, 2001.
- COSTA, J.R.; COSTA, J.L. da S. Efeito da aplicação de fungicidas no solo sobre a germinação carpogênica e miceliogênica de escleródios de *Sclerotinia sclerotiorum*. **Pesquisa Agropecuária Tropical**, v.34, p.133-138, 2004.
- DHINGRA, O.D. et al. Doenças e seu controle. In: SEDIYAMA, T. **Tecnologias de produção e usos da soja**. Londrina: Mecnas, 2009. p.133-156.
- FERRAZ, L.C.L. et al. Viabilidade de *Sclerotinia sclerotiorum* após a solarização do solo na presença de cobertura morta. **Fitopatologia Brasileira**, v.28, p.17-26, 2003. Available from: <http://www.scielo.br/scielo.php?script=sci_arttext&pid=S0100-41582003000100003>. Accessed: 01 out. 2014. doi: 10.1590/S0100-41582003000100003.
- GRAU, C.R. et al. Resistance of soybean cultivars to *Sclerotinia sclerotiorum*. **Plant Disease**, v.66, p.506-508, 1982. Available from: <http://www.apsnet.org/publications/PlantDisease/BackIssues/Documents/1982Articles/PlantDisease66n06_506.PDF>. Accessed: 01 out. 2014. doi: 10.1094/PD-66-506.
- HJELJORD, L.; TRONSMO, A. *Trichoderma* and *Gliocladium* in biological control: an overview. In: HARMAN, G.E.; KUBICEK, C.P. *Trichoderma and Gliocladium: enzymes, biological control and commercial applications*. Londres: Taylor & Francis, 2005. p.115-133.
- HUANG, H.; ERICKSON, R.S. Factors affecting biological control of *Sclerotinia sclerotiorum* by fungal antagonists. **Journal of Phytopathology**, v.156, p.628-634, 2008. Available from: <<http://eds.b.ebscohost.com/eds/pdfviewer/pdfviewer?sid=eccc9188->

- a316-4038-9c8c-c8f7edf55a5d%40sessionmgr198&vid=1&hid=117>. Accessed: 10 out. 2014. doi: 10.1111/j.1439-0434.2008.01423.x.
- LOBO JUNIOR, M.; ABREU M.S. de. Inibição do crescimento micelial de *Sclerotinia sclerotiorum* por metabólitos voláteis produzidos por alguns antagonistas em diferentes temperaturas e pH's. *Ciência e Agrotecnologia*, v.24, p.521-526, 2000.
- MUELLER, D.S. et al. Efficacy of fungicides on *Sclerotinia sclerotiorum* and their potential for control of Sclerotinia stem rot on soybean. *Plant Disease*, v.86, p.26-31, 2002. Available from: <<http://apsjournals.apsnet.org/doi/pdf/10.1094/PDIS.2002.86.1.26>>. Accessed: 20 out. 2014. doi: 10.1094/PDIS.2002.86.1.26.
- MUELLER, D.S. et al. Application of thiophanate-methyl at different host growth stages for management of Sclerotinia stem rot in soybean. *Crop Protection*, v.23, p.983-988, 2004. Available from: <http://www.ars.usda.gov/SP2UserFiles/ad_hoc/54000000WhiteMoldResearch/PublicationPDFs/Applicationofthiophanate.pdf>. Accessed: 20 out. 2014. doi: 10.1016/j.cropro.2004.02.013.
- PAULA JÚNIOR, T.J. et al. White mold intensity on common bean in response to plant density, irrigation frequency, grass mulching, *Trichoderma* spp., and fungicide. *Summa Phytopathologica*, v.35, p.44-48, 2009. Available from: <<http://www.scielo.br/pdf/sp/v35n1/v35n1a07.pdf>>. Accessed: 20 out. 2014. doi: 10.1590/S0100-54052009000100007.
- PICININI, E.C.; GOULART, A.C.P. Novos fungicidas para tratamento de sementes. *Revisão Anual de Patologia de Plantas*, v.10, p.33-66, 2002.
- REIS, E.M. et al. *Manual de fungicidas*: guia para o controle químico de doenças de plantas. 6.ed. Passo Fundo: Universidade de Passo Fundo, 2010.
- SARMA, B.K. et al. Use of non-conventional chemicals as an alternative approach to protect chickpea from Sclerotinia stem rot. *Crop Protection*, v.26, p.1042-1048, 2007.
- TU, J.C. Management of white mold of white bean in Ontario. *Plant Disease*, v.73, p.281-285, 1989.
- WRATHER, J.A.; KOENNING, S.R. *Effects of diseases on soybean yields in the United States 1996 to 2007*. Available from: <<http://www.plantmanagementnetwork.org/pub/php/Research/2009/yields/>>. Accessed: 20 out. 2014. doi: 10.1094/PHP-2009-0401-01-RS.