Leobet Lunkes, Vinicius; Tonin, Alexandre Alberto; Machado, Gustavo; Corbellini, Luis Gustavo; Nogueira Diehl, Gustavo; Carboneiro dos Santos, Lucila; de Sousa Bezerra, Camila; Santos de Azevedo, Sérgio; Fernandes Pequeno, Nebson; Moraes da Silva, Adriana; Weiblen, Rudi; Furtado Flores, Eduardo

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Universidade Federal de Santa Maria
Santa Maria, Brasil

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Antibodies against vesicular stomatitis virus in horses from southern, midwestern and northeastern Brazilian States

Vinícius Leobet Lunkes1 Alexandre Alberto Tonin1 Gustavo Machado1 Luis Gustavo Corbellini1
Gustavo Nogueira Diehl3IV Lucila Carboneiro dos Santos3IV Camila de Sousa Bezerra4
Sérgio Santos de Azevedo1 Vinícius Leobet Lunkes1 Alexandre Alberto Tonin1 Gustavo Machado1 Luis Gustavo Corbellini1
Rudi Weiblen4 Eduardo Furtado Flores5

ABSTRACT

Vesicular stomatitis virus (VSV) is the agent of a vesicular disease that affects many animal species and may be clinically confounded with foot-and-mouth disease in ruminant and swine. Horses are especially susceptible to VSV and may serve as sentinels for virus circulation. The present study investigated the presence of neutralizing antibodies against VSV Indiana III (VSIV-3) in serum samples of 3,626 horses from six states in three Brazilian regions: Southern (RS, n = 1,011), Midwest (GO/DF, n = 1,767) and Northeast (PB, PE, RN and CE, n = 848) collected between 2013 and 2014. Neutralizing antibodies against VSIV-3 (titers ≥40) were detected in 641 samples (positivity of 17.7%; CI95%: 16.5-19.0%), being 317 samples from CE (87.3%; CI95%: 83.4-90.5%), 109 from RN (65.7%; CI95%: 57.8-72.7%); 124 from PB (45.4%; CI95%: 39.4-51.5%); 78 from GO/DF (4.4%; CI95%: 3.3-5.5%) and nine samples of RS (0.9%; CI95%: 0.4-1.7%). Several samples from the Northeast and Midwest harbored high neutralizing titers, indicating a recent exposure to the virus. In contrast, samples from RS had low titers, possibly due to a past remote exposure. Several positive samples presented neutralizing activity against other VSV serotypes (Indiana I and New Jersey), yet in lower titers, indicating the specificity of the response to VSIV-3. These results demonstrated a relatively recent circulation of VSIV-3 in northeastern Brazilian State, confirming clinical findings and demonstrating the sanitary importance of this infection.

Key words: serology, differential diagnosis, foot-and-mouth disease, zoonosis, vesicular disease.

INTRODUCTION

Vesicular stomatitis virus (VSV) is a viral agent belonging to the order **Mononegavirales**, and is the agent of a disease that affects many animal species and may be clinically confounded with foot-and-mouth disease in ruminant and swine. Horses are especially susceptible to VSV and may serve as sentinels for virus circulation. The present study investigated the presence of neutralizing antibodies against VSV Indiana III (VSIV-3) in serum samples of 3,626 horses from six states in three Brazilian regions: Southern (RS, n = 1,011), Midwest (GO/DF, n = 1,767) and Northeast (PB, PE, RN and CE, n = 848) collected between 2013 and 2014. Neutralizing antibodies against VSIV-3 (titers ≥40) were detected in 641 samples (positivity of 17.7%; CI95%: 16.5-19.0%), being 317 samples from CE (87.3%; CI95%: 83.4-90.5%), 109 from RN (65.7%; CI95%: 57.8-72.7%); 124 from PB (45.4%; CI95%: 39.4-51.5%); 78 from GO/DF (4.4%; CI95%: 3.3-5.5%) and nine samples of RS (0.9%; CI95%: 0.4-1.7%). Several samples from the Northeast and Midwest harbored high neutralizing titers, indicating a recent exposure to the virus. In contrast, samples from RS had low titers, possibly due to a past remote exposure. Several positive samples presented neutralizing activity against other VSV serotypes (Indiana I and New Jersey), yet in lower titers, indicating the specificity of the response to VSIV-3. These results demonstrated a relatively recent circulation of VSIV-3 in northeastern Brazilian State, confirming clinical findings and demonstrating the sanitary importance of this infection.

Palavras-chave: sorologia, diagnóstico diferencial, febre aftosa, zoonose, doença vesicular.
family *Rhabdoviridae*, genus *Vesiculovirus*. Besides its economical importance for the livestock, VSV represents a sanitary threat since it causes a disease that is clinically similar to foot and mouth disease in cattle and pigs (RIET-CORREA et al., 1996). VSV naturally infects a variety of mammals including horses, cattle, swine, wild mammals and man. VSV infection is endemic in the Americas and seems to be restricted to western hemisphere (LETCHWORTH et al., 1999). The disease is characterized by the development of vesicular lesions in mouth, tongue, teats and coronary bands of cattle, horses and pigs (RIET-CORREA et al., 1996). In most cases, the disease is self-limiting and the clinical course lasts approximately two to three weeks (REIS JR et al., 2009).

VSV isolates belong to two antigenically distinct serogroups: New Jersey (VSNJV) and Indiana (VSIV). Viruses of the VSNJV serogroup are disseminated in the southern-central United States. Serogroup VSIV contains three subtypes: Indiana I (VSIV-1- classical strains), Indiana II (VSIV-2 Cocal and Argentina) and Indiana III (VSIV-3 - Alagoas). Subtypes VSIV-2 and VSIV-3 have been occasionally isolated in Brazil, being the serotype VSIV-3 more restricted to the northeastern States (CARGNELUTTI et al., 2014). According to the ICTV (2014), there are the least 20 additional serotypes to be characterized.

Vesicular stomatitis (VS) usually presents a seasonal pattern, whose incidence is usually higher in summer or in rainy seasons (MASON et al., 1978; OIE, 2010). This behavior has led to the hypothesis of dissemination by winds, birds and insect vectors (TESH et al., 1970). In this sense, the virus has been isolated from mosquitoes *Phlebotomus* and *Aedes*, indicating their possible role in virus transmission (HAYEK et al., 1998). According to OIE (2010), horses are particularly susceptible to VSV infection compared to cattle and pigs. Animal gathering in fairs, races, artificial insemination centers and other events seem to facilitate virus spread among susceptible animals (OKUDA et al., 2003).

VS in clinically undistinguishable from foot-and-mouth disease (FMD), making critical its prompt differential diagnosis. VS diagnosis may be performed by virus isolation or serological tests as virus-neutralization (VN), immunoenzimatic assays and complement-fixation (FERRIS & DONALDSON, 1988; ALONSO FERNANDEZ et al., 1991).

In Brazil, positive serology to VSV has been detected in several states and different animal species. By comparing two serological tests, ALLENDE & GERMANO (1993) analyzed 305 cattle, horse and pig sera, detecting 300/305 (98.40%) samples containing VN antibodies to VSIV-3. CUNHA et al. (2009) found that 21% of samples were positive to VSIV-2 (Cocal) and 5% to VSIV-3 (Alagoas) in the São Paulo State.

VSV was fist isolated in Brazil in 1964, in Alagoas State, from sick horses and named VSIV-3 Alagoas, due to differences in serogroups VSIV-1 and VSIV-2 (ANDRADE et al., 1980). In Minas Gerais, ARAÚJO et al. (1977) reported the isolation of serotype VSIV-3 from cattle. In Sergipe (1984), ALONSO FERNANDES & SÖNDAHL (1985) isolated a serotype VSIV-3 from horses.

In the last years, several outbreaks of VS have been reported in midwestern and northeastern (NE) Brazilian States (ROSENDO et al., 2013; CARGNELUTTI et al., 2014). Reports from the PANAFTOSA (2015) have also indicated viral activity in NE states in the last years. In 2013, outbreaks of VS were reported in horses and cattle in states of Paraiba and Rio Grande do Norte, with the identification of VSIV-3 (CARGNELUTTI et al., 2014).

Considering the sanitary and economical importance of the disease, this study was designed to investigate circulation of VS-3, through serology, in horses from three Brazilian regions: Southern, Midwestern and Northeast.

**MATERIALS AND METHODS**

The present study used 3,626 serum samples from horses of three Brazilian regions: southern (Rio Grande do Sul [RS]), midwest (Goiás [GO] and Federal District [DF]) and northeast (Pernambuco [PE], Paraiba [PB], Rio Grande do Norte [RN] and Ceará [CE]). Samples from northeast (n=848) and midwest (n=1,767) were from the official diagnosis of equine infectious anemia virus (EIAV) collected between 2013 and 2014. RS samples (n=1,011) were collected in 2013, as a part of an official serological survey of EIA.

Serum samples were submitted to VN test for detection of antibodies to VSV, according to the OIE (2010) protocol, using the isolate VSIV-3 2013 SaoBento/ParaibaE (CARGNELUTTI et al., 2014). After complement inactivation, serum samples were diluted 1:40 and incubated with 400-500 TCID₅₀ of the isolate VSIV-3 2013 SaoBento/ParaibaE for 1h at 37°C, followed by addition of a suspension of Vero cells and incubation at 37°C with 5% CO₂. The cultures were monitored for citopathic effect (cpe) for 72h. Samples not presenting cpe were considered positive for VSV antibodies at the
used dilution. Then, positive samples were submitted to a quantitative VN test, in which a fixed dose of virus (400-500 TCID_{50}) was incubated with serial 2-fold dilutions of sera, starting at 1:40. In this test, each sample was tested against three VSV strains/isolates: isolate VSIV-3 2013SaoBento/Paraiba E, strain Indiana (VSIV-1) and VSNJV. After 72h, the cultures were monitored for cpe and the VN titers were considered as the reciprocal of the highest serum dilution capable to prevent cpe. Virus titers were transformed in GMT log_2 (THRUSFIELD, 1986), considering 1:40 as the lowest dilution, and graded as low GMT (log_2 ≤3.0), moderate (3.0 < GMT log_2 ≤ 4.0) and high (GMT log_2 > 4.0).

RESULTS AND DISCUSSION

Results of VN assays revealed the presence of neutralizing antibodies reacting with isolate VSIV-3 2013 SaoBento/Paraiba E, in varied frequency and titers, in all three studied regions. The overall rate of seropositivity was 17.7% (CI_{95%}: 16.5-19.0%), with the highest values observed in NE states (Table 1). High percentages of positive samples were observed in CE (87.3%: CI_{95%}: 83.4-90.5%) and RN (65.7%: CI_{95%}: 57.8-72.7%); moderate rates in PB (45.4%: CI_{95%}: 39.4-51.5%); low levels in PE (8.7%), GO/DF (4.4%: CI_{95%}: 3.5-5.5%) and very low prevalence rates in RS (0.9%; CI_{95%}: 0.4-1.7%). Since no commercial VSV vaccines are available in Brazil (SINDAN, 2015), positive serological response is obviously due to a previous exposure to the virus, reflecting different levels of virus circulation in the three regions. These results indicated that CE, RN and PB States presented a broader viral activity, in contrast with the viral circulation in the Midwest and Southern regions, whose immunological reaction probably reflects a low frequency and remote viral activity.

The high prevalence of VSV-3 antibodies in most NE states is compatible with the study by ALLENDE & GERMANO (2003), who detected 91.5% (300/328) of seropositivity. Based upon these results, however, is difficult to estimate the real prevalence of VSV antibodies in the region since the Centro Panamericano de Febre Aftosa (PANAFTOSA) reports only cases notified from 2013 to the present. Nevertheless, several outbreaks/cases of vesicular disease have been notified in the NE States in the last 10 years (OIE, 2010; CARGNELUTTI et al., 2014), from which several were confirmed as VS. Information by PANAFTOSA (2015) also indicated recent virus circulation in NE in the last years.

Moderate to high antibody prevalence in horses from NE states likely reflects the environmental conditions, especially the climate, which favors the maintenance of large populations of insects (especially mosquitoes), probable VSV vectors (BENNITT et al., 2008). Likewise, low frequency of VSV antibodies in RS (0.9%) may be attributed, in part, to the climatic conditions that disfavor the maintenance of abundant insect populations. Low levels of antibodies in horses from RS likely reflect a timely remote exposure to VSIV-3 or related viruses. Similar findings have been reported for other arboviruses, such as Dengue, Bluetongue and EIAV, whose frequency in RS is markedly lower than that observed in central and northern States (MELLOR & LEAKE, 2000). Regardless, VS cases have been reported in Santa Catarina, a southern state (LOPES et al., 1999).

After the initial screening with isolate VSIV-3 2013SaoBento/ParaibaE, positive samples were tested against VSIV-1 (Indiana) and VSNJV, trying to confirm the viral serotype involved. In RS,

<table>
<thead>
<tr>
<th>State</th>
<th>Number</th>
<th>Positive (%)*</th>
<th>Neutralizing titers**</th>
<th>log_2</th>
</tr>
</thead>
<tbody>
<tr>
<td>Rio Grande do Sul</td>
<td>1,011</td>
<td>9 (0.9)</td>
<td>40-160</td>
<td>2.1</td>
</tr>
<tr>
<td>Goiás/Distrito Federal</td>
<td>1,767</td>
<td>78 (4.4)</td>
<td>40-10240</td>
<td>3.0</td>
</tr>
<tr>
<td>Pernambuco</td>
<td>46</td>
<td>4 (8.7)</td>
<td>40-1280</td>
<td>3.0</td>
</tr>
<tr>
<td>Paraíba</td>
<td>273</td>
<td>124 (45.4)</td>
<td>40-10240</td>
<td>4.5</td>
</tr>
<tr>
<td>Rio Grande do Norte</td>
<td>166</td>
<td>109 (65.7)</td>
<td>40-10240</td>
<td>3.8</td>
</tr>
<tr>
<td>Ceará</td>
<td>363</td>
<td>317 (87.3)</td>
<td>40-10240</td>
<td>4.4</td>
</tr>
<tr>
<td>Total</td>
<td>3,626</td>
<td>641 (17.7)</td>
<td>40-10240</td>
<td>4.1</td>
</tr>
</tbody>
</table>

Neutralizing titers > 40.

*Reciprocal of the highest serum dilution capable to prevent the production of cytopathic effect.
Unfortunately, we could not test the samples against VSV Cocal because we could not obtain a virus of this serotype. Thus, it can not be discarded that part of the samples that reacted with VSIV-3 2013 SaoBento/ParaibaE could be result from exposure to VSIV-2 Cocal, since there are reports indicating the circulation of this serotype in Brazil (REIS JR et al., 2009). In this sense, circulation of VSIV-2 seems to be more abundant in the Southeastern and Midwestern regions (LÓPEZ et al., 1996-1997).

Figure 1 presents the frequency and distribution of neutralizing titers by state. In NE states - PE is an exception - moderate titers (160-1280) were more frequent, but high titers were also observed (2560, 5120, >10240). Moderate titers suggest a not recent exposure since high titers are usually observed within weeks or months after VSV infection (CARGNELUTTI et al., 2014). Moreover, high titers (2560 a ≥ 10240) indicate recent exposure. These findings agree with data by

9 samples were also positive for VSIV-1 (titers from 40 to 160) and none reacted with VSNJV. In GO/DF, 10 samples reacted with VSIV-1 (titers 40 to 80) and none with VSNJV. In NE states, 164 samples reacted with VSIV-1 (40 to 1280) and 7 reacted also with VSNJV (titer of 40). In spite of the antigenic differences between VSV serotypes, variable levels of cross-neutralization are observed among VSIV-1, 2 e 3 (PAUSZEK et al., 2011). In contrast, cross-neutralization between VSIV and VSNJV occurs at very low levels due to their antigenic differences (CARTWRIGHT & BROWN, 1972). Positivity to VSNJV is probably to this cross-reactivity because VSNJV is considered exotic in Brazil (OIE, 2010; PANAFTOSA, 2015). In summary, results of VN tests indicated that the neutralizing antibodies detected were probably produced in response to infection by viruses antigenically related to VSIV-3 (VSIV-3 2013SaoBento/ParaibaE), confirming the frequent circulation of this serotype in the region.
PANAFTOSA (2015), which reported confirmed cases/outbreaks of VS in the region in the last years, including those reported by CARGNELUTTI et al. (2014). A similar distribution of virus titers, with predominant moderate titers (160-1280) and some high titers (≥2560) were observed in GO/DF, also suggesting recent exposure.

VSV infection is considered endemic in Brazilian northeastern States, where is probably under notified. The present results validate this status because they indicated the circulation of VSV - likely serotype 3 - in horses from this region. In addition, our work indicates a viral activity of lower intensity also in GO/DF. Residual antibodies were detected, probably reflecting remote past exposure to the virus in RS.

Because cattle is the most important host species for VSV infection – which it can be confounded with FMD, leading to serious sanitary consequences -, it would be interesting to investigate the presence and distribution of VSV antibodies in cattle of the studied regions. In this sense, a report by CARGNELUTTI et al. (2014) described VS cases predominantly in horses but also in cattle in northeastern States. In agreement with the latter work, our data indicates that VSV is already circulating in these regions.

CONCLUSION

Sero logic data indicated the circulation of VSIV-3 (related to VSIV-3 2013saoBento/ ParaíbaE) in horses in NE States, presence in low levels in GO/DF and residual levels of antibodies in RS, probably reflecting a timely remote virus circulation. Our results reinforce the sanitary importance of this infection, contributing for its understanding, notification and control.

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