

Ciência Rural

ISSN: 0103-8478

cienciarural@mail.ufsm.br

Universidade Federal de Santa Maria Brasil

Cecagno, Diego; Anghinoni, Ibanor; Valadão Gigante de Andrade Costa, Sérgio Ely;
Martins Brambilla, Daniel; Posselt Martins, Amanda; Cavazini Magiero, Emanuelle;
Bagatini, Tatiane; Assmann, Joice Mari; Nabinger, Carlos
Long-term nitrogen fertilization in native pasture with Italian ryegrass introduction - Effects
on soil health attribute indicators
Ciência Rural, vol. 47, núm. 5, 2017, pp. 1-6
Universidade Federal de Santa Maria
Santa Maria, Brasil

Available in: http://www.redalyc.org/articulo.oa?id=33150130015



Complete issue

More information about this article

Mana information object this outin

relalyc.arg

Scientific Information System

Network of Scientific Journals from Latin America, the Caribbean, Spain and Portugal Non-profit academic project, developed under the open access initiative

Key words: carbon and nitrogen stocks, microbial biomass, natural pasture.



ISSNe 1678-4596 SOIL SCIENCE

Long-term nitrogen fertilization in native pasture with Italian ryegrass introduction - Effects on soil health attribute indicators

Diego Cecagno^{1*} Ibanor Anghinoni¹ Sérgio Ely Valadão Gigante de Andrade Costa¹ Daniel Martins Brambilla² Amanda Posselt Martins¹ Emanuelle Cavazini Magiero¹ Tatiane Bagatini¹ Joice Mari Assmann¹ Carlos Nabinger²

ABSTRACT: Native pastures are of great importance for cattle and sheep nutrition in the Pampa biome. However, due to its low productivity, the Italian ryegrass introduction and the nitrogen (N) fertilization are alternatives proposed to intensify livestock production in a sustainable manner. The objective of this study was to evaluate the effects of long-term N application on soil health indicators in a native pasture with Italian ryegrass introduction in southern Brazil. The experiment consists of a secondary native pasture under continuous grazing and constant herbage allowance. In 1996 experimental area was broadcast limed and the experiment was initiated, testing three N topdressing rates (0, 100 and 200kg N ha⁻¹ year⁻¹). In 2010 soil of experimental and reference area of non grazed native grassland was sampled in the soil layers of 0-20 and 20-40cm. Total, particulate and mineral associated carbon (C) and N stocks were evaluated. Soil microbiological attributes were evaluated in 0-5 and 5-10cm soil layers. The long-term N fertilization in soils with native pasture and Italian ryegrass introduction did not affect total C and N stocks. However, increases in N particulate fraction were seen with 100kg ha⁻¹ year⁻¹ of N rate of fertilization. Furthermore, the increase in N rates increased N microbial biomass and respiration.

Fertilização nitrogenada de longo prazo em campo nativo com introdução de azevém anual – Efeitos sobre atributos indicadores da saúde do solo

RESUMO: A pastagem nativa é uma importante fonte de forragem para produção de bovinos e ovinos no bioma Pampa. A introdução do azevém e a fertilização nitrogenada são alternativas que visam intensificar a exploração pecuária de forma mais sustentável, podendo causar alterações em vários atributos do solo. Este trabalho teve por objetivo avaliar os efeitos de longo prazo de fertilização nitrogenada sobre atributos indicadores da saúde do solo em pastagem nativa com introdução de azevém anual. O experimento consistiu de uma sucessão secundária da pastagem natural submetida ao pastejo contínuo por bovinos e ovinos, com oferta de forragem constante. Em 1996, anteriormente ao inicio do experimento, a área experimental foi calcareada para correção da acidez do solo. O experimento foi então iniciado, sendo constituído da aplicação de três doses de N em cobertura, correspondendo a 0, 100 e 200kg ha¹ ano¹ de N. Em 2010, amostras de solo foram coletadas nas camadas de 0-20 e 20-40cm para análise de C e N total, particulado e associado aos minerais, além de uma área de referência (campo nativo sem pastejo). Análises microbiológicas foram conduzidas nas camadas de 0-5 e 5-10cm. A adubação nitrogenada de longo prazo em solos de campo nativo com introdução de azevém não altera os estoques totais de C e N, mas aumenta a fração particulada de N na dose de 100kg ha¹ ano¹. Em relação à atividade microbiana, o aumento na dose do fertilizante aumenta a respiração dos microrganismos, assim como o N da sua biomassa. Palavras-chave: estoques de carbono e nitrogênio, biomassa microbiana, pastagem natural.

INTRODUCTION

Pampa's biome native pasture is the main nutritional source used to feed sheep and beef cattle in the Rio Grande do Sul State (SEBRAE/SENAR/FAR-SUL, 2005). However, incorrect anthropogenic actions have been degrading native pastoral ecosystems from this region, by decreasing the occurrence of desirable forage species and animal load bearing capacity of the native pasture. Excessive stocking rates and improper fertilization used in pastoral environments has caused reduction in plant biomass production and soil organic matter (SOM) content and lability as well as soil microbial biomass (SOUZA et al., 2010; CONTE et al, 2011).

The use of nitrogen (N) fertilizer is required for intensifying the meat production on pasture since N is an essential nutrient required in large amounts by grass species (SARMENTO et al., 2008). In addition, summer species predominate in the Pampas biome, becoming necessary the introduction during winter season of species with high yields such as Italian ryegrass (*Lolium multiflorum* Lam.) (CONTERATO et al., 2016). Both nitrogen fertilization and Italian ryegrass introduction in native grassland can be an alternative for sustainable Pampa biome exploration. However, these factors affect the carbon (C) dynamics in soil due to their effects on chemical, physical and biological properties (CARTER, 2002). Considering the existing coupling between C and

¹Departamento de Solos, Universidade Federal do Rio Grande do Sul (UFRGS), Av. Bento Gonçalves, 7712, 91540-000, Porto Alegre, RS, Brasil. E-mail: deceagno@hotmail.com. *Corresponding author.

²Departamento de Plantas Forrageiras e Agrometeorologia, Universidade Federal do Rio Grande do Sul (UFRGS), Porto Alegre, RS, Brasil.

Cecagno et al.

N in the soil system, monitoring C dynamics in these conditions is necessary to monitor the environmental impacts over time.

According to SOUSSANA & LEMAIRE (2014), plants coupling atmospheric C and mineral N in the photosynthesis process are an ongoing process. Thus, these compounds are added to the soil in a coupled form, accumulating both simultaneously. However, since plants normally have a higher C:N ratio compared to the soil, constant N inputs in the soil can increase C sequestration (FORNARA & TILMAN, 2012).

Both microbial biomass and soil organic fraction are indicators of soil quality and sustainability of production systems, being affected by soil management (NANNIPIERI et al., 2003; BODDEY et al., 2010). Thus, over time, N fertilization can increase crop production and soil C stocks (COSTA et al., 2008). However, OLSON et al. (2005) observed a decrease in C stocks with N fertilization, due to its potential to increase soil microbial activity. There may be both positive (MAJUMDER et al., 2007) and negative effects (YU et al., 2016) in microbial biomass by nitrogen fertilization.

Thus, looking for a strategy to manage native grasslands and avoid pasture and environmental degradation, this study aimed to evaluate the long-term N fertilization impacts in an Ultisol under native grassland with Italian ryegrass introduction, by measuring some soil attributes that are considered as soil health indicators.

MATERIALS AND METHODS

The experiment was carried out since 1996 at the Agronomic Experimental Station of the Federal University of Rio Grande do Sul, in Eldorado do Sul, Brazil, on a Rhodic Paleudult clay loam soil. In the 0-20 and 20-40cm soil layers clay contents were 130 and 200g kg¹, respectively. The climate is subtropical with warm summer weather (Cfa), according to the Köppen classification. The historical annual average from 1970 to 2009 for air temperature, rainfall and reference evapotranspiration (ETo) are 18.8°C, 1455 and 1161mm, respectively (BERGAMASCHI et al., 2013).

In 1996, the soil was fertilized with 500kg ha⁻¹ of 05-20-20 fertilizer and limed to reach pH 6.0 with 3.0Mg ha⁻¹ of lime broadcast applied. In this year, grazing began with beef cattle and sheep. The stocking rate was controlled to keep the forage supply at the level of 12% (12kg dry mass offered / 100kg live weight / day). Some years had beef cattle grazing, while others sheep. Treatments consisted of topdressed N annual rates of 0, 100 and 200kg ha⁻¹ yr⁻¹ (called in

the current study as "without N", "moderate N" and "high N", respectively) conducted since 1996. The N source was always urea and the rates were splitted in two applications (50% in May and 50% in July). The experiment is characterized by a secondary succession of native grassland spanning an area of 3.11 hectares, and was carried out in a randomized block design with two replicates. Experimental plots size ranged from 0.3961 to 0.6587ha. The Italian ryegrass introduction occurred in 2007, when soil was again limed and fertilized with 300kg ha⁻¹ of 12-52-00 fertilizer. During winter and summer, Italian ryegrass and Paspalum notatum and Cynodon dactylon were the predominant species. In 2010, N was applied July 30th and November 2nd, with rainfall around 15mm (nine and seven days after application). The average forage accumulation rate in 2010 was 7918, 9657 and 14144kg ha-1 of dry matter, and grazing pressures of 757, 895 and 1167kg ha-1 of live weight were recorded for N rates of 0, 100 and 200kg ha⁻¹, respectively. To estimate the dry matter accumulation rate (kg ha-1) three exclusion cages per plot using the double-pairing technique were used. The accumulation rate was obtained by the difference between the forage mass outside the cage at measurement i-1 and the forage mass from within the cage at measurement i, after approximately 28 days.

The average soil chemical attributes in the 0-20cm layer from shovel sampling in August 28th of 2010 were, for SOM, available phosphorus (P Mehlich 1) and potassium (K Mehlich 1), exchangeable calcium and magnesium (KCl 1mol L⁻¹): 18.2g kg⁻¹, 18.6mg dm⁻³, 0.31cmol_c kg⁻¹, 1.45cmol_c kg⁻¹ and 0.83cmol_c kg⁻¹, respectively.

Total (TOC, TN), particulate (POC, PN) and mineral-associated carbon and nitrogen stocks (MAC, MAN) were determined for combined soil samples (ten subsamples) of the 0-20 and 20-40cm soil layers sampled during August of 2010. A native grassland area without grazing located about 500m from the experiment was also sampled. This area was grazed with traditional cattle management until approximately 1980 when grazing has been suspended. SOM fractionation was performed according to CAMBARDELLA & ELLIOT (1992). Total organic carbon and POC were analyzed by dry combustion using a Shimadzu TOC-V CSH analyzer. TN and PN were determined by Kjeldahl method. MAC and MAN were calculated by the difference between total and particulate stocks. Carbon and N stocks were calculated using the soil equivalent mass according to ELLERT & BETTANY (1995). For stock calculations, soil bulk density was estimated using volumetric rings (270 cm³); soil density values were 1.50 and 1.56kg dm⁻³ for the 0-20 and 20-40cm layers, respectively.

Combined soil samples from five subsamples per plot sampled from the 0-5 and 5-10cm soil layers were used to determine soil microbial biomass. Soil microbial biomass carbon (SMBC), soil microbial biomass nitrogen (SMBN), microbial respiration and metabolic quotient were determined according to VANCE et al. (1987), BROOKES et al. (1985), ALEF & NANNIPIERI (1995) and ANDERSON & DOMSH (1993), respectively.

Results were submitted to analysis of variance (ANOVA) and, when significant (P<0.10), averages were compared by Tukey test (P<0.10) using the SISVAR software. The following statistical model was used for the ANOVA:

 $Y_{ijk} = \mu + B_i + N_j + \text{Errora}(ij) + L_k + \text{Errorb}(ik) + N_j L_k + \text{Errorc}(ijk)$ where $\mu = \text{the overall experiment average}; B = \text{the blocks}$ (i = 1, 2); N = the nitrogen rates (j = 1, 2, 3); L = the soil layer (k = 1, 2); and Error = the experimental error.

RESULTS AND DISCUSSION

C stocks and C fractions were not affected by topdressed nitrogen fertilization over time, being similar to those observed in the reference area (Table 1). TOC stocks in 0-20cm were around 25% lower than those reported by CONTE et al. (2011) in similar climate, soil and herbage conditions. Conversely, POC stocks

were similar, demonstrating the role of Italian ryegrass introduction to maintain soil quality.

Similar C stocks under different N rates can be explained by increased microbial respiration with increasing availability of this nutrient (Table 2). Even with an increase in dry matter production with increasing nitrogen fertilization (2.39 to 4.68Mg ha⁻¹ yr⁻¹), there was a higher CO₂ emission by microorganisms (Table 2) leading to organic compounds oxidation, as described by KHAN et al. (2007).

The climate is another factor that explains the similarity in C stocks, regardless of N fertilizer rate. The average temperatures observed on this trial (18.8°C) are moderates based on the growth curve of plants, which is not low or high enough to be a limiting factor for growth/development. This factor associated with high annual rainfall distributed throughout the year contributes to soil organic matter oxidation. Under such environmental conditions microbial activity is intense (Table 2), decomposing faster the soil organic residues. Despite the increase in dry matter production with increasing N rate, there is a balance between matter and energy outputs and inputs in the system, keeping the C stocks under treatments and the reference area at similar levels.

Table 1 - Carbon and nitrogen stocks and fractions in an Ultisol with continuous grazing in native pasture with Italian ryegrass introduction, under different nitrogen topdressing rates.

Stocks	Fractions	Soil layer	Reference area	Nitrogen rate – kg ha ⁻¹				
				0	100	200	Average	
		cm			Mg ha ⁻¹			
Carbon	Total*	0 - 20	25.77±2.28	25.88±3.06	30.95±2.55	29.42±1.24	28.01±1.72a	
		20 - 40	18.37±1.71	21.12±0.48	24.93±4.23	20.16±0.42	21.15±1.07b	
	Particulate**	0 - 20	6.72±0.52	5.08±0.30	5.67±1.17	4.42±0.10	5.47±0.40a	
		20 - 40	2.23±0.31	1.73±0.07	2.03±0.48	1.32±0.37	1.83±0.23b	
	Mineral- associated*	0 - 20	19.06±1.76	20.79±2.76	25.27±1.38	25.00±1.14	22.53±1.32a	
		20 - 40	16.14±1.65	19.40±0.41	22.90±3.74	19.33±0.78	19.44±0.84b	
Nitrogen	Total**	0 - 20	2.46±0.15	2.42±0.25	2.97±0.16	2.91±0.02	2.69±0.11a	
		20 - 40	1.65±0.22	1.79±0.24	1.79±0.36	1.65±0.05	1.72±0.17b	
	Particulate**	0 - 20	0.62±0.04 aAB	0.41±0.06 aC	0.64±0.03aA	0.55±0.03aB	0.56 ± 0.03	
		20 - 40	0.35±0.02 bA	0.22±0.00 bB	0.22±0.02bB	0.25±0.03bB	0.26 ± 0.01	
	Mineral- associated*	0 - 20	1.84±0.11	2.01±0.20	2.33±0.13	2.36±0.00	2.14±0.08a	
		20 - 40	1.30±0.20	1.57±0.24	1.57±0.35	1.40±0.02	1.46±0.15b	

Values are mean and standard error (n=2) and those followed by the same capital letter in the lines and lower case letter in the columns do not differ according to Tukey test (* = P<0.10; ** = P<0.05; *** = P<0.01).

4 Cecagno et al.

Table 2 - Microbial activity in an Ultisol, with continuous grazing in native pasture with Italian ryegrass introduction, under different nitrogen topdressing rates.

Soil attribute	Soil layer (cm)	Nitrogen rate – kg ha ⁻¹			Average
		0	100	200	
	0 - 5	255±8.1	285±0.6	305±11.5	281±0.9 a
Soil microbial biomass carbon* (µg g ⁻¹)	5 – 10	171±24.3	169±13.7	224±1.9	188±13.3 b
	Average	213±8.1	227±6.5	264±6.7	
	0 - 5	35±1.3 aC	44±1.4 aB	49±3.0 aA	43±1.9
Soil microbial biomass nitrogen**** (μg g ⁻¹)	5 – 10	24±0.4 b	25±1.4 b	24±0.6 b	24±0.1
	Average	29±0.5	34±1.4	37±1.2	
	0 - 5	1630±50	2090±78	2490±355	2070±161 a
Microbial respiration** (μg C-CO ₂ g ⁻¹ soil h ⁻¹)	5 – 10	740±223	800±100	870±93	800±139 b
	Average	1190±137 B	1450±89 AB	1680±224 A	
	0 - 5	6.40 ± 0.02	7.35 ± 0.20	8.18±1.07	7.31±0.42 a
Metabolic quotient** (μgC-CO ₂ μg Cmic g ⁻¹ soil h ⁻¹)	5 – 10	4.32 ± 0.60	4.72 ± 0.06	3.89 ± 0.26	4.31±0.27 b
	Average	5.36±0.29	6.04 ± 0.07	6.04 ± 0.66	

Values are mean and standard error (n=2) and those followed by the same capital letter in the lines and lower case letter in the columns do not differ according to Tukey test (* = P<0.10; ** = P<0.05; *** = P<0.01).

Total nitrogen stocks also were not affected by N fertilization (Table 1). Grazing recycles (as manure, urine and plant senescent material) a large soil N fraction of soil or fertilizer-N. Residue decomposition increases N inorganic availability becoming available to leaching, denitrification and ammonia volatilization processes (PARDON et al., 2016).

Rainfall events with at least 5mm within two days after urea application led N volatilization losses of less than 20% of the applied N (WHITEHEAD, 1995). Nitrogen applications under this trial were always carried under low soil moisture conditions. In the evaluated year (2010), rainfall only occurred nine and seven days after the first and second application, respectively, leading to potential NH₃ losses. Therefore, a fraction of the applied N rates was potentially lost even before soil absorption.

Cation exchange capacity (CEC), pH, buffering capacity and SOM content are important factors that influence ammonia losses by volatilization (KNOBLAUCH et al., 2012). The surrounding urea granule region may rise up to 3 pH units, creating favorable conditions for NH₃ volatilization even in acidic soils (WHITEHEAD, 1995). In low buffering capacity soils and low CEC_{pH7.0}, as in the present trial, volatilization may have occurred a long time after fertilization (FRENEY et al., 1983).

The N losses by nitrate leaching occurs from 10 to 30% of the fertilized N (MEISINGER et al., 2008) and may differ according to the soil texture, N rates and the water percolation in the soil profile.

In average, historically, there is an annual water surplus of 300mm (BERGAMASCHI et al., 2013), that can be leached into the soil profile. As a result, the lack of increase in N stocks even after long-term N fertilization may occur for two main reasons: 1) ammonia volatilization; and 2) soil texture and climate of the area favoring leaching losses (WHITEHEAD, 1995; MEISINGER et al., 2008).

The TN as well as the TOC stock was higher in the 0-20cm layer (Table 1). Such pattern results from surface residue deposition, higher biological activity due to higher root growth and exudates production (HAFNER et al., 2014). Despite N rates not affecting POC, the PN at 0-20cm layer was higher (P<0.10) under moderate rate, being similar to the reference area (Table 1). Such behavior can be explained by the higher microbial activity under high N rates. GAJDA (2010) also observed particulate organic matter losses in soils under high microbial activity.

Microbial activity responds faster to different grazing management processes than the TOC and TN, being more intense in the 0-5cm layer (Table 2). Despite the evaluated soil layer, the N rate effects only affected (P<0.10) the microbial respiration and, in the 0 to 5cm layer, the SMBN. SMBN was higher under the highest N rates and higher microbial respiration occurred under all N rates (Table 2). Similar N fertilization impacts in the long-term conditions were observed by HATCH et al. (2000), which can be explained by the increase in available N fraction

and microbial protein synthesis as well as increasing TOC and microbial biomass. As observed in this trial, PEREZ et al. (2005) reported no N addition effect in the SMBN in the 5-10cm layer. This occurs due to residues accumulation on the soil surface leading to an increase in microbial respiration (PEÑA et al., 2005). However, microbial respiration is high in all treatments compared with the results of SILVA et al. (2010). Even under conventional tillage and in a clayey soil, these authors reported lower values due to the higher organic matter physical protection resulted from clay-organic matter interactions.

CONCLUSION

The long-term nitrogen fertilization in a native pasture with Italian ryegrass does not change total soil carbon and nitrogen stocks. Yet, rates of 100kg N ha⁻¹ yr⁻¹ increase soil nitrogen in its particulate fraction. The increase in N fertilization rates up to 200kg N ha⁻¹ yr⁻¹ led to increases in the soil microbial biomass nitrogen as well as microbial respiration. Thus, the rate of 100kg ha⁻¹ yr⁻¹ increases in soil health, increasing N in the more labile organic matter fraction and enhancing N cycling in the soil system.

REFERENCES

ALEF, K.; NANNIPIERI, P. Methods in applied soil microbiology and biochemistry. London: Academic, 1995. 576p.

ANDERSON, J.P.E.; DOMSCH, K.H. The metabolic quotient (qCO₂) as a specific activity parameter to assess the effects of environmental conditions, such as pH, on the microbial biomass of forest soils. **Soil Biology and Biochemistry**, v.25, p.393-395, 1993. Available from: http://www.sciencedirect.com/science/article/pii/0038071793901407>. Accessed: Feb. 19, 2017.

BERGAMASCHI, H. et al. **Boletins agrometeorológicos da Estação Experimental Agronômica da UFRGS**: série histórica 1970 - 2012. Porto Alegre, RS, 2013. Available from: www.ufrgs.br/agronomia/joomla/files/EEA/Srie_Meteorolgica_da_EEA-UFRGS.pdf Accessed: Feb. 19, 2017.

BODDEY, R.M. et al. Carbon accumulation at depth in Ferralsols under zero-till subtropical agriculture. **Global Change Biology**, v.16, p.784-795, 2010. Available from: http://onlinelibrary.wiley.com/doi/10.1111/j.1365-2486.2009.02020.x/abstract. Accessed: Feb. 19, 2017.

BROOKES, P.C. et al. Chloroform fumigation and the release of soil nitrogen: a rapid direct extraction method to measure microbial biomass nitrogen in soil. **Soil Biology and Biochemistry**, v.17, p.837-842, 1985. Available from: http://www.sciencedirect.com/science/article/pii/0038071785901440. Accessed: Feb. 19, 2017.

CAMBARDELLA, C.A.; ELLIOT, E.T. Particulate soil organicmatter changes across a grassland cultivation sequence. **Soil Science Society of America Journal**, v.56, p.777-783, 1992. Available from: https://dl.sciencesocieties.org/publications/sssaj/abstracts/56/3/SS0560030777. Accessed: Feb. 19, 2017.

CARTER, M.R. Soil quality for sustainable land management: organic matter and aggregation interactions that maintain soil functions. **Agronomy Journal**, v.94, p.38-47, 2002. Available from: https://dl.sciencesocieties.org/publications/aj/abstracts/94/1/38>. Accessed: Feb. 19, 2017.

CONTE, O. et al. Soil density, aggregation and carbon fractions of an Alfisol under natural pasture and different herbage allowance. **Revista Brasileira de Ciência do Solo**, v.35, p.579-587, 2011. Available from: www.scielo.br/pdf/rbcs/v35n2/v35n2a27.pdf. Accessed: Feb. 19, 2017.

CONTERATO, I.F. et al. Agronomic behavior of annual ryegrass (*Lolium multiflorum L.*) in Rio Grande do Sul state. **Boletim de Indústria Animal**, v.73, p.198-205, 2016. Available from: <www.iz.sp.gov.br/pdfsbia/1475172230.pdf>. Accessed: Feb. 19, 2017.

COSTA, K.A.P. et al. Nitrogen doses and sources in Marandu pasture. I - Changes in soil chemical properties. **Revista Brasileira de Ciência do Solo**, v.32, p.1591-1599, 2008. Available from: http://www.scielo.br/pdf/rbcs/v32n4/a23v32n4.pdf>. Accessed: Feb. 19, 2017.

ELLERT, B.H.; BETTANY, J.R. Calculation of organic matter and nutrients stored in soils under contrasting management regimes. **Canadian Journal of Soil Science**, v.75, p.529-538, 1995. Available from: http://pubs.aic.ca/doi/pdf/10.4141/cjss95-075. Accessed: Feb. 19, 2017.

FORNARA, D.A.; TILMAN, D. Soil carbon sequestration in prairie grasslands increased by chronic nitrogen addition. **Ecology**, v.93, p.2030-2036, 2012. Available from: https://www.ncbi.nlm.nih.gov/pubmed/23094375. Accessed: Feb. 19, 2017.

FRENEY, J.R. et al. Volatilization of ammonia. In: FRENEY, J.R.; SIMPSON, J.R. **Gaseous loss of nitrogen from plant-soil systems**. The Hague: Martinus Nijhoff, 1983. p.1-32.

GAJDA, A.M. Microbial activity and particulate organic matter content in soils with different tillage system use. **International Agrophysics**, v.24, p.129-137, 2010. Available from: https://www.old.international_agrophysics/IntAgr_2010_24_2_129.pdf>. Accessed: Feb. 19, 2017.

HAFNER, S. et al. Spatial distribution and turnover of root-derived carbon in alfalfa rhizosphere depending on top- and subsoil properties and mycorrhization. **Plant and Soil**, v.380, p.101-115, 2014. Available from: http://link.springer.com/article/10.1007%2Fs11104-014-2059-z- Accessed: Feb. 19, 2017.

HATCH, D.J. et al. Nitrogen mineralization and microbial activity in permanent pastures amended with nitrogen fertilizer or dung. **Biology and Fertility of Soils**, v.30, p.288-293, 2000. Available from: http://link.springer.com/article/10.1007%2Fs003740050005. Accessed: Feb. 19, 2017.

KHAN, S.A. et al. The myth of nitrogen fertilization for soil carbon sequestration. **Journal of Environmental Quality**, v.36, p.1821-1832, 2007. Available from: https://dl.sciencesocieties.org/publications/jeq/abstracts/36/6/1821. Accessed: Feb. 19, 2017.

KNOBLAUCH, R. et al. Volatilização de amônia em solos alagados influenciada pela forma de aplicação de ureia. **Revista Brasileira de Ciência do Solo**, v.36, p.813-821, 2012. Available from: <www.scielo.br/pdf/rbcs/v36n3/12.pdf>. Accessed: Feb. 19, 2017.

6 Cecagno et al.

MAJUMDER, B. et al. Soil organic carbon pools and productivity relationships for a 34-year-old rice-wheat-jute agroecosystem. **Plant and Soil**, v.297, p.53-67. 2007. Available from: http://link.springer.com/article/10.1007%2Fs11104-007-9319-0. Accessed: Feb. 19, 2017.

MEISINGER, J.J. et al. Soil nitrogen budgets. In: SCHEPERS, J.S.; RAUN, W.R. **Nitrogen in agricultural systems**. Madison: American Society of Agronomy, 2008. p.505-562.

NANNIPIERI, P. et al. Microbial diversity and soil functions. **European Journal of Soil Science**, v.54, p.655-670, 2003. Available from: http://onlinelibrary.wiley.com/doi/10.1046/j.1351-0754.2003.0556.x/abstract. Accessed: Feb. 19, 2017.

OLSON, K.R. et al. Soil organic carbon changes after 12 years of notillage and tillage of Grantsburg soils in southern Illinois. **Soil and Tillage Research**, v.81, p.217-225, 2005. Available from: http://www.sciencedirect.com/science/article/pii/S0167198704001977. Accessed: Feb. 19, 2017.

PARDON, L. et al. Quantifying nitrogen losses in oil palm plantations: models and challenges. **Biogeosciences**, v.13, p.5433-5452, 2016. Available from: www.biogeosciences.net/13/5433/2016/bg-13-5433-2016.pdf>. Accessed: Feb. 19, 2017.

PEÑA, M.L.P. et al. Respiração microbiana como indicador da qualidade do solo em ecossistema florestal. **Floresta**, v.35, p.117-127, 2005. Available from: http://ojs.c3sl.ufpr.br/ojs/index.php/floresta/article/viewArticle/2435. Accessed: Feb. 19, 2017.

PEREZ, K.S.S. et al. Nitrogênio da biomassa microbiana em solo cultivado com soja, sob diferentes sistemas de manejo, nos cerrados. **Pesquisa Agropecuária Brasileira**, v.40, p.137-144, 2005. Available from: http://www.scielo.br/pdf/pab/v40n2/23820.pdf>. Accessed: Feb. 19, 2017.

SARMENTO, P. et al. Atributos químicos e físicos de um argissolo cultivado com *Panicum maxicum* Jacq. cv. IPR-36 Milênio, sob lotação rotacionada e adubado com nitrogênio. **Revista Brasileira**

de Ciência do Solo, v.32, p.183-193, 2008. Available from: http://www.scielo.br/pdf/rbcs/v32n1/18.pdf>. Accessed: Feb. 19, 2017.

SEBRAE/SENAR/FARSUL. Diagnóstico de sistemas de produção de bovinocultura de corte no estado do Rio Grande do Sul. Porto Alegre: SENAR, 2005. 265p.

SILVA, R.R. et al. Biomassa e atividade microbiana em solo sob diferentes sistemas de manejo na região fisiográfica Campos das Vertentes—MG. **Revista Brasileira de Ciência do Solo**, v.34, p.1585-1592, 2010. Available fom: www.scielo.br/pdf/rbcs/v34n5/11.pdf>. Accessed: Feb. 19, 2017.

SOUSSANA, J.F.; LEMAIRE, G. Coupling carbon and nitrogen cycles for environmentally sustainable intensification of grasslands and crop-livestock systems. **Agriculture, Ecosystems and Environment**, v.190, p.9-17, 2014. Available from: http://www.sciencedirect.com/science/article/pii/S0167880913003526. Accessed: Feb. 19, 2017.

SOUZA, E.D. et al. Biomassa microbiana do solo em sistema de integração lavoura-pecuária em plantio direto, submetido a intensidades de pastejo. **Revista Brasileira de Ciência do Solo**, v.34, p.79-88, 2010. Available from: http://www.scielo.br/pdf/rbcs/v34n1/a08v34n1.pdf>. Accessed: Feb. 19, 2017.

VANCE, E.D. et al. An extraction method for measuring microbial biomass C. **Soil Biology and Biochemistry**, v.19, p.703-707, 1987. Available from: http://www.sciencedirect.com/science/article/pii/0038071787900526>. Accessed: Feb. 19, 2017.

WHITEHEAD, D.C. Volatilization of ammonia. In: WHITEHEAD, D.C. **Grassland nitrogen**. Wallingford: CAB International, 1995. p.152-179.

YU, H. et al. Decreasing nitrogen fertilizer input had little effect on microbial communities in three types of soils. **Plos One**, v.11, p.1-12, 2016. Available from: http://journals.plos.org/plosone/article?id=10.1371/journal.pone.0151622. Accessed: Feb. 19, 2017.