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# Methylxanthine and catechin content of fresh and fermented cocoa beans, dried cocoa beans, and cocoa liquor

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## Abstract

The theobromine and catechin content can affect the quality of cocoa liquor and is influenced by cacao variety, production area (PA), and fermentation, as well as the method of drying beans (FDB) and cocoa liquor production (CLP). This study examined variations in methylxanthine and catechin levels in fresh and fermented cocoa beans, dried cocoa grains, and in cocoa liquor from Trinitario, Criollo, and Forastero cacao varieties. A total of 123 cocoa bean samples from three Peruvian PAs at different altitudes, Tingo María (TM), San Alejandro (SA), and Curimana (CU), were evaluated. The theobromine (Tb) and caffeine (Cf) contents in fresh cocoa beans were affected by both cocoa type and PA. The caffeine content was higher in Trinitario cacao than in Criollo and Forastero varieties ( $p \leq 0.05$ ). The Tb and CF contents decreased in dry cocoa grain and was affected by FDB ( $p \leq 0.05$ ) ( $1.449 \pm 0.004$  to  $1.140 \pm 0.010$  and  $0.410 \pm 0.03$  to  $0.165 \pm 0.02$  g Tb and C, respectively, per 100 g dry weight). Cocoa beans from Tingo María, which has the highest altitude, had higher Tb and CF contents than those from other PAs. The catechin (C) and epicatechin (EC) contents were affected by the FDB and CLP, and were highest in fresh cocoa beans from the Tingo María area (range:  $0.065 \pm 0.01$  to  $0.020 \pm 0.00$  g C/100 g). The C and EC contents decreased during FDB and CLP ( $0.001$  g C/100 g of cocoa liquor). Taken together, these results show that higher concentrations of Tb, Cf, C, and EC are present in fresh cocoa beans. Moreover, the cocoa variety influenced cocoa liquor quality. Overall, cocoa from the Tingo María PA had the most desirable chemical composition.

**Keywords:** HPLC; methylxanthines; theobromine; caffeine; catechin; epicatechin; cocoa.

## 1. Introduction

The post-harvest handling of cocoa is of critical importance to product quality, and if performed according to industry and market standards, can result in a better market price (Quarmin *et al.*, 2012). Cocoa is a popular foodstuff worldwide and has high acceptance by consumers (Sarriá *et al.*, 2015). Cocoa beans are the main cocoa product, and can act as a stimulant because of their theobromine and caffeine content (Araujo *et al.*, 2014). Multiple health benefits of cocoa can be attributed to its enrichment in polyphenols, which can modulate oxidative effects associated with the development of cardiovascular and cerebrovascular diseases,

as well as certain types of cancer (Giacometti *et al.*, 2016; Ioannone *et al.*, 2015).

The three major genetic groups of cocoa are Criollo, Forastero and Trinitario, which is a hybrid of Criollo and Forastero (Saltini *et al.*, 2013). Criollo cocoa trees are relatively rare and less robust compared to other varieties. The crown of the Criollo tree is round, and its pale green leaves are small and oval. Cocoa beans are ivory white. Criollo cocoa is characterized by elongated cobs that are green and red when the beans are immature, and as they ripen they turn yellow and orange-red. Cocoa beans from Criollo trees are desirable because of their nutty and fruity flavor.

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Criollo beans require between two and three days to ferment, and are very aromatic. Based on these qualities, Criollo beans have the commercial designation “fine cocoa” (Navia and Pazmiño, 2012). Meanwhile, Forastero, or Amazon, cocoa trees have violet staminodes, with cobs that turn yellow upon ripening. The cobs are grooved and slightly rough with smooth and rounded ends. The shells of Forastero cocoa beans are thick with a lignified mesocarp, and the beans themselves are flattened with purple cotyledons. Chocolate made from Forastero cocoa has a basic flavor. Trinitario cocoa is a polymorphic hybrid that carries Criollo intermediates and a Forastero strain that has relevant disjunction with Trinitario descendants. Among the tested strains, Trinitario cocoa has intermediate quality (Navia and Pazmiño, 2012).

The chemical composition of cocoa beans and their byproducts is complex and varies depending on the bean type, geographical area and maturity, as well as the fermentation and drying method (Bortolini *et al.*, 2016). Cocoa contains methylxanthine alkaloids with a purine base (e.g., theobromine, caffeine and theophylline); these compounds have stimulatory activity. Theobromine ( $C_7H_8N_4O_2$ , 3,7-dimethylxanthine or 3,7-dihydro-3,7-dimethyl-1H-purine-2,6-dione) is the main methylxanthine in cocoa that represents ~4% of cocoa butter, or between 0.8 and 2% of total cocoa bean dry weight (Carrillo *et al.*, 2014). The quantity of the xanthine alkaloid caffeine in cocoa averages around 0.2%, and varies depending on bean type, plant species, and degree of fermentation (Carrillo *et al.*, 2014). Meanwhile, theophylline in cocoa is present at very low amounts. Menguy *et al.* (2009) reported that fresh cocoa had theobromine and caffeine contents of 27 and 6.1 mg g<sup>-1</sup>, respectively, whereas fermented cocoa had 21 and 4 mg g<sup>-1</sup>, respectively. Although alkaloids do not undergo chemical transformations during fermentation, about 30% of alkaloids are eliminated by diffusion and migration to the outside of the bean

(Camu *et al.*, 2008). The bitter taste of cocoa is mostly influenced by the purine content (theobromine and caffeine), and, by phenolic compounds (Kongor *et al.*, 2016; Brunetto *et al.*, 2007).

Catechin is a component of cocoa tannins (Pedan *et al.*, 2015) which contribute to bean color and astringency, and is soluble in ethanol. Cocoa beans contain four types of catechins, of which (-)-epicatechin is the main type (92% of total).

Cocoa beans also contain a variety of bioactive compounds, including polyphenols, the content of which can differ substantially between beans and cocoa derivatives. Among polyphenols present in cocoa are quercetin, isoquercitrin (quercetin 3-O-glucoside), hyperoside (quercetin 3-O-galactoside), naringenin, luteolin, and apigenin. Cocoa beans are rich in flavonoids, mainly flavanols, as well as flavones, anthocyanins, and other phenolic compounds (Torres, 2012).

Most initial pre-industrial processing of cocoa beans, including fermentation and sun drying, is performed at harvest. These processes are important to develop characteristic aromas and aroma precursors in chocolate. After harvesting the cob, cocoa beans are removed and carried to wooden boxes where fermentation takes place for either 2 to 3 days for Criollo cocoa beans, or 5 to 7 days for Forastero cocoa beans (Gil, 2012). During cocoa bean fermentation, polyphenols diffuse from cellular compartments and are oxidized to produce insoluble high molecular weight tannins. These oxidation reactions are catalyzed by polyphenol oxidase, which can be inactivated strongly during the first day of fermentation, such that the enzymatic activity falls from 50% full activation to 6% between days 1 and 2. The main objective of drying is to reduce grain humidity (to 5-7%) to decrease the likelihood of fungal contamination and facilitate storage and transport.

To our knowledge, there are no studies on the chemical composition of cocoa or cocoa byproducts from different cocoa varieties grown in Peru. Such information

would be important given that loss of catechins and others polyphenols that occurs during post-harvest, pre-industrialization, and industrialization processes can lessen product quality (Giacometti *et al.*, 2015).

This study sought to assess variations in methylxanthine and catechin contents of fresh cocoa beans, fermented cocoa grains, dry cocoa grains and cocoa liquor. The samples were from Tingo María (TM), Huánuco department, as well as San Alejandro (SA) and Curimana (CU) in the Ucayali department in Peru. Tingo María has the highest altitude, and Curimana has the lowest.

## 2. Materials and methods

### 2.1 Cocoa beans and reagents

A total of 82 samples of cocoa pods were collected in the Tingo María area (TM) at 660 meters above sea level (MASL) and 09° 17' 08" south latitude, 75° 59' 52" west latitude, whereas 21 samples were from the San Alejandro area (SA) at 217 MASL and 08° 49' 44" south latitude, 75° 12' 44" west latitude. Twenty samples from the Curimana area (CU) were taken at 180 MASL and 08° 27' 51" south latitude, at 75° 09' 03" north latitude. Samples were collected with the support of the Cooperativa Agraria Cafetalera Divisoria Ltda., using information from farmers about trees that produce grains with special flavors and aromas. Theobromine  $\geq$  99.0%, anhydrous caffeine, catechin  $\geq$  98%, and methanol (Sigma, St. Louis, MO, USA) were used as standards. The following compounds isolated from green tea and were also purchased from Sigma: epicatechin  $\geq$  98%, epigallocatechin gallate  $\geq$  80% (HPLC), epigallocatechin  $\geq$  95%, and epicatechin gallate  $\geq$  98%. Ethanol (99.99%), glacial acetic acid, and deionized distilled water were from Merck.

### 2.2 Preparation of cocoa sample and extract

Samples from fresh cocoa pods were immediately transported to the laboratory where grain samples were cracked and extracted before drying at 35° C for 8 h

and at 65 °C for 18 hours. The final humidity of the grains was 7%. Grains were stored in polyethylene bags at -20 °C for later use. Dried fresh beans were peeled manually and ground in a grinder before theobromine, caffeine, and catechins were extracted from 0.02 g ground cocoa in 5 ml of acetic acid 0.3%, and HPLC grade methanol (85:15). The mixture was vortexed for 15 min and a 500 µg/mL solution was prepared, centrifuged, and passed through a microfilter prior to injection into an HPLC (see below).

Fresh, fermented, and dry samples were also examined. To obtain fermented beans micro-fermentation was performed for different time periods (48, 96 and 120 hours). For this procedure, 2 kg cocoa grain was separated from the cocoa slime mass in a nylon bag or plastic netting. The nylon bag maintained the "inserted product" as a separate sample but was exposed to the same conditions as the remaining grains. The bag was then inserted in the top (30 cm) of a fermentation box that contained a larger amount (200 kg) of cocoa slime. The "inserted product" was inverted at the same time as the rest of the cocoa mass in the slime (48 and 96 hours), and the total fermentation time was 120 hours. During micro-fermentation we took samples (10 grains) at 0, 48, 96 and 120 hours for evaluation. These samples were placed in polyethylene bags and transported to the laboratory in a container with ice.

Dried cocoa samples were obtained by sun drying to 7% humidity. Fresh, fermented (48, 96 and 120 hours), and dried beans were peeled manually, milled in a grinder, centrifuged, filtered and extracted for analysis of theobromine, caffeine and catechin compounds.

### 2.3 HPLC analysis of cocoa bean chemical contents

Chromatographic analysis was performed using reverse phase liquid chromatography on a LC-10 AT VP Scientific apparatus (Shimadzu, MD, USA) that included: a Shimadzu DGU degasser-14A; a manual Rheodyne 7725i injector; a management

pack solvent that included a quaternary pump, a Shimadzu LC-10 AT and one column oven (Shimadzu CTO-10AS). A Shimadzu UV-Vis SPD\_10AV detector was used to determine the chromatographic peak purity at SCL interface 10AV. Peak identification and integration was performed using Shimadzu software Class-VP version 6.13 SP2. Chromatographic separations were performed with a guard ultra-cartridge C18 column (20 mm x 4 mm, Restek, Catalog: 917 450 220) and an Ultra C18 column (150 mm x 4.6 mm x 5  $\mu$ m, Restek, Catalog: 9174565). The column was maintained at 35° C. The mobile phase was a mixture of 24.9% methanol, 75.0% distilled deionized water, acetic acid (100:0.1), and the flow rate was 1 ml/min. Solutions were filtered with a 0.2  $\mu$ m nylon microfilter prior to injection of a 20  $\mu$ l sample volume. Simultaneous chromatographic analysis of theobromine, caffeine, and catechins in selected samples was performed at a wavelength of 210 nm, whereas the selected mobile phase was 20% methanol: 80% water, acetic acid (100:0.1).

## 2.4 Determination of methylxanthine and catechin content in cocoa

The HPLC apparatus described above was also used to quantify theobromine (Tb), caffeine (Cf) (Menguy *et al.*, 2009), catechin (C), epicatechin (EC), epigallocatechin (EGC), epicatechin gallate (ECG) and epigallocatechin gallate (EGCG) (Meng *et al.*, 2009) in three samples for fresh, fermented (48, 96, and 120 hours) grain, and liquor samples according to previously described methods.

### 2.4.1 Preparation of theobromine and caffeine standards

Before injecting samples into the HPLC instrument, calibration with theobromine standards was performed. 1 mM and 3 mM stock solution was diluted in HPLC grade deionized distilled water to prepare 3, 6, 12, 24, 36, 48, and 60  $\mu$ M and 1.5, 3, 6, 12, 18, 24, and 30  $\mu$ M theobromine and caffeine solutions, respectively, which were used to generate standard curves. Aliquots of theobromine and caffeine stock

solutions were stored at -20° C to preserve their stability and quality, and solutions were filtered prior to HPLC injection.

### 2.4.2 Preparation of catechin standards

Stock solutions (500 ppm) were used for catechin, epicatechin, epicatechin gallate, and epigallocatechin gallate, whereas the epigallocatechin stock solution was 418 ppm. The stock solutions were stored at -20 °C. A mixture of standards was prepared to generate curves simultaneously. Five concentrations were measured in triplicate such that the final solution concentrations were: 1.72, 17.23, 25.84, 51.68, 103.36  $\mu$ M (catechin); 2.07, 4.13, 41.34, 82.68, 165.36  $\mu$ M (epicatechin); 2.26, 15.82, 31.65, 63.30, 126.59  $\mu$ M (epicatechin gallate); 1.09, 10.91, 16.36, 32.72, 76.36  $\mu$ M (epigallocatechin gallate); and 1.63, 8.16, 16.33, 48.98, 97.95  $\mu$ M (epigallocatechin).

## 2.5 Cocoa genotyping

Cocoa genotype selection was based on a comparison of the Tb/Cf ratio with Cf, which indicates the cocoa genotype, and was calculated using previously described methodology (Carrillo *et al.* 2014). Cocoa index of Tb/Cf < 3, 3-9, and >9 is consistent with Criollo type, Trinitario type, and Forastero type, respectively.

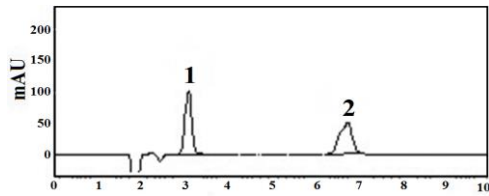
## 2.6 Statistical analysis

Tb, Cf, and catechin content in fresh, fermented, and dried cocoa grains, as well as cocoa liquor were statistically evaluated, and a completely randomized design was used with three replications (Rios, 2012). Statistical differences were evaluated by a Tukey test. All statistical analyses were performed using STATGRAPHICS Centurion XV software version 2.15.06.

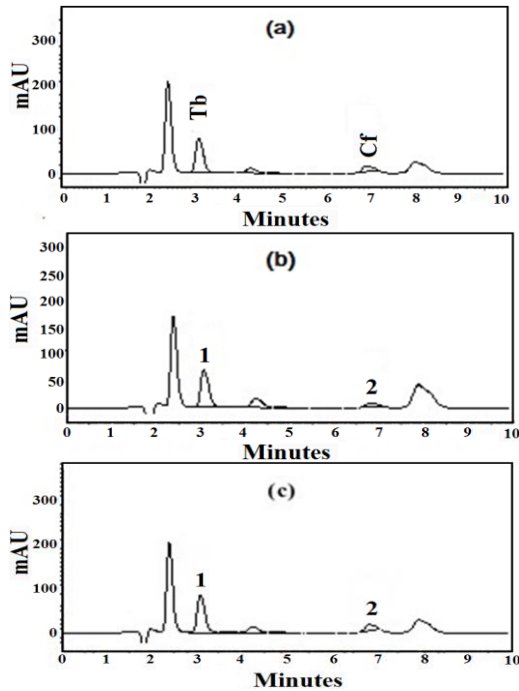
## 3. Results and discussion

### 3.1 Cocoa genotyping

Theobromine and caffeine contents of fresh cocoa beans were analyzed by HPLC at a wavelength of 210 nm. To quantify theobromine and caffeine contents we first generated standard curves for both compounds. These curves showed that theobromine (Tb) eluted first, followed by caffeine (Cf) (Fig. 1).



**Figure 1.** Chromatographic profile of theobromine (1) and caffeine (2f) standards.



**Figure 2.** Chromatographic profile of fresh cocoa beans from Tingo María (a), San Alejandro (b), and Curimana (c) areas; Theobromine (1) and caffeine (2).

Standard curves for Tb and Cf with seven points and three replications were generated using the equations below:

$$\text{Area} = 29441 \mu\text{M Tb} + 18936, r^2 = 0.9962$$

$$\text{Area} = 33519 \mu\text{M Cf} + 5963, r^2 = 0.9962$$

**Table 1**

Tb and Cf contents and Tb/Cf ratios of cocoa samples from Tingo María, San Alejandro, and Curimana areas

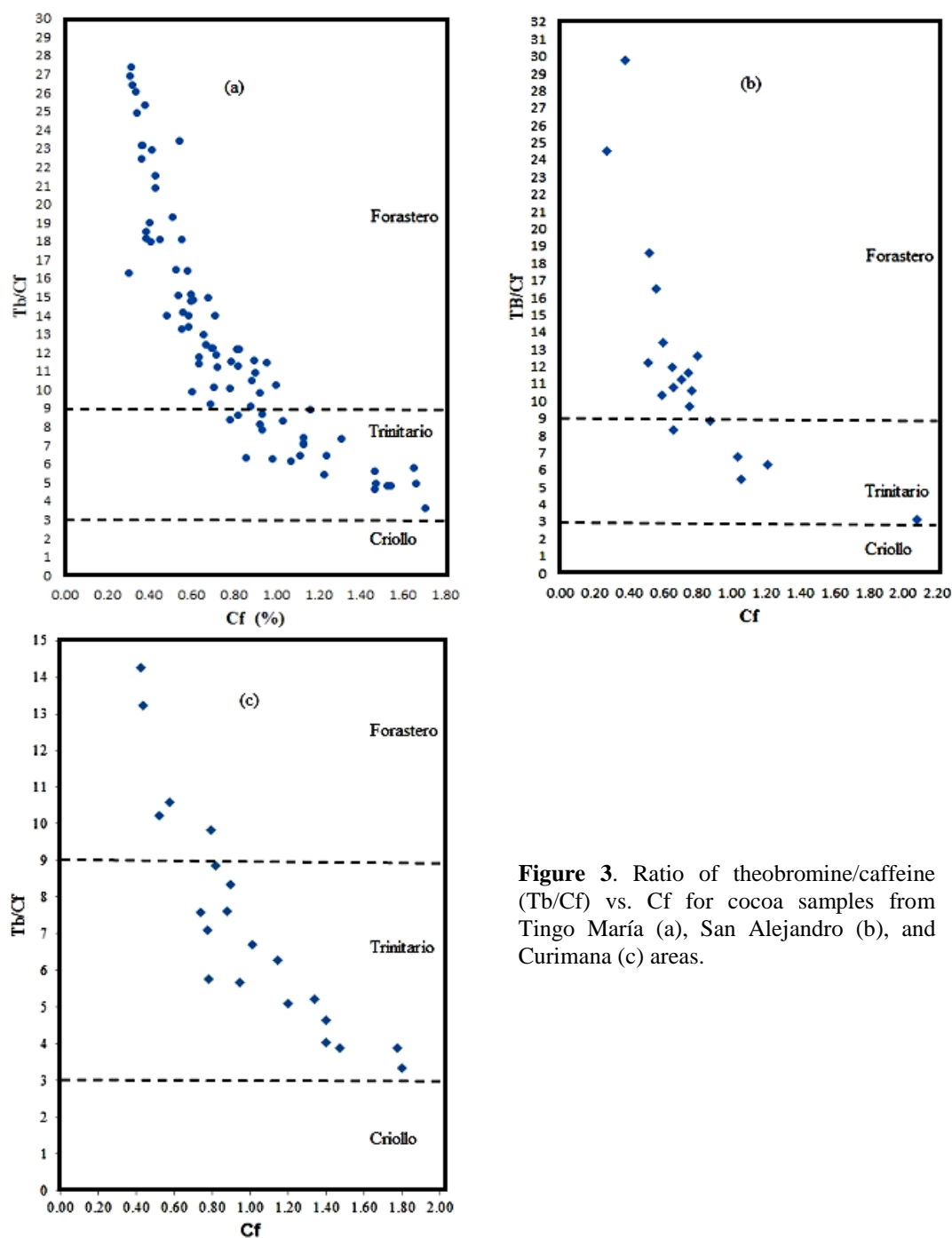
Place of Production	Code	Tb	Cf	Index of
		(g/100 g of cocoa)	(g/100 g of cocoa)	Tb/Cf
Tingo María	OC1	$6.195 \pm 0.189$	$1.706 \pm 0.108$	$3.65 \pm 0.132$
	JC0	$6.811 \pm 0.129$	$1.465 \pm 0.051$	$4.66 \pm 0.149$
	TN2	$7.359 \pm 0.203$	$1.526 \pm 0.045$	$4.83 \pm 0.137$
San Alejandro	TMG1	$6.460 \pm 0.029$	$2.083 \pm 0.025$	$3.10 \pm 0.053$
	CCM1	$5.774 \pm 0.109$	$1.055 \pm 0.016$	$5.47 \pm 0.020$
	GCZ6	$7.606 \pm 0.043$	$1.208 \pm 0.018$	$6.30 \pm 0.057$
Curimana	VCM4	$6.025 \pm 0.217$	$1.804 \pm 0.054$	$3.34 \pm 0.108$
	VCM2	$6.879 \pm 0.125$	$1.778 \pm 0.014$	$3.87 \pm 0.051$
	CMA4	$5.735 \pm 0.09^a$	$1.476 \pm 0.020$	$3.89 \pm 0.058$

Values represent the average of  $\pm$  SEM, n= 3 20  $\mu\text{L}$  injections of HPLC.

Figure 2 shows chromatographic profile of Theobromine (1) and caffeine (2) of fresh cocoa beans from Tingo María (a), San Alejandro (b), and Curimana (c) areas. Table 1 shows the theobromine (Tb) and caffeine (Cf) content together with the Tb/Cf ratio. The index relationship of Tb/Cf vs. Cf in beans from Tingo María (TM), San Alejandro (SA), and Curimana (CU) areas ranged between 3.10 and 6.30 (Fig. 3). Values between three and nine were previously shown to correspond to Trinitario cocoa (Fig. 3), whereas values less than three and greater than nine are representative of Criollo and Forastero cocoa, respectively (Carrillo *et al.*, 2014; Zambrano *et al.*, 2010).

Criollo cocoa is less bitter, which is likely due to its lower concentration of theobromine relative to other genetic groups. Different types of cocoa are known to have different methylxanthine contents, and these differences are useful for differentiating cocoa genotypes. Our statistical analysis showed that the methylxanthine content was indeed affected by cocoa type and production area ( $p \leq 0.05$ ). Tests based on average values for a given cocoa growth area showed that the theobromine content in samples was lower for Trinitario cocoa, whereas this type had high caffeine content (Table 1). The Tb/Cf ratio is an important parameter for cocoa, since lower ratios are typical of Criollo cocoa as opposed to Trinitario or Forastero cocoas. High levels of caffeine are characteristic of high-quality Criollo cocoa (Trognitz *et al.*, 2013).



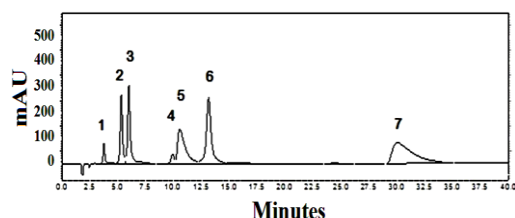


**Figure 3.** Ratio of theobromine/caffeine (Tb/Cf) vs. Cf for cocoa samples from Tingo María (a), San Alejandro (b), and Curimana (c) areas.

### 3.2 Tb, Cf, C and EC standard curve

A chromatographic profile of standard solutions showed an order of elution that is consistent with earlier reports, although there was a change in retention time (Fig. 4). The composition of the mobile phase (80:20) was acetic acid (0.1%) and methanol.

A mixture of 5 concentrations for each standard was evaluated with 3 replicates for each concentration. The  $r^2$  values ranging between 0.9984 and 0.9999 indicated a strong and almost perfect positive relationship. The minimum and maximum concentration of an analyte could be calculated within the linear range of the standard curve.



**Figure 4.** Chromatograms of standards for theobromine (1), epigallocatechin (2), catechin (3), caffeine (4), epigallocatechin (5), epicatechin (6), and epicatechin gallate (7).

**Table 2**

Equation standards to quantify the compounds detected in samples

Standards	Equations*	r <sup>2</sup>
Theobromine	Y = 155708X - 5953.8	0.9995
Catechin	Y = 175948X - 15985	0.9999
Caffeine	Y = 153598X + 26094	0.9984
Epicatechin	Y = 167567X + 104559	0.9990

\*Y = area of curve; X = ppm of compound.

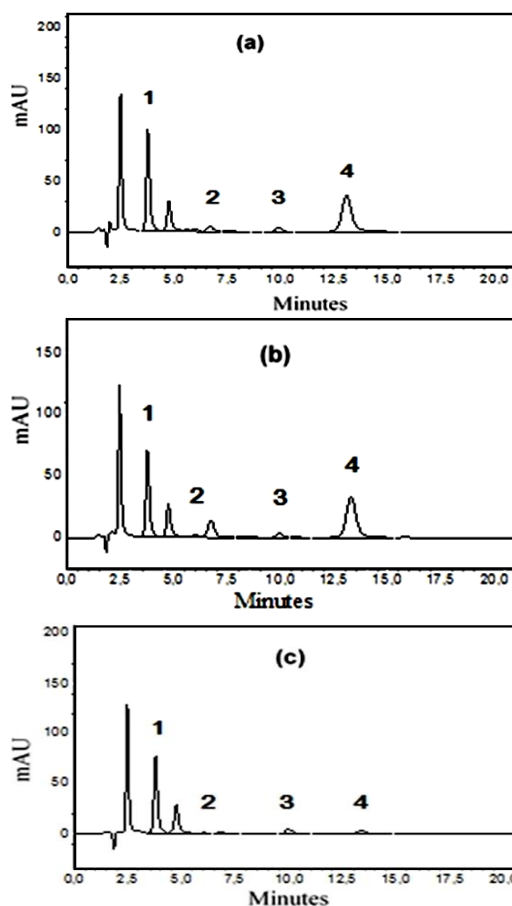
### 3.3 Theobromine, caffeine, catechin and epicatechin content

The bitter taste of cocoa is largely influenced by the methylxanthine (theobromine and caffeine) content, and to a lesser degree, by phenolic compounds. Indeed, Criollo cocoa has a lower bitter index due to its low theobromine content (Trogitz *et al.*, 2013). As such, we used HPLC to analyze methylxanthine contents of fresh cocoa beans from the Tingo María, San Alejandro, and Curimana PAs (Fig. 5a-c). The chromatograms showed high theobromine contents, and were similar to previous studies on cocoa beans (Fig. 5; Pereira-Caro *et al.*, 2013). Caffeine and theobromine are the most abundant methylxanthines in cocoa and their physiological effects are notable. The health-promoting benefits of cocoa are so pronounced that the therapeutic properties of chocolate have been the subject of several studies (Franco *et al.*, 2013).

### 3.4 Theobromine and caffeine

Measurements of theobromine (Tb) and caffeine (Cf) content in samples of fresh, fermented, and dried beans, as well as cocoa liquor showed that, relative to other production stages, fresh cocoa grains have

higher Tb contents, with values that varied between 1.088 and 1.449 g Tb/100 g (Table 3). Fresh cocoa grain from Tingo María had the highest Tb content, which was higher than those from an earlier investigation that reported values between 0.8622 and 0.8909 g Tb/100 g. This difference may be due to the extraction conditions used (Lo Coco *et al.*, 2007).



**Figure 5.** Chromatographic profiles of fresh cocoa beans,  $\lambda = 210$  nm, from Tingo María (a), San Alejandro (b), and Curimana (c) PAs. Chromatographic peaks for theobromine (1), catechin (2), caffeine (3) and epicatechin (4) are shown.

Analysis of Tb results revealed that processed cocoa and liquor had a statistically significant difference ( $p \leq 0.05$ ) in Tb contents. Moreover, a Tukey test showed that fresh cocoa beans had a higher theobromine content and that there was no statistical difference between the dry fermented samples and cocoa liquor.



**Table 3**

Theobromine (Tb) and caffeine (Cf) content in fresh cocoa beans, fermented cocoa (48, 96, and 120 hours), dried grains, and liquor from the Tingo María (TM) San Alejandro (SA), and Curimana (CU) PAs. Expressed in dried base<sup>1</sup>

Processing stage	TM	SA	CU	TM	SA	CU
	(g Tb/100 g of cocoa)			(g Cf/100 g of cocoa)		
Fresh	1.449 ± 0.004	1.088 ± 0.009	1.324 ± 0.074	0.410 ± 0.03	0.204 ± 0.01	0.350 ± 0.01
48 h	1.376 ± 0.029	1.047 ± 0.020	1.182 ± 0.020	0.317 ± 0.01	0.168 ± 0.01	0.312 ± 0.01
96 h	1.282 ± 0.003	1.002 ± 0.019	1.236 ± 0.081	0.277 ± 0.02	0.155 ± 0.00	0.182 ± 0.01
120 h	1.241 ± 0.011	0.952 ± 0.022	1.063 ± 0.043	0.210 ± 0.01	0.100 ± 0.00	0.139 ± 0.01
Dried	1.156 ± 0.005	0.885 ± 0.012	0.854 ± 0.011	0.200 ± 0.01	0.088 ± 0.00	0.087 ± 0.00
Liquor	1.140 ± 0.010	0.836 ± 0.011	0.940 ± 0.106	0.165 ± 0.02	0.077 ± 0.00	0.091 ± 0.01

<sup>1</sup> Values represent the average of ± SEM, n=3 injection of HPLC. The volume of each injection was 20 µL.

During processing that occurs between harvest and liquor production, the theobromine content decreases (Gil, 2012), but the chemical quality improves through reductions in alkaloid and polyphenol contents that influence the organoleptic characteristics of cocoa (Trognitz *et al.*, 2013). During fermentation, the theobromine content in cocoa from the Tingo María, San Alejandro, and Curimana PAs decreased by 21.33, 23.16 and 29%, respectively, relative to fresh beans. Similarly, previous reports showed that theobromine content is reduced by 20% during fermentation, and this reduction decreases the bitterness of the beans (Trognitz *et al.*, 2013; Brunetto *et al.*, 2007).

Meanwhile, fresh cocoa beans had a higher Cf content that varied between 0.204 ± 0.01 and 0.410 ± 0.03 g Cf/100 g. Fresh beans from Tingo María had the highest Cf content. Relative to earlier studies, the Cf contents found here were higher, and ranged from 0.0827 to 0.0908 g Cf/100 g. These differences may again be attributed to different extraction conditions (Lo Coco *et al.*, 2007).

Analysis of Cf results showed a statistically significant difference between post-harvest cocoa and cocoa liquor ( $p \leq 0.05$ )

Analysis of Cf results showed a statistically significant difference between post-harvest cocoa and cocoa liquor. A Tukey test established that fresh cocoa beans had a significantly higher Cf content than the other processing stages, and that there was

no significant difference between fermented and dried cocoa, or cocoa liquor. The caffeine content of cocoa from Tingo María, San Alejandro, and Curimana areas fell by 40.24%, 37.75%, and 26% from fresh cocoa beans, respectively. Meanwhile, the methylxanthine content is generally higher during the first days of fermentation, and gradually decreases as fermentation proceeds (Brunetto *et al.*, 2007).

The methylxanthine (theobromine and caffeine) content was affected by the FDB factor, and highlights that fresh beans from the Tingo María area have a higher content of Tb and Cf relative to other areas. These contents decreased during processing and liquor production to range from 1.449 ± 0.004 to 0.140 ± 0.010 g Tb/100 g of sample and 0.410 ± 0.03 to 0.165 ± 0.02 g Cf/100 g of sample. There was a statistically significant difference between the fresh beans and cocoa liquor.

### 3.5 Catechin and epicatechin

The catechin (C) and epicatechin (EC) contents were next measured for fresh, fermented, and dried cocoa, as well as cocoa liquor from the different areas (g per 100 g sample; Table 4). Fresh cocoa beans from the Tingo María area had a higher content of C (0.065 ± 0.01–0.013 ± 0.0 g C/100 g). These values were higher than those found earlier (0.0388–0.0434 g C/100 g of sample), the difference may be related to the presence of procyanidin (Rabadan-Chávez *et al.*, 2016).

**Table 4**

Catechin (C) and epicatechin (EC) contents in cocoa beans: fresh (0 hours), fermented (48, 96, and 120 hours), dry grains and cocoa liquor from cocoa from Tingo María (TM) San Alejandro (SA), and Curimana (CU) PAs. Expressed as dried base<sup>1</sup>

Processing stage	TM	SA	CU	TM	SA	CU
	(g C/100 g of cocoa)			(g EC/100 g of cocoa)		
Fresh	0.065 ± 0.01	0.020 ± 0.00	0.026 ± 0.00	1.855 ± 0.20	1.291 ± 0.01	0.055 ± 0.01
48 h	0.053 ± 0.01	0.007 ± 0.0	0.023 ± 0.00	1.180 ± 0.16	0.760 ± 0.04	0.041 ± 0.01
96 h	0.027 ± 0.00	0.003 ± 0.00	0.017 ± 0.00	0.931 ± 0.17	0.393 ± 0.08	0.043 ± 0.01
120 h	0.016 ± 0.00	0.003 ± 0.00	0.011 ± 0.00	0.496 ± 0.13	0.375 ± 0.05	0.029 ± 0.01
Dried	0.014 ± 0.00	0.002 ± 0.00	0.010 ± 0.00	0.504 ± 0.06	0.377 ± 0.05	0.033 ± 0.02
Liquor	0.013 ± 0.00	0.001 ± 0.00	0.004 ± 0.00	0.139 ± 0.01	0.086 ± 0.01	0.022 ± 0.01

<sup>1</sup> These values represent the average ± SEM, n=3 injection of HPLC. The volume of each injection was 20 µL.

Statistical analysis of C results indicated that the C content differed with processing stage. A Tukey test showed that fresh cocoa beans had higher C contents than the other stages, and there was no statistically significant difference ( $p \leq 0.05$ ) between fermented, dried, and liquor samples. This result is consistent with those presented in a study by Nazaruddin *et al.* (2006) that reported a decrease in the C content from  $0.58 \pm 0.10$  mg/g in mature beans to  $0.07 \pm 0.00$  in fermented and dried beans. In another study, the C content increased as the epicatechin content decreased. Specifically, during fermentation there was a marked increase (68%, 0.29 to 0.49 mg/g) in C that decreased to 0.24 mg/g for sun dried grains (Gil, 2012). EC contents also varied according to processing stage. A Tukey test showed that EC contents in cocoa beans from the Tingo María, San Alejandro, and Curimana PAs were higher in fresh beans relative to other processing stages ( $1.855 \pm 0.20$ ;  $1.291 \pm 0.01$ , and  $0.055 \pm 0.01$  g EC/100 g of sample, respectively). Values for beans from the San Alejandro PA were similar to that seen in a study by Gil (2012, 1.27 g EC/100 g). There was no statistically significant difference between beans and cocoa liquor, nor were there marked differences between the EC content of fermented and dried cocoa beans.

The EC content decreased during drying, reaching values between  $0.022 \pm 0.01$  and  $0.139 \pm 0.01$  g EC/100 g of cocoa liquor

(Table 4); although the 0.377 g EC/100 g value is almost equal to that reported earlier (Gil, 2012).

#### 4. Conclusions

In the higher altitude Tingo María and San Alejandro PAs, Forastero and Trinitario cocoa are the most prevalent varieties, whereas in the Curimana PA, which has the lowest altitude, Forastero cocoa is the most common. Statistical analyses determined that cocoa type and PA affected the theobromine and caffeine contents ( $p \leq 0.05$ ). Fresh cocoa beans had the highest theobromine and caffeine content, and the content decreased during fermentation and drying. Fresh beans from the PA with the highest altitude, Tingo María, had high theobromine and caffeine contents, suggesting that altitude may affect the chemical content of cocoa beans. The amount of theobromine and caffeine both decreased during fermentation, drying, and cocoa liquor production, (from  $1.449 \pm 0.004$  and  $1.140 \pm 0.010$ , and  $0.410 \pm 0.03$  and  $0.165 \pm 0.02$  g/100 g of theobromine and caffeine, respectively); there was also a statistical difference between the contents in fresh beans and cocoa liquor. C and EC contents also differed according to processing stage, and were highest in fresh cocoa beans. Cocoa from the high altitude Tingo María PA had the highest C content that varied between  $0.065 \pm 0.01$  and  $0.020 \pm 0.00$  g C/100 g of beans. This content decreased during fermentation, drying, and

liquor production, and reached values between  $0.001 \pm 0.0$  and  $0.013 \pm 0.0$  g C/100 g of liquor. Meanwhile EC concentrations varied between  $0.055 \pm 0.01$  and  $1.855 \pm 0.20$  g EC/100 g for fresh beans and  $0.022 \pm 0.01$  and  $0.139 \pm 0.01$  g EC/100 g of liquor, with no statistical difference seen between fermented and dried samples. Together these results show that Tb, Cf, C, and EC contents were affected by processing stage and PA, and were the highest in fresh cocoa beans harvested in the high altitude Tingo María area. The presence of polyphenols in cocoa samples and cocoa liquor indicate that cocoa products can have health benefits. Our results provide a basis for investigating the chemical characteristics of cocoa plantations and exploring how the functionality of cocoa can be improved in cocoa producing areas.

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