



Jornal Brasileiro de Patologia e Medicina Laboratorial

ISSN: 1676-2444

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Sociedade Brasileira de Patologia  
Clínica/Medicina Laboratorial  
Brasil

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Jornal Brasileiro de Patologia e Medicina Laboratorial, vol. 53, núm. 6, noviembre-diciembre, 2017, pp. 397-399

Sociedade Brasileira de Patologia Clínica/Medicina Laboratorial  
Rio de Janeiro, Brasil

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# Spontaneous chromatid break as clonal evolution in myelodysplastic syndrome patients

## *Quebra espontânea de cromátides como evolução clonal em pacientes com síndrome mielodisplásica*

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### ABSTRACT

Myelodysplastic syndrome (MDS) is a clonal hematopoietic stem cell disorder characterized by peripheral cytopenias due to ineffective erythropoiesis and an increased risk for evolving into acute myeloid leukemia (AML). Chromosomal abnormalities represent the most important marker of risk stratification for AML transformation. Chromatid break (chtb) is a discontinuity of a single chromatid. We report the case of a patient with MDS whose cytogenetic analysis showed spontaneous chromatid breakage (chrb): 46,XY,add(13)(q34),chtb(15)(q24)[3]/47,XY,chtb(2)(q22),del(5)(q35),del(7)(q32),+8,del(11q)(q23),del(q22)[cp17]. He was considered a high-risk patient due to the complex karyotype and the presence of chtb. We suggest that this chromosomal abnormality may be considered as a marker of genomic instability in MDS.

**Key words:** chromosomal breakage; myelodysplastic syndromes; chromosomal instability; genomic instability.

### INTRODUCTION

Myelodysplastic syndrome (MDS) is a clonal hematopoietic stem cell malignancy characterized by peripheral cytopenias due to ineffective erythropoiesis and an increased risk of evolving into acute myeloid leukemia (AML)<sup>(1)</sup>. The identification of chromosomal abnormalities by conventional cytogenetics (G-banding) is important to confirm diagnosis and crucial to determine prognosis and therapeutic approach to MDS. Chromosomal abnormalities are detected in 40% to 60% of MDS patients and represent the most important marker for risk stratification for AML transformation<sup>(2)</sup>.

The presence of mutation in deoxyribonucleic acid (DNA) was detected in up to 80% of the MDS patients<sup>(3)</sup>, many of these DNA lesions are due to increased oxidative stress, which is a common problem in MDS due to the inefficient erythropoiesis and the transfusion dependence<sup>(4-7)</sup>. Excess of iron is toxic and causes oxidative damage, which predisposes to genomic instability<sup>(7,8)</sup>.

Unfolding of chromatin structure occurs after DNA damage, so that repair enzymes can access the sites of the lesions. However, these modifications result in a mechanical stress of the chromatin and the chromosome may collapse at specific sites, leading to

chromatids break<sup>(9)</sup>. Chromatid break (chtb) is a discontinuity of a single chromatid in which there is a clear misalignment of one of the chromatids, originating acentric fragments and, consequently generating a misalignment of the chromosome that occurs, mostly, in later S phase and G2 of the cell cycle<sup>(10-14)</sup>.

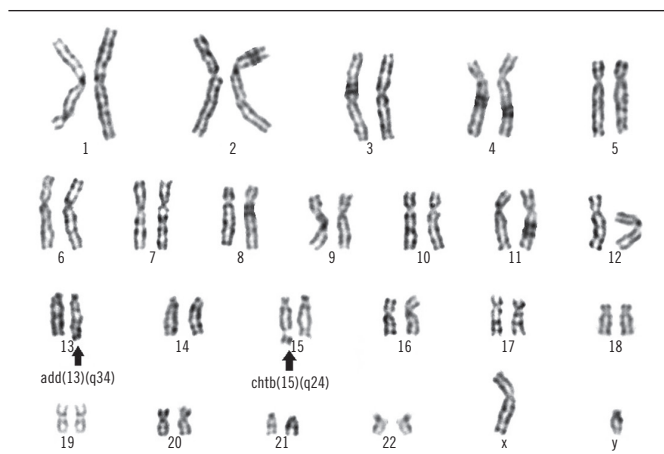
This type of aberration can be observed in rare autosomal recessive disorder such as Bloom syndrome<sup>(10-12)</sup> and following exposure to clastogenic drugs such as mitomycin C and bleomycin<sup>(10, 15)</sup>. The aim of this report is to present a rare case of MDS patient who showed spontaneous chromatid break (chtb) during clonal evolution, and to discuss the possible impact of this finding.

### CASE REPORT

A 77-year-old-man sought for medical attendance for malaise and fever. At physical examination, the patient was pale. The patient also reported a history of occupational exposure to solvent. Peripheral blood in the presentation showed hemoglobin 8.1 g/dl, mean corpuscular volume (MCV) = 96 fl, leukocytes 2000/μl (neutrophils 1000/μl) and platelets =  $52 \times 10^3/l$ . Bone marrow aspirate showed

erythroid dysplasia (> 10%) and micromegakaryocytes. Bone marrow biopsy demonstrated hypercellular marrow with erythroid dysplasia (50% of cells). Cytogenetic analysis showed: 46,XY,del(5)(q31),del(11)(q23)[7]/46,XY[5]. The diagnosis of refractory cytopenia with multilineage dysplasia [(RCMD) – MDS] was established according to the World Health Organization (WHO) classification<sup>(16)</sup> and the Revised International Prognostic Scoring System (IPSS-R)<sup>(17)</sup> was intermediate.

The patient has lost follow-up for one year and then presented hemoglobin 5.4 g/dl, leukocytes 2200/μl, neutrophils 1100/μl and platelets  $24 \times 10^3/l$ , and became transfusion dependent. At this time, a new cytogenetic analysis was performed and revealed: 46,XY,add(13)(q34),cthb(15)(q24)[3]/47,XY,cthb(2)(q22),del(5)(q35),del(7)(q32),+8,del(11q)(q23),del(q22)[cp17] (**Figure**). He was considered a high-risk patient due to the complex karyotype [according to the International System for Human Cytogenomic Nomenclature (ISCN) 2016] and the presence of spontaneous cthb; he progressed to death after four months.



**FIGURE** – Cytogenetic analysis after one year without follow-up

Cytogenetic analysis showed clonal evolution and the presence of spontaneous chromatid break.

## DISCUSSION

The chromosomal abnormality detected by G-banding is the most important marker of prognosis in MDS. Patients with complex karyotype and clonal evolution has a high risk for AML transformation<sup>(18)</sup>. Greenberg *et al.* (2012)<sup>(17)</sup> confirmed this finding by demonstrating that the cytogenetic abnormality is more predictive of AML evolution than blast counts. The most frequent chromosomal abnormalities in MDS are deletions and aneuploidy involving chromosomes 5, 7 and 8<sup>(17, 19, 20)</sup>. The reports of chromosomal breakage in MDS have been limited to clastogenic exposure or therapy related MDS<sup>(21)</sup>.

Korgaonkar *et al.* (2008)<sup>(15)</sup> evaluated 145 MDS cases using cultures of peripheral blood lymphocyte stimulated with phytohemagglutinin and treated with mitomycin C. They reported an increased sensitivity to chromosomal breakage after exposure to clastogenic (mitomycin). We found only one report demonstrating chromosomal breakage in MDS, but limited to centromeric region and detected only by multicolor fluorescence *in situ* hybridization (M-FISH). In this study, the authors attempted to correlate these findings with poor prognosis<sup>(21)</sup>. To the best of our knowledge, this is the first case of spontaneous chromatid break (cthb) detected by G-banding in MDS.

Vilalba-Campos *et al.* (2016)<sup>(22)</sup> evaluated the chromosomal abnormalities (by G-band) and the DNA damage (by comet assay) of the peripheral blood lymphocytes from 24 workers exposed to solvents and paint thinners and found a high frequency of chrB among these workers. Chromosomal abnormalities were 3.6 higher in the exposed group when compared to the non-exposed group, indicating a deleterious effect of the aromatic hydrocarbons, such as benzene, present in these solvents<sup>(21)</sup>, suggesting that the presence of chrB is related to the increase of DNA damage.

Our patient became transfusion dependent during the progression of the disease. Massive blood transfusion leads to an iron overload. Iron acts as a cofactor in the production of the hydroxyl radical ( $\cdot OH$ ), presenting a high reactivity with the DNA molecule, which may induce genomic instabilities and chromosomal breakage<sup>(23)</sup>. Bryant *et al.* (2010)<sup>(24)</sup> suggested that the excess of reactive oxygen species (ROS) in leukemic cells leads to the activation of the enzyme Topoisomerase II $\alpha$ , which is involved in chromatid break formation. In this experiment, they showed that the frequency of chromatid breaks was reduced with reduced expression of Topoisomerase II $\alpha$ <sup>(24, 25)</sup>.

It is possible that the occupational exposure of solvents and the transfusion dependency that leads to an iron overload and, consequently, ROS increase, may have led to the chromatid break after one-year period. As shown, chromatid break is strongly correlated with the presence of various DNA damages. We suggest that this abnormality is part of a clonal evolution of this patient and, together with the clinical manifestations, led to a poor evolution and death of the patient.

## ACKNOWLEDGEMENT

We were supported by Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES), Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq) and Fundação Cearense de Apoio ao Desenvolvimento Científico e Tecnológico (Funcap).

## RESUMO

*A síndrome mielodisplásica (SMD) é uma desordem clonal das células-tronco hematopoiéticas caracterizada por citopenias periféricas devido à hematopoiese ineficaz e pelo aumento do risco de evolução para a leucemia mieloide aguda (LMA). As alterações cromossômicas representam o marcador mais importante da estratificação de risco para a transformação de LMA. Quebra das cromátides (chb) é uma descontinuidade de uma única cromátide. Relatamos o caso de um paciente com SMD, cuja análise citogenética mostrou chb espontâneo: 46,XY,add(13)(q34),chb(15)(q24)[3]/47,XY,chb(2)(q22),del(5)(q35),del(7)(q32),+8,del(11q)(q23),del(q22)[cp17]. O paciente foi considerado de alto risco devido ao cariótipo complexo e à presença de chb. Sugerimos que essa anormalidade cromossômica possa ser considerada como marcador de instabilidade.*

*Unitermos: quebra cromossômica; síndrome mielodisplásica; instabilidade cromossômica; instabilidade genômica.*

## REFERENCES

1. Bejar R, Levine R, Ebert BL. Unraveling the molecular pathophysiology of myelodysplastic syndromes. *J Clin Oncol*. 2011; 29: 504-15.
2. Dolatshad H, Pellagatti A, Fernandez-Mercado M, et al. Disruption of SF3B1 results in deregulated expression and splicing of key genes and pathways in myelodysplastic syndrome hematopoietic stem and progenitor cells. *Leukemia*. 2015; 29(8): 1798.
3. Papaemmanuil E, Gerstung M, Malcovati L, et al. Clinical and biological implications of driver mutations in myelodysplastic syndromes. *Blood*. 2013; 122(22): 3616-27.
4. Kao J, Rosenstein BS, Peters S, Milano MT, Kron SJ. Cellular response to DNA damage. *Ann N Y Acad Sci*. 2005; 1066: 243-58.
5. Gattermann N. Pathophysiological and clinical aspects of iron chelation therapy in MDS. *Curr Pharm Des*. 2012; 18(22): 3222-34.
6. Ghoti H, Fibach E, Westerman M, Gordana O, Ganz T, Rachmilewitz EA. Increased serum hepcidin levels during treatment with deferasirox in iron-overloaded patients with myelodysplastic syndrome. *Br J Haematol*. 2011; 153(1): 118-20.
7. Saigo K, Takenokuchi M, Hiramatsu Y, et al. Oxidative stress levels in myelodysplastic syndrome patients: their relationship to serum ferritin and hemoglobin values. *J Int Med Res*. 2011; 39(5): 1941-5.
8. de Souza GF, Ribeiro Jr. HL, de Sousa JC, et al. HFE gene mutation and oxidative damage biomarkers in patients with myelodysplastic syndromes and its relation to transfusional iron overload: an observational cross-sectional study. *BMJ Open*. 2015; 5: e006048.
9. Terzoudi GI, Hatzl VI, Bakoyianni CD, Pantelias GE. Chromatin dynamics during cell cycle mediate conversion of DNA damage into chromatid breaks and affect formation of chromosomal aberrations: biological and clinical significance. *Mutat Res*. 2010; 711: 174-86.
10. Payne M, Hickson ID. Genomic instability and cancer: lessons from analysis of Bloom's syndrome. *Biochem Soc Trans*. 2009; 37(3): 553-9.
11. Taylor AM. Chromosome instability syndromes. *Best Pract Res Clin Haematol*. 2001; 14(3): 631-44.
12. Wechsler T, Newman S, West SC. Aberrant chromosome morphology in human cells defective for Holliday junction resolution. *Nature*. 2011; 471(7340): 642-6.
13. Ludwig LB, Valiati VH, Palazzo PR, et al. Chromosome instability and oxidative stress markers in patients with ataxia-telangiectasia and their parents. *Bio Med Res Int*. 2013; 2013: 762048.
14. Singh RR, Bansal D, Shah AS. Genetic study in 50 cases of carcinoma breast: a prospective study. *Int Surg J*. 2011; 3(4): 1812-5.
15. Korgaonkar S, Babu VR, Kerketta L, Ghosh K. Chromosomal breakage in myelodysplastic syndrome. *Asian Pac J Cancer Prev*. 2008; 9(1): 151-4.
16. Swerdlow SH, Campo E, Harris NL, Jaffe ES, Pileri SA, Stein H, et al. WHO Classification of Tumours of haematopoietic and lymphoid tissues. Lyon, France: IARC Press, 2008.
17. Greenberg PL, Tuechler H, Schanz J, et al. Revised International Prognostic Scoring System (IPSS-R) for myelodysplastic syndromes. *Blood*. 2012; 120: 2454-65.
18. Greenberg P, Cox C, LeBeau MM, et al. International scoring system for evaluating prognosis in myelodysplastic syndromes. *Blood*. 1997; 89(6): 2079-88.
19. Cazzola M, Porta MGD, Malcovati L. The genetic basis of myelodysplasia and its clinical relevance. *Blood*. 2013; 122(25): 4021-34.
20. Visconte C, Selleri C, Maciejewski JP, Tiu RV. Molecular pathogenesis of myelodysplastic syndromes. *Translational Med*. 2014; 8(4): 19-30.
21. Andersen MK, Christiansen DH, Pedersen Bjergaard J. Centromeric breakage and highly rearranged chromosome derivatives associated with mutations of TP53 are common in therapy MDS and AML after therapy with alkylating agents: an M-FISH study. *Genes Chromosomes Cancer*. 2005; 42(4): 358-71.
22. Vilalba-Campos M, Chuaire-Noack L, Sánchez-Corredor MG, Rondón-Lagos M. High chromosomal instability in workers occupationally exposed to solvents and paint removers. *Mol Cytogenet*. 2016; 9: 46.
23. Invernizzi R. Iron overload, oxidative damage and ineffective erythropoiesis in myelodysplastic syndromes. *Eur Haematol*. 2010; 4: 34-8.
24. Bryant PE, Riches AC, Terry SYA. Mechanism of the formation of radiation-induced chromosomal aberrations. *Mutat Res*. 2010; 701: 23-6.
25. Terry SY, Riches AC, Bryant PE. A role for topoisomerase II alpha in the formation of radiation-induced chromatid breaks. *Br J Cancer*. 2008; 99(4): 670-4.

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