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Quality of mussels cultivated and commercialized in Ubatuba, SP, Brazil - monitoring *Bacillus cereus* and *Staphylococcus aureus* growth after post-harvest processing

*Tratamento térmico de mexilhões *Perna perna* como forma de assegurar a qualidade - avaliação do crescimento de *Bacillus cereus* e de *Staphylococcus aureus**

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Abstract

Aiming at improving the quality of *Perna perna* mussels cultivated and commercialized in Ubatuba, SP, Brazil, the growth and elimination of *Staphylococcus aureus* and *Bacillus cereus* artificially inoculated in mussels were studied. The inoculation was carried out in "in natura" and pre-cooked mussels for 30 min, and after that the mussels were kept for 10 hours at room temperature ($25 \pm 1^\circ\text{C}$) and under refrigeration ($7 \pm 1^\circ\text{C}$). Six thermal treatments were evaluated: three using steam (5, 10 and 15 minutes) and three in boiling water (5, 10 and 15 minutes), in order to find the best time/temperature binomial to provide pathogenic control. Yield and physical-chemical and sensory characteristics were evaluated. All thermal treatments were efficient to eliminate microorganisms in 2 logarithmic cycles. However, the boiling water treatments presented better results than the steam treatments. The physical-chemical and sensory analyses did not show statistical differences among the thermal treatments studied. The best performances were reached in the shortest times of heat exposure. Overall, the treatments in boiling water presented better results than the steam treatments.

Keywords: microorganisms; mussel farming; processing.

Resumo

Buscando a qualidade do mexilhão *Perna perna* cultivado e comercializado no município de Ubatuba, SP, foram estudados o crescimento e o controle de *Staphylococcus aureus* e *Bacillus cereus* inoculados artificialmente em mexilhões in natura e pré-cozidos, mantidos por 10 horas à temperatura ambiente ($25 \pm 1^\circ\text{C}$) e sob refrigeração ($7 \pm 1^\circ\text{C}$). Foram estabelecidos seis tipos de tratamentos térmicos, sendo três sob vapor (5, 10 e 15 minutos) e três por imersão em água (5, 10 e 15 minutos), buscando estabelecer o binômio que proporcionasse a diminuição da contagem microbiana. Posteriormente, foram avaliados o rendimento, os aspectos físico-químicos e sensoriais. Todos os tratamentos térmicos foram eficientes, pois reduziram a contagem microbiana em pelo menos 2 ciclos logarítmicos. Porém, o tratamento térmico por imersão em água com ebulição apresentou resultados melhores que os tratamentos a vapor. As análises físico-químicas e sensoriais não mostraram diferença estatística entre os tratamentos térmicos estudados. Os melhores desempenhos foram alcançados nos menores tempos de exposição ao calor, e os tratamentos em água com ebulição apresentaram resultados mais satisfatórios que os tratamentos a vapor.

Palavras-chave: microrganismos; mitilicultura; beneficiamento.

1 Introduction

Aquaculture has been growing rapidly over the last decades, with an average 10% growth a year since 1984 (FAO, 2005). Mollusk farming accounts for 24% of the total international aquacultural production and mussels represent about 13.3% of this production (OSTRENSKY; BORGHETTI; PEDINI, 2000; STREIT et al., 2002).

In Brazil, marine aquaculture is represented basically by the farming of crustaceans and mollusks. Brazilian production of mollusks is about 6,690 tons a year, which represents 48% of all the mussels produced in Latin America (SUBASINGHE et al., 2001). According to Borghetti and Ostrensky (2000), *Perna perna* mussels, two species of oysters and one of scallops are produced in Brazil.

Mussel farming is one of the most productive activities of aquaculture, reaching up to 30 tons of meat per hectare/year, which represents the highest amount obtained with a modality that is not submitted to artificial feeding. Besides several favorable biological aspects, other factors such as the lower cost of installations, ease of handling, and localization of the culture in the sea contributed towards the international expansion of this activity (GELLI et al., 1998; PEREIRA et al., 2005).

Due to their distribution in the seashore and estuary, mollusks are constantly submitted to sewage pollution, especially near large urban centers. The presence of bacteria and pathogenic virus are the main dangers associated to pollution, which becomes a Public Health problem, affecting the reputation of the

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food and the population. The consumption of fresh or slightly-cooked mollusks makes this problem worse (RODRIGUES, 1999; RODRIGUES-ARIZA et al., 1992).

Both *B. cereus* and *S. aureus* are frequently involved in the outbreak of food-borne diseases and are consequently isolated in human beings. According to data from the United States Department of Agriculture (USDA), from 1993 to 1997, 14 outbreaks and 691 cases, and 41 outbreak and 1413 cases (one death) caused by *B. cereus* and *S. aureus*, respectively, were reported in the United States (LANCIOTTI et al., 2001).

The potential risk of developing a food-borne disease caused by *B. cereus* and *S. aureus* through the consumption of mussels cultivated in Ubatuba, SP, Brazil, has been identified (GALVÃO et al., 2006).

The storage of mollusks is limited if they are commercialized in their natural condition; the valves occupy a lot of space, making storage difficult. One of the advantages of processing this product is the possibility of presenting it in a more attractive condition than the traditional one, as well as allowing storage for longer periods, making consumption more flexible (BEIRÃO et al., 2000).

Thermal processing is the key area in mussel processing in order to obtain the required sensory properties. It also inactivates bacteria and enzymes with the objective of producing a safer and more stable food. An adequate shelf life is essential for the demand in the exporting market, and for the expansion of the domestic market (SKIPNES et al., 2002; WOOD, 1979).

Fish is consumed because of its higher nutritional value and typical and varied taste. These characteristics must be qualitatively and quantitatively present, not only in the processed food, such as in surimi-based products, but also in the original foods, such as farmed fish and mussels, which may have a milder taste than those in their wild stage (HAARD, 1992).

The main purpose of the present investigation was to verify the growth of *Bacillus cereus* and *Staphylococcus aureus* inoculated in fresh and pre-cooked mussel samples stored for a period of 10 hours at room temperature (25 ± 1 °C) and under refrigeration (7 ± 1 °C) and to establish the time-temperature binomial to control these microorganisms through six different thermal treatments, besides evaluating the effect of these treatments on the physical, chemical and sensory characteristics of this mollusk.

2 Materials and methods

2.1 Sample collection

Perna perna mussel samples were collected from a commercial farm located in Costão da Praia do Cedro, in Ubatuba, SP ($23^{\circ} 33' 45.5''$ S, $45^{\circ} 09' 32.9''$ W), from November 2003 to March 2004.

The mussels collected were submitted to cleaning, in order to remove barnacles and parasites. They were then washed with sea water, packed in raffia bags, placed in Styrofoam boxes with ice in a ratio of 1:1, with no direct contact with the ice, to avoid the death of the mussels.

2.2 Growth of *S. aureus* and *B. cereus*

Inoculum Preparation

IZ 1974 and CCT 0198 strains were used as *Staphylococcus aureus* and *Bacillus cereus* inoculums, respectively. Both were kept at room temperature in tubes with inclined nutrient agar (NA) (Merck).

After reactivation in Brain Heart Infusion broth (BHI) (Oxoid), for *S. aureus*, and nutrient broth (NB) (Merck), for *B. cereus*, for 24 hours, at 36 ± 1 °C, the growth curve was determined, where the period necessary to obtain an inoculum with 10^8 CFU g⁻¹ was determined.

Sample and inoculum preparation

The mussel shells were washed under running water with the aid of a sterile brush and distributed in paper towels to remove the washing water (VANDERZANT; SPLITTSTOESSER, 1992). The batch was separated into two sub-batches, one for analysis in the natural condition and the other for analysis of the cooked and shell-less material.

After cleaning, one of the sub-batches was pre-cooked in a water bath up to boiling temperature, for about 5 minutes, until the shells opened. Then, they were immersed in water and ice in order to cool rapidly and reach an internal temperature of 10 °C to be removed from the shells.

After a 24-hours reactivation of the cultures, 1 mL was inoculated in 100 mL BHI and incubated for 6 hours at 37 °C. Then, an aliquot of 1 mL was suspended into 10 L of a 0.7% saline solution, where 1 kg of fresh or shell-less cooked mussels was inoculated for 30 minutes. After this period, mussels were drained for 15 minutes and placed in sterile containers.

The two treatments were divided once again into three samples: one for immediate analysis; one stored under refrigeration (7 ± 1 °C) and the other stored at room temperature (25 ± 1 °C) for 10 hours.

Microbiological analysis

The fresh mussels were forced open around the byssus area with the aid of a knife, the water was drained, and the meat was completely removed and placed in a sterile beaker (VANDERZANT; SPLITTSTOESSER, 1992).

For the presumptive count of both microorganisms, we used the direct plate count method, in accordance with Vanderzant; Splittstoesser (VANDERZANT; SPLITTSTOESSER, 1992), in which 1 mL (0.3, 0.3, 0.3 and 0.1 mL) of the dilution of 10^{-3} and 0.1 mL of the dilutions of 10^{-4} and 10^{-5} were inoculated in the fresh mussels, and 1 mL (0.3, 0.3, 0.3 and 0.1 mL) of the dilution of 10^{-5} and 0.1 mL of the dilutions of 10^{-6} and 10^{-7} were inoculated in the cooked mussels.

Bacillus cereus count

Inoculums were spread in duplicates with a Drigalsky spreader on Petri dishes with Mannitol Yolk Polymyxin Agar

(MYP A) (Merck), previously added of yolk emulsion (Oxoid) and *Polymyxin* B (Oxoid), until excess liquid was absorbed.

After drying, the dishes were inverted and incubated at $36 \pm 1^\circ\text{C}$ for 20 to 24 hours. After this period, typical colonies (large, with no specific form, pink, surrounded by an opacity zone) were selected for confirmative Gram and catalase tests. Some typical colonies were inoculated in a BHI broth and incubated at $36 \pm 1^\circ\text{C}$, for 24 hours, in order to carry out other identification tests: motility, nitrate reduction, tyrosine decomposition, rhizoid growth and Voges-Proskauer (VP) tests.

Staphylococcus aureus count

Inoculums were spread in duplicates with a Drigalsky spreader on Petri dishes with Baird-Parker Agar (BPA) (Merck), previously added of yolk tellurite (Oxoid), until excess liquid was absorbed. After drying, the dishes were inverted and incubated at $36 \pm 1^\circ\text{C}$, for 48 hours. After this period, typical colonies (black, bright, circular, plain, convex, and surrounded by a transparent halo) were selected for confirmative Gram, catalase and coagulase tests.

2.3 Mussel thermal treatments

After hygienization with the aid of a sterile brush, the mussel batch was separated into 6 samples (1 kg each), which were submitted to the following cooking times: 5, 10 and 15 minutes in steam, and 5, 10 and 15 minutes in a water bath. For cooking, the mussels were placed in wire baskets (20 cm top diameter x 16.5 cm bottom diameter x 10 cm height), suspended in a lidded stainless steel container (24.5 cm diameter x 14 cm height).

The cooking temperature was monitored by a group of thermocouples with 5 calibrated sensors. They were distributed as follows: at the bottom of the recipient, in the middle, on the surface, and in the center of the mussel mass. The fifth thermocouple was placed in the cooling recipient, where the mussels were immersed in water with ice at the rate of 3:1, until their internal temperature reached 10°C .

Microbiological analysis

After cooking, the mussels were removed from their shells and placed in a sterile recipient. Microbiological analysis was carried out as described for the fresh and pre-cooked mussel analysis.

Physical-chemical analysis

Moisture, protein, lipids, carbohydrates and ash analyses were carried out according to the Association of Official Analytical Chemists (A.O.A.C., 1995).

A Digimed digital potentiometer was used to determine the pH. A sample (meat + intravalvar liquid) triturated with water, at the rate of 1:1, was measured.

Yield

Yield calculation was carried out to verify possible losses in mussel yield depending on the cooking period. Before each

treatment, the fresh samples were weighed and their shells were removed. After that, the meat and shells with residues were weighed separately. The yield was obtained according to the following formulas (Equations 1 and 2):

$$\text{Total weight (in\%)} = \frac{\text{cooked meat weight}}{\text{fresh mussel weight}} \times 100 \quad (1)$$

$$\text{Edible part (in\%)} = \frac{\text{cooked meat weight}}{(\text{fresh mussel weight} - \text{shell weight})} \times 100 \quad (2)$$

Biometry constituted in the measurement of the valves of 100 individuals from each collection with the help of a pachymeter.

Sensory analysis

An ordering test was carried out using 4 samples (5 and 10 minutes in water, and 5 and 10 minutes in steam), which were presented to panelists to be organized in an increasing order, according to their general appreciation of the product. The team was formed by 50 untrained panelists who, nevertheless, were acquainted with the product.

The results were calculated by the sum of the ordering of each sample and statistically evaluated by Friedman's test, using the data from Newell; MacFarlane's table (A.O.A.C., 1995).

2.4 Statistical Analysis

The results of the physical-chemical analyses were submitted to analysis of variance using the SAS software (SAS INSTITUTE, 1999). Means were compared by the Tukey test, adopting a significance level of 5%.

3 Results and discussion

3.1 Inoculation

Table 1 presents the results of the growth of *Staphylococcus aureus* inoculated in fresh and pre-cooked mussels kept at room temperature ($25 \pm 1^\circ\text{C}$) or under refrigeration ($7 \pm 1^\circ\text{C}$).

As can be observed in Table 1, microbial growth occurred both in fresh or pre-cooked mussels kept at room temperature or under refrigeration. The largest growth of *S. aureus* was observed in cooked mussels after 10 hours storage at room temperature, when counts of 5.3 to 6.1×10^7 CFU g^{-1} were observed, with an increase of 3 logarithmic cycles. This is an important aspect, once *S. aureus* is related to food-borne diseases. It is well known that a density of 10^6 - 10^8 CFU g^{-1} is associated to enterotoxin production in a toxic level (SILVA, 2000).

According to Ayulo, Machado and Scussel (1994), fish and mussels produced and commercialized in the state of Santa Catarina were positive for *S. aureus* in 20% of the analyzed samples and also 60% of the bivalve samples presented *S. aureus*, but only 9 out of 109 strains of *S. aureus* produced enterotoxin, including enterotoxins A (4), D (1) and B (4).

A different behavior was observed for *B. cereus* inoculated in fresh and pre-cooked mussels kept at room temperature ($25 \pm 1^\circ\text{C}$) and under refrigeration ($7 \pm 1^\circ\text{C}$) (Table 2). In fresh mussels kept at room temperature, a growth of 1 logarithmic cycle was observed 10 hours after inoculation, whereas for cooked mussels kept at room temperature ($25 \pm 1^\circ\text{C}$) a large microbial growth of 10^4 to 10^8 was observed.

Johnson, Nelson and Busta (1983) observed that in a Trypticase Soy broth, *B. cereus* growth was observed between 15 and 50°C , being optimal between 35 to 40°C . Mead et al. (1999) presented the possibility of growth at 4 to 50°C for this microorganism. This is an important data related to spore production, since from 10^7 to 10^8 cells g^{-1} of food are needed for the incidence of food-borne gastroenteritis (FORSYTHE, 2002).

Ola and Oladipo (2004) have also studied the storage period of the cassava croaker (*Pseudolithus senegalensis*) in ice and at room temperature, and observed that the total count of microorganisms increased from 10^3 to 10^6 CFU g^{-1} after 12 hours storage, resulting in rejection by consumers after sensory analysis. In this study, it was also observed that *Bacillus* was the most important contribution to the increase in counts during storage. On the

other hand, when stored under refrigeration, *Bacillus* presented a slight decrease after 20 days storage.

Rawles et al. (1995), studying the growth of *Listeria monocytogenes* in cooked blue crab meat under refrigeration, obtained 7 logarithmic cycles at the end of 21 days of storage at 5°C . This growth became more intense after the 10th day of storage. Also, Reily and Hackney (1985) observed the growth of *Vibrio cholerae* in inoculation-infected oysters kept for 21 days under refrigerated storage ($\pm 7^\circ\text{C}$) and obtained a reduction of 3 logarithmic cycles in the first 7 days of storage, which remained unaltered in the next 14 days.

In this paper, refrigeration proved to be an efficient process in controlling the growth of both microorganisms and shows the importance of refrigerating this product, mostly after thermal treatment and without the valve, when the product can be easily infected.

Inoculation was more efficient in pre-cooked mussels than in fresh mussels, which shows the importance of the shell in mussel protection against microbial infection.

According to epidemiological data, between 1986 and 1995, bacterial pathogens were referred to as the main causes of several outbreaks in Taiwan, Republic of China (555 out of 852 outbreaks, 65%), being *S. aureus* (169 out of 555 outbreaks, 30%) and *B. cereus* (104 out of 555 outbreaks, 18%) two of the main causative agents of these outbreaks (PAN et al., 1997).

Table 1. Counts of *Staphylococcus aureus* inoculated in fresh and pre-cooked mussels, after 10 hours storage at room temperature or under refrigeration.

	Initial ¹	After 10 hours storage	
		Room temperature (25 ± 1 °C)	Refrigeration (7 ± 1 °C)
Fresh mussels ¹			
Sample 1	5.0 × 10 ³	2.5 × 10 ⁵	1.0 × 10 ⁴
Sample 2	6.0 × 10 ³	4.0 × 10 ⁵	2.0 × 10 ⁴
Sample 3	3.0 × 10 ³	3.0 × 10 ⁵	1.0 × 10 ⁴
Mean	4.6 × 10 ³	3.2 × 10 ⁵	1.3 × 10 ⁴
Cooked mussels ¹			
Sample 1	5.6 × 10 ⁴	5.3 × 10 ⁷	1.7 × 10 ⁵
Sample 2	6.0 × 10 ⁴	6.1 × 10 ⁷	1.3 × 10 ⁵
Sample 3	6.4 × 10 ⁴	5.8 × 10 ⁷	1.7 × 10 ⁵
Mean	6.0 × 10 ⁴	5.7 × 10 ⁷	1.6 × 10 ⁵

¹Mean of duplicates, expressed in CFU g^{-1} of meat.

Table 2. *Bacillus cereus* inoculated in fresh and pre-cooked mussels, after 10 hours storage at room temperature and under refrigeration.

	Initial	After 10 hours storage	
		Room temperature (25 ± 1 °C)	Refrigeration (7 ± 1 °C)
Fresh mussels ¹			
Sample 1	4.0 × 10 ³	3.1 × 10 ⁴	4.0 × 10 ³
Sample 2	6.0 × 10 ³	3.0 × 10 ⁴	5.0 × 10 ³
Sample 3	5.0 × 10 ³	3.1 × 10 ⁴	6.0 × 10 ³
Mean	5.0 × 10 ³	3.1 × 10 ⁴	5.0 × 10 ³
Cooked mussels ¹			
Sample 1	2.0 × 10 ⁴	6.6 × 10 ⁸	5.3 × 10 ⁴
Sample 2	2.1 × 10 ⁴	4.5 × 10 ⁸	4.3 × 10 ⁴
Sample 3	1.8 × 10 ⁴	7.0 × 10 ⁸	5.6 × 10 ⁴
Mean	1.9 × 10 ⁴	6.0 × 10 ⁸	5.0 × 10 ⁴

¹Mean of duplicates, expressed in CFU g^{-1} of meat.

3.2 Thermal treatment

Figure 1 shows the average temperatures reached at the end of the different cooking periods.

All thermal treatments reached temperatures above 60°C , the differences registered in temperature ranges among the thermocouples in the same system did not exceed 10°C , except in steam cooking for 5 minutes. This suggests a uniform distribution of the heat in the conditions of the experiment. In thermal treatment studies carried out by Antonioli (1999), there were no variations in heat distribution, as observed in our study.

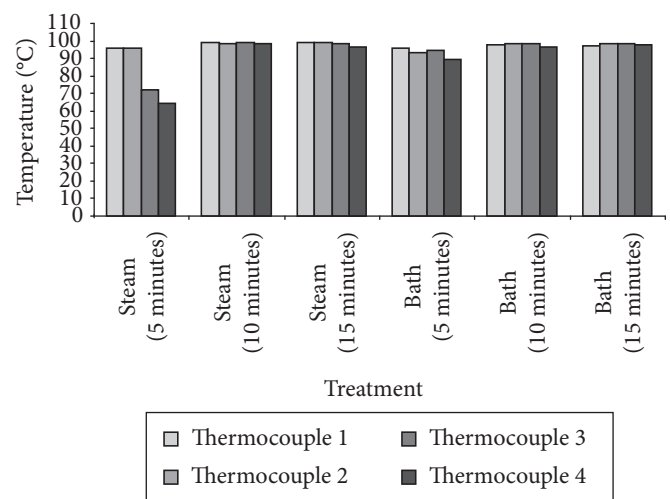


Figure 1. Average temperatures measured by the thermocouples after different cooking periods.

According to Pigott and Tucker (1990), pathogenic microorganisms are destroyed at temperatures under 100 °C, and the effect of increasing the internal food temperature to 66 °C is enough to make the food safe.

This evidence can be observed in Figure 2, where the values of the Most Probable Number (MPN) for *Staphylococcus aureus* and *Bacillus cereus* in all treatments applied are presented. Both microorganisms presented similar behavior, and were reduced by at least 2 logarithmic cycles in all treatments. It is evident that the thermal treatments applied were efficient to decrease the microorganisms inoculated considerably. It was also noticed that treatments with boiling water were more efficient in microbial reduction, if compared to treatments with steam.

Antoniolli (1999), studying the shelf life of processed and refrigerated *Perna perna* mussels, used different cooking periods, from 15 to 30 minutes, obtaining final temperatures vary-

ing from 80 to 96 °C, and observed that all thermal treatments applied were satisfactory to decrease microorganism counts. Also, Chai et al. (1991) studied the thermal processing of oysters and suggested the treatment at 75-76 °C, for 8 minutes, aiming a product of better quality.

Another means of reducing microorganisms was studied by Juneia, Marmer and Eblen (1999), who showed that the addition of NaCl protects the pathogenic agent, in this case, *E. coli* O157:H7, against the lethal effect of heat. Kotrola and Conner (2002) showed that the survival of *E. coli* in poultry meat was significantly increased (longer period of decimal reduction) by the use of additive elements, such as sodium lactate, sodium chloride and polyphosphates, in the preparation of turkey meat. In all cases, the percentage of water was reduced, as well as the water mobility in the environment.

Ama, Hamdy and Toledo (1994) studied the effects of temperature and pH in the inactivation of *V. vulnificus* and observed that for higher initial counts of microorganisms ($\sim 10^7$ per mL or g), the thermal treatments are not effective. This evidence is important because, despite the fact that thermal treatment is effective in eliminating microorganisms, it cannot be used for this purpose when the raw material is of poor quality.

3.3 Centesimal composition and pH

The results obtained for the centesimal composition and pH of fresh and cooked mussels (different thermal treatments) are presented in Table 3.

Mean values of 82.82; 11.78; 1.55, and 2.37 g 100 g⁻¹ for moisture, protein, lipids and ash, respectively, were obtained for the fresh mussels. These data are in accordance with several food composition tables, such as that from the Santa Catarina Aquaculture Association (ACAQ, 2005), which presents values of 83.00; 12.00; 1.5; 1.9 and 3.00 g 100 g⁻¹, respectively, for the same shellfish studied in this research, and the same parameters; and Franco's table (2004), which presents values of 14.40 and 2.30 g 100 g⁻¹, respectively, for protein and lipids.

Pedrosa and Cozzolino (2001) evaluated the centesimal composition of fresh mussels (*Anomalocardia brasiliensis*), and found values of 81.8; 12.7; 1.0 and 2.2 g 100 g⁻¹ for moisture, protein, lipids and ash, respectively. Also, Bindu et al. (2002) observed the nutritional composition of fresh *Perna viridis* mus-

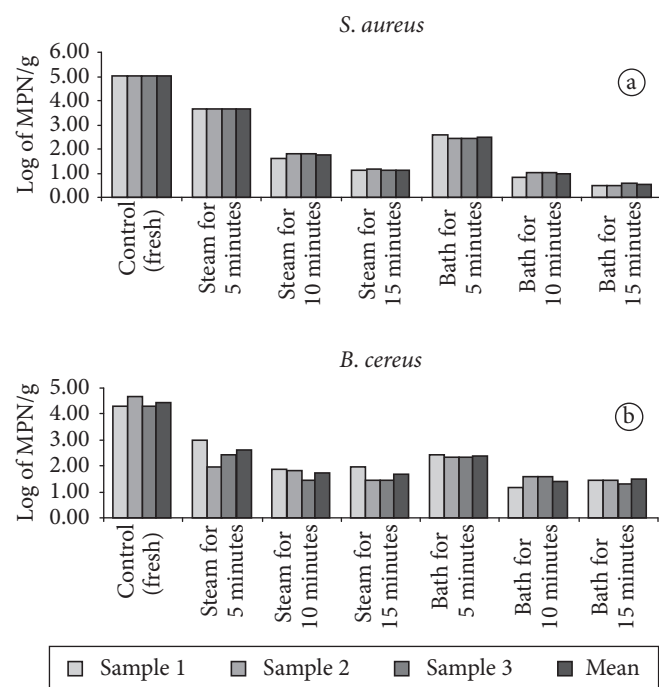


Figure 2. *Staphylococcus aureus* and *Bacillus cereus* inoculated in mussel samples and submitted to different thermal treatments.

Table 3. Centesimal composition and pH of the mussels submitted to different thermal treatments.

Thermal Treatment ^{1,2}	Moisture	Protein	Lipids	Ash	Carbohydrates	pH
	g 100 g ⁻¹					
Control (Fresh)	82.82 ^{a3}	11.78 ^a	1.55 ^a	2.37 ^a	1.98 ^a	6.46 ^a
Steam						
5 minutes	79.85 ^a	15.01 ^a	1.79 ^a	2.03 ^a	1.32 ^a	6.87 ^a
10 minutes	75.57 ^a	17.52 ^a	2.33 ^a	2.05 ^a	2.53 ^a	6.97 ^a
15 minutes	75.13 ^a	17.34 ^a	2.57 ^a	2.01 ^a	2.95 ^a	7.07 ^a
Water bath up to boiling point						
5 minutes	80.03 ^a	14.73 ^a	2.03 ^a	1.49 ^a	1.72 ^a	6.92 ^a
10 minutes	77.11 ^a	16.80 ^a	2.61 ^a	1.61 ^a	1.87 ^a	7.08 ^a
15 minutes	76.23 ^a	16.59 ^a	2.81 ^a	1.56 ^a	2.81 ^a	7.11 ^a

¹Mean of triplicates; ²period measured after boiling point; ³means followed by the same letters in columns are not statistically different (p < 0.05).

sel and found values of 76.9 g 100 g⁻¹ for moisture, 2.57 g 100 g⁻¹ for lipids, 12.55 g 100 g⁻¹ for protein, and 2.06 g 100 g⁻¹ for ash. These values are close to the ones found in this study.

Different values from those found in this research were presented by Tavares et al. (1998), who evaluated the variation of the composition of *Perna perna* mussels along the year in Ubatuba/SP and obtained lower values for moisture (72.12 g 100 g⁻¹), and higher values for protein (20.5 g 100 g⁻¹) and lipids (3.24 g 100 g⁻¹).

The nutritional composition of green crab claws (*Carcinus maenus*) was studied by Skonberg and Perkins (2002). No differences were observed in the centesimal composition of the raw or cooked claws after twelve minutes in steam, as observed in our research for mussels.

Gokoglu, Yerlikaya and Cengiz (2004), studying the effects of different cooking methods on the centesimal composition and the mineral content of rainbow trout (*Oncorhynchus mykiss*), observed that trout cooked in boiling water for 5 minutes, did not differ significantly in their protein and fat content, but presented reduced values for moisture and ash.

Moisture contents varied from 75.13 to 82.82 g 100 g⁻¹, and the more intense the thermal treatment, the lower the moisture values. Similar values were found in Schramm's (1993) research, who obtained moisture contents between 75 and 82 g 100 g⁻¹ for mussels pre-cooked in boiling water for 5 minutes, and Oetterer (2003), who obtained values between 78.30 and 89.77 g 100 g⁻¹ for fresh mussels.

The protein contents found in this work ranged from 11.78 to 17.34 g 100 g⁻¹ and were directly related to the thermal treatment applied. Higher protein contents were observed in the treatments with a longer period of exposure to high temperatures and in the steam treatments. Our data were higher than those presented by Gelli (1992), from 6.7 to 9.5 g 100 g⁻¹.

For lipids, the values obtained ranged from 1.55 g 100 g⁻¹ for fresh mussels to 2.81 g 100 g⁻¹ for those submitted to 15 minutes in boiling water. In a study by Ackman (1999), shellfish lipids content ranged from 1 to 2 g 100 g⁻¹, and this reduced lipid content was related to the storage of energy in the glycogenic form by bivalves.

The pH values found in this paper ranged from 6.46 to 7.11. An increase in pH was observed, regardless of the cooking treatment. These data were similar to those found by Oetterer (2003) for *Perna perna* mussels collected in Ubatuba, which, according to the author, ranged from 5.8 to 6.9 in fresh mussels.

3.4 Yield

The values obtained for the yield of the edible part and of the total weight of mussels submitted to different thermal treatments are presented in Figure 3.

The yield determination was carried out with the objective of verifying possible losses in mussel yield due to the cooking period. Two forms of determination were adopted: one emphasizing the total weight and the other considering only the edible part of the mussels.

The yield varied from 45.72 to 69.24% with the treatment. Higher yields were obtained with shorter cooking times. Although differences were observed, all treatments had a high total weight yield, according to the Commercial Condition Index proposed by Gomes, Marenzi and Manzoni (1998), in which average and maximum yields are respectively 21.64 and 40.32%.

In studies carried out by Beirão (2002), yield values of about 11% were found for the ratio shell/meat. These values cannot be compared because the evaluation occurred using different methods, once the author evaluated only the tissue weight.

3.5 Sensory Analysis

Table 4 shows the differences in the sum of the ordering of each sample, being statistically evaluated by Friedman's test using data from Newell and MacFarlane's (1987) table.

The critical value for the significance level of 5%, according to Newell and MacFairlane's (1987) table is 34.

The results obtained in this paper show that even with longer thermal treatment periods, sensory losses did not take place, and the best treatment in the panelists' opinion was the 10-minutes treatment in boiling water. The use of boiling water or steam did not influence the final product, resulting in good-quality products in both treatments.

In studies carried out by Cambero et al. (1998), who observed the effects of different cooking methods on the aromatic compounds of shrimp (*Parapenaeus longirostris*) broth, the best shrimp extract was obtained with the treatment at 55 °C for 120 minutes and, the higher the temperature applied, the

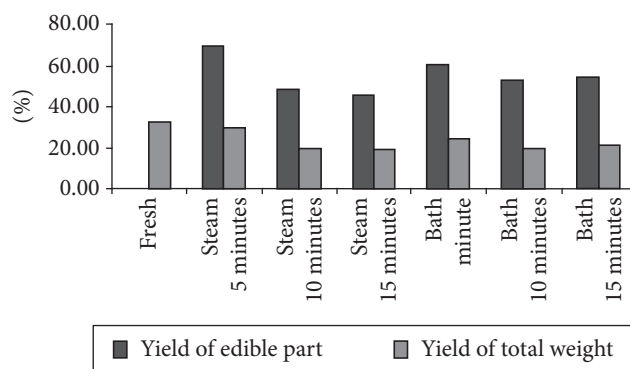


Figure 3. Yield of edible part and of total weight (%) of mussels submitted to different thermal treatments.

Table 4. Differences in the sum of the ordering of each sample submitted to the ordering test.

Thermal treatments	Samples			
	Steam		Water bath	
	5 minutes	10 minutes	5 minutes	10 minutes
Steam 5 minutes	-	0	1	9
Steam 10 minutes	-	-	1	9
Water bath 5 minutes	-	-	-	10
Water bath 10 minutes	-	-	-	-

shorter the times needed to obtain the best extracts, such as, for example, 95 °C for 15 and 30 minutes.

Antoniolli (1999), studying the shelf life of processed and refrigerated *Perna perna* mussels, used different cooking times, ranging from 15 to 30 minutes, and observed that a 30-minutes cooking of the mussels with a final temperature of 96 °C resulted in the best sensory characteristics and improvement of the mussel shells removal process.

4. Conclusions

The present investigation allowed us to conclude that a microbial growth of *S. aureus* and *B. cereus* took place both in fresh and cooked mussels, along the 10 hours of observation. The mussels kept at room temperature (25 ± 1 °C) had a higher growth compared to those stored under refrigeration (7 ± 1 °C), a fact that emphasizes the need of refrigeration of artisan mussels.

All thermal treatments applied were efficient, reducing both microorganisms by at least 2 logarithmic cycles. The boiling water bath treatments were more efficient than the steam treatments.

In the physical-chemical and sensory analysis, variations were not observed among any of the thermal treatments applied. The yield was higher for shorter periods of exposure to heat, and higher in water bath treatments than in steam treatments.

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