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Restructuring Passiflora cincinnata fruit pulp: influence of hydrocolloids

Estruturação de polpa de Passiflora cincinnata: influência de hidrocoloides

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Abstract

This study evaluated the effect of mixtures of alginate, low methoxy pectin and gelatin on characteristics of *P. cincinnata* fruit gels, containing pulp with high soluble solids content (50 °Brix). The results of a central composite design showed that the models obtained, except for water activity and pH, were predictive. Gelatin was an important factor affecting firmness and colour parameters since higher concentrations of this hydrocolloid, combined with alginate concentrations greater than 1.3% and pectin quantity up to 1.26%, could be used to obtain clear yellow products with firmness greater that 1.2 kg.

Keywords: alginate; fruits; gelatin; pectin.

Resumo

Este trabalho avaliou o efeito de misturas de alginato, pectina de baixa metoxilação e gelatina nas características de géis de *P. cincinnata*, contendo polpa com alto teor de sólidos solúveis (50 °Brix). Os resultados do planejamento experimental central composto mostraram que os modelos obtidos, exceto para a atividade de água e pH, foram preditivos. A gelatina foi o mais importante fator de influência na firmeza e nos parâmetros de cor, sendo que altas concentrações desse hidrocoloide, combinadas com concentrações de alginato e pectina superiores a 1,3% e 1,26%, respectivamente, podem ser utilizadas para obter um produto amarelo claro com firmeza superior a 1,2 kg. *Palavras-chave:* alginato; frutas; gelatina; pectina.

1 Introduction

The botanic genus *Passiflora* consists of approximately 400 species, among which over 150 are native to Brazil (BRUCKNER; PICANÇO, 2001). Although most studies on the exploitation of this genus have dealt with the *P. edulis* species, there are other species with great agronomic potential, such as *P. cincinnata*, which is distributed spontaneously in the semi-arid region of northeast Brazil (CERVI, 1997; NUNES; QUEIROZ, 2001).

The fruit production still comes from extractivism and from areas cultivated in a domestic scale, but it has been increasingly growing with the production of homemade jelly and jams by a local agroindustry (Araújo et al., 2006).

Gelled sweet products are mainly represented by traditional jams and jellies gel specially with high methoxy pectins. However, other products such as fruit bars, desserts, snacks, and restructured fruit are also formulated with fruit, sugar, and combinations of one or more polysaccharides to get the desired texture, along with other functional and sensory characteristics (GLICKSMAN, 1982; TENN, 1985; OLIVIER; GUIGOU; BOUILLETTE, 1988; GRIZOTTO et al., 2005).

The production of restructured fruit with high contents of fruit pulp using hydrocolloids as binding agents could open up a new market alternative for the semi-arid native fruit and would enlarge the already existing market resulting in higher added value, and in products that can be used in many food formulations, such as in dairy and baking industry.

The objective of this work was to evaluate the effects of mixtures of alginate, pectin, and gelatin on the characteristics of fruit gels containing pulp with an elevated soluble solids level (50 °Brix).

2 Materials and methods

2.1 Plant material

The native fruit (*P. cincinnata* Mast.) was obtained from the Embrapa Semi-Arid Experimental Field at Petrolina, Pernambuco State (Brazil). The fruits were washed, cleaned, and the pulp was extracted with the seeds being separated using a home grinder, packed in 1 kg plastic bags and immediately frozen at –18 °C.

2.2 Restructuring process

The technological co-adjutants to prepare the restructured *P. cincinnata* fruit were commercial sugar (Usina União,

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160

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Primavera, Brazil), sodium alginate (Vetec Química Fina, Rio de Janeiro, Brazil), low methoxy pectin (CP Kelco, Limeira, Brazil), and 180 Bloom gelatin (Rebière Gelatinas, Amparo, Brazil) such as food grade hydrocolloids, glycerol ($C_3H_5(OH)_3$) (Vetec Química Fina, Rio de Janeiro, Brazil), as the solute to suppress the water activity, and anhydrous calcium hydrogen phosphate (CaHPO $_4$) (Vetec Química Fina, Rio de Janeiro, Brazil, analytical grade), as the source of calcium.

The proportions of hydrocolloids used were in accordance to an experimental design, as presented in Table 1.

Initially, glycerol was added to the fruit pulp at a rate of 10 g.100g⁻¹ (or 10% of the pulp weight) and, based on the soluble solids content, the amount of sugar required to reach 50 °Brix was calculated. This mixture, previously heated to 60 °C, was transferred to a plastic beaker and the dry mixture of hydrocolloids (alginate+pectin+gelatin) plus sugar were added by mixing using a laboratory mixer (Nova Técnica, NT 137, Piracicaba, Brazil). After mixing for 10 minutes, 2 g of CaHPO₄ suspended in 5 mL of distilled water was added and mixed for 5 more minutes. With the aid of 5 cm-diameter Petri dishes (depth 1 cm), the restructured fruits were molded into the shape of a solid cylinder and maintained under refrigeration at 10 °C for 24 hours to complete the gelling process.

2.3 Physicochemical analysis

The samples of the fruit pulp were submitted to the following physicochemical analysis, according to AOAC (2000): moisture (wet basis), pH, total soluble solids (°Brix), total titrable acidity (expressed as citric acid, g.100 g⁻¹), and water activity. For the restructured fruit samples, it was determined: water activity, pH, firmness, and color. For total soluble solids and pH determinations, a bench digital

refractometer (Cambridge Instruments Inc., Reichert-Jung Abbe Mark II, Buffalo, USA) and a potentiometer (Tecnal, Tec-3MP, Piracicaba, Brazil) were used, respectively. Firmness was measured using a fruit penetrometer (Effegi, FT 327, Milan, Italy) and water activity using a portable hygrometer (Decagon, PawKit, Pullman, USA). Color parameter (L*, a* and b*) measurements were carried out using a portable colorimeter (ColorTec, PCM, Clinton, USA).

2.4 Experimental design and statistical analysis

The restructuring process for *P. cincinnata* was optimized using the Response Surface Methodology (RSM), as described by Rodrigues and Iemma (2005). The proportions of alginate, pectin and gelatin, along with their codified and real values and following a central composite design, composed of a 2^3 factorial design with a central point (0) and axial points (± 1.68 , which correspond to the star design), are shown in Table 1. The central point was repeated three times to provide an estimate of the experimental error, and a total of 17 trials were performed. The following polynomial model was fitted to the data (Equation 1):

$$y = \beta_0 + \beta_1 x_1 + \beta_2 x_2 + \beta_3 x_3 + \beta_{11} x_1^2 + \beta_{22} x_2^2 + \beta_{13} x_1^2 + \beta_{12} x_1 x_2 + \beta_{13} x_1 x_3 + \beta_{23} x_2 x_3$$
(1)

where β_n are constant regression coefficients; y is the response or dependent variable (water activity, pH, firmness, L*, a* and b*); and x_1 , x_2 , and x_3 are the independent variables (alginate, pectin and gelatin concentrations).

The statistically significant model parameters were determined, as well as the significance of the model regression investigation by the analysis of variance (ANOVA) and the

Table 1. Results for the central composite design trials for soluble water activity (a_w) , pH, firmness (F), and colour parameters (L^*, a^*, b^*) of the restructured fruit with different concentrations of alginate (A), pectin (P), and gelatin (G)

Trial	A (%)	P (%)	G (%)	a _w	pН	F (g)	L*	a*	b*
1	0.50 (-1)	0.80 (-1)	10.00 (-1)	0.83	3.48	199.58	27.52	1.77	0.66
2	1.5 0(+1)	0.80 (-1)	10.00 (-1)	0.82	3.52	163.29	28.63	1.61	1.01
3	0.50(-1)	2.20 (+1)	10.00 (-1)	0.82	3.51	181.44	28.38	1.55	1.26
4	1.50 (+1)	2.20 (+1)	10.00 (-1)	0.82	3.55	190.51	28.91	1.13	1.93
5	0.50 (-1)	0.80 (-1)	20.00 (+1)	0.81	3.75	839.15	27.73	0.15	1.02
6	1.50 (+1)	0.80 (-1)	20.00 (+1)	0.80	3.77	1834.36	27.99	0.97	3.10
7	0.50 (-1)	2.20 (+1)	20.00 (+1)	0.80	3.73	907.19	29.66	1.51	2.02
8	1.50 (+1)	2.20 (+1)	20.00 (+1)	0.82	3.79	721.21	29.62	1.46	2.70
9	1.00(0)	1.50(0)	15.00(0)	0.81	3.700	725.75	24.92	0.17	0.70
10	1.00(0)	1.50(0)	15.00(0)	0.81	3.68	725.75	25.07	0.14	0.72
11	1.00(0)	1.50(0)	15.00(0)	0.79	3.61	734.82	24.27	0.18	0.69
12	0.16 (-1.68)	1.50(0)	15.00(0)	0.83	3.67	508.02	24.42	0.56	-1.88
13	1.84 (+1.68)	1.50(0)	15.00(0)	0.81	3.56	526.16	24.54	0.69	-0.85
14	1.00(0)	0.32 (-1.68)	15.00(0)	0.82	3.64	616.88	26.04	-1.00	0.38
15	1.00(0)	2.68 (+1.68)	15.00(0)	0.82	3.68	607.81	30.40	-0.98	0.65
16	1.00(0)	1.50(0)	6.60 (-1.68)	0.82	3.43	99.79	30.13	2.10	3.14
17	1.00(0)	1.50(0)	23.4 (+1.68)	0.79	3.47	925.33	44.53	1.90	3.64

Alginate, pectin, and gelatin are expressed in % (grams per 100 grams of passion fruit pulp). Values in parenthesis are the coded levels of these variables.

generation of three-dimensional graphs using the Statistic 5.0 software (STATSOFT, 1995).

3 Results and discussion

P. cincinnata pulp presented the following physicochemical characteristics: moisture = 84.47%; total titrable acidity = $3.34\,\mathrm{g.100\,g^{-1}}$; total soluble solids = $11.5\,^{\circ}$ Brix; pH = 3.1; and water activity = 0.86. The acidity was low in comparison to the value obtained by Araújo et al. (2002) for this same fruit specie and to the value obtained by Laboissière et al. (2007) for yellow passion fruit pulp, whereas the pH was similar to the values found by these authors. The soluble solids content was similar to that reported by Araújo et al. (2002), but lower than the value reported by Laboissière et al. (2007).

After the addition of glycerol to the pulp (10 g.100 g⁻¹ of fruit pulp), the total soluble solids content of the mixture was 17.5 °Brix requiring the addition of 71.5 g of sucrose per 100 g of pulp in order to reach 50 °Brix. The quantity of sugar used in this study was similar to the quantity needed to restructure papaya (GRIZOTTO et al., 2005) and mango pulps (MOUQUET; DUMAS; GUILBERT, 1992).

Table 1 presents the results for water activity (a_w) , pH, firmness (F), and colour parameters (L*, a*, b*) of the restructured *P. cincinnata* obtained from the central composite design trials. The modeling and the statistical analysis of each of these response values are discussed bellow.

3.1 Water activity

The obtained model did not show a significant regression (95% confidence level), indicating that it was not capable of describing the variations in the water activity (a_w). Both the standard error calculation and ANOVA showed that there were no significant terms implying that the hydrocolloids concentrations have no influence on the a_w of the structured fruit. The mean and the standard deviation (SD) for trials 1-8 and 12-17 were 0.82 ± 0.01 , whereas for the three-fold replicate center point was 0.80 ± 0.01 . The SD for the replicate points is the same as that for trials 1-8 and 12-17, for which the factors level varied extensively, hence the SD for the factorial and star design can simply be explained by the experimental error.

The observed $a_{\rm w}$ values were within the intermediate moisture range of 0.65 to 0.90, as reported by Chirife and Buera (1994).

3.2 pH

In contrast to the $a_{_W}$ data, the pH model showed a significant regression, but the multiple correlation coefficient of 0.64 was not satisfactory. The statistical analysis showed that only the linear and quadratic terms of gelatin were significant (p \leq 0.05). The average and SDs, 3.61 \pm 0.12 for trials 1-8 and 12-17, and 3.66 \pm 0.04 for the three-fold replicate center point showed much larger variations in pH when the factor levels were varied than for the replicate trials.

The result obtained in experiment 6 (Table 1) showed that the structured fruit with the highest firmness value (1.8 kg) presented pH value around 3.77 and were obtained using high quantities of alginate and gelatin. Thus, the 3.7 pH value can be taken as a minimum value for structuring acid pulps when the mixtures of these hydrocolloids are used.

3.3 Firmness

The statistical analysis at a 95% confidence level revealed that all terms (linear, quadratic, and cross terms) were significant for this response. The significance of the interactions indicates the existence of a synergic effect between hydrocolloids, as pointed out by Grizotto et al. (2007) and Mancini and McHugh (2000). The fitted model is described by Equation 2.

$$F(g) = -1961.73 + 533.69A - 216.73A^{2} + 985.65P - 41.67P^{2}$$

$$+154.21G - 2.23G^{2} - 405.65AP + 41.82AG - 37.65PG$$
(2)

(where F is firmness, g; A is alginate concentration, % or g.100 g $^{-1}$; P is pectin concentration, % or g.100 g $^{-1}$; and G is gelatin concentration, % or g.100 g $^{-1}$) had a significant regression and explained 84.63% of the variability in firmness.

The generated surfaces are shown in Figure 1. It can be seen that the firmness of the restructured fruit pulp was most influenced by gelatin, and as its concentration is increased, the firmness values increase. According to Grizotto et al. (2007), gels presented a good firmness when a value around 1.5 kg was obtained for restructured concentrated pineapple pulp, whereas a firmness around 1.3 kg was considered satisfactory for concentrated papaya pulp (GRIZOTTO et al., 2005). For the *P. cincinnata* pulp, higher values of gelatin (>19%) combined with values of alginate greater than 1.3% (Figure 1a) and pectin values up to 1.26% (Figure 1b) could be used in order to achieve higher firmness (>1.2 kg).

3.4 Colour parameters

The Hunter colour parameters L*, a* and b* have been widely used to describe color changes during the processing of fruit and vegetable products. The colour variables have been related to the types and quantities of some components present in foods (AMENY; WILSON, 1997; SASS-KISS et al., 2005). The major coloured compounds of *P. cincinnata* are carotenoids and the fruit pulp were initially yellowish-green (a* = -2.79; b* = 16.96).

L* values represents the lightness index of a product. Like pectin, for the *P. cincinnata* restructured fruit, gelatin showed a significant effect both in linear and quadratic terms. Alginate and the variables' interactions with each other did not show significant effect. The fitted model is described by Equation 3, which had a significant regression and explained 79.42% of the variability in L*.

$$L^* = 59.01 - 4.70P + 1.99P^2 - 4.67G + 0.17G^2$$
 (3)

The generated surfaces for this response are presented in Figure 2. The quadratic effect was more pronounced for the gelatin concentration, and some values of this parameter can be observed, where minimum L* values can be observed. For example, lower L* values, which indicates darker samples,

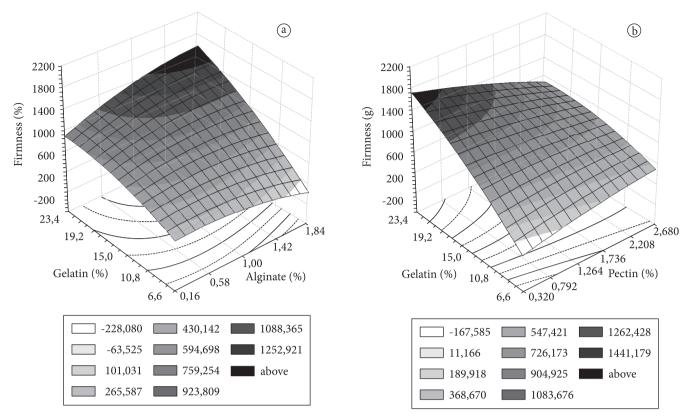


Figure 1. Response surfaces for firmness at: a) P = 1.5%; b) A = 1.0%.

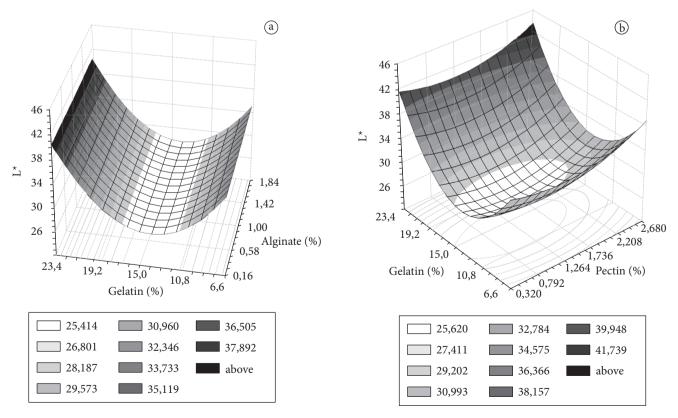


Figure 2. Response surfaces for L* at: a) P = 1.5%; b) A = 1.0%.

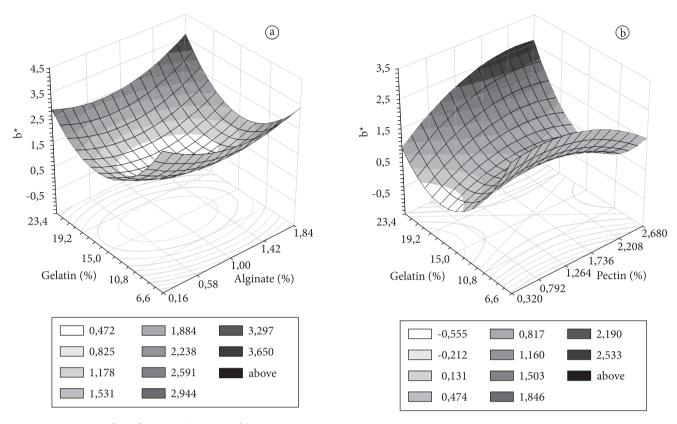


Figure 3. Response surfaces for a^* at: a) P = 1.5%; b) A = 1.0%.

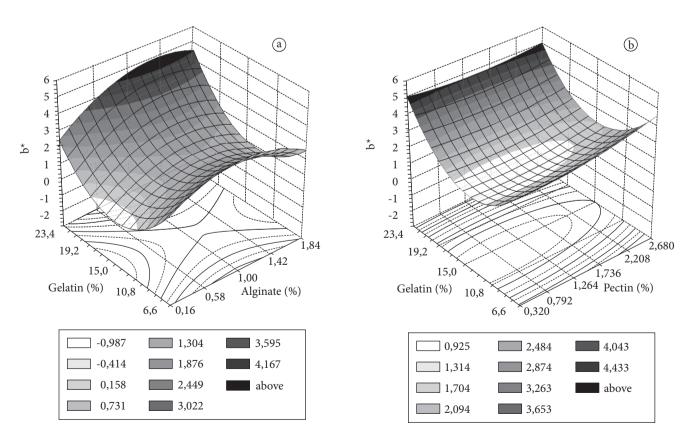


Figure 4. Response surfaces for b^* at: a) P = 1.5%; b) A = 1.0%.

can be obtained when gelatin concentration ranged from 10.8 up to 17%, combined to pectin concentrations up to 1.9% (Figure 2b), independent of the quantity of alginate used (Figure 2a). On the other hand, L* values similar to those of the fresh pulp (L* = 35.38) could be obtained using higher gelatin concentration (\geq 20%), independent of the quantity of pectin and alginate.

The a* values represent greenness and redness of a product. The statistical analysis for such response indicated that all the terms were significant at a 95% confidence level. The fitted model was described by Equation 4, which had a significant regression and explained more than 87% of the variability in a^* .

$$a^* = 10.21 - 2.90A + 1.28A^2 + 0.71P - 0.52P^2 - 1.21G + 0.03G^2 - 0.40AP + 0.07AG + 0.09PG$$
(4)

The generated surfaces for this response are shown in Figure 3. It can be observed that when intermediate concentrations of both gelatin and alginate were used (Figure 3a), the lowest values of a^* were obtained and the product were closer to the fresh pulp colour. On the other hand, higher gelatin concentration (\geq 20%) combined with pectin concentrations higher than 0.5% or lower gelatin concentrations (up to 10%), independent of the pectin concentration used, gave higher a^* values (Figure 3b).

The b* values represent yellowness and blueness of a product. As for the a* response, the statistical analysis indicated that all the terms were significant at a 95% confidence level. The fitted model was described by Equation 5, which had a significant regression and explained more than 89% of the variability in b*.

$$b^* = 7.04 + 4.42A - 2.17A^2 + 0.40P + 0.25P^2 - 1.34G$$

+ $0.05G^2 - 0.39AP + 0.09AG + 0.03PG$ (5)

The generated surfaces as a function of the hydrocolloids are shown in Figure 3. Alginate concentration greater than 0.3%, combined with lower (\leq 8%) or greater (\geq 20%) gelatin concentrations gave higher b* values (Figure 4a), therefore the restructured fruit color could be closer to color of the fresh pulp. On the other hand, gelatin concentrations ranging from 11 to 16% and pectin concentrations up to 1.9% resulted in lower b* values (Figure 4b).

The type and concentration of the hydrocolloids induced changes in colour, and thus it can be observed that their addition and the processing of the pulp resulted in color changes (a* values increased, b* values decreased, whereas the L* values either increased or decreased). Studies on to the colour of restructured fruits and the use of the three hydrocolloids (alginate, pectin and gelatin) were not available in the literature, but Gill et al. (2004) reported, for mango pulp texturized using sodium alginate, that the L* values decreased with the increase of this hydrocolloid concentration from 0 to 1%, whereas a* and b* values progressively increased with the increase of alginate. However, comparisons between this study with the study carried out by Gill et al. (2004) is not adequate since the native fruit was restructured with mixtures of hydrocolloids and the synergism between them has to be considered.

4 Conclusions

The empirical models obtained for a_w and pH were not considered predictive, but it was observed that the samples a_w values ranged within the intermediate moisture range, and that a pH around 3.7 can be taken as a minimum value for structuring the fruit pulp, when mixtures of alginate, low methoxy pectin, and gelatin are used. For firmness and colour parameters, the models were predictive. Restructured fruits with greater firmness and clear yellow color could be obtained by employing higher values of gelatin, combined to alginate concentrations higher than 1.3% and pectin quantities up to 1.26%.

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