



Ciência e Tecnologia de Alimentos

ISSN: 0101-2061

revista@sbcta.org.br

Sociedade Brasileira de Ciência e  
Tecnologia de Alimentos  
Brasil

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Ciência e Tecnologia de Alimentos, vol. 31, núm. 3, julio-septiembre, 2011, pp. 714-722  
Sociedade Brasileira de Ciência e Tecnologia de Alimentos  
Campinas, Brasil

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# Moisture sorption isotherms of fresh and blanched pumpkin (*Cucurbita moschata*)

*Isotermas de sorção de umidade de abóbora (Cucurbita moschata) fresca e branqueada*

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## Abstract

Moisture desorption isotherms of fresh and heat blanched pumpkins (*Cucurbita moschata*) were determined at three temperatures (30, 50 and 70 °C), using the standard, static-gravimetric method. The GAB, Oswin, BET, Halsey, and Henderson models were tested and, with the exception of the Henderson model, showed satisfactory fits to the experimental data. The GAB model was used to analyze the fitting ability to describe the isotherm type. The shape of the desorption isotherms of fresh and blanched pumpkin at 30 and 50 °C was intermediate to types II and III, and at 70 °C it was of type II for the blanched pumpkin and close to type II for the fresh sample. The influence of blanching on the decrease in equilibrium moisture was very small compared to the fresh samples and it was related to the loss of soluble solids during the pre-treatment. The isosteric heat of sorption measures indicated that a larger amount of heat was required to remove the water from the fresh samples than from the blanched ones.

**Keywords:** blanching; pumpkin; desorption isotherms; isosteric heat; GAB model.

## Resumo

Isotermas de sorção de umidade de abóboras frescas e branqueadas termicamente foram determinadas em três temperaturas (30, 50 e 70 °C), utilizando o método estático gravimétrico padrão. Os modelos de GAB, Oswin, BET, Halsey e Henderson foram testados e, com exceção do modelo de Henderson, mostraram ajuste satisfatório aos dados experimentais. O modelo de GAB foi usado para analisar a habilidade do ajuste na descrição do tipo de isoterma. A forma das isotermas de dessorção de abóbora fresca e branqueada determinadas a 30 e 50 °C foi intermediária aos tipos II e III e a 70 °C foi do tipo II para abóbora branqueada e próxima ao tipo II para abóbora fresca. A influência do branqueamento sobre a redução da umidade de equilíbrio das amostras foi muito pequena em comparação com amostras frescas e foi relacionada à perda de sólidos solúveis durante o pré-tratamento. Os valores de calor isostérico de sorção para tecido de abóbora indicaram que maior calor foi requerido para remover água das amostras frescas do que das amostras branqueadas.

**Palavras-chave:** branqueamento; abóbora; isotermas de dessorção; calor isostérico; modelo de GAB.

## 1 Introduction

Pumpkins, which are widely cultivated in Brazil, are a good source of carotenoids, mainly  $\alpha$ -carotene and  $\beta$ -carotene, components that present pro-vitamin A activity (RODRIGUEZ-AMAYA; KIMURA; AMAYA-FARFAN, 2008). The consumption of carotenoid-rich foods has been associated with the prevention of cancer, macular degeneration and other degenerative diseases, mostly related to their antioxidant and free-radical scavenging properties (KRINSKY, 1993).

Drying is one of the most widely used primary methods of food preservation, aiming to reduce the water content to a level at which microbial and deterioration reactions are greatly minimized (DOYMAZ, 2007; AKPINAR; BICER, 2005). Drying also contributes to a reduction in production losses and to an extension in storage life, thus making seasonal foods available throughout the year. However, physical, chemical and biochemical transformations occurring during air-drying represent one of the main problems that may lead to product quality depreciation (MUJUMDAR, 1997), since the maximum

temperatures used in food drying are generally not high enough to inactivate the enzymes. The application of heat blanching to fruits and vegetables before air-drying is, therefore, aimed at stopping enzymatic activity and avoiding undesirable changes to the sensory and nutritional properties during drying and storage, enhancing product quality (CHANTARO; DEVAHASTIN; CHIEWCHAN, 2008; WOLFE; LUI, 2003).

Drying processes and dryer design require knowledge of the relationship between water content and vapor pressure equilibrium, since the vapor pressure gradients are the essential driving forces during these processes (VAN DEN BERG; BRUIN, 1981). Due to its importance in dehydration processes, rehydration conditions, as well as in packaging and in quality changes during storage, the sorption of water vapor by foods has received much attention (VEGA-GÁLVEZ et al., 2009; MOREIRA; CHENLO; TORRES, 2009; MORAES; ROSA; PINTO, 2008; PARK; YADO; BROD, 2001). The sorption isotherm consists of a graphic representation of the relationship

Received 28/11/2009

Accepted 5/6/2010 (004521)

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between the water content and the equilibrium relative humidity at constant temperature. The knowledge of the moisture sorption isotherm data as a function of two or more temperatures is important for the thermodynamic analysis and modeling of drying kinetics, and it is also useful to evaluate food stability during storage (VEGA-GÁLVEZ et al., 2007). The knowledge of the thermodynamic properties can also provide information about the food microstructure as well as allowing for the theoretical interpretation of physical phenomena occurring at the water-food interface (RIZVI, 1995).

Water affects the physical nature and properties of food in a complex way due to interactions with their solutes, colloids and dispersed particles (PARK; YADO; BROD, 2001). The water activity of food reflects the availability of water for deteriorative reactions and microbiological growth and it is one of the major control variables in food preservation technology (VAN DEN BERG; BRUIN, 1981).

Many theoretical, partially theoretical and empirical models can be found in the literature for the mathematical description of food system moisture sorption (VAN DEN BERG; BRUIN, 1981). Boquet, Chirife and Iglesias (1978) evaluated the usefulness of eight two-parameter isotherms in describing the moisture sorption isotherms of 39 different nature foods and found that the most versatile models were those of Halsey and Oswin. In addition, the Henderson model has found many practical applications, and it is one of the only equations including a direct temperature term (VAN DEN BERG; BRUIN, 1981).

The BET model, based on the multi-layer adsorption of water, is considered the best model for interpreting sorption isotherms (MATHLOUTHI; ROGÉ, 2003). Of all the multi-layer models available for sorption, this model acts as a useful compromise between theory and practice, since the equation parameters have physical significance and the model has the ability to fit sigmoidal sorption isotherms up to an  $a_w$  of about 0.4 (VAN DEN BERG; BRUIN, 1981). The GAB model was considered an important improvement on the BET equation, and its application has shown a good fit up to an  $a_w$  of 0.9 (VAN DEN BERG; BRUIN, 1981; LEWICKI, 1997).

The calculation of the energy consumption during drying demands knowledge of the enthalpy of water sorption throughout the entire range of moisture contents (TSAMI, 1990). The enthalpy of sorption yields a figure for the theoretical minimum amount of heat required to remove a given amount of water (RIZVI; BENADO, 1984).

The purpose of this work was to evaluate the influences of temperature and heat blanching on the behavior desorption isotherms and on the isosteric heat of sorption of pumpkins.

## 2 Materials and methods

### 2.1 Materials

Mature pumpkins (*Cucurbita moschata*) of the 'Rajada Seca Melhorada' cultivar, with a total weight of approximately 40 kg, were obtained from the local market

(CEAGESP - São José do Rio Preto, State of São Paulo, Brazil) and stored at room temperature (25 °C). Saturated aqueous salt solutions were prepared using NaOH, LiCl,  $\text{CH}_3\text{COOK}$ ,  $\text{MgCl}_2$ ,  $\text{K}_2\text{CO}_3$ , NaBr,  $\text{NaNO}_2$ , NaCl, KCl and  $\text{BaCl}_2$  (all analytical grade) and commercial formaldehyde (40%) was used for cleaning the material.

### 2.2 Samples

The pumpkins were cut into portions transversally to their axes, and the extremities were discarded. Each portion was cut into longitudinal pieces, peeled, seeded and sliced ( $4 \times 10^{-3}$  m thickness). To standardize the samples, circles were cut from the slices using a cork borer with a  $2.8 \times 10^{-2}$  m diameter, and then a  $2.2 \times 10^{-2}$  m plug was removed from the center of each circle and discarded. The resulting rings were put in a plastic bag, mixed and divided in two portions, one for fresh and one for blanched sampling.

### 2.3 Blanching

The pumpkin rings (0.4 kg) were placed in a wire basket, immersed in boiling water ( $2.0 \times 10^{-2}$  m<sup>3</sup>, 98,3 °C) for 60 seconds, cooled with tap water for a further minute, drained and dried with absorbent paper.

### 2.4 Sorption isotherms

The gravimetric static method (RIZVI, 1995) was employed to determine the equilibrium moisture of fresh and blanched pumpkin at 30, 50 and 70 °C. Saturated salt solutions were prepared in glass flasks previously cleaned with formaldehyde (40%) to avoid microbial growth. The relative humidity ranged from 5 to 90%, obtained using ten salts: NaOH, LiCl,  $\text{CH}_3\text{COOK}$ ,  $\text{MgCl}_2$ ,  $\text{K}_2\text{CO}_3$ , NaBr,  $\text{NaNO}_2$ , NaCl, KCl and  $\text{BaCl}_2$ . Five gram samples (in triplicate) of fresh and blanched pumpkin were weighed in plastic baskets and placed in the glass flasks on top of acrylic tripods arranged in the flasks to maintain the samples above the solutions. Fresh and blanched samples were characterized with respect to total solids and sugars in independent assays. Each flask, containing three fresh and three blanched samples, was hermetically closed and placed in a temperature-controlled incubator. A BOD incubator was used at 30 and 50 °C (sensor PT100,  $\pm 0,1^\circ\text{C}$  precision and  $\pm 1^\circ\text{C}$  homogeneity). An oven with air circulation was used at 70 °C (type J sensor,  $\pm 2^\circ\text{C}$  precision and  $\pm 4^\circ\text{C}$  homogeneity). The samples were weighed periodically until constant weight. The total solids contents were then determined, as well as the average equilibrium moisture contents related to the corresponding water activity.

### 2.5 Analytical methods

The solids contents of the fresh and blanched samples were determined gravimetrically by drying to constant weight in a vacuum oven at 60 °C and 10 kPa. The reducing and total sugar contents were determined by oxy-reduction titration (ASSOCIATION..., 1970).

**Table 1.** Water sorption isotherm models.

Model	Equation
BET	$X = \frac{X_m \cdot C \cdot a_w}{(1 - a_w)(1 + (C - 1)a_w)} \quad (1)$
GAB	$X = \frac{X_m \cdot C \cdot K \cdot a_w}{(1 - K \cdot a_w)(1 - K \cdot a_w + C \cdot K \cdot a_w)} \quad (2)$
Halsey	$X = \left[ \frac{-a}{\ln(a_w)} \right]^{\frac{1}{b}} \quad (3)$
Henderson	$X = \left[ \frac{\ln(1 - a_w)}{a} \right]^{\frac{1}{b}} \quad a = kT, \quad T \text{ em } ^\circ Ra \quad (4)$
Oswin	$X = a \left[ \frac{a_w}{(1 - a_w)} \right]^b \quad (5)$

Where  $C$ ,  $K$ ,  $a$  and  $b$  are constants;  $a_w$  is the water activity;  $X$  represents the moisture content on a dry weight basis (kg water/kg dry matter) and  $X_m$  is the monolayer of water (kg water/kg dry matter).

## 2.6 Mathematical modeling

### Sorption isotherms

BET, GAB, Halsey, Henderson and Oswin models (Table 1) were fitted to the food system moisture sorption.

### Net isosteric heat of sorption

The net isosteric heat of sorption of water ( $q_{st}$ ) can be determined from Equation 6, which is derived from the Clausius – Clapeyron equation, applied to food and pure water (RIZVI, 1995).

$$\left. \frac{\partial \ln(a_w)}{\partial (1/T)} \right|_X = -\frac{Q_{st} - \lambda}{R} = -\frac{q_{st}}{R} \quad (6)$$

where  $Q_{st}$  is the integral isosteric heat of sorption ( $\text{kJ} \cdot \text{mol}^{-1}$ ),  $\lambda$  the molar enthalpy of vaporization of pure water ( $\text{kJ} \cdot \text{mol}^{-1}$ ),  $R$  the gas constant ( $\text{kJ} \cdot \text{mol}^{-1} \cdot \text{K}^{-1}$ ) and  $T$  the temperature (K). The net isosteric enthalpy is obtained by subtracting the value obtained for the corresponding equation for pure water.

The net isosteric heat of sorption can be calculated from Equation 6 by plotting the sorption isostere as  $\ln(a_w)$  against  $(1/T)$  for a specific moisture content of the material and determining the slope, which equals  $(Q_{st}/R)$  (TSAMI et al., 1990). This procedure is repeated for different moisture contents in order to determine the dependence of  $(q_{st})$  on the moisture content ( $X$ ). The method presents an overall error due to plotting graphical differentiation (KAYMAK-ETERKIN; GEDIK, 2004).

An empirical exponential relationship between the net isosteric heat of sorption ( $q_{st}$ ) and the moisture content ( $X_{eq}$ , dry basis) was proposed by Tsami et al. (1990) in the following form (Equation 7):

$$q_{st} = q_0 \exp(-X / X_0) \quad (7)$$

where  $q_0 = q_{st}$  at  $X = 0$  and, at  $X = X_0$ ,  $q_{st} = 0.368q_0$ . The constant  $X_0$  is the characteristic moisture content (kg water  $\text{kg}^{-1}$  dry matter) of a food material in which the net isosteric heat of sorption has been reduced by 63%. Due to exponential decay, the net isosteric heat of sorption becomes very small at high moisture contents.

## 2.7 Statistic methods

The efficiency of fit was evaluated based on the adjusted coefficient of determination ( $R^2$ ) and on the mean relative modulus,  $P$ , defined by Equation 8 (LOMAURO; BAKSHI; LABUZA, 1985),

$$P(\%) = \frac{100}{n} \sum_{i=1}^n \frac{|y^{\text{exp}} - y^{\text{calc}}|}{y^{\text{exp}}} \quad (8)$$

where  $y^{\text{exp}}$  represents the experimental value,  $y^{\text{calc}}$  the calculated value and  $n$  is the number of the observations.

The graphic software ORIGIN v. 5.0 (MICROCAL SOFTWARE, 1997), which adjusts non-linear functions, was used to determine the adjustment constants of the isotherms (Equation 1 to 5) and Equation 7.

## 3 Results and discussion

### 3.1 Sorption isotherms

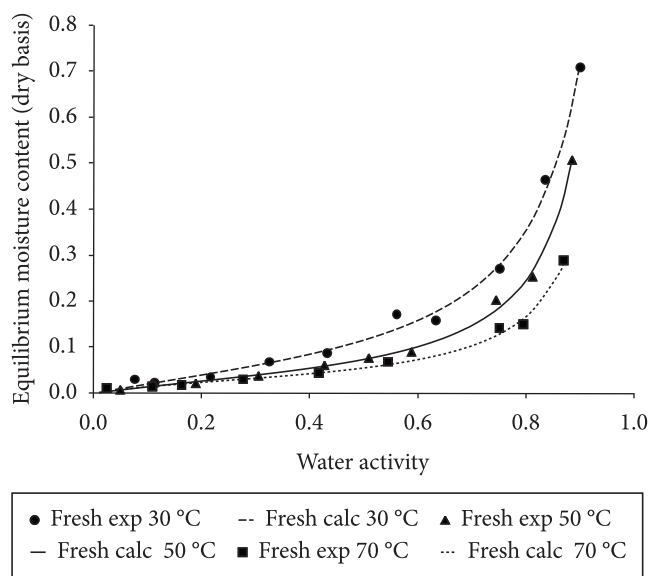
The equilibrium moisture contents experimentally determined at temperatures 30, 50 and 70 °C were compared to the values calculated according to the GAB equation (Equation 2) for fresh (Figure 1) and blanched (Figure 2) pumpkin. The desorption curves showed that, at constant water activities, the values for the equilibrium moisture content decreased with as the temperature increased, following the same tendency of most agricultural products (RESENDE et al., 2006). The influence of blanching on the pumpkin isotherms

was small, showing a behavior quite similar to that of fresh pumpkin isotherms. Nevertheless, the blanched samples showed slightly lower values for the moisture equilibrium than the fresh ones at 30 and 50 °C (Figures 3 and 4, respectively). Moreover, as the temperature increased, the difference between fresh and blanched curves diminished at low and intermediate moisture levels and even showed a small inversion at 70 °C (Figure 5).

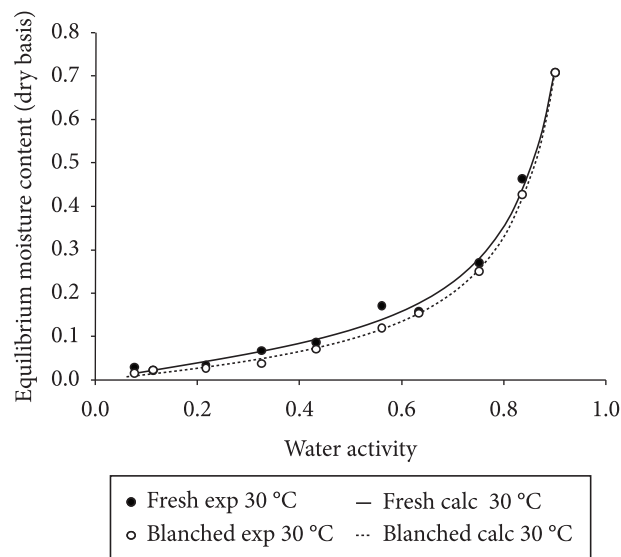
Independent tests were carried out to characterize the fresh and blanched samples (Table 2). The total sugar loss during

blanching was approximately 15%, which can explain the lower equilibrium moisture content of the blanched samples when compared to the fresh ones, since sugars retain a larger amount of water (CHIRIFE; FERRO FONTAN; BENMERGUI, 1980) than cellulosic compounds (PAPADAKIS et al., 1993) and proteins (BULL, 1944), which, together with the sugars, are the major pumpkin components (UNIVERSIDADE..., 2006).

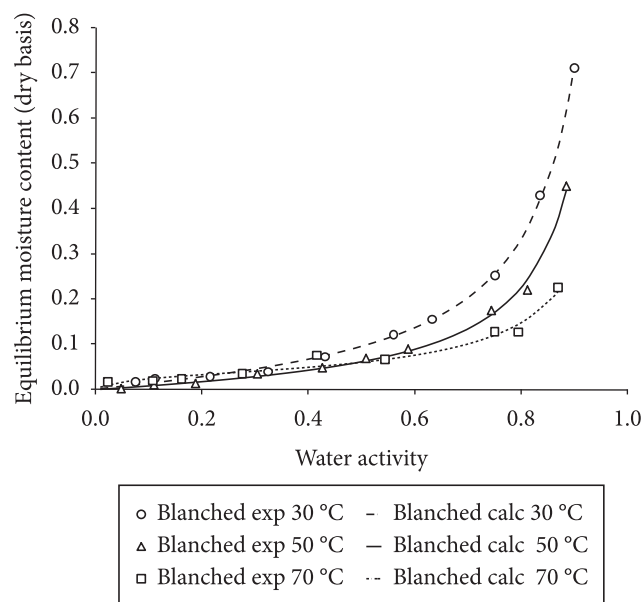
Sorption models (Table 1) were tested with experimentally defined equilibrium moisture data (Table 3) to describe fresh



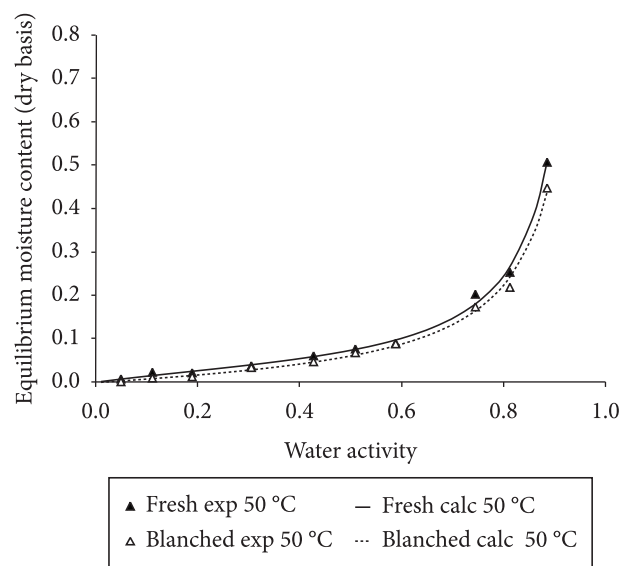
**Figure 1.** Comparison between experimental and calculated (GAB model) equilibrium moisture contents (dry weight basis, dwb) for fresh pumpkin at 30, 50 and 70 °C.



**Figure 3.** Comparison between experimental and calculated (GAB model) equilibrium moisture contents (dry weight basis, dwb) for fresh and blanched pumpkin at 30 °C.



**Figure 2.** Comparison between experimental and calculated (GAB model) equilibrium moisture contents (dry weight basis, dwb) for blanched pumpkin at 30, 50 and 70 °C.



**Figure 4.** Comparison between experimental and calculated (GAB model) equilibrium moisture contents (dry weight basis, dwb) for fresh and blanched pumpkin at 50 °C.

**Table 2.** Water content and total sugar content, on both wet and dry weight bases, of fresh and blanched pumpkin.

	Water content kg.kg <sup>-1</sup> wet matter	Reducing sugar	Non-reducing sugar	Total sugar content kg.kg <sup>-1</sup> wet matter	Total sugar content kg.kg <sup>-1</sup> dry matter
Fresh	0.9347	0.0163	0.0170	0.0333	0.5087
Blanched	0.9565	0.0089	0.0099	0.0189	0.4325

**Table 3.** Equilibrium moisture content (dry weight basis) of fresh and blanched pumpkin at 30, 50 and 70 °C, and corresponding water activity ( $a_w$ )

Temperature	Salt	Water activity $a_w$	Equilibrium content $X_{eq}$ (dwb)* Fresh pumpkin	Equilibrium content $X_{eq}$ (dwb)* Blanched pumpkin
30 °C	NaOH	0.0758	0.0300 ± 0.0072	0.0162 ± 0.0013
	LiCl	0.1128	0.0222 ± 0.0002	0.0239 ± 0.0012
	CH <sub>3</sub> COOK	0.2160	0.0350 ± 0.0054	0.0281 ± 0.0017
	MgCl <sub>2</sub>	0.3244	0.0678 ± 0.0007	0.0389 ± 0.0027
	K <sub>2</sub> CO <sub>3</sub>	0.4317	0.0870 ± 0.0054	0.0716 ± 0.0032
	NaBr	0.5603	0.1716 ± 0.0555	0.1209 ± 0.0053
	NaNO <sub>2</sub>	0.6330	0.1581 ± 0.0013	0.1547 ± 0.0022
	NaCl	0.7509	0.2704 ± 0.0200	0.2516 ± 0.0028
	KCl	0.8350	0.4640 ± 0.0795	0.4278 ± 0.0052
	BaCl <sub>2</sub>	0.9000	0.7097 ± 0.0224	0.7094 ± 0.0084
50 °C	NaOH	0.0494	0.0073 ± 0.0007	0.0021 ± 0.0008
	LiCl	0.1110	0.0226 ± 0.0090	0.0106 ± 0.0017
	CH <sub>3</sub> COOK	0.1890	0.0213 ± 0.0021	0.0135 ± 0.0017
	MgCl <sub>2</sub>	0.3050	0.0376 ± 0.0028	0.0347 ± 0.0027
	K <sub>2</sub> CO <sub>3</sub>	0.4270	0.0606 ± 0.0040	0.0480 ± 0.0031
	NaBr	0.5090	0.0765 ± 0.0030	0.0688 ± 0.0015
	NaNO <sub>2</sub>	0.5880	0.0899 ± 0.0026	0.0892 ± 0.0142
	NaCl	0.7440	0.2030 ± 0.0086	0.1742 ± 0.0095
	KCl	0.8120	0.2537 ± 0.0130	0.2192 ± 0.0086
	BaCl <sub>2</sub>	0.8850	0.5083 ± 0.0123	0.4478 ± 0.0134
70 °C	NaOH	0.0230	0.0109 ± 0.0024	0.0159 ± 0.0017
	LiCl	0.1073	0.0144 ± 0.0021	0.0187 ± 0.0031
	CH <sub>3</sub> COOK	0.1620	0.0174 ± 0.0012	0.0233 ± 0.0010
	MgCl <sub>2</sub>	0.2760	0.0299 ± 0.0037	0.0344 ± 0.0045
	K <sub>2</sub> CO <sub>3</sub>	0.4160	0.0446 ± 0.0023	0.0754 ± 0.0385
	NaBr	0.4970	nd**	nd**
	NaNO <sub>2</sub>	0.5440	0.0682 ± 0.0029	0.0662 ± 0.0031
	NaCl	0.7506	0.1423 ± 0.0034	0.1280 ± 0.0008
	KCl	0.7950	0.1498 ± 0.0040	0.1267 ± 0.0035
	BaCl <sub>2</sub>	0.8700	0.2894 ± 0.0127	0.2258 ± 0.0053

\*dwb: dry weight basis; \*\*non-determined.

and blanched pumpkin sorption isotherm curves. With the exception of Halsey's equation when fitted to 50 °C data of blanched tissue and Henderson's equation, in general, all the models showed satisfactory adjustments to the experimental data (Table 4). For fresh tissue data, the best fitting was obtained through BET and GAB equations, since the values for  $R^2$  were always higher than 0.98 and  $P$  values were lower than 13.3%. The blanched tissue showed a satisfactory fit to GAB, BET, Halsey and Oswin models only at 30 °C ( $R^2 > 0.99$  and  $P < 12.6\%$ ). For 50 °C isotherm, BET, GAB and Oswin models were acceptable

( $R^2 > 0.99$  and  $P < 17.5\%$ ), while for 70 °C isotherm, BET, GAB, Halsey and Oswin models showed  $R^2 > 0.95$  and  $P < 17.9\%$ .

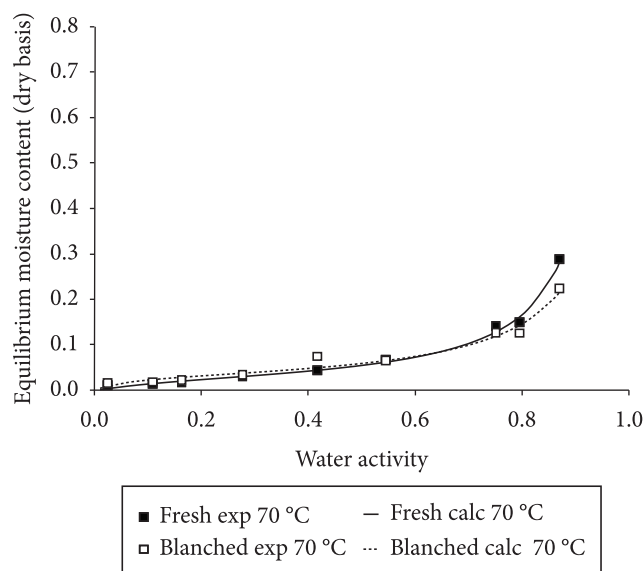
$P$  modulus value below 10% is an indicative of a good fit for practical purposes (LOMAURO; BAKSHI; LABUZA, 1985). However, the  $P$  presented values are justified by the low equilibrium moisture contents obtained in relative humidity around 5%, since the  $P$  calculation is based on the relative residual (Equation 8), which amplifies the deviations when the moisture content is quite low.



Five types of isotherms are described by Brunauer (BRUNAUER et al., 1940; INTERNATIONAL..., 1985) The GAB model describes sigmoidal type isotherms (type II) well and fulfils the requirements of the BET model when the parameters are kept within the following regions:  $0.24 < K \leq 1$  and  $5.67 \leq C \leq \infty$ . Outside these regions the isotherm is either no longer sigmoidal or the monolayer capacity is estimated with an error greater than  $\pm 15.5\%$  (LEWICKI, 1997).

Table 4 shows that only the fresh and blanched pumpkin curves prepared at 70 °C complied with the requirement for parameter C, and that only the blanched one complied with parameter K. The other isotherms, adjusted using the GAB model, presented parameter C outside the band, showing mixed behavior between types II and III (slightly sigmoidal). Mayor et al. (2005) also found forms of fresh pumpkin parenchyma isotherms (cv *Cucurbita Pepo* L.) between types II and III at 5, 25 and 45 °C.

Figures 6 and 7 show the curves for the neperian logarithm of water activity ( $\ln(a_w)$ ) as a function of the inverse values of the absolute temperature ( $1/T$ ) for specific values of equilibrium moisture (dwb) for fresh and blanched pumpkin, respectively.

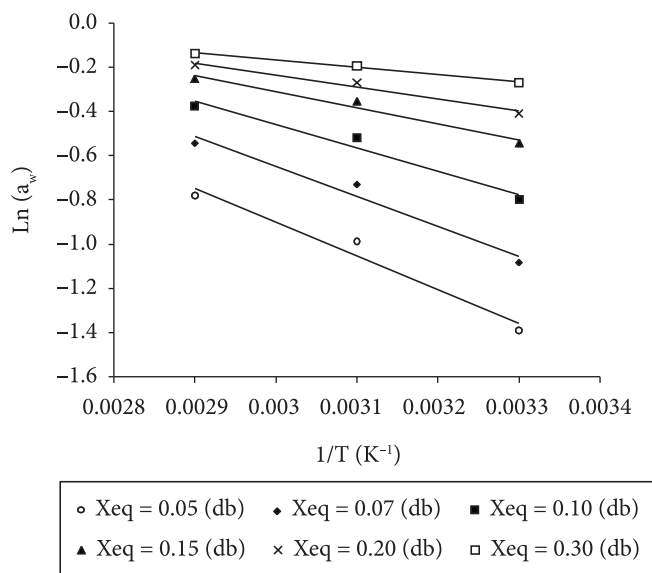


**Figure 5.** Comparison between experimental and calculated (GAB model) equilibrium moisture contents (dry weight basis, dwb) for fresh and blanched pumpkin at 70 °C.

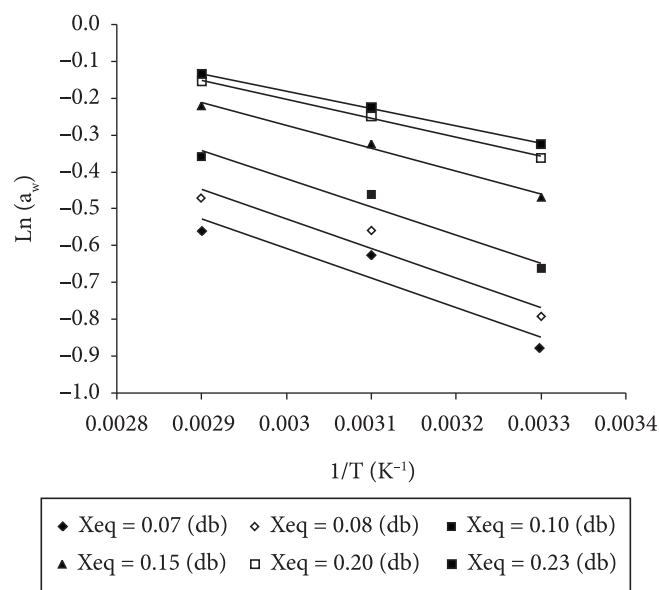
**Table 4.** BET, GAB, Halsey, Henderson and Oswin model fitting constants for fresh and blanched pumpkins, determined at 30, 50 and 70 °C; corresponding R2 and RMSR.

Sample	Model	T (°C)	R <sup>2</sup>	P (%)	Parameters				
					a	b	C	K	X <sub>m</sub>
Fresh	BET	30	0.9849	12.98	–	–	3.0505	–	0.0759
		50	0.9877	11.40	–	–	1.9177	–	0.0584
		70	0.9816	13.32	–	–	4.2217	–	0.0366
	GAB	30	0.9853	12.68	–	–	2.6575	0.9916	0.0809
		50	0.9927	8.52	–	–	3.2653	1.0303	0.0460
		70	0.9846	13.30	–	–	6.3183	1.0267	0.0312
	Halsey	30	0.9815	13.75	0.0796	0.9646	–	–	–
		50	0.9957	15.88	0.0668	0.8796	–	–	–
		70	0.9908	9.01	0.0405	0.9740	–	–	–
	Henderson	30	0.9570	24.10	3.1919	0.7245	–	–	–
		50	0.9503	29.20	3.7861	0.6635	–	–	–
		70	0.9426	25.56	6.9871	0.8525	–	–	–
Blanched	Oswin	30	0.9852	19.50	0.1169	0.8189	–	–	–
		50	0.9854	12.74	0.0756	0.9001	–	–	–
		70	0.9769	13.27	0.0611	0.7654	–	–	–
	BET	30	0.9926	12.59	–	–	1.6106	–	0.0766
		50	0.9922	16.35	–	–	1.3596	–	0.0541
		70	0.9510	17.46	–	–	22.4103	–	0.0296
	GAB	30	0.9926	12.57	–	–	1.5902	0.9993	0.0771
		50	0.9926	17.45	–	–	1.5621	1.0114	0.0495
		70	0.9598	17.89	–	–	15.5818	0.9818	0.0322
	Halsey	30	0.9906	11.55	0.0813	0.8775	–	–	–
		50	0.9657	41.44	0.0672	0.7822	–	–	–
		70	0.9615	12.02	0.0213	1.2316	–	–	–
	Henderson	30	0.9703	24.16	3.0064	0.6233	–	–	–
		50	0.9738	19.80	4.0644	0.6540	–	–	–
		70	0.9240	20.39	15.7155	1.1978	–	–	–
	Oswin	30	0.9927	11.96	0.0952	0.9142	–	–	–
		50	0.9915	16.58	0.0618	0.9491	–	–	–
		70	0.9534	16.32	0.0671	0.5789	–	–	–

Figure 8 shows the net isosteric heat of sorption values ( $q_{st}$ ) for fresh and blanched pumpkins according to the equilibrium moisture values (dwb) calculated for different equilibrium moisture contents at 30, 50 and 70 °C, using Equation 6. The



**Figure 6.** Neperian logarithm of the water activity ( $\ln(a_w)$ ) as a function of the inverse values of absolute temperature ( $1/T$ ) for the fresh samples.

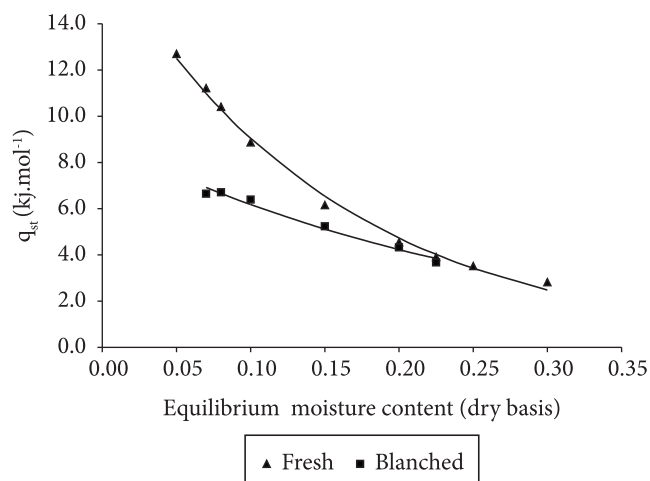


**Figure 7.** Neperian logarithm of the water activity ( $\ln(a_w)$ ) as a function of the inverse values of absolute temperature ( $1/T$ ) for the blanched samples.

net isosteric heat of sorption is represented by the slopes of the lines obtained (Figures 6 and 7), and decreases as the product moisture content increases. Such a behavior can be explained by considering that the sorption of the first layers of water occurs at the most active sites available, originating high energy of interaction (MAZZA; LE MAGUER, 1980). Table 5 shows the values obtained for the constants  $q_0$  and  $X_0$ , as well as the determination coefficient ( $R^2$ ) established by fitting Equation 7 to the values of  $q_{st}$  and  $X_{eq}$  shown in Figure 8.

The net isosteric heat of sorption was inversely proportional to the moisture content (Table 5), what was expected due to the higher degree of binding energy of the water to the surface food material for the lower moisture contents (GABAS; MENEGALLI; TELIS-ROMERO, 2000). It can be seen that the heat of sorption of the fresh samples was higher than that of the blanched samples when the moisture content was low. This means that a larger amount of heat is required to remove the water from the fresh sample as compared to the blanched sample. These results were related to the higher sugar contents and the presence of more polar sites on the surface of the fresh tissue.

However, the heat sorption difference decreases as the moisture content increases until this difference is minimal, nearly 0.25 (dwb) (Figure 8). As the moisture increases, the presence of water in the free form increases, which diminishes the binding energy degree and its influence on the net isosteric heat of sorption.



**Figure 8.** Comparison between the net isosteric heat of sorption values determined by the slopes of the lines obtained (Figures 6 and 7) and calculated (continuous line: Equation 7), as a function of equilibrium moisture content for fresh and blanched pumpkin.

**Table 5.** The constants  $q_0$  and  $X_0$  and the determination coefficient ( $R^2$ ) established by fitting Equation 7 to the values of  $q_{st}$  and  $X_{eq}$  shown in Figure 8.

	$q_0$ (kJ.mol <sup>-1</sup> )	$X_0$ (dry basis)	$R^2$
Fresh	17.29	0.1544	0.9975
Blanched	9.02	0.2645	0.9899



## 4 Conclusions

The blanching process had a slight influence on the equilibrium moisture content of the samples, which decreased compared to the fresh samples, this being associated with the loss of soluble solids during the pre-treatment. GAB, Oswin, BET and Halsey models, in general, showed satisfactory fits to the experimental data. The GAB model was used, since its fitted parameters allowed for the analysis of curve types. The isotherms of the fresh and blanched samples determined at 30 and 50 °C showed behaviors between types II and III. At 70 °C, only the isotherm of the blanched sample clearly showed type II behavior. The behavior of the fresh sample was very similar to that of type II, but one of the parameters did not meet the requirements. The isosteric heats of sorption were quite similar to each other in the moisture levels around 0.25 (dwb). For lower moisture levels, the isosteric heat values indicated that the interactions between solids and water in the fresh samples were stronger than in the blanched ones.

## Acknowledgements

The authors are grateful to FAPESP ('Fundação de Amparo à Pesquisa do Estado de São Paulo') for the financial support (Proc 07/07586-0) and for the studentships (Proc 07/00809-3; Proc. 04/15550-7).

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