Silva de MORAES, Kessiane; FAGUNDES, Cristiane; MELO, Maria Carolina; ANDREANI, Pamela; Rodriguez MONTEIRO, Alcilene

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Conservation of *Williams* pear using edible coating with alginate and carrageenan

Conservação de pera *Williams* utilizando coberturas comestíveis compostas de alginato e carragena

Kessiane Silva de MORAES¹, Cristiane FAGUNDES¹, Maria Carolina MELO¹, Pamela ANDREANI¹, Alcilene Rodriguez MONTEIRO*¹

Abstract

The aim of this study was to evaluate the physical and chemical parameters of *Williams* pear, stored at 25 °C for 15 days, with and without edible coating. Edible coatings prepared with alginate 2% and carrageenan 0.5% were tested. The analyses carried out on the samples were: weight loss, pH, soluble solids, firmness, and color. The edible coatings were characterized in terms of mechanical properties, permeability, thickness, and opacity. The results show that the application of edible coatings with carrageenan and alginate in pears influenced physical and chemical characteristics such as weight loss, pH, total soluble solids, color, and firmness of the fruit. However, the alginate coating showed the best results on pear conservation since it had lower water vapor permeability and greater tensile strength, and therefore it can be used as a protective film on these fruits.

*Keywords:* storage; room temperature; physicochemical analysis.

1 Introduction

Pears are climacteric fruits, in which the major negative changes are the results of maturation. During ripening, some changes are observed in firmness, color, acidity, sugar content, and development of aroma (ALPALHAO et al., 2006).

In order to slow down the changes that occur during ripening, studies on the use of edible coatings in order to prolong the shelf life of fruits and vegetables such as strawberries (RIBEIRO et al., 2007), grapes (FAKHOURI et al., 2007), and apples have been conducted (LEE et al., 2003; OLIVAS; MATTINSON; BARBOSA-CÁNOVAS, 2007).

Edible films are used to inhibit migration of moisture, oxygen, carbon dioxide, aromas, and lipids and to improve the mechanical integrity or handling characteristics of the food (KROCHTA; DE MULDER-JOHNSTON, 1997). Edible films should have adequate mechanical strength and extensibility to maintain integrity and withstand the external stress that occurs during processing, handling, and storage (YANG; PAULSON, 2000).

These films can be obtained from different types of materials, but the most used ones are made of polysaccharides, proteins, and lipids (CHEN, 1995). Alginate is a product of interest for the preparation of films and coatings due to its colloidal properties, which include thickener, stabilizer, gelling, and suspending and film capacities (FABRA et al., 2008).

The use of carrageenan as edible films and coatings has been explored by various fields of the food industry such as its application on fresh and frozen meat, poultry and fish to prevent superficial dehydration (SHAW; SECRISt; TUOMY, 1980), ham or sausage casings (MACQUARRIE; SCHUPP; TAYLOR, 2004), granulation-coated powders, dry solid foods, and oily foods (NINOMIYA; SUZUKI; ISHII, 1997). According to Olivas and Barbosa-Cánovas (2005), some polysaccharides such as carrageenan have been used successfully in the coverage of minimally processed fruits.

Most fruits available in Brazil are marketed at room temperature justifying the interest in evaluating the quality of whole *Williams* pears stored at 25 °C using edible coatings as a conservation technique. The objective of this study was to characterize edible coatings through the evaluation of mechanical properties, permeability, thickness, and opacity.

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¹ Laboratório de Propriedades Físicas de Alimentos, Departamento de Engenharia Química e Engenharia de Alimentos, Universidade Federal de Santa Catarina – UFSC, Campus Trindade, CP 476, CEP 88040-900, Florianópolis, SC, Brasil, e-mail: alcilene@enq.ufsc.br
*Corresponding author

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in addition to evaluating the changes in the physicochemical parameters of Williams pear, with or without edible coating, stored at room temperature (25 °C) for 15 days.

2 Material and methods

2.1 Material

Williams Pears (Pyrus communis) were obtained from a local market at the commercial ripening stage (12 to 14 °Brix and green skin color). The coatings were made from carrageenan from Sigma Aldrich (St. Louis, USA), sodium alginate from Sigma Aldrich (Gillingham, UK), glycerol (Nuclear), and sodium hypochlorite (Vetec).

2.2 Edible coating characterization

25 mL of film solutions were added to acrylic plates 14 cm in diameter and dried in an oven (TE-394/2, Tecnal, Piracicaba, Brazil) at 25 °C for 24 hours. The gain of mass in each diffusion cell was determined by successive weighing on an analytical balance (AY 220, Shimadzu, São Paulo, Brazil) in intervals of 1 hour for 12 hours; after that it was determined 24 hours from the beginning of the experiment. Water vapor permeability was determined by the gravimetric method ASTM E96-00 described by Sarantopulos et al. (2002). The coverage thickness was measured using a micrometer (MDC 25P, Mitutoyo, Tokyo, Japan), as an average of ten random measurements. The tensile tests were performed on a digital texture analyzer (TAXT2i, Stable Micro System, Godalming, UK) with 6 samples (25 mm × 100 mm) conditioned for 48 hours at 58% RH at 25 °C. The test was conducted based on the D882-00 method (AMERICAN... 1996) with initial distance from the clutches of 50 mm and tensile speed of 0.8 mm/s. The color parameters of the films were determined using a colorimeter (Miniscan EZ, HunterLab, Reston, USA), and the opacity was calculated according to Fakhouri et al. (2007).

2.3 Preparation and application of edible coatings

For the carrageenan coating (Sigma Aldrich, St. Louis, USA), a 0.5% aqueous solution and 0.15 g of glycerol was prepared according to Larotonda (2007). For the sodium alginate (Sigma Aldrich, Gillingham, UK) coating, a 2% solution was prepared according to Meneghel, Benassi and Yamashita (2008) with modifications. The pears were sanitized in sodium hypochlorite (100 ppm/20 min). Next, they were divided into two batches of 15 units. The control batch (without coating) was immersed in distilled water, and the fruits of the second batch were immersed for 5 seconds in one of the filmogenic solutions (500 mL) at 25 °C and kept at room temperature to allow the coating to dry. The control samples and the samples with edible coatings were stored at 25 °C for 15 days in an incubator at relative humidity of 78%.

2.4 Characterization of pears

Pears were characterized by physical and chemical analysis (weight loss, pH, and soluble solids), firmness (penetration force), and color. All tests were performed in triplicate on the 1st, 4th, 6th, 8th, 11th, and 15th days of storage.

The weight loss was determined gravimetrically using an analytical balance (AY 220, Shimadzu, São Paulo, Brazil) during the storage period (PEREIRA et al., 2006).

The pear juice was obtained by mixing 10 g of the sample and 100 mL of water with a mixer (Turbonix, Moulinex). The pH of pear juice was determined using a digital pH meter (Q400 A, Quimis, Diadema, Brazil).

The soluble solids content was determined after the compression of the fruit pulp to extract juice using a digital refractometer (AR 200, Reichert Analytical Instruments, New York, USA).

The firmness of the samples was measured by penetration using a digital texture analyzer (TAXT2i, Stable Micro System, Godalming, UK) with a load cell of 50 kg and a 2 mm probe, test speed of 3.3 mm/s, penetration of 2 mm, and time of 30 seconds. Ten measurements were performed for each day of analysis.

To determine the color of the samples, 8 measurements were performed in the fruit skin using a colorimeter (Miniscan EZ, HunterLab, Reston, USA) with a D65/10° lighting system. Lightness (L), green to red (a*), and blue to yellow color (b*) were evaluated using the Hunter scale.

2.5 Statistical analysis

The results were evaluated by analysis of variance (ANOVA) with Statistica 6.0 (Statsoft Inc., USA) software, and the factors that showed significant differences (p ≤ 0.05) were submitted to Tukey’s test.

3 Results and discussion

3.1 Edible coating characterization

Table 1 shows the thickness, opacity, tensile tests, and water vapor permeability (WVP) of edible coatings. The thickness of the films were 0.042 mm and 0.039 mm for the alginate and carrageenan films, respectively. Larotonda (2007), who studied carrageenan coatings produced by casting, found film thickness values of 0.09 mm. The values were different due to the amount of filmogenic solution added to the plates in order to obtain the films. Zactiti (2004) found that calcium alginate films had a thickness of 0.097 mm. According to the author, thickness has a great influence on the barrier properties of films. The opacity values of alginate and carrageenan showed a significant difference (p ≤ 0.05). In the present study, the alginate and carrageenan films showed opacities of 4.52% and 6.75%, respectively. The results obtained by Da Silva (2009) for pure alginate films showed 7% opacity.

The alginate edible coating showed higher tensile strength, elongation, and elastic modulus when compared to the carrageenan coating (p ≤ 0.05). Oliveira et al. (2006) showed elongation values of 1.9% for sodium alginate films. Pranoto, Salokhe and Raksh (2005) evaluated the mechanical properties
of alginate-based edible film incorporated with garlic oil and found rupture tension values of 66.12 MPa and elongation of 4.05% for films without the addition of garlic oil. Larotonda (2007) evaluated the mechanical properties of carrageenan film in different concentrations (16.7, 33.3, 44.4, and 100%) and found rupture tension values between 10 and 55 MPa, elongation between 1 and 45 %, and Young’s Modulus between 10 and 25 MPa. The results show that the alginate coating had lower permeability and showed a significant difference (p ≤ 0.05) from carrageenan coating. According to Hambleton et al. (2008), the water vapor permeability values of the alginate film were lower than those of the carrageenan film. This is probably due to ionic crosslinking in alginate films reducing the segmental mobility of the polymer.

### 3.2 Characterization of pears

The fruit weight loss values are presented in Figure 1. The loss of water in fruits is mainly related to the sweating, breathing, and storage time, and it results in wrinkling, softening of tissues, and loss of brightness making fruits more susceptible to deterioration (KADER, 1992).

The weight loss was greater for the control samples, followed by the samples with carrageenan 0.5% and alginate 2% (Figure 1). It was found that 2% alginate used in the coatings was sufficient to minimize the loss of water to the environment. These results indicate that weight loss was the lowest for the samples with alginate due to the lower water vapor permeability of this coating. Miguel et al. (2009) evaluated the effect of the film concentration on the weight loss of grapes and found that the grapes treated with 1% alginate showed the lowest losses (10.16%), followed by concentrations of 0.75% (11.72%), 0.50% (11.79%), 0.25% (14.14%), and the control sample (16.43%). Larotonda (2007) studied the influence of carrageenan coating on the weight loss of cherries and found that the coating significantly reduced (p ≤ 0.05) weight loss of the fruits during storage when compared to that of the control sample.

The pH of fruit is presented in Table 2. During storage, there was a significant increase (p ≤ 0.05) in the pH of the control samples and the fruits coated with alginate and carrageenan 0.5%. The coated samples differed significantly (p ≤ 0.05) of the control sample on all days evaluated. Since the control sample showed an increase in pH greater than that of the coated samples, this indicates that the use of coating slows down changes in the pH of the fruit. Comparing the edible coatings used, alginate showed smaller changes in the pH of the pears analyzed from the 1st to the 15th day of storage.

According to Chitarra and Chitarra (2005), the organic acid content decreases with maturation due to the breathing process or its conversion into sugars. Therefore, the pH of the fruit increases with a decrease in acidity, and thus it is used as an indicator of this variation. Fontes et al. (2008) used an edible coating of alginate on minimally processed apples stored at 2 °C for 13 days and found that the pH of the fruits remained unchanged during storage. Bico et al. (2009) observed a significant increase (p ≤ 0.05) in pH values of fresh-cut bananas stored at 5 °C for 5 days treated with a coating of carrageenan and a preservative solution under controlled atmosphere.

The soluble solids content of the fruits are presented in Table 2. The soluble solids (SS) content increased significantly (p ≤ 0.05) during storage for all samples. A significant difference (p ≤ 0.05) between the control and coated samples was observed. Samples with edible coatings showed less variation in soluble solids between the first and last days of storage when compared to that of the control sample. The application of an edible coating of 2% sodium alginate on pears may have slowed down

<table>
<thead>
<tr>
<th>Coating characteristics (1)</th>
<th>ALG2</th>
<th>C05</th>
</tr>
</thead>
<tbody>
<tr>
<td>Thickness (mm)</td>
<td>0.04±0.001&lt;sup&gt;a&lt;/sup&gt;</td>
<td>0.02±0.001b&lt;sup&gt;a&lt;/sup&gt;</td>
</tr>
<tr>
<td>Opacity (%)</td>
<td>5.42±0.03&lt;sup&gt;b&lt;/sup&gt;</td>
<td>6.75±0.02&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>Tensile strength (MPa)</td>
<td>60.96±6.20&lt;sup&gt;b&lt;/sup&gt;</td>
<td>23.42±4.07&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>Elongation (%)</td>
<td>2.49±0.25&lt;sup&gt;a&lt;/sup&gt;</td>
<td>1.10±0.26&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>Elastic modulus (MPa)</td>
<td>37.32±2.81&lt;sup&gt;b&lt;/sup&gt;</td>
<td>21.83±2.65&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>WVP (g/m²Pa.h) × 10&lt;sup&gt;-7&lt;/sup&gt;</td>
<td>3.42±0.12&lt;sup&gt;b&lt;/sup&gt;</td>
<td>4.33±0.10&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
</tbody>
</table>

(1) Same letters in the same line indicate that the samples did not differ significantly (p ≥ 0.05).

| Table 2. pH values of control pear and pears with edible coating of alginate 2% (ALG2) and carrageenan 0.5% (C05) during 15 days of storage. |
|-----------------------------|------|-----|
| Days | Control | pH (1) | ALG2 | C05 |
| 1   | 4.48±0.01<sup>c</sup> | 4.15±0.01<sup>c</sup> | 4.41±0.01<sup>d</sup> |
| 4   | 4.84±0.05<sup>d</sup> | 4.21±0.01<sup>c</sup> | 4.67±0.01<sup>c</sup> |
| 6   | 4.86±0.03<sup>d</sup> | 4.26±0.02<sup>c</sup> | 4.75±0.04<sup>c</sup> |
| 8   | 5.24±0.08<sup>c</sup> | 4.28±0.01<sup>c</sup> | 4.86±0.04<sup>b</sup> |
| 11  | 5.46±0.11<sup>c</sup> | 4.34±0.01<sup>c</sup> | 4.91±0.06<sup>b</sup> |
| 15  | 5.85±0.02<sup>c</sup> | 4.62±0.01<sup>c</sup> | 5.21±0.03<sup>c</sup> |

(1) Same capital letters in the same line and same lowercase letters in the same columns indicate that the samples did not differ significantly (p ≥ 0.05).

![Figure 1. Weight loss of the control pear and pears coated with alginate 2% (ALG2) and carrageenan 0.5% (C05) during 15 days of storage.](image-url)
the metabolic activities of the fruit reducing the degradation of polysaccharides and changes in the soluble solids content.

Similar results were found by Fontes et al. (2008), who studied the application of coatings of alginate on fresh-cut apples stored at 2 °C for 13 days. The author describes that the content of soluble solids in the control sample and samples treated with alginate showed significant differences (p ≤ 0.05) during storage. Bico et al. (2009) observed an increase in soluble solids content of fresh-cut bananas stored at 5 °C for 5 days treated with coating of carrageenan and a preservative solution under controlled atmosphere.

The firmness of fruit is presented in Figure 3. The penetration force values of the control sample decreased significantly (p ≤ 0.05) from the 1st to the 6th day. After this period, no significant difference (p ≥ 0.05) was observed. Samples with an edible coating of 0.5% carrageenan varied significantly (p ≤ 0.05) during the 15 days of storage. The results of penetration force for the samples with alginate 2% did not differ significantly (p ≥ 0.05) during the storage period indicating that the edible coating used maintained the firmness of the stored pears for 15 days.

Olivas, Mattinson and Barbosa-Cánovas (2007) studied sliced apples treated with alginate stored at 5 °C for 10 days. The authors showed that the coating maintained the firmness of the apples during the storage period. According to Larotonda (2007), who analyzed cherries with an edible coating of carrageenan stored at 5 °C for 18 days, the values of firmness of the samples with and without coating were similar indicating that the coating did not influence the strength of the product.

Table 3 shows the results obtained for the color of pears. Externally, the yellowing of the skin is the greatest change that occurs during ripening (VILAS BOAS et al., 2001). Therefore, the color result is an important indication of the shelf life of fruits and vegetables. The maturation stage can be characterized subjectively by the level of skin color, which is an important parameter to predict the shelf life of fruits.

The L, a*, and b* values for the control sample varied significantly (p ≤ 0.05) during the storage period indicating color variation from green to yellow. The control samples

<table>
<thead>
<tr>
<th>Sample</th>
<th>Color</th>
<th>Storage time (days)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>1</td>
</tr>
<tr>
<td>Control</td>
<td>L</td>
<td>74.1 ± 0.85&lt;sup&gt;a&lt;/sup&gt;</td>
</tr>
<tr>
<td>ALG2</td>
<td>a*</td>
<td>−5.5 ± 0.98&lt;sup&gt;d&lt;/sup&gt;</td>
</tr>
<tr>
<td>ALG2</td>
<td>b*</td>
<td>48.3 ± 1.41&lt;sup&gt;c&lt;/sup&gt;</td>
</tr>
<tr>
<td>C05</td>
<td>L</td>
<td>70.1 ± 0.42&lt;sup&gt;a&lt;/sup&gt;</td>
</tr>
<tr>
<td>C05</td>
<td>a*</td>
<td>−2.6 ± 1.13&lt;sup&gt;c&lt;/sup&gt;</td>
</tr>
<tr>
<td>C05</td>
<td>b*</td>
<td>42.1 ± 0.43&lt;sup&gt;c&lt;/sup&gt;</td>
</tr>
</tbody>
</table>

<sup>1</sup>Same capital letters in the same columns for each parameter and same lowercase letters in the same line indicates that the samples did not differ significantly (p ≥ 0.05).
showed greater variation in the parameters $a^*$ and $b^*$ when compared to those of the coated samples; this result justifies the more intense yellow color of the control samples at the end of the storage period. According to Chitarra and Chitarra (2005), chlorophyll is abundant in green products and the loss of this pigment is due to its structural decomposition, mainly by changes in pH, caused by the accumulation of organic acids in vacuoles, the activation of the chlorophyllase enzyme, and the presence of oxidizing systems. The values of $L$, $a^*$, and $b^*$ of the samples with 0.5% carrageenan coating varied significantly ($p \leq 0.05$) during storage. For the sample with 2% alginate, $L$ values decreased significantly ($p \leq 0.05$) during storage. In the alginate samples, the values of $a^*$ and $b^*$ varied significantly ($p \leq 0.05$), but this variation was lower than that observed for the control samples and the samples coated with carrageenan indicating that the coating with 2% alginate showed a better result in maintaining the green color of the fruit. Investigating cherries, Larotonda (2007), the lightness ($L$) values of the control sample and the samples coated with carrageenan tended to decrease during storage under refrigeration. The control sample and the coated fruits showed similar behavior indicating that the application of carrageenan coating did not slow down the senescence of cherries.

4 Conclusions

The application of an edible coating with sodium alginate minimized the weight loss of samples. The results showed that the use of coatings on pears reduced changes in the pH of the fruit. The use of alginate coating showed smaller changes in pH and soluble solids of the pears analyzed. Similar results were obtained for the color of pears. The penetration force results indicated that the alginate coating maintained the firmness of pears during the 15 days of storage at 25 °C. Alginate coating had the lowest water vapor permeability and highest tensile strength. Therefore, it can be used as a protective barrier on fruits. With regard to films made with carrageenan, new experiments must be carried out in order to evaluate the suspension concentrations different from those studied in this research.

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Conservation of Williams pear using edible coating


