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## Quality evaluation of red wines produced from the Isabella and Ives cultivar (*Vitis labrusca*): physicochemical parameters, phenolic composition and antioxidant activity

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### Abstract

The objective of this study was to evaluate the quality of commercial red wines (Isabella and Ives [IsB]; Isabella, Ives, Seibel and Concord [IsSC]; and Isabella and Ives [IsBb]) produced from Isabella and Ives grapes grown in southern Brazil. The parameters required by Brazilian law were evaluated, and the chemical parameters of the phenolic compounds, organic acids, anthocyanins and antioxidant capacity were quantified for red wine samples containing the Isabella and Ives grape. All of the wine parameters were consistent with Brazilian legislation. There were significant differences ( $p < 0.05$ ) among the wines in phenolic compound content, flavonoids, antioxidant activity measured by the oxygen radical absorbance capacity (ORAC) and 2,2-azino-bis(3-ethyl benzothiazoline-6-sulfonic acid) (ABTS) methods, anthocyanin levels and colour parameters. The IsSC wine was the clearest; it had the lowest anthocyanin content (88.76 mg/L) and colour  $a^*$  (11.28) parameter values. Statistical analysis revealed significant correlations between antioxidant activity and some of the identified phenolic compounds. The differences among the wines confirm the distinctive effects of the wine-producing region on each sample and on the varieties that make up each wine.

**Keywords:** American grapes; antioxidants; fermented beverage; phenolic compounds; legislation.

**Practical Application:** Physicochemical parameters, phenolic composition and antioxidant activity in red wine is very important, for directly influences the quality of the wine and its acceptance and rejection of the consumer by drink. It is important to emphasize that these results contribute to enhancing scientific literature on quality wines produced with the cultivar Isabella and Ives.

## 1 Introduction

Production region, climatic and soil variations, enological practices and the use of different grape varieties are factors that affect the characteristics of wines (Lingua et al., 2016). Jackson (2008) reported that the understanding of the chemical nature of grapes and wines that has been gained in recent decades has helped to drive wine production practices toward higher quality and more consistent wines.

The phenolic compounds present in wine are among the most important quality parameters due to their direct influence on sensory characteristics such as colour, flavour, bitterness and astringency (Ivanova-Petropulos et al., 2015). In addition to these properties, phenolic compounds have a beneficial health action, mainly because of their antioxidant, bactericidal and vitaminic properties, which aid in the prevention of cardiovascular diseases and cancer (Garrido & Borges, 2011; Gris et al., 2013; Nixford & Hermosín-Gutiérrez, 2010).

Brazilian wine production uses several grape varieties and is characterised by its diversity. According to current Brazilian legislation, grapes of the *Vitis vinifera* variety are used in the preparation of “viniferous table wines”, called fine table

wine, whereas American grape varieties (*Vitis labrusca* and *Vitis bourquina*) and/or hybrids are used in the preparation of “American table wines”, called ordinary table wine, which are also called “ordinary wine” (Brasil, 2014).

The production and consumption of ordinary wines in Brazil is greater than the production of wines produced with viniferous grapes, and represents 80% of the wines produced in the country. This production results from the high production of *Vitis labrusca* grapes in the South of Brazil, for instance in 2013 were produced 102,744 tons of Ives grape and 232,259 tons of Isabella grape, according the Winemakers Gaucha Association (Biasoto et al., 2014; Brasil, 2014; Machado & Guedes, 2015; Uliana et al., 2015).

Production in the Southern region of Brazil is concentrated on ordinary wines. According to data from the Brazilian Wine Institute (Instituto Brasileiro de Vinhos), production of these wines has reached a volume of 196.07 million litres. Of wine sales in the southern region totalling 206.0 million litres, 177.9 million litres of ordinary red wines were sold (Instituto Brasileiro do Vinho, 2014; Mello, 2003).

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'Isabel' is the main *Vitis labrusca* variety cultivar, followed by Ives variety cultivar, both are used in the development of ordinary wines. It is a red grape that are very rustic and highly fertile (Assis et al., 2011) that proliferated in Brazil due to its easy adaptation to varying weather conditions and high productivity (Castilhos et al., 2016; Rombaldi et al., 2004)

Given its potential for table wine production and the lack of studies on the physicochemical qualities of this wine, the objective of this study was to evaluate the quality of red wines produced from the Isabella cultivar originating from southern Brazil, which has a tradition of producing ordinary wines. The parameters required by Brazilian law were evaluated (Brasil, 2014), and the wine chemical parameters of the phenolic compounds, organic acids, anthocyanins and antioxidant capacity were quantified.

## 2 Materials and methods

### 2.1 Wine samples

The physicochemical quality, phenolic composition, antioxidant capacity and phenolic profiles of three dry red wines made from Isabella grapes from traditional wineries in southern Brazil were evaluated (Table 1). The data were derived from five separate wine collections belonging to the same lot. Tests were performed in triplicate for each collection. According to data provided by the producers, the wines were made with grapes harvested from December to March, which is the harvest period in Serra Gaucha. Fermentation occurs in stainless steel tanks followed by bottling and storage.

### 2.2 Chemicals

All standards and solvents were of HPLC grade with a purity (>99%) and Water was Milli-Q quality. For HPLC analyzes standards were obtained from Sigma-Aldrich, UK (siringic acid, p-coumaric acid and elagic acid), from Sigma-Aldrich, China (p-hydroxybenzoic acid, protocatechuic acid, vanilic acid, trans-cinnamic acid, caffeic acid, rutin), and from Sigam Aldrich, France (catechin, myricetin, kaempferol, quercetina), others were purchased from Sigma-Aldrich, USA (Salicylic acid, Gentisic acid, Ferulic acid, sinapic acid and Resveratrol). The standards used in the analysis of organic acid (tartaric acid, malic acid, succinic acid, lactic acid and acetic acid) by

HPLC, antioxidants and total phenolic were purchased from Sigma-Aldrich, USA (Folin-Ciocalteu, gallic acid, catechin, DPPH-(2,2-diphenyl-1-picrylhydrazyl radical reduction), (ABTS•+) 2,2-azino-bis(3-ethylbenzothiazoline-6-sulfonic acid), TEAC- (6-hydroxy-2,5,7,8-tetramethylchroman-2-carboxylic acid), APPH [2,2'-azobis (2'-metilpropionamidine) dihydrochloride]).

### 2.3 Physicochemical parameters required under Brazilian law

The wines were subjected to analysis to determine the density, alcohol content, total, volatile and fixed acidity, pH, reducing sugars, dry extract, reduced dry extract, ash, ash alkalinity, alcohol in weight/reduced dry extract (RDE) ratio, total sulphur dioxide, free sulphur dioxide, sulphates and chlorides. This was achieved using the physicochemical methods established by the Ministry of Agriculture, Livestock and Supply's (Ministério da Agricultura, Pecuária e Abastecimento - MAPA) current legislation (Brasil, 2005).

### 2.4 Instrumental colour evaluation

Colorimetric parameter measurements were made using a CR400 portable Konica Minolta colorimeter (Osaka, Japan) after calibration with a porcelain plate (CR-A43). The machine was programmed to perform readings considering the standard observer D65 illuminant and 2 (corresponding to daylight). The adopted colour space was the CIELAB system; in this system, L\* represents lightness (0 (black) to 100 (white)); a\* represents a colour coordinate that varies from +a (red) and -a (green), and b\* represents another colour coordinate ranging from +b (yellow) to -b (blue). The coordinate C\*, which represents chromaticity, was also determined, as well as h, the parameter identifying hue.

### 2.5 Spectrophotometric determination of the wines

*Determination of total phenolics, total flavonoids and monomeric anthocyanins*

Total phenolics were measured using the Folin-Ciocalteu method proposed by Roesler (2007). Methanolic extracts of the sample (500 µL) was added with 2.0 mL of 7.5% sodium carbonate and 2.5 mL of aqueous solution of 10 % Folin-Ciocalteu. Subsequently, the mixture was incubated for 5 minutes in a water bath at 50 °C for the colour formation. The absorbance was determined in a spectrophotometer (BECKMAN DU640) at 760 nm, using a blank sample as reference. For quantification of the wine's total phenols, a standard curve (10 to 90 µmL<sup>-1</sup>) prepared with gallic acid (Sigma, Brazil) was constructed and expressed as mg of gallic acid equivalents (GAE) per litre of sample.

The method proposed by Zhishen et al. (1999) was used to determine total flavonoids. The reaction system was formed by 100 µL of samples and 0.15 mL of NaNO<sub>2</sub> waiting reacting for 5 minutes, followed by addition of 0.15 mL of AlCl<sub>3</sub> and reaction time of 6 minutes. Finally it was added 1.0 mL NaOH and 1.2 mL H<sub>2</sub>O. The measure of colour formation was determined by absorbance readings in a spectrophotometer (BECKMAN DU640) at 510 nm. Quantification of total flavonoids of the

**Table 1.** Dry red wine characterisation, production origin and price per bottle.

Region	Codes <sup>1</sup>	Varietals <sup>2</sup>	Geographical coordinates <sup>3</sup>	Price <sup>4</sup>
South	IsB	Isabella, Ives	29° 1' 50" S; 51° 11' 30" W	8.65 (2.20)
	IsSC	Isabella, Ives, Seibel and Concord	29° 1' 50" S; 51° 11' 30" W	7.79 (1.98)
	IsBb	Isabella, Ives	29° 10' 26" S; 51° 31' 7" W	8.45 (2.16)

<sup>1</sup>Encoding of analysed wines; <sup>2</sup>Varietals making up each analysed wine; <sup>3</sup>Geographic coordinates of the cities that produce each wine, expressed as south latitude (S) and west longitude (W); <sup>4</sup>Production price in R\$ and price in US\$ in parentheses (dollar exchange rate R\$ 3.918 on 30/09/2016) for a 750-mL bottle.

sample was performed by standard curve (10 to 150 mg.L<sup>-1</sup>) prepared with catechin (Sigma, Aldrich) at 0.2%, and expressed as catechin equivalents.

Monomeric anthocyanins were determined using the differential pH method (Wrolstad, 2006). Initially, the dilution factor of the sample was determined by diluting the sample with 0.0025M potassium chloride buffer, pH 1.0, until the absorbance of the sample at  $\lambda_{\text{vis-max}}$  (510 nm) reached a linear range (less than 1.2) in the spectrophotometer readings of absorbance. The dilution value (DF) was obtained by dividing the final sample volume by the initial volume. Aiming to do not exceed the buffer capacity, the sample was kept in volume less than 20% of the total volume. Two dilution of each sample were prepared, one containing 0.025M sodium chloride and the other sodium acetate buffer 0.4M at pH 4.5, the pH was adjusted with concentrated hydrochloric acid. Thereafter, each sample was diluted in buffer and diluting each of the predetermined dilution factor; following they were leaved to stand until the dilutions come to equilibrium. Readings were taken in a spectrophotometer (UV Microproce 0798U, Quimis, São Paulo, Brazil) at wavelengths of 510 nm and 700 nm, against a ultra-pure water (Equation 1).

$$A = (A_{\lambda_{\text{vis-max}}} - A_{700})_{\text{pH } 1.0} - (A_{\lambda_{\text{vis-max}}} - A_{700})_{\text{pH } 4.5} \quad (1)$$

The concentration of monomeric anthocyanins in the original sample was calculated using Equation 2, where MW is the molecular weight of cyanidin-3-glucoside (MW = 449.2), DF is the dilution factor and  $\epsilon$  is the molar absorptivity (26900 mol/L).

$$\text{Monomeric anthocyanin (mg/L)} = (A \times \text{MW} \times \text{DF} \times 1000) / (\epsilon \times l) \quad (2)$$

#### Antioxidant activity

The DPPH tests (2,2-diphenyl-1-picrylhydrazyl radical reduction) were performed according to Roesler (2007) with some modifications. Preparation of A solution of DPPH • (0.004% w/v) with a range of between 0.8 and 1.2 absorbance at 517 nm and from which the solution was prepared daily and stored in a refrigerator until the moment of use. Sample preparation took place using methanol at a concentration of 10 µL.mL<sup>-1</sup>. An aliquot of 200 µL was transferred to a tube and added with 1000 µL of DPPH solution, stirred, and after 30 minutes the amount of DPPH was recorded in UV-Visible spectrometer (UV Microproce 0798U, Quimis, São Paulo, Brazil) at a wavelength of 517 nm. The results were expressed in Trolox equivalent, where the standard curve was determined by the same procedure for the sample, using readings obtained from the calibration curve of Trolox curve:  $A = 0.4335 C + 0.4532$ , and  $r^2$  correlation coefficient: 0.9954, where  $a$  is the absorbance at 517 nm and  $C$  is the concentration of Trolox (6.25 to 250 mg mL<sup>-1</sup>).

The Trolox equivalent antioxidant capacity (TEAC) - 2,2-azino-bis(3-ethylbenzothiazoline-6-sulfonic acid) radical capture (ABTS•+) was determined according to the method described by Re et al. (1999). The sample was prepared in distilled water at a concentration of 10 µL/mL. Through testing with the ABTS • +, obtained by reacting 5 ml of ABTS (7 mM) with 88µL potassium persulfate 140mM (final concentration of 2.45 mM), where the system was kept to stand at room temperature for a period of 12 to 16 hours in absence of light. Subsequently, the

sample absorbance was reading at 734 nm after the reaction mixture containing 200 µL sample and 1000 uL ABTS • + solution, leaving to rest for 6 minutes. Reading the prepared blank was performed according to the procedure described, without the addition of the sample. It was built the Trolox calibration curve (10 -250 mM) and is plotted as the percentage (%) of inhibition versus concentration of Trolox:  $A = 2.1824 \cdot C - 4.1649$ , and the linear correlation coefficient  $r^2 = 0.9904$ , where  $a$  is the absorbance at 734 nm and  $C$  is the concentration of Trolox. The result of the activity of the tested compounds was expressed in TEAC values, defined in micromol equivalents Trolox.g-1.

The oxygen radical absorbance capacity (ORAC) test procedures were based on the methods of Prior et al. (2003) and Dávalos et al. (2004). AAPH (2,2'-azobis(2-amidinopropane) dihydrochloride) was used as a peroxy radical generator and Trolox as a standard. All readings were taken using a NOVOstar (Labtech® BMG, Offenburg, Germany) microplate reader (96 wells) together with the *MARS Data Analysis* software version 1.3 (BMG Labtech®, Offenburg, Germany). Fluorescence intensity (485 nmEx/520 nmEm) was checked at every 60-second cycle for 80 cycles in the microplate reader. The same procedure was adopted for the reference standards. Stock solutions of each sample were prepared at a concentration of 10 mg/ml (w/v) with 75 mM potassium phosphate buffer, pH 7.4, they were called "mother solution". The final ORAC value was calculated based on the area of liquid under the fluorescence decay curve and expressed in µmol TE L<sup>-1</sup>.

#### 2.6 Chromatographic analysis of the wines

##### Quantification of the class of phenolic compounds

For better identification of phenolic compounds (phenolic acids, flavonóis compounds and resveratrol), prior extraction was performed according to Malovaná et al. (2001). The phenolic compounds were separated and quantified in an LC-20 AT high-performance liquid chromatography (HPLC) system (Shimadzu Corporation, Japan) equipped with a Vydac 218TP C18 column (25 cm x 4.6 mm x 5µm) and diode array detector. The samples were eluted in a gradient system consisting of solvent A (2% v/v acetic acid) and solvent B (acetonitrile:methanol 2:1 v/v). The gradient system progressed from 90% A at 0 min to 80% A at 10 min, 70% A at 15 min, 60% A at 25 min, 50% A at 30-40 min, 75% A at 42 min and 90% A at 44 min. The flow rate was 1 ml/min, the column temperature was maintained at 25° C, and the sample injection volume was 20 µL. The phenolic compound peaks were monitored at 280 nm (Prasad et al., 2009). The identification and quantification of compounds was given by injecting standard curves at concentrations of 1; 0.5; 0.25; 0.1; 0.05; 0.04; 0.03; 0.02; 0.01 mg. mL<sup>-1</sup>.

##### Analysis of organic acids

The organic acids present in the wine were determined by injecting 20 µL of the filtered sample into a Waters 2690 HPLC system (Varian, California, USA) equipped with a binary solvent system, a Rheodyne valve (20 µL capacity) coupled with an Agilent Hi-Plex H column (7.7 x 300 mm, 8 µ) at 65° C, a diode array detector (Varian 330) set at 220 to 275 nm wavelength, a



pump system with high pressure gradient setting (Varian 230) and Galaxie Chromatography Data System processing software. The mobile phase used was 0.009 M sulphuric acid at a flow rate of 0.7 ml/min. The run duration was 30 minutes (Zeppa et al., 2001). Organic acids (tartaric, malic, acetic, lactic and succinic acids) were quantified by injecting a standard curve.

## 2.7 Statistical analysis

The data were analysed statistically using analysis of variance (ANOVA) and Tukey's means test at a 5% significance level, using SAS 9.3 (SAS Institute Inc., 2011). The data were also subjected to Pearson correlation analysis using GraphPad Prism 6.0. Principal Component Analysis (PCA) was applied to the physicochemical parameters, phenolic compound and antioxidant datasets using MVSP 3.13 (Kovach Computing Services, 2006).

## 3 Results and discussion

### 3.1 Physicochemical quality of red wines produced from *Isabella* grapes

The wines produced in the southern region (Isabella wine and Ives [IsB]-South, Isabella wine, Ives, Seibel and Concord [IsSC]-South and Isabella and Ives [IsBb]-South) were in accordance with the limits recommended by the Brazilian legislation for wines (Brasil, 2014; Table 2). The wines differed significantly ( $p < 0.05$ ) in alcohol content, pH, total and fixed acidity, reduced dry extract, reducing sugars, ash and ash alkalinity. It may be assumed that the differences in these parameters among the wines are caused by differences in the wine-making processes and by the use of different grape varieties for wine production in the southern region (Castilhos & Del Bianchi, 2012).

Differences observed between table wines related to the full and fixed acidities, can be explained by the fact that wines were not produced by just one type of grape, so the formation of the characteristic acids present in the fixed acidity resulted from the mixture of different acids, such as tartaric, malic, succinic, etc.

So, as the winemaking process varies from winery to winery, this fact has a direct influence on the production of these acids, promoting the statistical difference observed in the results (Castilhos & Del Bianchi, 2011).

In addition, the results of the dry extract ratios were expected, dry wine with a dry extract content of less than 20 g/L tend to present a mild taste; however, dry wine with 30g/l or more is considered to a bond taste. In this context, the table wines analyzed showed dry matter content of less than 30 g/L, thus suggesting that the high quantity of dry matter resulted from the largest solubilization of solid substances of the grape during the maceration stage and the stabilization period of the wines (Castilhos & Del Bianchi, 2011).

The reducing sugar content (4.00 g/L) found in IsB wine, the maximum amount permitted by law, probably resulted from the addition of sucrose during the chaptalisation stage (Oliveira et al., 2011).

With regard to  $\text{SO}_2$  levels, the wines did not exceed the maximum allowable amount of 0.35 g/L dictated by the law.  $\text{SO}_2$  is used as a preservative, and it acts as an antimicrobial and antioxidant agent in wine production. Reports suggest that it may be associated with allergic reactions in some consumers (Danilewics, 2007).

### 3.2 Phenolic compounds, flavonoids, anthocyanins and antioxidant capacity of red wines derived from *Isabella* grapes

Table 3 shows the concentrations of total phenolic compounds, total flavonoids, monomeric anthocyanins and antioxidant capacity of the red table wines.

The total phenolic content of the wines ranged from 830.51 to 846.90 mg GAE.L<sup>-1</sup>, values that are in the range reported by Amerine & Ough (1980) for red wines. The phenolic composition of wine depends on the type of grape used for wine

**Table 2.** Physicochemical characterisation of red wines produced from *Isabella* grapes (parameters required by Brazilian law) (Brasil, 2014).

Parameters	IsB	IsSC	IsBb	Legislation
Density at 20°C	0.9962±0.00 <sup>a</sup>	0.9935±0.00 <sup>a</sup>	0.9943±0.00 <sup>a</sup>	
Alcohol content 20°C	10.60±0.06 <sup>a</sup>	10.10±0.06 <sup>b</sup>	10.10±0.06 <sup>b</sup>	10 to 14°GL
pH	3.75±0.04 <sup>b</sup>	3.56±0.01 <sup>c</sup>	3.99±0.00 <sup>a</sup>	
Total acidity (meq.L <sup>-1</sup> )	79.33±0.58 <sup>b</sup>	70.00±1.00 <sup>c</sup>	88.00±1.00 <sup>a</sup>	40 to 130
Volatile acidity (meq.L <sup>-1</sup> )	16.67±1.53 <sup>a</sup>	15.33±1.53 <sup>a</sup>	17.00±1.73 <sup>a</sup>	Max 120
Fixed acidity (meq.L <sup>-1</sup> )	62.00±1.15 <sup>b</sup>	56.00±2.31 <sup>c</sup>	69.00±0.58 <sup>a</sup>	
Dry extract (g.L <sup>-1</sup> )	27.31±1.61 <sup>a</sup>	26.14±0.15 <sup>a</sup>	28.80±0.47 <sup>a</sup>	
RDE (g.L <sup>-1</sup> )	24.27±1.58 <sup>ab</sup>	23.37±0.18 <sup>c</sup>	27.36±1.47 <sup>a</sup>	
RDE/Alcohol content	2.88±0.02 <sup>a</sup>	2.19±0.15 <sup>b</sup>	2.65±0.17 <sup>a</sup>	Max 4.8
Reducing sugars (g.L <sup>-1</sup> )	4.00±0.03 <sup>a</sup>	3.77±0.01 <sup>b</sup>	2.44±0.02 <sup>c</sup>	Max 4.0
Ash (g.L <sup>-1</sup> )	2.13±0.07 <sup>c</sup>	2.59±0.09 <sup>a</sup>	2.39±0.08 <sup>b</sup>	Min 1.5
Ash alkalinity (g.L <sup>-1</sup> )	0.02±0.23 <sup>c</sup>	0.04±0.46 <sup>a</sup>	0.03±0.69 <sup>b</sup>	Max 1.0
Sulphates (g.L <sup>-1</sup> )*	<1.0	<1.0	<1.0	Max 1.2
Free sulphur dioxide (mg.L <sup>-1</sup> )	22.19±0.74 <sup>a</sup>	19.2±1.28 <sup>a, b</sup>	17.07±1.96 <sup>b</sup>	
Total sulphur dioxide (g.L <sup>-1</sup> )	0.07±2.41 <sup>a</sup>	0.06±1.21 <sup>b</sup>	0.07±1.21 <sup>a</sup>	Max 0.35
Chlorides (g.L <sup>-1</sup> )	0.02±3.38 <sup>a</sup>	0.02±3.38 <sup>a</sup>	0.01±5.85 <sup>a</sup>	Max 0.20

Mean ± standard deviation followed by the same letters in the same rows do not differ at 5% according to Tukey's test. \*Semi-quantitative method – Marty's approximate test. RDE: Reduced dry extract; IsB (Isabella and Ives); IsSC (Isabella, Ives, Seibel and Concord); IsBb (Isabella and Ives).

**Table 3.** Quantification of total phenolic compounds, total flavonoids, anthocyanins and antioxidant activity by spectrophotometric methods and quantification of profile phenolic compounds and organic acids by chromatographic methods in red wines produced from the Isabella and Ives cultivar.

Compound		IsB <sup>1</sup>	IsSC <sup>1</sup>	IsBb <sup>1</sup>
Total phenolics <sup>2</sup>		837.28±0.15 <sup>b</sup>	846.90±1.88 <sup>a</sup>	830.51±3.52 <sup>c</sup>
Total flavonoids <sup>3</sup>		824.92±9.35 <sup>a</sup>	453.49±11.47 <sup>c</sup>	590.79±11.48 <sup>b</sup>
Anthocyanins <sup>4</sup>		196.07±5.54 <sup>b</sup>	88.76±6.67 <sup>c</sup>	258.88±8.84 <sup>a</sup>
Compound	Class			
Protocatechuic acid	Phenolic acid <sup>5</sup>	35.00±4.50 <sup>b</sup>	36.50±1.55 <sup>b</sup>	52.20±5.80 <sup>a</sup>
<i>p</i> -hydroxybenzoic acid	Phenolic acid	34.67±3.51 <sup>a</sup>	36.50±2.04 <sup>a</sup>	34.00±4.00 <sup>a</sup>
Salicylic acid	Phenolic acid	4.77±1.16 <sup>a</sup>	3.44±0.47 <sup>b</sup>	4.80±0.45 <sup>a</sup>
Syringic acid	Phenolic acid	0.74±0.05 <sup>b</sup>	1.43±0.30 <sup>a</sup>	1.31±0.07 <sup>a</sup>
Gentisic acid	Phenolic acid	72.86±3.25 <sup>b</sup>	111.33±7.41 <sup>a</sup>	76.77±1.51 <sup>b</sup>
Vanillic acid	Phenolic acid	nd	1.60±0.08 <sup>a</sup>	1.70±0.10 <sup>a</sup>
<i>p</i> -coumaric acid	Phenolic acid	3.84±0.99 <sup>b</sup>	14.22±2.84 <sup>a</sup>	5.14±0.92 <sup>b</sup>
Sinapic acid	Phenolic acid	1.48±0.29 <sup>a</sup>	2.00±0.57 <sup>a</sup>	1.51±0.05 <sup>a</sup>
Trans-cinnamic acid	Phenolic acid	tr	tr	tr
Caffeic acid	Phenolic acid	1.00±0.35 <sup>b</sup>	3.32±0.49 <sup>a</sup>	1.29±0.03 <sup>b</sup>
Ferulic acid	Phenolic acid	2.50±0.40 <sup>a</sup>	3.70±0.62 <sup>a</sup>	3.38±0.45 <sup>a</sup>
Ellagic acid	Phenolic acid	0.20±0.01 <sup>b</sup>	0.22±0.01 <sup>b</sup>	0.34±0.03 <sup>a</sup>
Rutin	Flavonoid	3.03±0.81 <sup>b</sup>	6.18±0.60 <sup>a</sup>	1.80±0.11 <sup>b</sup>
Kaempferol	Flavonoid	1.38±0.29 <sup>a</sup>	0.64±0.02 <sup>b</sup>	0.86±0.08 <sup>b</sup>
Myricetin	Flavonoid	18.95±9.45 <sup>b</sup>	45.50±0.41 <sup>a</sup>	49.00±1.00 <sup>a</sup>
Quercetin	Flavonoid	11.82±1.54 <sup>a</sup>	8.15±0.20 <sup>b</sup>	11.70±0.80 <sup>a</sup>
Catechin	Flavonoid	236.03±1.27 <sup>a</sup>	145.17±3.12 <sup>c</sup>	216.50±2.50 <sup>b</sup>
Resveratrol	Stilbene	<b>6.70±0.26<sup>a</sup></b>	<b>2.30±0.65<sup>b</sup></b>	6.67±0.91 <sup>a</sup>
Tartaric acid	Organic acid <sup>6</sup>	<b>1.80±0.01<sup>a</sup></b>	<b>1.20±0.01<sup>b</sup></b>	<b>2.90±0.01<sup>a</sup></b>
Malic acid	Organic acid	<b>0.60±0.02<sup>a</sup></b>	nd	<b>0.30±0.02<sup>a,b</sup></b>
Succinic acid	Organic acid	<b>1.50±0.02<sup>a</sup></b>	nd	<b>1.60±0.03<sup>a</sup></b>
Lactic acid	Organic acid	<b>1.10±0.01<sup>a</sup></b>	<b>1.20±0.01<sup>a</sup></b>	<b>1.50±0.01<sup>a</sup></b>
Acetic acid	Organic acid	nd	nd	nd
Antioxidant Activity <sup>7</sup>				
ABTS		629.41±0.01 <sup>b</sup>	1420.47±0.01 <sup>a</sup>	1419.07±0.01 <sup>a</sup>
DPPH		40.78±1.56 <sup>a</sup>	44.57±0.07 <sup>a</sup>	<b>45.60±0.09<sup>a</sup></b>
ORAC		39159.72±1.52 <sup>a</sup>	30882.33±1.51 <sup>b</sup>	38411.91±3.00 <sup>a</sup>

<sup>1</sup>IsB (Isabella and Ives); IsSC (Isabella, Ives, Seibel and Concord); IsBb (Isabella and Ives); <sup>2</sup>Quantified as mg catechin L<sup>-1</sup>; <sup>3</sup>Quantified as mg L<sup>-1</sup>; <sup>4</sup>and <sup>5</sup>Quantified as mg L<sup>-1</sup>; <sup>6</sup>Quantified as g L<sup>-1</sup>; <sup>7</sup>The results of antioxidant activity measurements are expressed as μM TE<sup>+</sup>.mL<sup>-1</sup> (Trolox equivalent). Mean ± standard deviation followed by the same letters in the same rows do not differ at 5% according to Tukey's test; nd: compound not detected; tr: traces of the compound.

making, on juice extraction and wine production processes and on the chemical reactions that occur during aging of the wine (Granato et al., 2011).

Increased levels of total flavonoids were found in the IsSC wine (824.92 mg CAT.L<sup>-1</sup>), higher ( $p < 0.05$ ) than those of the other wines analysed. Flavonoids are responsible for a wine's flavour and astringency (Abe et al., 2007; Ali et al., 2010).

The reduced anthocyanin content of the IsSC wine (88.76 mg.L<sup>-1</sup>) is probably due to the different grape varieties used to prepare the wine. According to Gris et al. (2013), anthocyanins are effective antioxidants, and the anthocyanin composition of grapes varies according to species, cultivar, ripeness, seasonal conditions and solar radiation levels. Cabrita et al. (2003) found similar values in red wine produced with Trincadeira grapes, i.e. total phenolics content of 1568.0 mg EC. L<sup>-1</sup>, total flavonoid content of 1535.0 EC mg. L<sup>-1</sup>, and anthocyanins content of 773 mg.L<sup>-1</sup>.

Resveratrol is often associated with antioxidant reactions, and it was found in the Isabella grape red wines in concentrations ranging from 2.30 mg.L<sup>-1</sup> (IsSC) to 6.70 mg.L<sup>-1</sup> (IsB). Resveratrol is synthesised in the grape skin; in wines, typical resveratrol values range from 0.1 to 5.65 mg.L<sup>-1</sup> (Abe et al., 2007).

Trans-cinnamic acid was identified among the phenolic acids in the wines, but it was present only in trace amounts. Vanillic acid was not identified in the IsB wine. Protocatechuic acid (10.51 to 52.20 mg.L<sup>-1</sup>), *p*-hydroxybenzoic acid (25.40 to 36.50 mg.L<sup>-1</sup>) and gentisic acid (49.62-111.33 mg.L<sup>-1</sup>) were found in higher concentrations in the wines analysed than the remaining phenolic acids. Other acids such as *p*-coumaric (-14.22 3.84 mg.L<sup>-1</sup>), caffeic acid (1.00 to 3.32 mg.L<sup>-1</sup>), vanillin (1.60-1,70 mg.L<sup>-1</sup>) and ferulic (2.50 to 3.70 mg.L<sup>-1</sup>) are under study in red wines from different grape varieties (Granato et al., 2011).

Phenolic acids, which are derived from benzoic and cinnamic acids, play an important role in the sensory characteristics of wines. Hydroxycinnamic acids and their tartaric esters are

involved in the browning reactions of must and wine; that is, they affect the colour of wine and are precursors to volatile phenols (Gris et al., 2013).

Among the flavonoid class, catechin stood out due to its higher concentration and significant differences among wines, with values varying from 145.17 mg.L<sup>-1</sup> in the IsSC wine to 236.03 mg.L<sup>-1</sup> in the IsB wine. For other compounds of the class of flavonoids (kaempferol myricetin, quercetin, rutin) also are in agreement with results reported in the study developed by Granato et al. (2011). Individual flavonoid compounds such as rutin, quercetin, myricetin and kaempferol, compounds identified in the wines in this study, are known for their antioxidant activities (Sagratini et al., 2012).

No malic, succinic or acetic organic acids were detected in the IsSC wine. The presence of these acids in various concentrations in the wines and their absence from one of the tested wines may have been due to the use of different wine production techniques and the different grape varieties used.

Tartaric acid is specific to grapes and wines, and its concentration in wine varies from 2 to 8 g L<sup>-1</sup> (Jackson, 2000). It is a strong acid that directly affects the wine's pH. Malic acid is considered a weak acid and is poorly resistant to oxidation and high temperatures (Kliewer et al., 1967). Lactic acid is formed during malolactic fermentation, with resulting concentrations ranging from 1 to 3 g.L<sup>-1</sup> (Lima et al., 2010).

Acetic acid contributes to the wine's volatile fraction; it was not detected in the tested samples, indicating good control of the fermentation process. During alcoholic fermentation, small amounts of acetic acid, ranging from 0.2 to 0.4 g.L<sup>-1</sup>, are produced (Amerine et al., 1967; Zoecklein et al., 1994).

Acidity defines the structure and balance of wine; it is dependent on the concentration of acids such as tartaric and malic acids and to a lesser extent on the concentration of lactic and acetic acid (Lima et al., 2010). Rizzon et al. (2000) reported that the skin of the Isabella grape yields a higher concentration of organic acids than that of *Vitis vinifera* cultivars, possibly due to release of the skin's organic acids during maceration.

The highest concentration of phenolic compounds was found in the IsSC wine, whereas the highest concentration of flavonoids was found in the IsB wine, and the highest concentration of anthocyanins was found in the IsBb wine. In general, the levels of phenolic compounds, flavonoids and anthocyanins were reflected in the determination of antioxidant capacity using the ABTS and ORAC methods, with variations of 629.41 to 1420.47 µM TE. mL<sup>-1</sup> and 30,882.33 to 39,159.72 µM TE. mL<sup>-1</sup>, respectively. However, there was no significant difference in the measured amounts of DPPH radicals. Nixford & Hermosín-Gutiérrez (2010) evaluated the antioxidant activity of Isabella wines using the DPPH method and found antioxidant capacities of 26-63 µM TE. mL<sup>-1</sup>, values that are consistent with the results of this study.

### 3.3 Colour evaluation

Table 4 shows the CIELAB colour results of the red wines made from Isabella grapes. All of the measured parameters showed significant differences in the IsSC wine and the IsB and IsBb wines.

The IsSC wine had the highest L\* value (24.73) and therefore was the clearest wine. The IsSC wine also had the lowest values for the a\*, C\* and h parameters; that is, it was characterised as being less red and having lower saturation and hue. In contrast, it had a greater mean b\* (yellow) value, which can impart a slightly orange colour. These values reflect the reduced concentrations of anthocyanins and total flavonoids, which are the molecules responsible for wine colour.

Romero-Cascales et al. (2012) found that the maceration process increases the concentration of anthocyanins in wine. The fact that the IsSC wine has a less intense and less reddish colour than the other wines from the southern region may be explained by differences in the wine production process, as wine colour also depends on other factors such as grape variety. The Isabella grape has less colouring than is desirable for wines and often needs to be cut with other species that confer colour (Camargo et al., 2011).

### 3.4 Statistical analysis

Pearson correlation analysis revealed significant correlations ( $p < 0.05$ ) between some of the red wine parameters analysed in this study.

A strong positive correlation was observed between pH and anthocyanins ( $r = 0.99$ ), total acidity ( $r = 1.0$ ) and dry extract ( $r = 0.99$ ), indicating the effect of this parameter on the pigment concentration in the wines and on acidity.

There were also correlations with antioxidant activity measured by the ORAC method and gentisic acid ( $r = 0.99$ ), sinapic acid ( $r = 0.99$ ) and caffeic acid ( $r = 0.99$ ). Rice-Evans et al. (1996) found that phenolic acids also contribute to antioxidant activity in wine and that this effect depends mainly on the number of hydroxyl groups present in the molecules. The correlations between antioxidant activity, total phenolic compounds, total flavonoids, monomeric anthocyanins and other phenolic acids, flavonoids and resveratrol were weak and not significant ( $p > 0.05$ ).

In contrast to the findings of this study, Granato et al. (2011) found that compounds such as gallic acid, myricetin and quercetin, along with anthocyanins, were responsible for the differences in antioxidant activity among South American red wines. In another study conducted by Granato et al. (2010), it was found that non-flavonoids were the main compounds responsible for the antioxidant activity in Brazilian red wines.

**Table 4.** CIELAB colorimetric parameters in red wines produced from the Isabella cultivar.

Parameters	IsB	IsSC	IsBb
L*	21.9±0.52 <sup>b</sup>	24.7±0.54 <sup>a</sup>	20.4±0.39 <sup>b</sup>
a*	19.0±1.52 <sup>a</sup>	11.3±1.25 <sup>b</sup>	23.1±1.32 <sup>a</sup>
b*	2.1±0.40 <sup>b</sup>	7.5±0.76 <sup>a</sup>	2.2±0.31 <sup>b</sup>
C*	26.4±1.47 <sup>a</sup>	11.7±5.86 <sup>b</sup>	21.9±0.43 <sup>a</sup>
h	40.5±0.36 <sup>a</sup>	17.9±2.24 <sup>b</sup>	41.2±0.35 <sup>a</sup>

experimental n = 6

Mean ± standard deviation followed by the same letters in the same row do not differ significantly at 5% according to Tukey's test. L\* (lightness); a\* (red-green component); b\* (blue-yellow component); C\* (chromaticity); h (hue angle).

Significant and negative correlations were found between sinapic acid and resveratrol content ( $r = -0.99$ ) and between quercetin and gentisic ( $r = -0.99$ ) and sinapic acid ( $r = -0.99$ ), indicating that these compounds may affect each other's antioxidant activity.

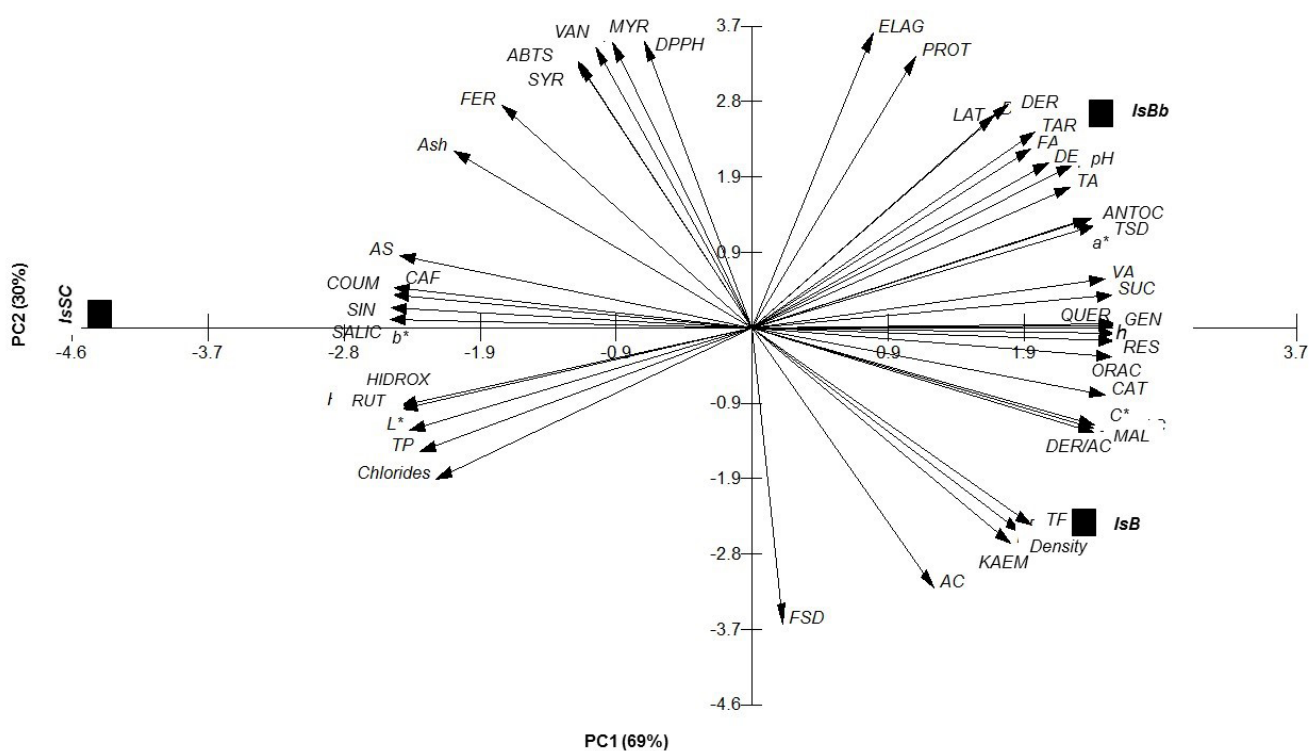
Principal component analysis was performed to evaluate the effects of the substances found in the studied wines (Figure 1) because it is believed that the potential of each wine depends on these substances and in particular that the antioxidant capacity of red wine is related to its content of phenolic compounds. Phenolic content, in turn, is thought to depend on the composition of nitrogenous compounds, the alcohol content, the composition of volatile and fixed acids and other parameters.

The two principal components of the PCA graph accounted for approximately 90% of the variance. Similar wines occupy nearby regions in the chart and are characterized by the vectors (parameters and compounds) that appear closer to them. It can be seen that the wines from the southern region are located in different regions of the chart. The IsB and ISBB samples presented similarity regarding the variables studied; this probably resulted

from the fact that the wines are made with the same types of grapes.

Certain parameters such as pH,  $a^*$ , total acidity, total anthocyanins and the amounts of some phenolic compounds such as ellagic, protocatechuic, lactic and tartaric acids and quercetin most affected the quality of the IsBb wine and most differentiated it from the others. Other compounds such as resveratrol, malic acid, kaempferol and catechin were important in differentiating the IsB wine from the others, and these phenolic compounds may have produced a greater antioxidant activity value in the wine according to the ORAC method. Compounds such as myricetin, rutin, syringic, ferulic, hydroxybenzoic, caffeic and vanillic acids were the variables that made the highest contributions to the IsSC wine, and these phenolic compounds probably influenced the antioxidant activity values measured for this wine using the DPPH and ABTS methods.

According to the PCA graph in general, the association between phenolic compounds and antioxidant activity found in this study was to be expected because phenolic compounds are the main components responsible for the antioxidant activity



**Figure 1.** Projection of the physical parameters, physicochemical parameters, antioxidant activity, phenolic compound and organic acids content, and colour in red wine samples produced with Isabella grapes in the first two principal components. Wines: IsB (Isabella); IsSC (Isabella, Ives, Seibel and Concord); IsBb (Isabella and Ives). Parameters: ACE: Acetic acid; AC: Alcohol content; AS: Ash alkalinity;  $a^*$ : red-green component; ANTOC: Anthocyanins; ANT: Antioxidant activity;  $b^*$ : blue-yellow component;  $C^*$ : chromaticity; CAF: Caffeic acid; CAT: Catechin; CINAN: Trans-cinnamic acid; COUM: *p*-coumaric; DE: Dry extract; DER: Reduced dry extract; ELAG: Ellagic acid; FER: Ferulic acid; FA: Fixed acidity; FSD: Free of sulphur dioxide; GEN: Gentisic acid;  $h$ : hue angle; HIDROX: *p*-hydroxybenzoic acid; KAEM: Kaempferol;  $L^*$ : lightness; LAT: Lactic acid; MAL: Malic acid; MYR: Myricetin; pH: hydrogeonic potential; PROT: Protocatechuic acid; QUER: Quercetin; RES: Resveratrol; RS: Reducing sugars; RUT: Rutin; SALIC: Salicylic acid; SIN: Sinapic acid; SUC: Succinic acid; SYR: Syringic acid; TAR: Tartaric acid; TA: Total acidity; TF: Total flavonoids; TP: Total phenolics; TSD: Total sulphur dioxide; VAN: Vanillic acid; VA: Volatile acidity; DPPH: 2,2-diphenyl-1-picrylhydrazyl, ABTS: 2,2'-azino-bis(3-ethylbenzothiazoline-6-sulphonic acid), DER/AC: Reduced dry extract/Alcohol content, ORAC: Oxygen radical absorbance capacity.



in red wine. A correlation or association between antioxidant activity and phenolic compounds has been found in many studies (Alén-Ruiz et al., 2009; Di Majo et al., 2008; Ferreira et al., 2010; Gris et al., 2011a, b). The antioxidant properties of phenolic compounds are probably primarily due to their chemical structures. However, physicochemical parameters and other substances present in the wine participate in the formation and stability of phenolics and in the stability and quality of the wine and may be important in the differentiation of wines.

## 4 Conclusion

The wines analysed in this study were produced in the Southern region of Brazil, a traditional red wine-producing region, had concentrations of phenolic compounds, flavonoids and anthocyanins similar to those reported in other studies. These values resulted in a higher antioxidant capacity of these wines as assessed using the ABTS and ORAC methods. The results clearly demonstrate the potential of wine produced from Isabella grapes. The observed variations in the physicochemical composition, phenolic composition and antioxidant activities of these wines are related to differences due to the use of different wine production conditions and especially to the grape variety used in the preparation of each wine.

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