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HS-SPME optimization and extraction of volatile compounds from soursop (*Annona muricata* L.) pulp with emphasis on their characteristic impact compounds

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Abstract

Aroma and taste are decisive factors in the selection of any food. The aim of this study was to extract the volatile compounds present in soursop (*Annona muricata* L.) pulp by Solid-phase microextraction (SPME) technique using 3 different fibers (DVB/CAR/ PDMS, CAR/PDMS and PDMS/DVB). An experimental design was set up to evaluate the best extraction conditions wherein the variables were adsorption temperature, ionic strength and pulp concentration. The separation of volatiles was performed in chromatographic columns of different polarity (polar and non-polar) while volatile compounds were identified by analysis in high resolution gas chromatography system coupled with mass spectrometry. The results obtained using 3 different fibers revealed the capture of about 40 compounds. The CAR/PDMS fiber was more efficient for the capture of esters and DVB/CAR/PDMS fiber for terpenes. The optimum conditions for capture of higher number of volatiles for polar column were 45 °C for extraction, 15% of ionic strength and 50% of pulp concentration which resulted in separation of 87 compounds. Among the principal character impact compounds from soursop are (*E*)-2-hexenoate, methyl hexenoate and linalool.

Keywords: *Annona muricata*; flavor; SPME; tropical fruits; volatile compounds.

Practical Application: The innovative nature of this work is that we have optimized the SPME conditions so as to capture a higher number of volatile compounds present in soursop pulp. There is no published work as yet on optimization of SPME conditions for soursop pulp. Thus if we need to monitor the aroma quality of soursop pulp, this work suggests the best conditions which should be followed for determination of volatile profile of this fruit.

1 Introduction

Tropical and subtropical fruits usually possess strong and pleasant aroma. Several studies have reported the analysis of volatile constituents of soursop fruit pulp (MacLeod & Pieris, 1981; Franco & Rodriguez-Amaya, 1983; Iwaoka et al., 1993; Wong & Khoo, 1993; Pelissier et al., 1994; Jirovetz et al., 1998; Augusto et al., 2000; Cheong et al., 2010, 2011; Cardozo et al., 2011, 2013). Solid-phase microextraction (SPME) technique was employed to capture volatile components from custard apple (*Annona reticulata* L.) pulp by Augusto et al. (2000) and they reported the presence of 21 volatile compounds mostly being esters.

Cheong et al. (2010) detected 118 volatile compounds in soursop using the CAR/PDMS fiber among which only 35 compounds were identified. In a later work, Cheong et al. (2011) reported the presence of 135 volatile compounds using the same coating of CAR/PDMS, however, only 37 compounds were identified from soursop. The main compounds detected in these studies were: methyl butanoate, methyl hexanoate, methyl (*E*)-2-butenate and methyl (*E*)-2-hexenoate (Cheong et al., 2011).

Cardozo et al. (2011) evaluated the performance on capture of volatile compounds from soursop in two types of columns: the RTX-5 (non-polar column) in which they could

identify 21 volatile compounds while in FFAP (polar column), 27 compounds were identified with the use DVB/CAR/PDMS fiber. In a later work (Cardozo et al., 2013) undertaken with the use of RTX-5 column, the authors reported the presence of 28 volatile compounds in soursop in which the main compounds were ethyl acetate, 2-methyl-4-pentanal and 3-hexenol.

SPME analysis allows a fast and very attractive extraction technique being easy to perform and it has an added advantage of direct application of analyte on to the GC column through thermal desorption whereby sample injection remains free of solvents and contaminants (Kozziel & Novak, 2002; Vesely et al., 2003; Deng et al., 2014). Additionally, SPME provides speed and practicality on the analysis of authentic chemical constituents of fruit flavor (Kataoka et al., 2000).

It could be concluded from the published articles on soursop that there is very little work done in identifying the volatile compounds obtained from SPME of soursop fruit pulp. Thus, this study was designed to perform various experiments varying the extraction parameters such as use of different coatings of fiber, adsorption temperature, ionic strength and concentration of the pulp sample in order to obtain a large number of volatile compounds in extracts as well as to evaluate their effects on

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capture of volatile compounds from soursop pulp so that the analytical conditions could be optimized for volatile constituents determination in soursop fruit pulp, having an emphasis of character impact compounds.

2 Materials and methods

2.1 Fruit and chemical standards

Soursop (*Annona muricata* L.) fruits were purchased from the Central Market, situated in the city of Aracaju in the northeast region of Brazil. At the laboratory, the fruits were washed with chlorinated water (10 ppm of available chlorine), followed by washing in running water and later only ripe fruits were selected for this study. The pulp was extracted in a blender (Waring 70115, Model WF2211214) and stored in airtight containers at -18 °C and thawed at room temperature (28 ± 3 °C) before analysis.

The organic standard compounds positively identified in this study were purchased from Sigma/Aldrich (St. Louis, USA) (such as ethyl acetate, 3-hexen-1-ol, methyl butanoate, methyl 2-butenate, methyl hexanoate, ethyl hexanoate, methyl octanoate, δ -limonene, linalool, 3-hexen-1-ol, pentanol, β -myrcene, β -caryophyllene, octanal, benzaldehyde and these were of pure grade (purity 99.9%) while the solvents utilized in this study were purchased from Merck (Brazil).

2.2 Sample preparation and SPME

The volatile components present in the pulp of ripe soursop fruit pulp were captured by SPME technique and separated by high resolution gas chromatography. Initially three different fibers - 85 μ M carboxen/polydimethylsiloxane (CAR/PDMS), 50/30 μ M divinyl benzene/carboxen/polydimethylsiloxane (DVB/CAR/PDMS)

and 65 μ M polydimethylsiloxane/divinylbenzene (PDMS/DVB), were tested to analyze their performance in capturing volatiles from soursop pulp.

The extraction conditions used for the selection of fiber were according to Cheong et al. (2011). Before each extraction the vial was placed in a water bath (40 °C) under magnetic stirring for 10 min (equilibrium time). Later the SPME fiber was inserted in the headspace of the solution for adsorption of volatiles for 30 minutes. After this exposure, the fiber was removed and later introduced into the injector of gas chromatograph (Varian, Model 4000) for thermal desorption of volatiles for 7 minutes. The pH was not altered as it was the natural pH (4.1) of the fruit.

Based on the results, the fiber was selected based on a factorial design of 2³ including 6 axial points and six repetitions at the central point amounting to a total of 20 experiments were performed. Table 1 presents the central composite design matrix which shows the coded and real values of different variables. The influence of adsorption temperature (minimum 28 °C, maximum 62 °C), amount of salt ionic strength (minimum 0, maximum 30.0%) and the concentration of the pulp (min 1, max 99.0%) on volatiles capture was evaluated .

2.3 GC-MS conditions

The separation was performed using a gas chromatography system (Varian Inc. Walnut Creek, California, USA) equipped with mass spectrometry (Model 4000). The chromatographic conditions were according to the methodology described earlier by Galvão (2011). Initial study on the selection of the fiber was conducted by using a non-polar column (VF-5ms). After optimization of analytical conditions, separations were performed

Table 1. Coded independent variables and their real values in the experimental design.

Experiment Number	Coded variables			Real values		
	X1	X2	X3	Adsorption temperature (°C)	NaCl (%)	Pulp concentration (%)
1	-1	-1	-1	35	6	21
2	+1	-1	-1	55	6	21
3	-1	+1	-1	35	24	21
4	+1	+1	-1	55	24	21
5	-1	-1	+1	35	6	79
6	+1	-1	+1	55	6	79
7	-1	+1	+1	35	24	79
8	+1	+1	+1	55	24	79
9	-1.68	0	0	28	15	50
10	+1.68	0	0	62	15	50
11	0	-1.68	0	45	0	50
12	0	+1.68	0	45	30	50
13	0	0	-1.68	45	15	1
14	0	0	+1.68	45	15	99
15	0	0	0	45	15	50
16	0	0	0	45	15	50
17	0	0	0	45	15	50
18	0	0	0	45	15	50
19	0	0	0	45	15	50
20	0	0	0	45	15	50

on two different columns: a Varian Factor Four Capillary VF-5ms analytical column (30 m x 0.25 mm i.d. x 0.25 mm film thickness; component: 5% diphenyl and 95% dimethylpolysiloxane) and CP-Wax52CB (30m x 0,25mm i.d. x 0,25µm film thickness; polyethylene glycol). The injector temperature was held at 220 °C. Helium (99.9999%) was used as a carrier gas at a flow rate of 1.0 mL/min. The mass spectrometer was operated in the electron ionization mode with an electrical energy of 70 eV and an ion source temperature of 250 °C. The mass spectrum was scanned between 33 and 550 amu at 0.3 s interval.

2.4 Compounds identification

The linear retention index (RI) values for unknowns were determined based on retention time data obtained by analyzing a series of normal alkanes (C8-C21). Volatile components were positively identified by matching their RI values and mass spectra with those of standards, also run under identical chromatographic conditions in the laboratory.

The identification was also based on matching an unknown mass spectrum with spectra available on the NIST (National Institute of Standards and Technology, USA) database, mass spectral data system or from the literature (Jennings & Shibamoto, 1980; Adams, 1995; Kondjoyan & Berdagué, 1996).

3 Results and discussion

3.1 Fiber selection – preliminary study

Initial tests were conducted using 3 different fibers (50/30 µm DVB/CAR/PDMS, 85 µm CAR/PDMS and 65 µm PDMS/DVB) in SPME of soursop pulp so as to evaluate the performance and effectiveness of each fiber coating material on the headspace extraction efficiency of volatile flavor compounds. As shown in Figure 1, semi-polar fiber coatings (CAR/PDMS and DVB/CAR/PDMS) appeared to be more efficient in extracting the soursop volatile compounds compared to bipolar fiber coating (PDMS/DVB). In the present study, CAR/PDMS was chosen due to its higher extraction efficiency for priority class of esters present in soursop pulp. About 90% of compounds

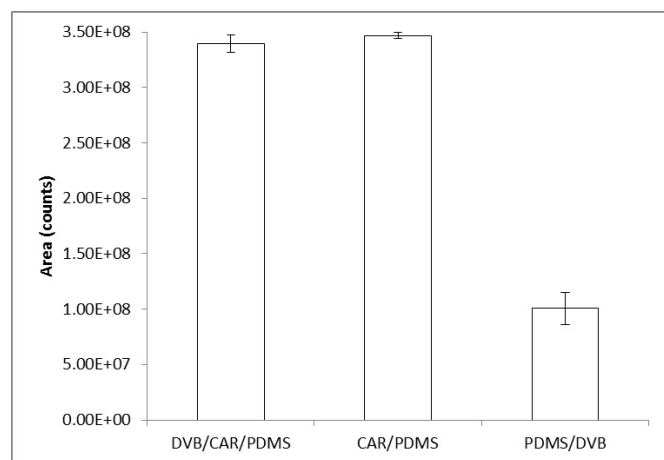


Figure 1. Effect of the SPME fiber coating on the extraction of the volatile compounds from soursop fruits.

captured by the fiber CAR/PDMS were esters, while the fibers DVB/CAR/PDMS and PDMS/DVB captured only 83 and 80% of this class, respectively. Roberts et al. (2000) reported CAR/PDMS fiber to be the most appropriate fiber coating to extract small molecules and acids. As soursop flavor is composed mainly of esters, the sensitivity of CAR/PDMS for smaller molecules, acids and non-polar compounds could enable the extraction of a wide range of volatile flavor compounds.

The use of 65 µm PDMS/DVB fiber permitted the extraction of only 35 volatile compounds when compared to that of 85 µm CAR/PDMS fiber in which 40 compounds were separated. This decrease could be expected as the amount of the coating of PDMS/DVB fiber results in a lower volume available for absorption of the analytes. Augusto et al. (2000) tested 4 different coatings (PDMS, CW/DVB, CAR/PDMS and PA) in fibers and reported that the most efficient fiber was PDMS which captured 16 compounds from soursop pulp while CAR/PDMS captured only 11 compounds. However, they also reported that the largest extraction capacity in area count and capture of lighter compounds such as ethyl acetate, 3-hexen-1-ol, ethyl butanoate, 1-butanol from soursop pulp was obtained by using CAR/PDMS fiber. Our data in this work corroborates this conclusion. However, the fruit used in their work was custard apple (*Anona reticulata* L.) while this study was done with soursop (*Annona muricata* L.). Cheong et al. (2011) also recommended the CAR/PDMS fiber based on their extraction of esters and their signal intensity in work with soursop.

Soares et al. (2015) suggested the CAR/PDMS fiber, in general, for extraction of highly volatile compounds when compared to other coatings such as PDMS/DVB and DVB/CAR/PDMS.

All the work published on soursop volatile compounds reports the predominant class being esters followed by terpenes (MacLeod & Pieris, 1981; Franco & Rodriguez-Amaya, 1983; Iwaoka et al., 1993; Wong & Khoo, 1993; Jirovetz et al., 1998; Augusto et al., 2000; Cheong et al., 2010, 2011; Cardozo et al., 2011, 2013), except for Pelissier et al. (1994) who analyzed essential oil of soursop. CAR/PDMS fiber was the most efficient for capturing esters (90% area) which represent major class of volatile compounds present in soursop pulp and hence this fiber was selected to conduct further experiments.

Table 2 presents data on volatile compounds identified in soursop pulp using SPME with different fibers while Figure 2 evaluates efficiency of different coatings on quantitative representation of several classes of organic compounds. Figure 3 shows the area obtained on volatiles capture using different fibers in SPME. The fibers DVB/CAR/PDMS, CAR/PDMS and PDMS/DVB captured 40, 40 and 35 compounds, respectively. The major classes of compounds were esters and terpenes. CAR/PDMS fiber was the most efficient fiber which resulted in higher total counts in the chromatogram as compared to the performance by other fibers (DVB/CAR/PDMS and PDMS/DVB) and hence this fiber was selected to conduct further experiments.

The identified compounds represent an area of more than 98% of the total area of the chromatogram which leads to conclude that most of the compounds present in soursop pulp could be separated and identified in this work. Figure 4 shows the profile

Table 2. Volatiles compounds identified in soursop pulp captured by use of different fibers in SPME and separated by GC on non-polar column.

Compounds	IR Non-polar	50/30 μm DVB/CAR/PDMS		85 μm CAR/PDMS		65 μm PDMS/DVB		Odor ⁹
		IR	Area (%) [*]	IR	Area (%)	IR	Area (%)	
Alcohol								
(Z)-3-hexen-1-ol ^{1,2,3,4,5,6,7,8}	836	830	1.25 ± 0.19			830	0.75 ± 0.19	grass
perillol	1289	1299	0.01 ± 0.00			1299	0.04 ± 0.00	
Sub-total			1.26				0.79	
Aldehydes								
acetaldehyde	400			<600	0.08 ± 0.01			
3-hexenal ³	771	778	0.57 ± 0.09	781	1.06 ± 0.12	781	0.70 ± 0.08	leaf, green
Sub-total			0.57		1.15		0.70	
Esters								
ethyl acetate ^{2,3,4,5}	625	605	0.12 ± 0.06	587	0.07 ± 0.03			pineapple
methyl butanoate ^{1,2,3,4,5,6,7,8}	728	733	7.44 ± 1.28	733	7.97 ± 0.91	733	6.19 ± 0.05	ether, fruity, sweet
methyl 2-butenate ^{3,4,5,6,7,8}	755	758	7.75 ± 0.48	758	15.57 ± 0.61			
methyl pentanoate	806			799	0.34 ± 0.12	800	0.29 ± 0.12	
methyl (E)-2-pentenoate ⁴	NE	844	0.80 ± 0.25	843	0.34 ± 0.01	844	0.37 ± 0.26	
methyl hexanoate ^{1,2,3,4,5,6,7,8}	905	899	32.28 ± 2.83	899	28.03 ± 3.26	899	33.27 ± 1.01	fruity, fresh, sweet
methyl (Z)-3-hexenoate ^{1,3,4,5,7}	916	907	0.69 ± 0.20	907	1.01 ± 0.02	907	0.37 ± 0.05	grass
methyl (E)-2-hexenoate ^{2,3,4,5,6,7,8}	939	940	31.05 ± 0.79	940	30.17 ± 1.41	940	32.56 ± 0.50	ester, fruity
methyl 2-hexenoate ¹	949	955	0.55 ± 0.07	955	1.83 ± 0.25			
ethyl hexanoate ^{1,2,3,4,5,6,7}	980			973	0.59 ± 0.05			fruity
methyl 2-heptenoate	NE	1039	0.07 ± 0.00	1039	0.12 ± 0.07	1039	0.04 ± 0.01	
methyl octanoate ^{1,3,4,5,7}	1105	1098	0.69 ± 0.08	1097	0.82 ± 0.04	1098	1.67 ± 0.27	orange
methyl (E)-2-octenoate ^{1,2,3,4,5,6,7}	1150	1141	2.01 ± 0.18	1141	1.98 ± 0.15	1141	4.84 ± 0.59	
cis-3-hexenyl butanoate	1166	1158	0.04 ± 0.00	1158	0.04 ± 0.01	1158	0.09 ± 0.01	
ethyl benzoate	1185	1178	0.02 ± 0.01			1178	0.04 ± 0.01	
methyl cinnamate ⁴	1365	1357	0.11 ± 0.02	1357	0.07 ± 0.01	1357	0.23 ± 0.03	strawberry
Sub-total			83.62		89.86		79.97	
Terpenes								
β-myrcene	966	966	0.41 ± 0.16					balsamic, must, spice
δ-limonene ^{2,4,6,7}	1002	1003	0.89 ± 0.13	1002	0.83 ± 0.32	1002	0.24 ± 0.01	citrus, mint
cis-β-ocimene	1013	1010	0.17 ± 0.03	1010	0.18 ± 0.04	1010	0.04 ± 0.01	
α-ocimene	1023	1019	0.19 ± 0.04	1019	0.26 ± 0.19	1019	0.04 ± 0.01	
linalool ^{2,3,4,5,6,7,8}	1079	1071	8.70 ± 0.63	1070	4.35 ± 0.72	1071	11.90 ± 1.45	flower, lavender
cis-allo-ocimene	1115	1114	0.06 ± 0.01	1113	0.02 ± 0.01	1113	0.04 ± 0.02	
α-terpineol ⁷	1195	1164	0.03 ± 0.00	1163	0.02 ± 0.00	1163	0.05 ± 0.02	
isocaryophyllene	1395	1395	0.05 ± 0.03			1395	0.03 ± 0.00	wood
β-selinene	1496	1497	0.02 ± 0.01					herb, spice
Sub-total			10.53		5.72		12.35	
Others								
styrene ¹	863			865	0.11 ± 0.01	864	0.15 ± 0.13	balsamic, gasoline
p-dichlorobenzene	988	986	3.04 ± 0.03	986	2.49 ± 0.29	986	5.03 ± 0.93	
γ-hexalactone	1159	1060	0.05 ± 0.02					oil, anise, mint
benzenepropanoic acid	1245	1249	0.02 ± 0.01					
Total			3.10		2.63		5.19	

^{*}Values expressed as Means ± Standard Deviation; ¹MacLeod & Pieris (1981) relates to Simultaneous Distillation and Extraction; ²Augusto et al. (2000) relates to SPME (CAR/PDMS);

³Cardozo et al. (2011) relates to SPME (DVB/CAR/PDMS); ⁴Cheong et al. (2011) relates to SPME (CAR/PDMS); ⁵Cardozo et al. (2013) relates to SPME (DVB/CAR/PDMS);

⁶Cheong et al. (2010) relates to SPME (CAR/PDMS); ⁷Jirovetz et al. (1998) relates to SDE; ⁸Iwaoka et al. (1993) relates to liquid-liquid extraction; ⁹Acree & Arn (2016) relates to characteristic odor of compounds.

of principal volatile compounds viz., methyl butanoate, methyl 2-butenate, methyl hexanoate, methyl 2-hexenoate, methyl (*E*)-2-hexenoate, ethyl hexanoate, methyl octanoate, methyl (*E*)-2-octenoate present in soursop pulp. The presence of these compounds has been reported in earlier work on soursop in spite of the extraction technique being different than of SPME. Most of these compounds characterize fruity odor note and thus demonstrate their importance in character impact compounds for soursop.

3.2 Experimental design

According to the experimental design (Table 1) which varied the key operational parameters (extraction temperature, % ionic strength and pulp concentration) in SPME, the data obtained on the number of peaks using the two columns (polar and non-polar) are assembled in Table 3.

After the selection of the CAR/PDMS fiber, a new experimental design was applied which focused on evaluation of the influence

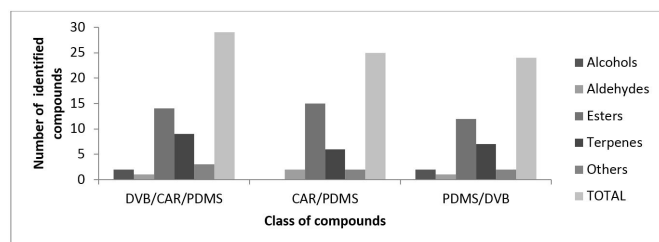


Figure 2. Number of volatile compounds of soursop of various chemical classes according to the use of different SPME fibers.

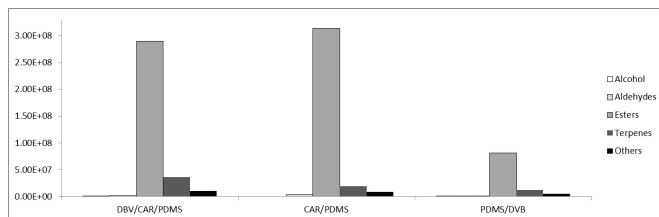


Figure 3. Area counts of total volatile compounds of soursop of various chemical classes according to the use of different SPME fibers.

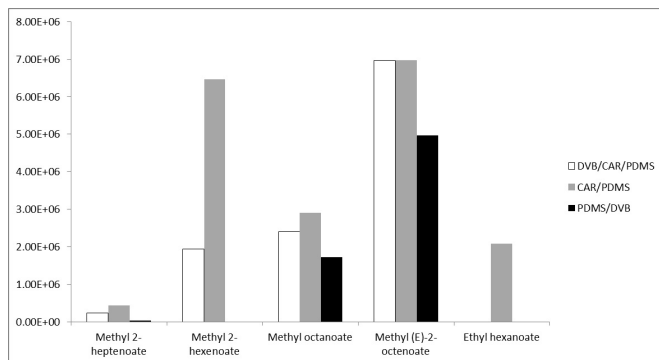


Figure 4. Effect of the SPME fiber coating on the area counts of principal aroma compounds from soursop fruits.

of pulp concentration in the volatile composition of soursop pulp. In these experiments on soursop pulp, more number of peaks were observed under the same extraction conditions (45 °C, 15% NaCl and 50% of pulp concentration) for both the columns (polar and non-polar). However, large number of peaks was found in the experiment number 15 for the non-polar column (57 compounds) and in experiment number 20 for polar column (87 compounds), both these points being the central points of the planned experiments.

According to the results presented in Table 3, it is clear that for each column the results were quite different. For non-polar column the design did not differ significantly at the level of $p < 0.05$ and the results are independent of its variables and thus no significant effect arose for their responses, revealing the data in the experiment number 15 with the largest number of peaks (57). This observation could also be confirmed through the data processing by STATISTIC 6.0 program, wherein the values for the main effects and interactions area are shown (Figure 5).

For polar column (Figure 6), it was evident that all parameters were significant ($p < 0.05$) and the temperature being the parameter that influenced most on the capture of volatile compounds.

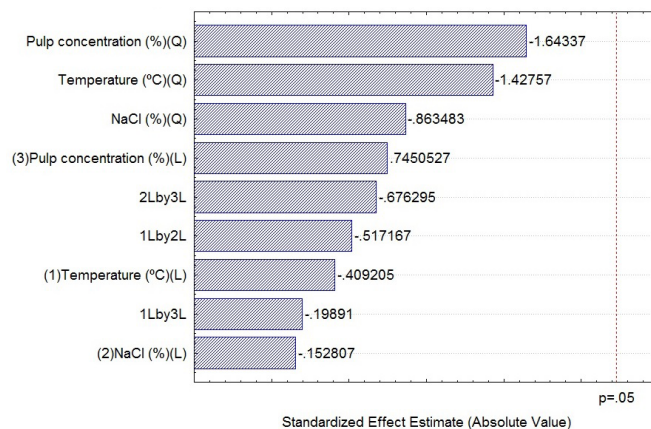


Figure 5. Pareto chart for the response (number of peaks) of the GC analysis of the HS-SPME-MS of soursop fruit for non-polar column.

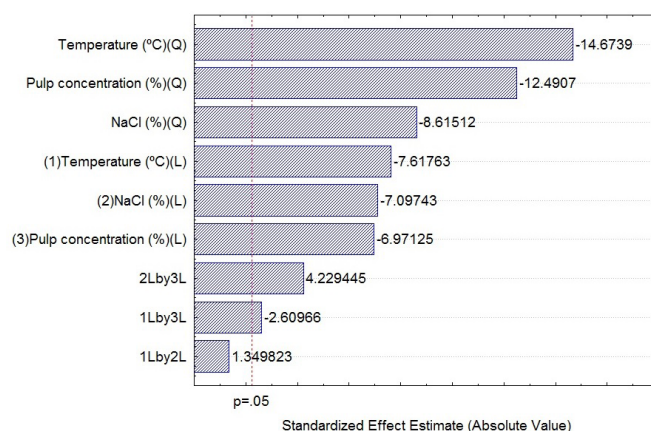


Figure 6. Pareto chart for the response (number of peaks) of the GC analysis of the HS-SPME-MS of soursop fruit for polar column.

Table 3. Number of volatile compounds present in soursop pulp captured by SPME and separated by GC on different columns.

Experiment Number	Temperature (°C)	NaCl (%)	Pulp concentration (%)	Number of peaks separated on columns	
				Non-polar	Polar
1	35	6	21	41	75
2	55	6	21	40	68
3	35	24	21	43	45
4	55	24	21	44	39
5	35	6	79	45	56
6	55	6	79	50	28
7	35	24	79	47	43
8	55	24	79	37	29
9	28	15	50	33	58
10	62	15	50	28	25
11	45	0	50	34	69
12	45	30	50	34	50
13	45	15	1	25	57
14	45	15	99	33	39
15	45	15	50	57	86
16	45	15	50	47	80
17	45	15	50	45	85
18	45	15	50	38	86
19	45	15	50	49	84
20	45	15	50	50	87

However, the effect was negative for all parameters (temperature, % NaCl and the % pulp concentration), i.e. at lower levels of these parameters there was an increased extraction of volatile compounds in SPME from soursop pulp. The interaction between % NaCl and % pulp concentration also influenced positively in the extraction process as higher values of this interaction revealed greater capture of volatile compounds.

3.3 Influence of extraction temperature

An increase in extraction temperature decreased the number of volatile compounds absorbed by CAR/PDMS from soursop pulp in headspace. One factor which may influence these results: higher temperatures affect adversely on the sorption of analyte on the coating due to thermodynamic reasons (reduced partition coefficient) and consequently, extraction efficiency decreases when the temperature increases (Setkova et al., 2007). It was possible to observe the influence of temperature, where higher temperatures (55 °C and 62 °C) resulted in detection of lower number of volatile compounds, being 68, 39, 28, 29 and 25 peaks for the experiments 2, 4, 6, 8 and 10, respectively, while for lower temperatures, the number of compounds were more, with higher number of compounds detected being 75, 45, 56, 43 and 58 peaks for the experiments 1, 3, 5, 7 and 9, respectively. However, the lowest temperature (28 °C) tried in SPME presented lower number of compounds, which may be because that the temperature was just insufficient for volatilization of organic compounds.

3.4 Influence of pulp concentration

The data presented in Table 3 reveal that an increase in pulp concentration over 50% has a negative effect on the absorption of volatile compounds present in pulp on to the fiber, i.e. higher

the concentration of soursop pulp, lower is the number of volatile compounds captured. However, the increase in pulp concentration was related with the increase in the concentration of added sodium chloride which led to the saturation of the pulp, thereby reducing the release of volatile compounds onto the headspace vial. Thus the pulp concentration of 79% and higher (24%) sodium chloride (ionic strength) decrease the release of these volatiles.

3.5 Influence of extraction ionic strength

The exposure of the fiber at a concentration of 79% pulp at 35 °C in which 6% and 24% of salt were used (experiment 5 and 7), separated 56 and 43 peaks, respectively in polar column (Table 3). However, when the extraction was performed at 55 °C having the same salt concentration (6 and 24%) in experiment numbers 6 and 8, 28 and 29 peaks were separated, respectively. Using 99% pulp at 45 °C having 15% ionic strength detected only 39 peaks indicating the negative effect of pulp concentration and temperature on the release of volatile compounds

Among all the experiments undertaken, the experiment number 10 which utilized higher temperature (62°C) for extraction but had lower salt (15%) and pulp concentration (50%) resulted in a lower number of 25 compounds in polar column separation which reveals that the interactions between these parameters were not significant, i.e. one parameter does not influence the other, demonstrating the detrimental effect of temperature and concentration of the pulp. The pareto chart (Figure 4) shows this effect at the significance level of $p < 0.05$.

Among all the experiments performed, the experiment number 20 which represented the central point of the experiments resulted in capture of a greater number of 87 compounds in

polar column separation. Thus the best conditions (50% pulp; 15% NaCl; Temp. 45°C) for SPME technique, proven through this experimental design, were defined as the conditions of this experiment which had the maximum number of volatile compounds separated in gas chromatography system coupled to high resolution mass spectrometry.

The exposure time of the fiber in injector of gas chromatograph for desorption of analytes is also an important factor. This would be another variable worth analyzing, although, it is known that desorption occurs in a very short period of time. The fiber exposure time in the injector for all experiments was 7 min, which is much higher than required for desorption. In addition, interspersing between the chromatographic analysis, fiber inserts were held in GC injector in order to check the “memory effect” of the fiber, i.e., if a small amount of analyte was not desorbed and still retained which may interfere in the subsequent analyses. This phenomenon was not observed in any of the tests performed, concluding that the 7 min exposure time was good enough for complete desorption.

Thus the optimized SPME conditions in soursop pulp were: use of 50% pulp, addition of 15% sodium chloride and volatiles capture realized in vial containing 10 mL aliquot at 45 °C for 30 min using CAR/PDMS fiber and separation performed in GC using polar column.

3.6 Volatile compounds identification in different columns

The volatile compounds were considered positively identified when the retention index and the spectra were similar to those of the standard run under identical analytical conditions as well as matching of spectra reported in the literature (Kondjoyan &

Berdagué, 1996; Adams, 2001) or with data from the database (National Institute of Standards and Technology, 2016). It is worth noting that the variation in the retention index for identification purposes was limited to an average of ± 4 units.

Table 4 lists the volatile compounds captured from soursop pulp by SPME technique using the conditions of experiment number 15 and GC analysis done on non-polar column along with the results obtained from experiment number 20 wherein GC separation was done using a polar column. In addition to the data of retention index and area (%) of each compound presented in Table 4, the information also includes their respective organic classes. Moreover the superscript numbers used in the name of the compounds substantiate whether the presence of the compound had been reported earlier in soursop pulp or identification is being reported for the first time in this study. Figures 7 and 8 present the total ion chromatograms of volatiles from soursop pulp under SPME in experiments numbers 15 and 20 for GC separation on non-polar and polar column, respectively.

A total of 57 and 87 compounds from SPME of soursop pulp were separated on GC analysis using non-polar and polar column, respectively. Among the 57 compounds separated by using non-polar column, 46 were identified by comparing the spectrum and linear retention index data based on literature (Kondjoyan & Berdagué, 1996; Acree & Arn, 2016) while 11 compounds could not be identified although 6 of these had a very high retention index which characterize lower volatility. However, when a polar column was used in GC separation, 87 compounds were detected and 53 of these were identified by comparing their spectra and linear retention indices while 34 compounds could not be identified.

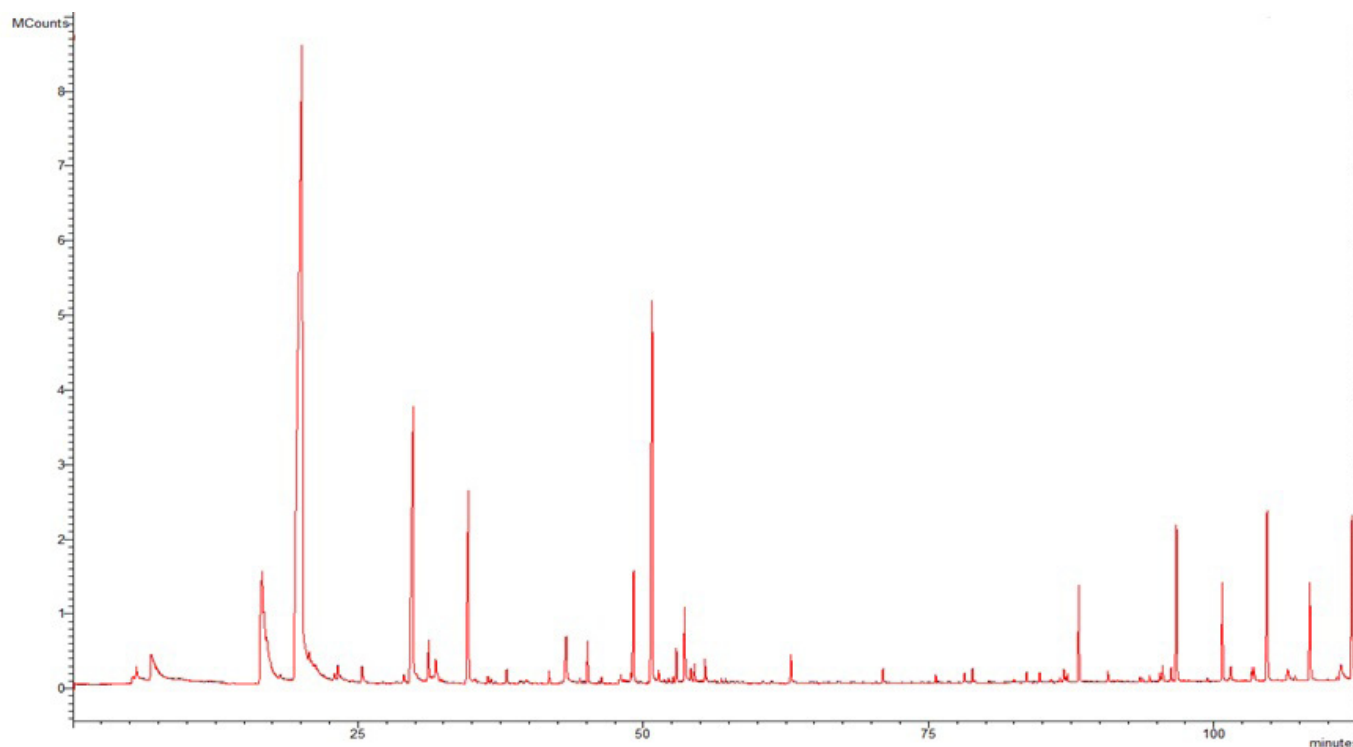


Figure 7. Total ion chromatogram of soursop fruit pulp by HS-SPME using non-polar column.

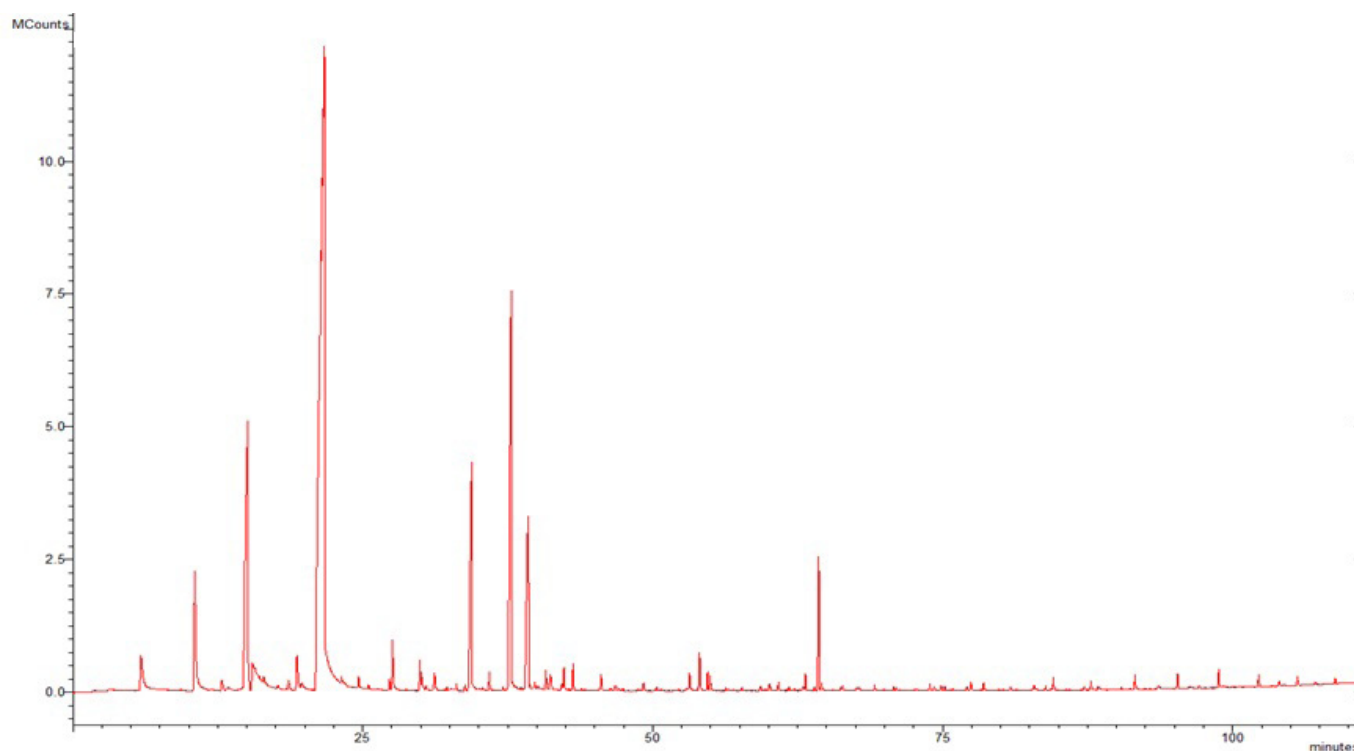


Figure 8. Total ion chromatogram of sour sop fruit pulp by HS-SPME using polar column.

Among the identified compounds based on usage of non-polar column in GC separation, the main classes of compounds were esters which constituted 20 compounds representing 84.18% of total area, 15 terpenes (10.63%), 8 alcohols (3.18%) and 1 aldehyde (0.58%). In the case of polar column used for separation, the major compounds identified were 24 esters (79.11%), 15 terpenes (16.55%), 10 alcohols (2.31%) and 1 aldehyde (0.11%). The major organic compounds identified in sour sop pulp using this column were methyl hexanoate (fruity, fresh, sweet odor) and methyl (*E*)-2-hexenoate (fruity odor), which represented about 60% area. The main difference in columns performance was that usage of polar column revealed a large number of esters and terpenes, which are very important in sour soap flavor fruits. Based on this comparison the decision to use a specific column for the analysis of sour sop pulp should be made giving emphasis on characteristic impact compounds of sour sop pulp.

The major compounds in relation to the area % in total ion chromatogram were the methyl (*E*)-2-hexenoate (31.45%), methyl hexanoate (30.56%), linalool (7.63%), methyl 2-butenate (26.7%), methyl butanoate (7.14%) for separation in non-polar column, while for the polar column, the major compounds were methyl (*E*)-2-hexenoate (48.14%), methyl hexanoate (20.63%), α -copaene (6.19%), linalool (5.54%), ethyl acetate (3.12%), methyl cinnamate (2.14%). Ten compounds viz. pentyl butanoate, ethyl dodecanoate, isopropyl benzoate, phenyl butanoate, 3-hexenyl salicylate, dodecanol, tetradecanol, β -pinene, benzaldehyde and β -ionone were detected just in the analysis employing polar column, while only 4 compounds viz. methyl octadecanoate, γ -elemene, 3-hexenal and 3-hexadecanone were detected when separation was performed in non-polar column.

This study indicated that esters were the most abundant compounds in sour sop pulp (Table 4). This observation is in agreement with previous studies on sour sop pulp (MacLeod & Pieris, 1981; Iwaoka et al., 1993; Wong & Khoo, 1993; Pelissier et al., 1994; Jirovetz et al., 1998; Augusto et al., 2000; Cheong et al., 2010, 2011; Cardozo et al., 2011, 2013).

Wong & Khoo (1993) determined the volatile compounds present in sour sop and identified compounds such as linalool, 1-hexanol and 2-pentanone to be of great importance from characteristic flavor point of view. Cardozo et al. (2011) reported the presence of (*Z*)-3-hexenol in ripe sour sop pulp. The (*Z*)-3-hexenol possesses the characteristic odor of green note along with methyl hexanoate and methyl (*E*)-2-hexenoate which are main volatile compounds in ripe fruit while compounds such as ethyl acetate and methyl butanoate, may be characterized as responsible indicators for initiation of the fermentation and thus loss in sensory quality of the fresh fruit. It is worth noting that all these compounds were identified in our study.

The volatile chemical composition reported here is consistent with the data reported by others. MacLeod & Pieris (1981) found 80% of TIC area belonging to esters, and the main esters were methyl hexanoate (31%) and methyl 2-hexenoate (27%) from fruits harvested in Sri Lanka. Iwaoka et al. (1993) working with unripe sour sop fruits grown in Hawaii found only 12 volatile compounds and reported high amount of (*Z*)-3-hexen-1-ol, whereas more ripe fruits had high levels of esters such as methyl (*E*)-2-hexenoate, methyl butanoate and methyl hexanoate.

Cheong et al. (2011) reported that compounds such as methyl butanoate, methyl hexanoate, methyl (*E*)-2-butenate and methyl (*E*)-2-hexenoate were four main compounds responsible for taste

Table 4. Volatile compounds captured by SPME from soursop pulp and analyzed in GC-MS system using polar and non-polar columns.

Compounds	Non-polar		Polar		Odor ⁹
	IR	Area (%) [*]	IR	Area (%) [*]	
Esters					
ethyl acetate ^{2,3,4,5}	604	0.12 ± 0.06	905	3.12 ± 0.16	pineapple-like
methyl butanoate ^{1,2, 3, 4,5,6,7,8}	733	7.14 ±1.28	978	1.17 ± 0.09	ether, fruity, sweet
methyl 2-butenate ^{3,4,5,6,7,8}	758	7.26 ± 0.48	1100	0.29 ± 0.05	-
methyl hexanoate ^{1,2,3,4,5,6,7,8}	899	30.56 ± 2.83	1190	20.63 ± 1.31	fruity, fresh, sweet
methyl (Z)-3-hexenoate ^{1,3,4,5,7}	907	0.70 ± 0.20	1284	0.13 ± 0.06	grass
methyl (E)-2-hexenoate ^{2,3,4,5,6,7,8}	940	31.45 ± 3.29	1302	48.14 ± 4.01	ester, fruity
ethyl hexanoate ^{1,2,3,4,5,6,7}	986	3.08 ± 0.03	1229	0.14 ± 0.03	fruity
butyl isobutanoate	990	0.13 ± 0.01	1150	1.13 ± 0.21	fresh, sweet, fruity
Isopropyl hexanoate	1039	0.07 ± 0.01	1230	0.10 ± 0.02	fresh
methyl octanoate ^{1,3,4,5,7}	1098	0.70 ± 0.08	1389	0.47 ± 0.08	orange
methyl (E)-2-octenoate ^{1,2,3,4,5,6,7}	1141	2.03 ± 0.18	1440	0.07 ± 0.03	-
ethyl benzoate	1178	0.02 ± 0.01	1597	0.36 ± 0.02	chamomile, flower, celery, fruity
butyl hexanoate	1249	0.02 ± 0.01	1544	0.06 ± 0.01	fruity
methyl cinnamate ⁴	1357	0.11 ± 0.02	2011	2.14 ± 0.08	strawberry
ethyl decanoate	1395	0.05 ± 0.03	1675	0.10 ± 0.04	grape
ethyl cinnamate	1420	0.12 ± 0.01	2084	0.07 ± 0.03	flower, honey
methyl tetradecanoate	1650	0.23 ± 0.01	1987	0.23 ± 0.09	orris
ethyl tetradecanoate	1700	0.01 ± 0.01	2052	0.04 ± 0.02	
methyl hexadecanoate	1920	0.21 ± 0.01	2200	0.03 ± 0.01	waxy
methyl octadecanoate	2049	0.17 ± 0.01			fat
pentyl butanoate			1345	0.14 ± 0.09	pineapple-like
ethyl dodecanoate			1823	0.24 ± 0.06	
isopropyl benzoate			1921	0.13 ± 0.02	
phenyl butanoate			1938	0.12 ± 0.01	
3-hexenyl salicylate			2226	0.06 ± 0.02	
Alcohols					
1-pentanol	766	0.19 ± 0.02	1170	0.38 ± 0.06	fruity
(Z)-3-hexen-1-ol ^{1,2,3,4,5,6,7,8}	830	1.27 ± 0.19	1392	0.83 ± 0.06	grass
2-hexenol ^{4,6}	844	0.81 ± 0.25	1212	0.19 ± 0.01	leaf, green, wine, fruit
2-heptanol ^{4,6}	984	0.56 ± 0.07	1316	0.06 ± 0.02	
2-ethyl-1-hexanol	1035	0.06 ± 0.01	1487	0.26 ± 0.05	rose, green
1-octanol	1079	0.24 ± 0.01	1452	0.09 ± 0.04	moss, nut, mushroom
isopulegol	1158	0.04 ± 0.01	1570	0.29 ± 0.02	
perillol	1299	0.01 ± 0.01	2003	0.05 ± 0.02	-
dodecanol			2016	0.10 ± 0.04	soapy, waxy
tetradecanol			2149	0.06 ± 0.02	coconut
Terpenes					
β-myrcene ²	966	0.42 ± 0.16	1159	0.85 ± 0.06	balsamic, must, spice
δ-limonene ^{2,4,6,7}	1003	0.90 ± 0.13	1176	0.36 ± 0.01	citrus, mint
cis-β-ocimene	1010	0.17 ± 0.03	1223	0.87 ± 0.12	sweet, herb
α-ocimene	1019	0.19 ± 0.04	1255	0.16 ± 0.01	fruity, wet cloth
linalool ^{2,3,4,5,6,7,8}	1071	7.63 ± 0.63	1506	5.54 ± 0.02	flower, lavender
cis-allo-ocimene	1114	0.06 ± 0.01	1350	0.84 ± 0.05	herb
α-terpineol ⁷	1164	0.03 ± 0.00	1794	0.24 ± 0.06	-
α-copaene	1379	0.04 ± 0.01	1462	6.19 ± 0.14	woody, spice
α-gurjunene	1409	0.12 ± 0.01	1741	0.03 ± 0.01	woody, balsamic
β-caryophyllene ^{3,4,6,7}	1419	0.17 ± 0.01	1594	0.10 ± 0.06	woody, spice
Z-β-farnesene ¹	1444	0.31 ± 0.01	1654	0.23 ± 0.01	citrus, green

^{*}Values expressed as Means ± Standard Deviation; ¹MacLeod & Pieris (1981) relates to Simultaneous Distillation and Extraction; ²Augusto et al. (2000) relates to SPME (CAR/PDMS); ³Cardozo et al. (2011) relates to SPME (DVB/CAR/PDMS); ⁴Cheong et al. (2011) relates to SPME (CAR/PDMS); ⁵Cardozo et al. (2013) relates to SPME (DVB/CAR/PDMS); ⁶Cheong et al. (2010) relates to SPME (CAR/PDMS); ⁷Jirovetz et al. (1998) relates to SDE; ⁸Iwaoka et al. (1993) relates to liquid-liquid extraction; ⁹Acree & Arn (2016) relates to characteristic odor of compounds.

Table 4. Continued...

Compounds	Non-polar		Polar		Odor ⁹
	IR	Area (%) [*]	IR	Area (%) [*]	
α -humulene	1456	0.34 \pm 0.01	1611	0.37 \pm 0.08	woody
γ -elemene ⁷	1466	0.12 \pm 0.01			green, woody, oily
α -guaiene	1471	0.11 \pm 0.01	1810	0.57 \pm 0.13	woody, balsamic
β -selinene	1497	0.02 \pm 0.01	1719	0.15 \pm 0.06	herb, spice
β -pinene ⁷			1039	0.05 \pm 0.02	pine, resin, turpentine
Aldehydes					
3-hexenal ³	778	0.58 \pm 0.12			leaf, green
benzaldehyde			1518	0.61 \pm 0.21	almond, burnt sugar
Sub-total					
Others					
γ -hexalactone	1060	0.05 \pm 0.02	1887	0.04 \pm 0.01	oily, anise, mint
3-hexadecanona	1316	0.16 \pm 0.01			-
β -ionone			1905	0.07 \pm 0.01	seaweed, violet, flower, raspberry

^{*}Values expressed as Means \pm Standard Deviation; ¹MacLeod & Pieris (1981) relates to Simultaneous Distillation and Extraction; ²Augusto et al. (2000) relates to SPME (CAR/PDMS); ³Cardozo et al. (2011) relates to SPME (DVB/CAR/PDMS); ⁴Cheong et al. (2011) relates to SPME (CAR/PDMS); ⁵Cardozo et al. (2013) relates to SPME (DVB/CAR/PDMS); ⁶Cheong et al. (2010) relates to SPME (CAR/PDMS); ⁷Jirovetz et al. (1998) relates to SDE; ⁸Iwaoka et al. (1993) relates to liquid-liquid extraction; ⁹Acree & Arn (2016) relates to characteristic odor of compounds.

and aroma of soursop, and that in our study all these compounds were identified except that the compound methyl (*E*)-2-butenate which could not be identified. Of the 21 volatile compounds identified by Augusto et al. (2000) working with custard apple (*Annona reticulata* L.) pulp reported the presence of esters such as ethyl acetate, butyl acetate and methyl 2-hexenoate which correlated well in our present study using soursop (*Annona muricata*, L.) pulp.

Jirovetz et al. (1998) reported the presence of compounds such as caryophyllene, limonene and linalool, which were also found in this study. Furfural and 2-dimethyl furfural identified in soursop pulp may have been formed by Amadori rearrangement of the reaction, the first involving carbonyl, usually reducing sugars and free amino groups (Yamaguchi et al., 1983).

4 Conclusions

Among several fibers used in SPME, the results reveal that the CAR/PDMS fiber was most efficient for the capture of esters from soursop pulp. The optimum SPME conditions were 45 °C for extraction, 15% of ionic strength and 50% of pulp concentration which resulted in a separation of 87 compounds on using polar column and 57 compounds when a non-polar column was used. The main compounds found in soursop fruit pulp were methyl (*E*)-2-hexenoate, methyl hexenoate and linalool. This detailed study optimizes the final extraction conditions of SPME on identification of a large number of volatile compounds from soursop fruit pulp.

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