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# Metazoan endoparasites of *Pygocentrus nattereri* (Characiformes: Serrasalminae) in the Negro River, Pantanal, Brazil

Endoparasitas metazoários de *Pygocentrus nattereri* (Caraciformes: Serrasalminae) no rio Negro, Pantanal, Brasil

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## Abstract

In the period of October 2007 to August 2008, 152 specimens of *Pygocentrus nattereri* were caught in the Negro River in the Nhecolândia region, central Pantanal wetland, State of Mato Grosso do Sul, Brazil. The specimens were necropsied and a total of 4,212 metazoan endoparasites were recovered, belonging to 10 taxons: *Procamallanus* (*Spirocamallanus*) *inopinatus*, Philometridae gen. sp., *Eustrongylides* sp., *Brevimulticaecum* sp., *Contracaecum* sp. (Nematoda), *Echinorhynchus paranensis* (Acanthocephala), *Leiperia gracile*, *Sebekia oxycephala*, *Subtriquetra* sp. 1 and *Subtriquetra* sp. 2 (Pentastomida). This is the first record of two parasite species from *P. nattereri*: *E. paranensis* and *L. gracile*.

**Keywords:** *Pygocentrus nattereri*, *Echinorhynchus paranensis*, *Leiperia gracile*, helminths, freshwater fishes, piranhas.

## Resumo

No período de outubro de 2007 a agosto de 2008, 152 espécimes de *Pygocentrus nattereri* foram capturados no rio Negro na região da Nhecolândia, parte central do Pantanal, Mato Grosso do Sul, Brasil. Os espécimes foram necropsiados e um total de 4.212 endoparasitas metazoários foram colhidos, pertencentes a 10 táxons: *Procamallanus* (*Spirocamallanus*) *inopinatus*, Philometridae gen. sp., *Eustrongylides* sp., *Brevimulticaecum* sp., *Contracaecum* sp. (Nematoda), *Echinorhynchus paranensis* (Acanthocephala), *Leiperia gracile*, *Sebekia oxycephala*, *Subtriquetra* sp. 1 e *Subtriquetra* sp. 2 (Pentastomida). Este é o primeiro registro de duas espécies de parasitas em *P. nattereri*: *E. paranensis* e *L. gracile*.

**Palavras-chave:** *Pygocentrus nattereri*, *Echinorhynchus paranensis*, *Leiperia gracile*, helmintos, peixes de água doce, piranhas.

## Introduction

*Pygocentrus nattereri* Kner, 1858 is a gregarious and piscivorous species (SAZIMA; MACHADO, 1990). Its wide geographical distribution includes the tropical and neotropical regions of South America (Argentina, Bolivia, Brazil, Colombia, Ecuador, Guyana, Paraguay, Peru and Uruguay), mainly in the Amazon

and Paraguay-Paraná river basins, coastal rivers in northeastern Brazil, and the Essequibo River basin (JÉGU, 2003).

Most information on its parasite fauna consists of records of 34 ectoparasite species (THATCHER, 2006; EIRAS et al., 2010). Only five described and five undescribed endoparasite species have been reported in *P. nattereri*: the nematodes *Eustrongylides ignotus* Jägerskiöld, 1909 larvae (MORAVEC, 1998), *Eustrongylides* sp. larvae (BARROS et al., 2010; EIRAS et al., 2010), *Contracaecum* sp. larvae (PAVANELLI et al., 2004), *Procamallanus* (S.)

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*inopinatus* Travassos, Artigas & Pereira, 1928 (THATCHER, 2006), *Brevimulticaecum* sp. larvae (VIEIRA et al., 2010), *Procamallanus* sp. (BARROS et al., 2010), *Philometra nattereri* n. sp. (CÁRDENAS et al., 2012); the acanthocephalan *Acanthocephala* fam. gen. sp. (EIRAS et al., 2010); the cestode *Proteocephalus serrasalmus* Rego & Pavanelli, 1990 (REGO; PAVANELLI, 1990); and larvae of the pentastomids *Leiperia* sp. (EIRAS et al., 2010) and *Sebekia oxycephala* Sambon, 1922 (REGO; EIRAS, 1989).

Because of the lack of knowledge regarding the endoparasite fauna in this host, the aim of this study was to report and describe the species of metazoan endoparasites found in *P. nattereri* in the Negro River, Pantanal wetland, State of Mato Grosso do Sul, Brazil.

## Materials and Methods

From October 2007 to August 2008, 152 specimens of *P. nattereri* were collected, pithed immediately after capture and preserved under refrigeration until the moment of necropsy. The necropsy, specimen collection, preparation and conservation of the endoparasites were performed according to Eiras et al. (2006).

Hosts measured  $19.89 \pm 3.11$  (11.4-24.8) cm in standard length and weighed  $376.36 \pm 179.35$  (49-853) g. The fish were caught with hooks and cast nets of different mesh sizes, in the main channel of the middle Negro River ( $19^{\circ} 34' 29.2''$  S and  $56^{\circ} 14' 37.1''$  W), a tributary of the Paraguay River in the Nhecolândia subregion of the central Pantanal wetland, state of Mato Grosso do Sul, Brazil.

Some parasite specimens were prepared for observation by means of scanning electron microscopy (SEM) according to Chiarini-Garcia (1997). Measurements of specimens in light microscopy (LM) were made with Leica (LAS Leica™) software. All measurements are in millimeters; means are followed by the range in parentheses.

These parasites were compared with specimens deposited in the Helminthological Collection of the Oswaldo Cruz Institute

(CHIOC). Representative specimens were deposited in the Zoological Reference Collection at the Universidade Federal do Mato Grosso do Sul, together with a host voucher specimen (ZUFMS-PIS No. 3087). Parasitological descriptors were calculated according to Bush et al. (1997). The mean values of descriptors are followed by the standard deviation ( $\pm$ ).

## Results and Discussion

Of 152 *P. nattereri* examined, 84% had at least one species of metazoan endoparasite: 35% were parasitized by one species, 24% by two, 16% by three, 7% by four, and 1% by five species. Ten species of endoparasites were represented by the total of 4,212 specimens.

### Nematoda

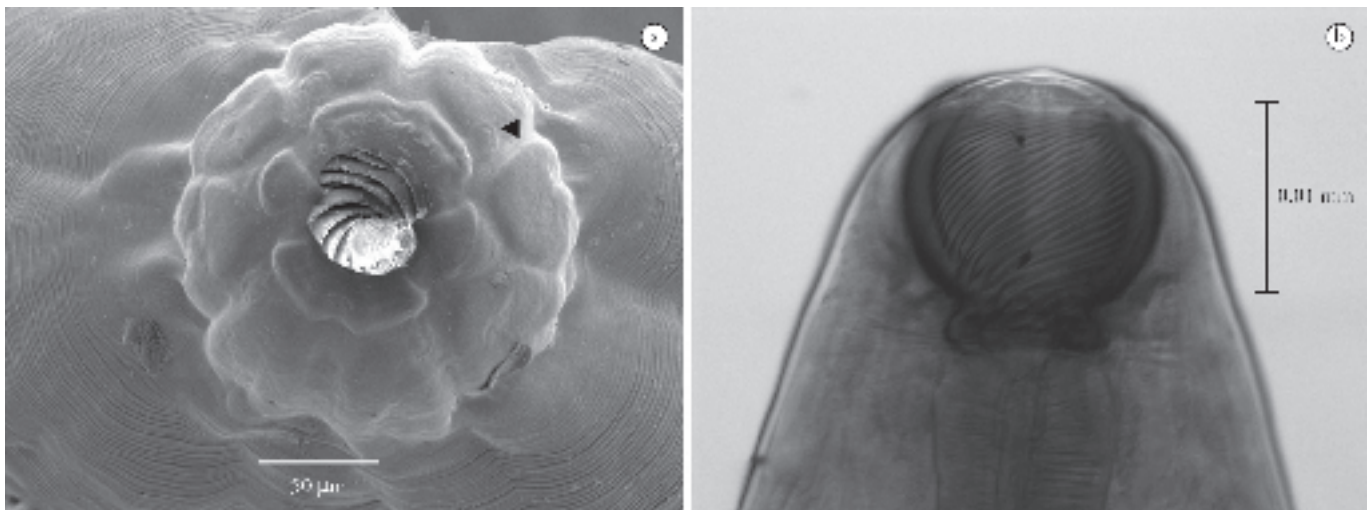
#### Camallanidae Railliet & Henry, 1915

##### *Procamallanus* Baylis, 1923

#### *Procamallanus* (*Spirocamallanus*) *inopinatus* Travassos, Artigas & Pereira, 1928 (ZUFMS-INV = 008)

Site of infection: cecum and intestine; prevalence 22.37%; mean abundance  $0.36 \pm 0.82$ ; mean intensity  $1.64 \pm 0.98$ ; range of variation: 1-5.

Based on 10 adult female specimens: Body 19.72 (12.35-24.28) long, 0.678 (0.546-0.806) largest width; oral opening surrounded by four cephalic papillae (Figure 1a) and two amphids; two median teeth (dorsal and ventral); buccal capsule including basal ring 0.141 (0.131-0.145) length and 0.16 (0.146-0.17) largest width, with 20-23 spiral thickenings in buccal capsule (Figure 1b); muscular-glandular oesophagus, short muscular portion 0.517 (0.49-0.59) length, 0.192 (0.17-0.21) largest width, oesophagus base 0.091 (0.08-0.1) width; glandular portion 0.869 (0.784-0.963) length, 0.241 (0.181-0.296) largest width, base 0.112 (0.095-0.143) width; nerve ring 0.276 (0.27-0.286) and excretory pore 0.365 (0.325-0.387) to anterior end; vulva 9.35



**Figure 1.** Micrographs of *Procamallanus* (*Spirocamallanus*) *inopinatus*: **A** – SEM micrograph of anterior end, apical view showing four cephalic papillae (black arrow). **B** – LM micrograph of lateral anterior end showing the spiral thickenings in the buccal capsule.

(7.562-10.48) to posterior end, equal to 45.54% (43.16-47.8%) of total body length; tail conical 0.243 (0.204-0.305) length.

**Remarks:** The number of spiral thickenings in the buccal capsule of the specimens examined is close to that reported by Moravec (1998). However, the number of spiral thickenings differs among several reports, and these differences are considered as intraspecific variability (MORAVEC et al., 1997). The specimens described by Petter and Dlouhy (1985) showed 14-25 spiral thickenings. Rodrigues et al. (1991) and Moravec et al. (1993, 1997) reported 15-19, 8-17 and 13-22 spiral thickenings, respectively.

Moravec et al. (1997) reported the presence of eight cephalic papillae arranged in two circles around the oral opening, differing from the present specimens which have only four cephalic papillae. Some specimens were compared to voucher specimen CHIOC 31.324, parasitic in *Leporinus* sp., from the Machado River, state of Rondônia, Brazil (GIESE et al., 2009). Similarities in the number of the spiral thickenings were observed, and although the voucher specimen was not measured, it was visibly longer and wider than our specimens. This voucher specimen is permanently mounted, and due to the impossibility of *en face* observation and the poor conservation state of the specimen, it was not possible to observe the number of cephalic papillae.

Several fish species were reported as being parasitized by *P. (S.) inopinatus* (MORAVEC, 1998; PAVANELLI et al., 2004), including *P. nattereri* (EIRAS et al., 2010).

#### **Philometridae Baylis & Daubney, 1926**

##### **Philometridae gen. sp. (ZUFMS-INV = 013)**

Site of infection: body cavity; prevalence 0.66%; mean abundance  $0.006 \pm 0.08$ ; intensity 1.

Based on one adult female specimen: Long and slender body, 15.39 length, 0.71 width; cuticle smooth, thin and fragile; mouth simple, buccal capsule absent; six cephalic papillae around oral opening; female gravid, viviparous, bulky uterus. Larvae 0.337 (0.189-0.452) length and 0.017 (0.014-0.230) width.

**Remarks:** Genus and species determination was not possible owing to the poor condition of this specimen. The family identification is according to the features of an adult female philometrid as described by Moravec (1998): anterior end rounded, peribuccal ring absent; mouth simple, without buccal capsule; oral opening surrounded by 6-8 cephalic papillae. The vulva and vagina were not observed in this specimen; Moravec (1998) mentioned that these structures are atrophied in gravid females. Recently, *Philometra nattereri* was described parasitizing *P. nattereri* in Amazonia (CÁRDENAS et al., 2012). Although the specimen recorded in the present study has not been identified, it differs in some aspects from the one described by Cárdenas et al. (2012), who reported a narrower range of widths (0.47-0.65) and 14 cephalic papillae. The infection site, also different, was reported as in the oculo-orbits and nasal cavity.

#### **Dioctophymatidae Railliet, 1915**

##### ***Eustrongylides* Jägerskiöld, 1909**

##### ***Eustrongylides* sp. (ZUFMS-INV 015)**

Site of infection: mesentery; prevalence 1.32%; mean abundance  $0.013 \pm 0.11$ ; mean intensity 1.

Based on two larval specimens: Body brownish, 58.27 (49.5-66.69) length, 0.70 (0.692-0.72) width; anterior end rounded; buccal cavity 0.057 (0.048-0.067) long, surrounded

by cephalic papillae arranged in circle; cylindrical oesophagus 3.4 (2.2-4.7) long; nerve ring 0.50 (0.09-0.11) from anterior end; anus posterior and terminal.

**Remarks:** The body length and width measurements are close to those reported for larval *Eustrongylides ignotus* Jägerskiöld, 1909, while the oesophagus length is close to those of larvae of *E. tubifex* (Nitzsch, 1819) (MORAVEC, 1998) (we found no report of the oesophagus length in *E. ignotus*). The buccal cavity is longer than that of *E. tubifex* (0.012) and closer to that reported for *E. ignotus* (0.060-0.097). Moravec et al. (1997), studying nematodes of freshwater fishes in Venezuela, described one larva of *Eustrongylides* sp., and reported a buccal cavity length (0.285) five times longer than in the present specimens.

It was not possible to count the cephalic papillae or to determine if the papillae of the outer circle were larger than the papillae of the inner circle, because of the condition of the specimens; these features enable the species determination of *Eustrongylides* according to Moravec (1998) and Thatcher (2006). Species determination of larval specimens belonging to the genus *Eustrongylides* Jägerskiöld, 1909 is problematic, and the surest way to identify the species is to obtain adult forms from experimental infection of birds, which are the definitive hosts (MORAVEC, 1998). Larval stages of this genus are recorded in a wide variety of fish species (MORAVEC, 1998; PAVANELLI et al., 2004), and larvae of *E. ignotus* were previously recorded in high prevalence in *P. nattereri* (= *Serrasalmus nattereri*) (MORAVEC, 1998).

#### **Acanthocheilidae Wülker, 1929**

##### ***Brevimulticaecum* Mozgovoy, 1951**

##### ***Brevimulticaecum* sp. larvae (ZUFMS-INV = 009)**

Site of infection: stomach tissue and mesentery; prevalence 19.08%; mean abundance  $22.36 \pm 71.24$ ; mean intensity  $117.24 \pm 125.91$ ; range of variation: 1-485.

Based on 10 larval specimens: Body 3.75 (3.07-4.1) length, 0.1 (0.072-0.137) width; ratio body length/width 2.6% (1.9-3.5); anterior end with four papillae, two subdorsal and two subventral and two tooth-like prominences; amphids between the lateral subdorsal and subventral papillae; oesophagus 0.401 (0.156-0.59) length, 0.018 (0.011-0.024) width at the base; comprising 10.48% (5.15-15.66) of body length; right ventricular lobes 0.06 (0.044-0.072) length, 0.041 (0.038-0.047) width; ratio right ventricular lobes length/oesophagus length 0.163 (0.103-0.322); left ventricular lobes 0.061 (0.043-0.075) length, 0.04 (0.036-0.044) width; ratio left ventricular lobes length/oesophagus length 0.165 (0.098-0.308); intestinal caecum 0.4 (0.329-0.486) length, 0.036 (0.032-0.04) width; nerve ring 0.183 (0.15-0.25) and excretory pore 0.198 (0.17-0.21) from anterior extremity; excretory nucleus situated about halfway along the intestinal caecum, 0.391 (0.28-0.48) from anterior extremity; conical tail 0.075 (0.041-0.08) length.

**Remarks:** The measurements indicate a wider range of values than reported by Moravec (1998). The first report of a fish parasitized by *Brevimulticaecum* sp. in the adult stage was in a stingray *Paratrygon motoro* (Müller & Henle, 1841) in the Salobra River, Mato Grosso by Rego (1979). This species was described as *Brevimulticaecum regoi* by Sprent (1990). In the Neotropical region, larvae of *Brevimulticaecum* sp. have been recorded in *Gymnotus carapo* Linnaeus, 1758 and *Loricariichthys brunneus* (Hancock, 1828) in Venezuela (MORAVEC et al.,



1997); in *Leporinus friderici* (Bloch, 1794), *L. lacustris* Amaral Campos, 1945, *L. obtusidens* (Valenciennes, 1837), *L. elongatus* Valenciennes, 1850 (GUIDELLI et al., 2006) and in *Potamotrygon falkneri* Castex & Maciel, 1963 (LACERDA et al., 2008) in the Upper Paraná River floodplain; and recently in the Upper Paraguay River basin, in *Gymnotus inaequilabiatus* (Valenciennes, 1839), *Hoplias* aff. *malabaricus* (Bloch, 1794), *Hemisorubim platyrhynchos* (Valenciennes, 1840), *Myleus levis* (Eigenmann & McAtee, 1844), *Pseudoplatystoma corruscans* (Spix and Agassiz, 1829), *P. nattereri* Kner, 1858 (VIEIRA et al., 2010) and *Serrasalmus marginatus* Valenciennes, 1837 (VIEIRA et al., 2010; VICENTIN et al., 2011).

Some of the present specimens were compared with the voucher specimen CHIOC 36.977, from *Potamotrygon falkneri* Castex & Maciel, 1963, collected in the Paraná River, State of Paraná, Brazil (LACERDA et al., 2008). In this specimen, the ventriculus could not be observed due to the overlapping of the body parts in the permanent mount.

#### **Anisakidae Railliet & Henry, 1912**

##### ***Contracaecum* Railliet & Henry, 1912**

##### ***Contracaecum* sp. (ZUFMS-INV = 010)**

Site of infection: stomach tissue, cecum and mesentery; prevalence 61.84%; mean abundance  $3.55 \pm 7.94$ ; mean intensity  $5.75 \pm 9.46$ ; range of variation: 1-74.

Based on 10 larval specimens: Body 20.19 (15.11-27.25) length, 0.695 (0.617-0.798) width; ratio body length/width 3.5% (2.5-4.08); transversely striated cuticle; larval tooth in anterior end below subventral lips; excretory pore in anterior end, between subventral lips; nerve ring 0.22 (0.16-0.28) from anterior extremity; narrow oesophagus 2.103 (1.7-2.93) long, comprising 10% (7-15) of body length; ventriculus 0.156 (0.114-0.243) length, 0.125 (0.076-0.169) width; ratio ventriculus/oesophagus length 0.07 (0.05-0.11); posterior ventricular appendix 0.51 (0.40-0.77) length; intestinal cecum 1.1 (1.42-2.74) length; ratio of ventriculus length and ventricular appendix length to intestinal cecum length 0.09 (0.05-0.14) and 0.29 (0.21-0.49), respectively; conical tail 0.20 (0.14-0.26).

**Remarks:** The measurements of *Contracaecum* sp. larvae in this study are close to those of *Contracaecum* sp. type 2 of Moravec (1998), with differences in the length and width of the ventriculus and in the length of the ventricular appendix (greater in the present specimens). Some specimens were compared with the voucher specimen CHIOC 35.521, from *Geophagus brasiliensis* (Quoy & Gaimard, 1824), collected in the Guandu River, State of Rio de Janeiro, Brazil (AZEVEDO et al., 2006), which is larger than the specimens found in the present study. *Contracaecum* sp. larvae have been reported in several fish species (MORAVEC et al., 1997; MORAVEC, 1998; MARTINS et al., 2005), including *P. nattereri* (EIRAS et al., 2010).

#### **Acanthocephala**

##### **Echinorhynchidae Cobbold, 1879**

##### ***Echinorhynchus* Zoega in Müller, 1776**

##### ***Echinorhynchus paranensis* Machado Filho, 1959 (ZUFMS-INV = 014)**

Site of infection: intestine; prevalence 0.66%; mean abundance  $0.006 \pm 0.08$ ; intensity 1.

Based on one immature adult female specimen: Trunk elongate, slender, 12.19 long, 2.31 largest width; anterior end wider than

posterior end; proboscis 1.38 long, 0.36 wide, armed with 14 longitudinal rows of 11 hooks each; proboscis receptacle double-walled, 1.56 length, 0.47 width; lemnisci 1.001 length; ovijector 0.89 length, selector apparatus 0.544 length.

**Remarks:** The dimensions of the trunk, proboscis and proboscis receptacle, number of rows and hooks, as well as the dimensions of the lemnisci and ovijector, are very similar to those reported for *E. paranensis* by Machado Filho (1959). According to Machado Filho (1959), the main difference observed between *E. paranensis* and other congeneric species is the presence of 14 longitudinal rows with 11 hooks per row on the proboscis. This specimen was considered adult, because it had well-developed reproductive structures, but non-gravid because it did not contain eggs. This is the first report of this species in *P. nattereri*.

#### **Pentastomida Diesing, 1836**

##### **Sebekidae Sambon, 1922**

##### ***Leiperia* Sambon, 1922**

##### ***Leiperia gracile* Diesing, 1835**

Site of infection: body cavity; prevalence 3.29%; mean abundance  $0.03 \pm 0.17$ ; intensity 1.

Based on 6 larval specimens: Body with anterior end rounded and tapering slightly at the posterior end, 21.53 (20.7-22.11) length, 0.89 (0.882-0.924) width, with 103 (100-107) distinct annuli with spines on their edges, spines 0.012 (0.009-0.014) length; anterior end with two pairs of double-hooks (Figure 2a), each with a principal hook and another smaller accessory; anterior pair of hooks with principal hook 0.24 (0.228-0.245) total length, and accessory hook 0.169 (0.144-0.178) total length; anterior pair of hooks, blade length (ab) = 0.201 (0.15-0.211), base length (ac) = 0.104 (0.091-0.13), base to extremity of hook blade (bc) = 0.84 (0.068-0.11) (Figure 2b); posterior pair of hooks, principal and accessory hooks measuring, respectively, 0.245 (0.24-0.26) and 0.141 (0.139-0.169) total length; blade length (ab) = 0.19 (0.18-0.234), base length (ac) = 0.098 (0.06-0.121), base to extremity of hook blade (bc) = 0.119 (0.103-0.134); oral cadre slightly chitinized, 0.253 (0.237-0.268) length, 0.131 (0.109-0.152) width, located between the pairs of hooks; anus posterior and terminal.

**Remarks:** The measurements of these specimens are in agreement with those reported for larval stages of *L. gracile* in *Salminus brevidens* (syn. = *S. brasiliensis*, Cuvier, 1819) and are slightly larger than those for larvae found in *H. malabaricus* and *Brachyplatystoma* sp. Bleeker, 1862 described by Rego and Eiras (1989). The principal hooks of these specimens are larger than the accessory hooks, differing from specimens found in *S. brevidens* and *H. malabaricus*, where the accessory hooks were larger than the principal hooks (REGO; EIRAS, 1989). Some individuals were compared to the voucher specimens CHIOC 32.446, parasitic in *Brachyplatystoma* sp., from the Salobra River, State of Mato Grosso, Brazil; CHIOC 30.353; parasitic in *H. malabaricus*, from the State of Espírito Santo, Brazil; and CHIOC 29.889a-b, parasitic in *Salminus brevidens* from the Salobra River (REGO; EIRAS, 1989).

Although some of these specimens were darkened and in permanent mounts, it was possible to observe similarities among

them, including the body length, number of annuli (more than 100) and the characteristics of the oral cadre, although the accessory hooks visually appeared to be smaller than the principal hooks in the voucher specimens. The report of this parasite in *P. nattereri* by Eiras et al. (2010) was based on the dissertation that reported the first record in this host and formed the basis of the present study, but Eiras et al. (2010) did not report any description of specimens, collection locality, or parasitological descriptors.

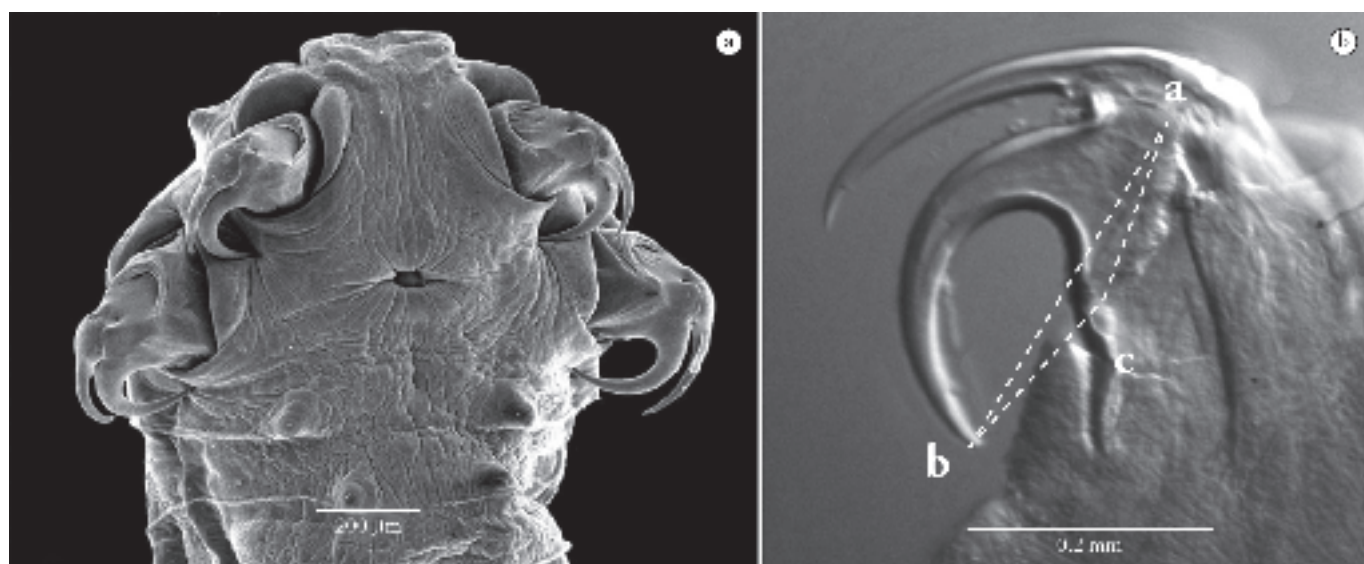
#### **Sebekidae Sambon, 1922**

#### ***Sebekia* Sambon, 1922**

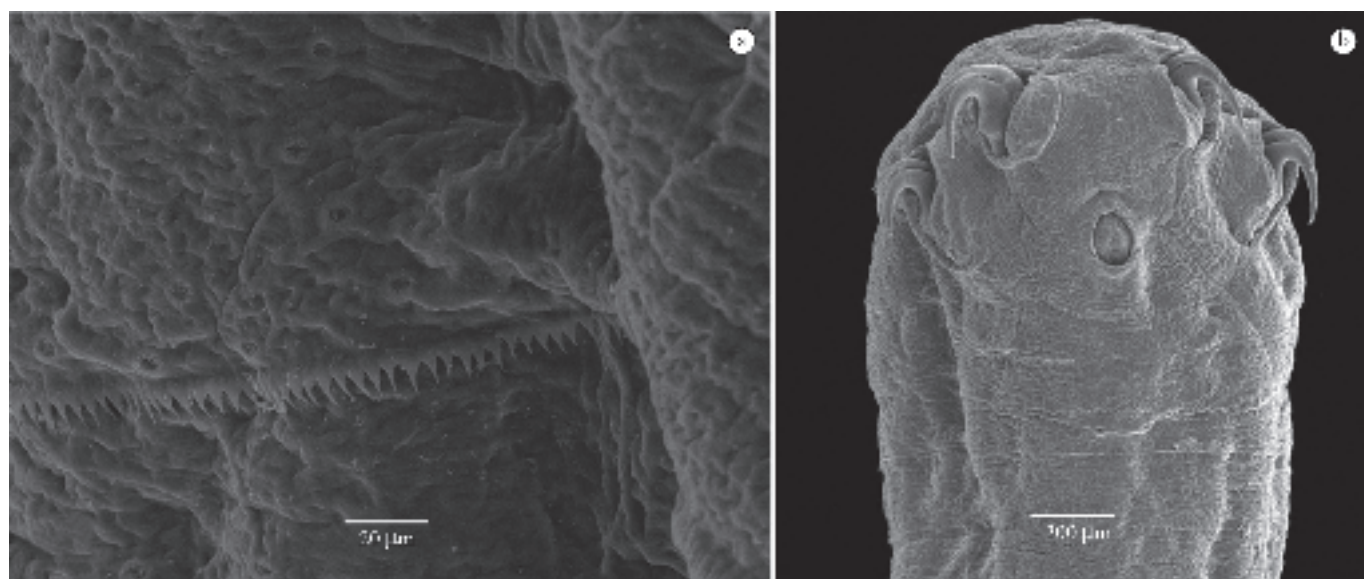
#### ***Sebekia oxycephala* Diesing, 1835 (ZUFMS-INV = 007)**

Site of infection: body cavity; prevalence 29.61%; mean abundance  $0.69 \pm 1.41$ ; mean intensity  $2.33 \pm 1.70$ ; range of variation: 1-8.

Based on 10 larval specimens: Body with anterior end rounded and tapering slightly at the posterior end, 6.7 (5.136-9.463) length, 0.832 (0.68-1.03) width, with 63 (56-75) distinct annuli with spines on their edges, spines 0.022 (0.017-0.025) length (Figure 3a); anterior end with two pairs of double-hooks, each with a principal hook and another smaller accessory (Figure 3b); anterior pair of hooks with principal hook 0.084 (0.061-0.125) total length and accessory hook 0.071 (0.048-0.113) total length; blade length (ab) = 0.073 (0.057-0.102), base length (ac) = 0.046 (0.028-0.079), base to extremity of hook blade (bc) = 0.036 (0.026-0.047); posterior pair of hooks with principal and accessory hooks measuring, respectively, 0.086 (0.06-0.121) and 0.067 (0.034-0.102) total length; blade length (ab) = 0.08 (0.059-0.102), base length (ac) = 0.048 (0.038-0.06), base to



**Figure 2.** Micrographs of *Leiperia gracile*: **A** – SEM micrograph of anterior end, ventral view. **B** – LM micrograph showing the main and accessory hooks; (ab) = length of hook; (ac) = hook base; (bc) = length from base to extremity of hook blade.



**Figure 3.** SEM micrographs of *Sebekia oxycephala*: **A** – detail of annulus with spines on its edges. **B** – anterior end, ventral view.



extremity of hook blade (bc) = 0.052 (0.03-0.088); oral cadre slightly chitinated, 0.134 (0.115-0.16) length, 0.066 (0.047-0.078) width, located between pairs of hooks; anus posterior and terminal.

**Remarks:** The measurements and some morphological features of *S. oxycephala* are close to those reported by Rego and Eiras (1989), mainly in relation to body length, hooks and number of annuli. In this study, specimens of *S. oxycephala* had the principal hooks larger than the accessory hooks, differing from the description by Rego and Eiras (1989) for this same species in *P. nattereri*. Some of the specimens were compared to voucher specimens CHIOC 32.445, parasitic in *P. nattereri*, from the Cuiabá River, State of Mato Grosso, Brazil and CHIOC 32.447, parasitic in *P. corruscans*, also from the Cuiabá River (REGO; EIRAS, 1989). These specimens were similar in the shape of hooks, oral cadre, body dimensions and number of annuli.

Despite the observed similarity between the larvae of *S. oxycephala* and *L. gracile*, these larvae can be distinguished mainly by the body size, approximately three times longer; by the number of body annuli, approximately twice as numerous in

*L. gracile*; and by the shape of the oral cadre, oval to elongate in *Sebekia* spp. and U-shaped in *Leiperia* spp. (OVERSTREET et al., 1985; REGO; EIRAS, 1989; JUNKER et al., 2000).

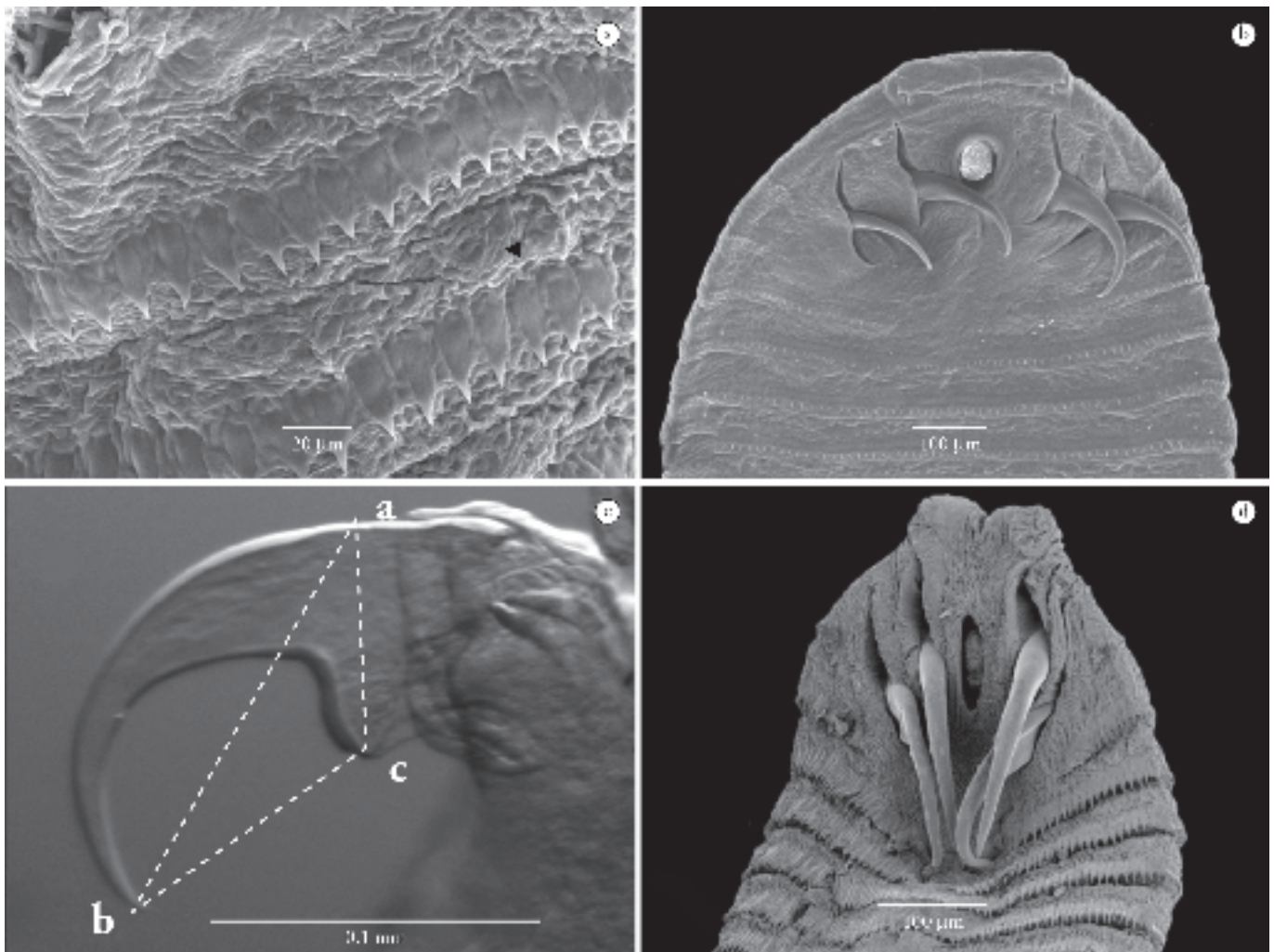
#### **Subtriquetrinae Fain, 1961**

#### **Subtriquetra Sambon, 1922**

#### **Subtriquetra sp. 1 (ZUFMS-INV = 011)**

Site of infection: swim bladder; prevalence 6.58%; mean abundance  $0.09 \pm 0.48$ ; mean intensity  $1.5 \pm 1.26$ ; range of variation: 1-5.

Based on 10 larval specimens: Larvae with bright red body color while in the host; body elliptical, ventrally flattened and dorsally convex; anterior end wider than posterior end; body 2.10 (1.562-2.528) length, 0.651 (0.556-0.779) largest width, with 30 (28-33) distinct annuli with spines on their edges, spines 0.025 (0.022-0.029) length, chloride cells arranged in single row in each annulus (Figure 4a); anterior end with two symmetrical cephalic papillae, and two pairs of single hooks (Figure 4b); anterior pair of hooks 0.164 (0.129-0.206) total length, blade length (ab) = 0.151 (0.111-0.201), base length (ac) = 0.109 (0.073-0.138),



**Figure 4.** Micrographs of *Subtriquetra* spp.: **A** – SEM micrograph showing rows of spines and chloride cells (black arrow). **B** – SEM micrograph of anterior end, ventral view of *Subtriquetra* sp. 1. **C** – LM micrograph showing measurements of simple hook (ab) = length of hook blade; (ac) = hook base; (bc) = length from base to extremity of hook blade. **D** – SEM micrograph of anterior end, ventral view of *Subtriquetra* sp. 2.

base to extremity of hook blade (bc) = 0.076 (0.054-0.092) (Figure 4c); posterior pair of hooks 0.174 (0.131-0.22) total length, blade length (ab) = 0.162 (0.122-0.213), base length (ac) = 0.107 (0.076-0.126), base to extremity of hook blade (bc) = 0.08 (0.057-0.107); oral cadre 0.128 (0.112-0.141) length, 0.068 (0.057-0.078) width, located between the first pairs of hooks.

#### ***Subtriquetra* sp. 2** (ZUFMS-INV = 012)

Site of infection: swim bladder; prevalence 22.37%; mean abundance  $0.55 \pm 1.69$ ; mean intensity  $2.5 \pm 2.86$ ; range of variation: 1-15.

Based on 10 larval specimens (Figure 4d): Larvae with whitish body while in the host; body elliptical, slightly flattened; anterior end wider than posterior; body 2.56 (2.193-3.21) length, 0.891 (0.781-1.08) largest width, with 30 (28-34) distinct annuli with spines on their edges, spines 0.029 (0.028-0.032) length; chloride cells arranged in single row in each annulus; anterior end with two symmetrical cephalic papillae, and two pairs of single hooks; anterior pair of hooks 0.211 (0.176-0.256) total length, blade length (ab) = 0.197 (0.164-0.231), base length (ac) = 0.14 (0.115-0.153), base to extremity of hook blade (bc) = 0.086 (0.077-0.101); posterior pair of hooks 0.199 (0.157-0.253) total length, blade length (ab) = 0.191 (0.154-0.237), base length (ac) = 0.133 (0.113-0.157), base to extremity of hook blade (bc) = 0.082 (0.066-0.098); oral cadre 0.153 (0.125-0.175) length, 0.097 (0.083-0.110) width, located between the first pairs of hooks.

**Remarks:** According to Junker and Boomker (2006), the features observed in these two morphospecies are in agreement with those of the genus *Subtriquetra* Sambon, 1922 including the elliptical and dorsally flattened body, and the rounded oral cadre located between two pairs of simple, slender and sharply pointed hooks.

Records of larval stages of species of *Subtriquetra* parasitizing fish are scarce, and are mainly from South America, which prevented the specific identification of this morphospecies. These two morphotypes were considered distinct because of the differences in the shape of the oral cadre, hooks and spines, structures that were larger in *Subtriquetra* sp. 2.

Some collected specimens were compared to some of the voucher specimens of the genus *Subtriquetra*, CHIOC 17.797, parasitic in *H. malabaricus*, from the Juparaná Lagoon, State of Espírito Santo, Brazil (TRAVASSOS; FREITAS, 1940) and CHIOC 11.424, parasitic in *P. nattereri*, from the Salobra River, State of Mato Grosso do Sul, Brazil (unknown publication). These parasites were similar in shape and dimensions of the body and oral cadre, and in the presence of ventral papillae on the anterior end of the body.

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