



Revista Brasileira de Parasitologia
Veterinária

ISSN: 0103-846X

zacariascbpv@fcav.unesp.br

Colégio Brasileiro de Parasitologia
Veterinária
Brasil

Schoenardie, Elizandra Roselaine; Scaini, Carlos James; Soares Pepe, Michele; Borsuk, Sibebe; Farias da Costa de Avila, Luciana; Villela, Marcos; Aires Berne, Maria Elisabeth

Vertical transmission of *Toxocara canis* in successive generations of mice

Revista Brasileira de Parasitologia Veterinária, vol. 22, núm. 4, octubre-diciembre, 2013,
pp. 623-626

Colégio Brasileiro de Parasitologia Veterinária
Jaboticabal, Brasil

Available in: <http://www.redalyc.org/articulo.oa?id=397841490029>

- How to cite
- Complete issue
- More information about this article
- Journal's homepage in redalyc.org

redalyc.org

Scientific Information System

Network of Scientific Journals from Latin America, the Caribbean, Spain and Portugal

Non-profit academic project, developed under the open access initiative

Vertical transmission of *Toxocara canis* in successive generations of mice

Transmissão vertical de *Toxocara canis* em gerações sucessivas de camundongos

Elizandra Roselaine Schoenardie¹; Carlos James Scaini²; Michele Soares Pepe¹; Sibe Borsuk³;
Luciana Farias da Costa de Avila¹; Marcos Villela¹; Maria Elisabeth Aires Berne^{1*}

¹Departamento de Microbiologia e Parasitologia, Instituto de Biologia, Universidade Federal de Pelotas – UFPEL, Campus Capão do Leão, Pelotas, RS, Brasil

²Programa de Pós-graduação em Ciências da Saúde, Universidade Federal do Rio Grande – FURG, Rio Grande, RS, Brasil

³Núcleo de Biotecnologia, Centro de Desenvolvimento Tecnológico, Universidade Federal de Pelotas – UFPEL, Pelotas, RS, Brasil

Received August 19, 2013

Accepted November 5, 2013

Abstract

Visceral toxocaríasis is a neglected zoonosis caused by *Toxocara canis* larvae in unusual hosts. In dogs, the definitive host, the infection occurs mainly through transplacental and transcolostral transmission. Studies on experimental models have shown that vertical transmission may result from acute infections. Considering that toxocaríasis is characterized as a chronic infection, with possible reactivation of larvae present in the brain, this study evaluated the presence of larvae in the brain of female BALB/c mice and their offspring with chronic infection during three successive pregnancies. ELISA-TES was used to evaluate the antibody levels. *T. canis* larvae were detected in the brain tissue of the mice during the three successive generations evaluated. The offspring's IgG level gradually decreased, and mean absorbance (ABS) above the cutoff point (0.070) was observed only at 30 (0.229) and 50 (0.096) days of age, while IgM was not detected. The infections in the offspring confirmed that vertical transmission of *T. canis* larvae occurred during chronic toxocaríasis in three successive generations of mice.

Keywords: Visceral larva *migrans*, *Toxocara canis*, vertical transmission, BALB/c mice, ELISA - TES.

Resumo

A toxocaríase visceral é uma zoonose negligenciada causada por larvas de *Toxocara canis* em hospedeiros não usuais. Em cães, os hospedeiros definitivos, a infecção ocorre normalmente por transmissão transplacentária e através do colostro. Estudos com modelos experimentais têm demonstrado a ocorrência de transmissão vertical durante a infecção aguda. Considerando que a toxocaríase é caracterizada como uma infecção crônica, com uma possível reativação das larvas presentes no cérebro, este estudo avaliou a presença de larvas no cérebro de camundongos Balb/C fêmeas e suas proles com infecção crônica durante três gestações sucessivas. Para avaliar os níveis de anticorpos foi utilizado ELISA-TES. Larvas de *T. canis* foram detectadas no encéfalo dos animais durante as três gerações sucessivas avaliadas. O nível de IgG das proles foi diminuindo gradualmente e as médias de absorbâncias (ABS) acima do ponto de corte (0,070) foram evidenciadas somente aos 30 (0,229) e 50 dias (0,096) de vida, enquanto que não foi detectada IgM. Infecções das proles confirmam a transmissão vertical de larvas de *T. canis* durante a toxocaríase crônica em três gerações sucessivas de camundongos.

Palavras-chave: Larva *migrans* visceral, *Toxocara canis*, transmissão vertical, camundongos BALB/c, ELISA - TES.

Visceral toxocaríasis, also known as visceral larva *migrans* (VLM) syndrome, is defined as the migration and persistence of helminthic larvae in tissues of unusual hosts (BEAVER, 1969),

and it is a zoonosis that presents a continuing public health risk. The nematode *Toxocara canis*, a small intestine parasite in dogs, is the etiological agent most commonly associated with this zoonosis. In dogs, the zoonotic profile is maintained by transplacental and transmammary infection (BURKE; ROBERSON, 1985), in which the young animals are more susceptible and also the main disseminators of eggs in the environment (GALLINA et al., 2011; OLIVEIRA-SEQUEIRA et al., 2002; VILLELA et al., 2009).

*Corresponding author: Maria Elisabeth Aires Berne
Departamento de Microbiologia e Parasitologia, Instituto de Biologia,
Universidade Federal de Pelotas – UFPEL, Campus Universitário, s/n.,
CP 354, CEP 96010-900, Pelotas, RS, Brasil
e-mail: bernemea@gmail.com

In humans, visceral toxocariasis is characterized by chronic infection that can persist for several years with possible reactivation and larval migration to the eyes and brain (PAWLOWSKI, 2001). The clinical symptoms of toxocariasis vary depending on the tissue parasitized, the number of eggs ingested and the immune response of the host (GLICKMAN et al. 1979). Epidemiological studies in various countries have shown that toxocariasis is common in humans (CHIEFFI et al., 2009; COLLI et al., 2010; SARIEGO et al., 2012; ALVARADO-ESQUIVEL, 2013). Accidental ingestion of embryonated *T. canis* eggs is the main infection path in humans, but is not the only route. Larvae present in undercooked meats may also cause infection (HOFFMEISTER et al., 2007; DUTRA et al., 2013).

Almost two decades ago, Anderson (1996) warned of the need to pay attention to pregnant women, because *T. canis* larvae could be transmitted to the fetus when the mother acquired an infection during pregnancy. A few years later, a case of congenital newborn infection was recorded in Argentina, in a premature infant with retinopathy (MAFFRAND et al., 2006). To simulate this form of transmission in the human hosts, mice have been used (REITEROVÁ et al., 2003; JIN et al., 2008), since they have the same migration behavior and develop the disease in the same way as in accidental hosts (CAMPAROTO et al., 2008; HOLLAND; COX, 2001). To assess the importance of this infection route, the present study evaluated vertical transmission of *T. canis* in BALB/c mice with chronic infections during three successive pregnancies.

T. canis eggs were collected from the uterine tubes of adult female parasites that had been obtained through treating young dogs with pyrantel pamoate (15 mg/kg). Afterwards, the eggs were incubated in a 2% formalin solution at 28°C for 30 days. Experimental infection was performed in five female BALB/c mice (at 8 weeks of age), which was done through intraperitoneal inoculation of 1,200 *T. canis* larvae. Ninety days after the inoculation (chronic infection), they were mated with males of the same age and observed for a period of 120 days. Blood was collected on the first mating day and from the three offspring sets at 30, 50, 70, 90, and 110 days of age. The presence of *T. canis* larvae in the brain tissue of the female mice was evaluated 120 days after mating (210 days after infection), and in their respective offspring (43 animals in total) at 110 days of age, from the three successive generations. To investigate the presence of *T. canis* infection in the female mice and their offspring, brain fragments were compressed between glass slides and examined under a microscope at 100× (KAYES; OAKS, 1976). The IgG and IgM levels were

evaluated using ELISA and TES antigen production, following the protocol of De Savigny (1975). This study was approved by the Ethics Committee for Animal Experimentation at the Federal University of Pelotas (CEEa no. 6554). The occurrence of vertical transmission was calculated. For statistical analysis, we used the Minitab software, version 15, and used Student's t test. P-value greater than 0.05 were considered to be not significant.

In this study, *T. canis* larvae was detected in all the three successive generations of mice evaluated (Table 1). However, the variation in the numbers of larvae diagnosed in the brain of each female mouse indicated that there was no statistically significant correlation between the number of mice born in the three generations and the number of larvae found in the brains of these offspring ($p > 0.05$). The larvae detected in the brain tissue (total of 141 larvae) were alive, thus confirming that there was chronic infection among the female mice. At the first mating, corresponding to 90 days of infection, the female mice had high levels of IgG and IgM ($p < 0.05$), and mean ABS of 1.280 and 0.685 respectively, which is typical of the chronic phase. According to Fan et al. (2003), IgG levels are maintained in sera from infected mice for long periods (up to 67 weeks post-infection).

In the offspring, the IgG levels decreased from the age of 30 day to 50 days, and greater reductions were detected at 70 days (Figure 1), thus characterizing vertical transfer of maternal antibodies (YAMASHITA et al., 2006). Moreover, the IgM levels were not significant, remaining lower than the cutoff from 30 to 110 days of age (Table 2).

Similar results were obtained by Bowman et al. (1987) who observed higher levels of IgM (between 42 and 70 days) and IgG (between 70 and 90 days) in female mice with chronic infection.

In the present study, out of the five female mice with chronic infections, three had *T. canis* larvae in their brain tissues and in their offspring's brain tissue: two from the first generation, one from the second generation and two from the third generation of offspring. All the larvae detected showed motility and no tissue reaction was found. Similar results have been obtained from mice 122 days after infection, in which viable *T. canis* larvae were detected in brain tissues with no correlated inflammatory reaction (DUNSMORE et al., 1983; FAN et al., 2003).

The re-emergence of *T. canis* larvae during pregnancy and vertical transmission in dogs are well known in relation to the biological cycle of this nematode (BURKE; ROBERSON, 1985). We demonstrated the importance of vertical transmission of *T. canis* in successive generations of mice with chronic infection.

Table 1. *Toxocara canis* larvae in brain tissue from female BALB/c mice and their offspring with chronic infection.

Infected female mice	No. of larvae	No. of mice born from three generations			No. of larvae in the brain of offspring in three generations		
		1 st	2 nd	3 rd	1 st	2 nd	3 rd
1	10	5	3	2	1	1	0
2	3	2	5	1	1	0	0
3	54	5	1	2	0	0	0
4	18	2	2	3	0	0	0
5	34	5	1	4	0	0	2
Total	141	19	12	12	2	1	2

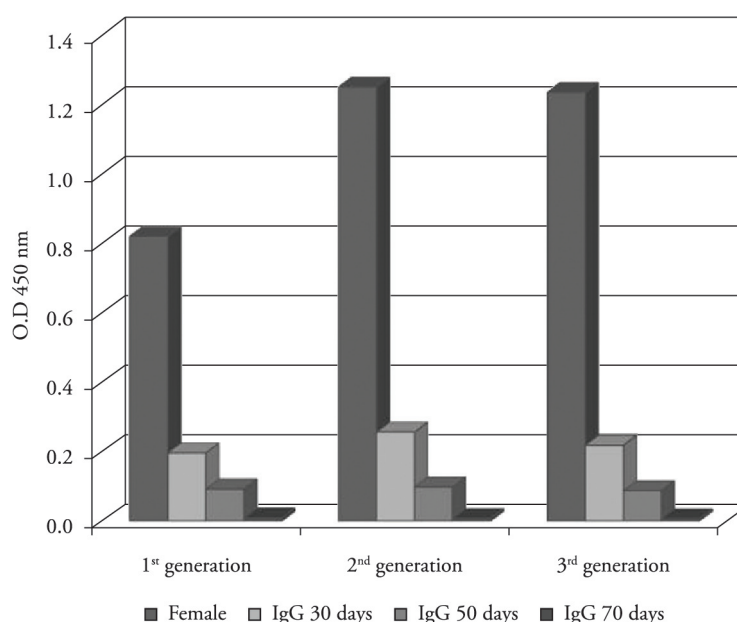


Figure 1. ELISA-TES on female BALB/c mice on the day of first mating, and their offspring at 30, 50 and 70 days of age, in three consecutive pregnancies.

Table 2. Immunoglobulin production against *Toxocara canis* detected by ELISA in offspring BALB/c mice from three successive generations, correlated with the presence or absence of *T. canis* larvae in the brain tissue. The values of IgG and IgM represent the mean value.

<i>T. canis</i> larvae	No. of animals	ELISA/IgG			ELISA/IgM		
		day 30	day 50	day 70	day 30	day 50	day 70
Presence	5	0.281	0.103	0.019	0.035	0.014	0.013
Absence	38	0.223	0.095	0.015	0.028	0.018	0.017

ELISA/IgG cutoff: 0.070 ELISA/IgM cutoff: 0.066.

Most studies on vertical transmission of *T. canis* have been conducted during the acute phase of infection (LEE et al., 1976; REITEROVÁ et al., 2003; JIN et al., 2008), although there is one record of vertical transmission of *T. canis* during the chronic phase in BALB/c mice (AVILA et al., 2009). With infections occurring during pregnancy, the likelihood that *T. canis* larvae will reach the fetus is greater because of their presence (during the acute phase) in liver and lung tissue (ABO-SHEHADA; HERBERT, 1984). In the chronic phase, which usually begins 40 days after infection, the accumulation of *T. canis* larvae in the brain favors vertical transmission of the parasite, since the larvae may remain viable in this tissue for months or even years (DUNSMORE, 1983). The infections described in the offspring demonstrated that vertical transmission occurred in these mice presenting chronic toxocariasis during successive pregnancies. However, determination of the transmission route, which could be either transplacental or transcolostral, and the related immunological responses, remains necessary.

References

- Abo-Shehadeh MN, Herbert IV. The migration of larval *Toxocara canis* in mice. II. Post-intestinal migration in primary infections. *Vet Parasitol* 1984; 17(1):75-83. [http://dx.doi.org/10.1016/0304-4017\(84\)90066-9](http://dx.doi.org/10.1016/0304-4017(84)90066-9)
- Alvarado-Esquivel C. Toxocariasis in Waste Pickers: A Case Control Seroprevalence Study. *PLoS One* 2013; 8(1): e54897. PMID:23349987 PMCID:PMC3551773. <http://dx.doi.org/10.1371/journal.pone.0054897>
- Anderson BC. Warning about potential for congenital neural larva migrans. *J Am Vet Med Assoc* 1996; 208(2):185. PMID:8567368.
- Avila LFC, Fonseca JSV, Furtado RD, Aguiar PS, Dutra GF, Telmo PL, et al. Registro da transmissão vertical em camundongos BALB/c com toxocarose crônica. *Vittalle* 2009; 21(1): 9-14.
- Beaver PC. The nature of visceral larva migrans. *J Parasitol* 1969; 55(1): 3-12. PMID:5812639. <http://dx.doi.org/10.2307/3277335>
- Bowman DD, Mika-Grieve M, Grieve RB. Circulating excretory-secretory antigen levels and specific antibody responses in mice infected with *Toxocara canis*. *Am J Trop Med Hyg* 1987; 36(1):75-82. PMID:3812886.
- Burke TM, Roberson EL. Prenatal and lactational transmission of *Toxocara canis* and *Ancylostoma caninum*: experimental infection of the bitch before pregnancy. *Int J Parasitol* 1985; 15(1): 71-75. [http://dx.doi.org/10.1016/0020-7519\(85\)90104-3](http://dx.doi.org/10.1016/0020-7519(85)90104-3)
- Camparoto ML, Fulan B, Colli CM, Paludo ML, Falavigna-Guilherme AL, Fernandez MA. Initial stage of development and migratory behavior of *Toxocara canis* larvae in BALB/c mouse experimental model. *Genet Mol Res* 2008; 7(2): 444-450. PMID:18551411. <http://dx.doi.org/10.4238/vol7-2gmr443>

- Chieffi PP, Santos SV, Queiroz ML, Lescano SA. Human toxocariasis: contribution by Brazilian researchers. *Rev Inst Med Trop Sao Paulo* 2009; 51(6): 301-308. PMID:20209265.
- Colli CM, Rubinsky-Elefant G, Paludo ML, Paludo ML, Falavigna DL, Guilherme EV, et al. Serological, clinical and epidemiological evaluation of toxocariasis in urban areas of south Brazil. *Rev Inst Med Trop Sao Paulo* 2010; 52(2): 69-74. PMID:20464126.
- De Savigny DH. *In vitro* maintenance of *Toxocara canis* larvae and a simple method for the production of *Toxocara* ES antigen for use in serodiagnostic tests for visceral larva migrans. *J Parasitol* 1975; 61(4): 781-782. <http://dx.doi.org/10.2307/3279492>
- Dunsmore JD, Thompson RC, Bates IA. The accumulation of *Toxocara canis* larvae in the brains of mice. *Int J Parasitol* 1983; 13(5): 517-521. [http://dx.doi.org/10.1016/S0020-7519\(83\)80017-4](http://dx.doi.org/10.1016/S0020-7519(83)80017-4)
- Dutra GF, Pinto NSF, Avila LFC, Telmo PL, Da Hora VP, Martins LHR, et al. Evaluation of the initial and chronic phases of toxocariasis after consumption of liver treated by freezing or cooling. *Parasitol Res* 2013; 112(6): 2171-2175. PMID:23494157. <http://dx.doi.org/10.1007/s00436-013-3376-5>
- Fan CK, Lin YH, Du WY, Su KE. Infectivity and pathogenicity of 14-month-cultured embryonated eggs of *Toxocara canis* in mice. *Vet Parasitol* 2003; 113(2):145-155. [http://dx.doi.org/10.1016/S0304-4017\(03\)00046-3](http://dx.doi.org/10.1016/S0304-4017(03)00046-3)
- Gallina T, Silva MA, Castro LL, Wendt EW, Villela MM, Berne ME. Presence of eggs of *Toxocara* spp. and hookworms in a student environment in Rio Grande do Sul, Brazil. *Rev Bras Parasitol Vet* 2011; 20(2): 176-177. PMID:21722496. <http://dx.doi.org/10.1590/S1984-29612011000200016>
- Glickman LT, Schantz PM, Cypess RH. Canine and human toxocariasis: review of transmission, pathogenesis, and clinical disease. *J Am Vet Med Assoc* 1979; 175 (12):1265-1269. PMID:528300.
- Hoffmeister B, Glaeser S, Flick H, Pornschlegel S, Suttorp N, Bergmann F. Cerebral toxocariasis after consumption of raw duck liver. *Am J Trop Med Hyg* 2007; 76(3):600-602. PMID:17360892.
- Holland CV, Cox DM. *Toxocara* in the mouse: a model for parasite-altered host behaviour? *J Helminthol* 2001; 75(2): 125-135. PMID:11520435.
- Jin Z, Akao N, Ohta N. Prolactin evokes lactational transmission of larvae in mice infected with *Toxocara canis*. *Parasitol Int* 2008; 57(4): 495-498. PMID:18664391. <http://dx.doi.org/10.1016/j.parint.2008.06.006>
- Kayes SG, Oaks JA. Effect of inoculum size and length of infection on the distribution of *Toxocara canis* larvae in the mouse. *Am J Trop Med Hyg* 1976; 25(4): 573-580. PMID:961975.
- Lee KT, Min HK, Soh CT. Transplacental migration of *Toxocara canis* larvae in experimentally infected mice. *J Parasitol* 1976; 62(3): 460-465. PMID:932920. <http://dx.doi.org/10.2307/3279158>
- Maffrand R, Avila-Vazquez M, Princich D, Alasia P. Congenital ocular toxocariasis in a premature neonate. *Ann Pediat* 2006; 64(6): 595-604.
- Oliveira-Sequeira TC, Amarante AF, Ferrari TB, Nunes LC. Prevalence of intestinal parasites in dogs from Sao Paulo State, Brazil. *Vet Parasitol* 2002; 103(1-2): 19-27. [http://dx.doi.org/10.1016/S0304-4017\(01\)00575-1](http://dx.doi.org/10.1016/S0304-4017(01)00575-1)
- Pawlowski Z. Toxocariasis in humans: clinical expression and treatment dilemma. *J Helminthol* 2001; 75(4): 299-305. PMID:11818044. <http://dx.doi.org/10.1017/S0022149X01000464>
- Reiterová K, Tomasovicová O, Dubinský P. Influence of maternal infection on offspring immune response in murine larval toxocariasis. *Parasit Immunol* 2003; 25(7): 361-368. <http://dx.doi.org/10.1046/j.1365-3024.2003.00642.x>
- Sariego I, Kanobana K, Junco R, Vereecken K, Núñez FA, Polman K, et al. Frequency of antibodies to *Toxocara* in Cuban schoolchildren. *Trop Med Int Health* 2012; 17(6): 711-714. PMID:22943301. <http://dx.doi.org/10.1111/j.1365-3156.2012.02996.x>
- Villela MM, Pepe MS, Ferraz ML, Moraes NCM, Araujo EB, Ruas JL, et al. Contaminação ambiental da orla da Laguna dos Patos (Pelotas, RS, Brasil), por parasitos com potencial zoonótico. *Vitalle* 2009; 21(2): 69-74.
- Yamashita T, Freigang S, Eberle C, Pattison J, Gupta S, Napoli C, et al. Maternal Immunization Programs Postnatal Immune Responses and Reduces Atherosclerosis in Offspring. *Circ Res* 2006; 99(7): E51-64. PMID:16946133. <http://dx.doi.org/10.1161/01.RES.0000244003.08127.cc>