



Revista Brasileira de Parasitologia
Veterinária

ISSN: 0103-846X

zacariascbpv@fcav.unesp.br

Colégio Brasileiro de Parasitologia
Veterinária
Brasil

Figueiredo Guimarães Hoshino, Maria Danielle; Rigôr Neves, Lígia; Tavares-Dias, Marcos
Parasite communities of the predatory fish, *Acestrorhynchus falcatus* and
Acestrorhynchus falcirostris, living in sympatry in Brazilian Amazon
Revista Brasileira de Parasitologia Veterinária, vol. 25, núm. 2, abril-junio, 2016, pp. 207-
216
Colégio Brasileiro de Parasitologia Veterinária
Jaboticabal, Brasil

Available in: <http://www.redalyc.org/articulo.oa?id=397846392011>

- How to cite
- Complete issue
- More information about this article
- Journal's homepage in redalyc.org

redalyc.org

Scientific Information System

Network of Scientific Journals from Latin America, the Caribbean, Spain and Portugal

Non-profit academic project, developed under the open access initiative

Parasite communities of the predatory fish, *Acestrorhynchus falcatus* and *Acestrorhynchus falcistrois*, living in sympatry in Brazilian Amazon

Comunidades de parasitos dos peixes predadores, *Acestrorhynchus falcatus* and *Acestrorhynchus falcistrois*, vivendo em simpatria na Amazônia brasileira

Maria Danielle Figueiredo Guimarães Hoshino¹; Lígia Rigôr Neves²; Marcos Tavares-Dias^{1,2,3*}

¹ Programa de Pós-graduação em Biodiversidade Tropical – PPGBIO, Universidade Federal do Amapá – UNIFAP, Macapá, AP, Brasil

² Programa de Pós-graduação da Rede de Biodiversidade e Biotecnologia da Amazônia Legal – Bionorte, Universidade Federal do Amapá – UNIFAP, Macapá, AP, Brasil

³ Embrapa Amapá, Macapá, AP, Brasil

Received March 10, 2016

Accepted May 3, 2016

Abstract

This study investigated the parasite communities of wild *Acestrorhynchus falcatus* and *Acestrorhynchus falcistrois* populations living in sympatry in Brazilian Amazon. In these two hosts, a total of 12 parasite species e 1-9 parasite species were found per fish, and 10 of these species are metazoans. Eight species of parasites were common to both host species and four of them exhibited differences in abundance and/or prevalence. Parasite communities of the hosts were taxonomically similar (83%) and composed of both ectoparasites and endoparasites, and characterized by high prevalence and high abundance of endoparasites and an aggregated dispersion pattern. For *A. falcistrois*, the dominant parasite was *Ichthyophthirius multifiliis*, and for *A. falcatus*, it was *Piscinoodinium pillulare*. Shannon diversity and Berger-Parker dominance were similar for both hosts, while the parasites species richness and evenness showed differences influenced by the ectoparasites species. These two populations of hosts that inhabited the same geographical area had different sizes, but were exposed to the same infective stages, and acquired qualitatively and quantitatively similar endoparasites community, thus indicating that the amounts and types of prey congeneric that they were eating were similar. Therefore, the overlap in the same occurrence area play an important role in the parasite communities to these phylogenetically related hosts.

Keywords: Aggregation, ectoparasites, freshwater fish, infection, helminths.

Resumo

Este estudo investigou as comunidades parasitárias de populações naturais de *Acestrorhynchus falcatus* e *Acestrorhynchus falcistrois*, peixes piscívoros filogeneticamente relacionados e vivendo em simpatria na Amazônia brasileira. Nessas duas espécies de hospedeiros foram encontradas um total de 12 espécies de parasitos, 1-9 espécies de parasitos por peixe, e desse total de espécies de parasitos, 10 são metazoários. Oito espécies de parasitos foram comuns às duas espécies de hospedeiros e quatro deles apresentaram diferenças na abundância e/ou prevalência. As comunidades parasitárias desses hospedeiros foram taxonomicamente similares (83%) e compostas de ectoparasitos e endoparasitos, mas foram caracterizados por elevada prevalência e elevada abundância de endoparasitos com padrão de dispersão agregado. Para *A. falcistrois*, *Ichthyophthirius multifiliis* foi o parasito dominante, mas para *A. falcatus* foi *Piscinoodinium pillulare*. O índice de diversidade de Shannon e a dominância de Berger-Parker foram similares para ambos hospedeiros, enquanto a riqueza de espécies de parasitos e uniformidade apresentaram diferenças influenciadas pelas espécies de ectoparasitos. Estas duas populações de hospedeiros congêneros que habitavam a mesma área geográfica tinham tamanhos diferentes, mas foram expostos aos mesmos estágios infectantes, adquirindo uma comunidade de endoparasitos qualitativa e quantitativamente similar, indicando assim que as quantidades e tipos de presa que eles estavam ingerindo foram similares. Portanto, a sobreposição em uma mesma área de ocorrência desempenhou um papel importante nas comunidades de parasitos para estes hospedeiros relacionados filogeneticamente.

Palavras-chave: Agregação, ectoparasitos, peixes de água doce, infecção, helmintos.

*Corresponding author: Marcos Tavares-Dias. Embrapa Amapá, Rodovia Juscelino Kubitschek, Km 5, 2600, CEP 68903-419, Macapá, AP, Brasil.
e-mail: marcos.tavares@embrapa.br

Introduction

Acestrorhynchus Eigenmann and Kennedy 1903, is the single genus of family Acestrorhynchidae Eigenmann 1912, which is formed by 15 valid species of Neotropical fish that are endemic to South America (TOLEDO-PIZA, 2007; PRETTI et al., 2009; FROESE & PAULY, 2016). The species of Acestrorhynchidae are of interest because they belong to a monophyletic taxon and presented a high degree of endemism to Amazon, Orinoco, São Francisco, Paraná, Paraguay basins and La Plata drainages (SILVA & GOITEIN, 2009; TOLEDO-PIZA, 2007; PRETTI et al., 2009; FROESE & PAULY, 2016). These species are found in a variety of habitats, but primarily live in lakes in areas near the shores of the Amazon River and Orinoco River systems, and the smallest species are especially found in small creeks (igarapés). Among the Amazonian species are *Acestrorhynchus falcatus* Bloch 1794, and *Acestrorhynchus falcistrois* Cuvier 1819, which are the fish of the present study, which are not listed by the IUCN as endangered species.

Acestrorhynchus falcatus and *A. falcistrois* are benthopelagic, sedentary and diurnal fish. They are piscivorous, but young individuals may also eat shrimps. *Acestrorhynchus falcatus* has a standard length of around 30 cm, while *A. falcistrois* may reach a standard length of around 40 cm. However, in both species, the females start their first sexual maturity when they reach a standard length of 14 cm. They spawn in batches, and reproduction occurs with greater intensity at the end of the dry period (December) and at the beginning of the flood period (May) in the Amazon region (SOARES et al., 2011). These host fish can therefore be infected by various taxa of endoparasites, since their diet includes invertebrate species and also small fish that feed on different invertebrates. The acestrorhynchids are not commercially important as food fish, but might be eventually found in aquarium shops.

Parasitism has been implicated as an important factor in the population dynamics of fish species (POULIN & FITZGERALD, 1987; ALCÂNTARA & TAVARES-DIAS, 2015; PANTOJA et al., 2016). The parasites can be found in host communities and coexists in the same individual of a single host fish species, or in different host fish species (MUÑOZ et al., 2006; TAVARES & LUQUE, 2008; ALARCOS & TIMI, 2012). Thus, explaining the drivers of species diversity is a major matter within ecological and evolutionary research. Host fish species that are phylogenetically close tend to present parasite diversity and species richness of greater similarity than those of unrelated host species (MUÑOZ et al., 2006; ALARCOS & TIMI, 2012; PANTOJA et al., 2016). Congeneric hosts mostly have similar ecology, and thus can overlap in space and time. The biggest problem is to determine where and how this overlapping occurs (TAVARES & LUQUE, 2008; ALARCOS & TIMI, 2012; PANTOJA et al., 2016). For host fish populations living in sympatry the similarity in parasite assemblages has been investigated (MUÑOZ et al., 2006; ALARCOS & TIMI, 2012; PANTOJA et al., 2016). However, this subject has not been addressed in relation to freshwater fish populations in the Amazon region, such as acestrorhynchid species.

In phylogenetically related species, host-parasite coevolution, exposure to parasites and the likelihood that hosts will encounter parasites are factors that influence the vulnerability of fish and the

levels of parasite infection (GUIDELLI et al., 2006; GONZÁLEZ & OLIVA, 2006; PANTOJA et al., 2016). Nevertheless, the main factors that control the structure of endoparasites communities and the levels of parasitism are generally ecological (age, size, habitat, trophic level, vagility, schooling behavior, reproductive season and population size, etc.). Thus, for parasites acquired through the trophic chain (endohelminths), the possibility of host-parasite encounters is the main factor determining the richness and diversity of parasite species (MUÑOZ et al., 2006; ALARCOS & TIMI, 2012; NEVES et al., 2013; PANTOJA et al., 2016). However, ectoparasites communities are affected by the habitat, behavior and density of the host, as well as by environmental characteristics (GONZÁLEZ & OLIVA, 2006).

One way of assessing the similarity of parasite community structures in fish and the ecological factors relating to them is to examine the effects of the ecology and phylogeny on parasite richness and abundance in phylogenetically related hosts that live in the same geographical area (POULIN & FITZGERALD, 1987; GUIDELLI et al., 2006; MUÑOZ et al., 2006; GONZÁLEZ & OLIVA, 2006; ALARCOS & TIMI, 2012; PANTOJA et al., 2016). Thus, to determine whether species of Acestrorhynchidae that live in the same environment and are phylogenetically related have similar parasite communities, we surveyed the ectoparasites and endoparasites of two closely related hosts, *A. falcistrois* and *A. falcatus*, which are congeneric sympatric fish inhabiting the eastern Amazon region (Northern Brazil). Thus, the aim of this study was to compare the parasite communities of *A. falcatus* and *A. falcistrois* population living in sympatry in Brazilian Amazon. These two hosts were captured in the Igarapé Fortaleza basin, an important tributary of the Amazon River system, in eastern Amazon (Brazil), which is highly influenced by the Amazon River system. It consists of linked fluvial physical systems that provide freshwater drainage, and is influenced by the high rainfall of the Amazon region and the daily tides of the Amazon River. The waters that periodically spread out across the floodplain are rich in nutrients, because of the rapid decomposition of grasses, animal remains and the humus layer of the forest. This leads to growth of new vegetation (Poaceae, Cyperaceae, Fabaceae, Onagraceae, Araceae, Asteraceae, Convolvulaceae and Lentibulariaceae) and invertebrates biomass (insects, zooplanktonic crustaceans and mollusks), which are used as food by the fish (GAMA & HALBOTH, 2004; TAKIYAMA et al., 2012).

Materials and methods

Study area and fish collection

Between April and December 2011, 61 specimens of *Acestrorhynchus falcatus* and 33 specimens of *Acestrorhynchus falcistrois* were caught in the basin of Igarapé Fortaleza (Figure 1), in the region of Macapá, state of Amapá (Brazil). The fish were caught in gillnets of different mesh sizes (ICMBio authorization number: 23276-1) and were then stored in insulated boxes containing ice and transported to the Laboratory for Aquatic Organism Health at Embrapa Amapá, for analyses on the parasites. This study was developed in accordance with the principles adopted by the Brazilian College of Animal Experimentation (COBEA).

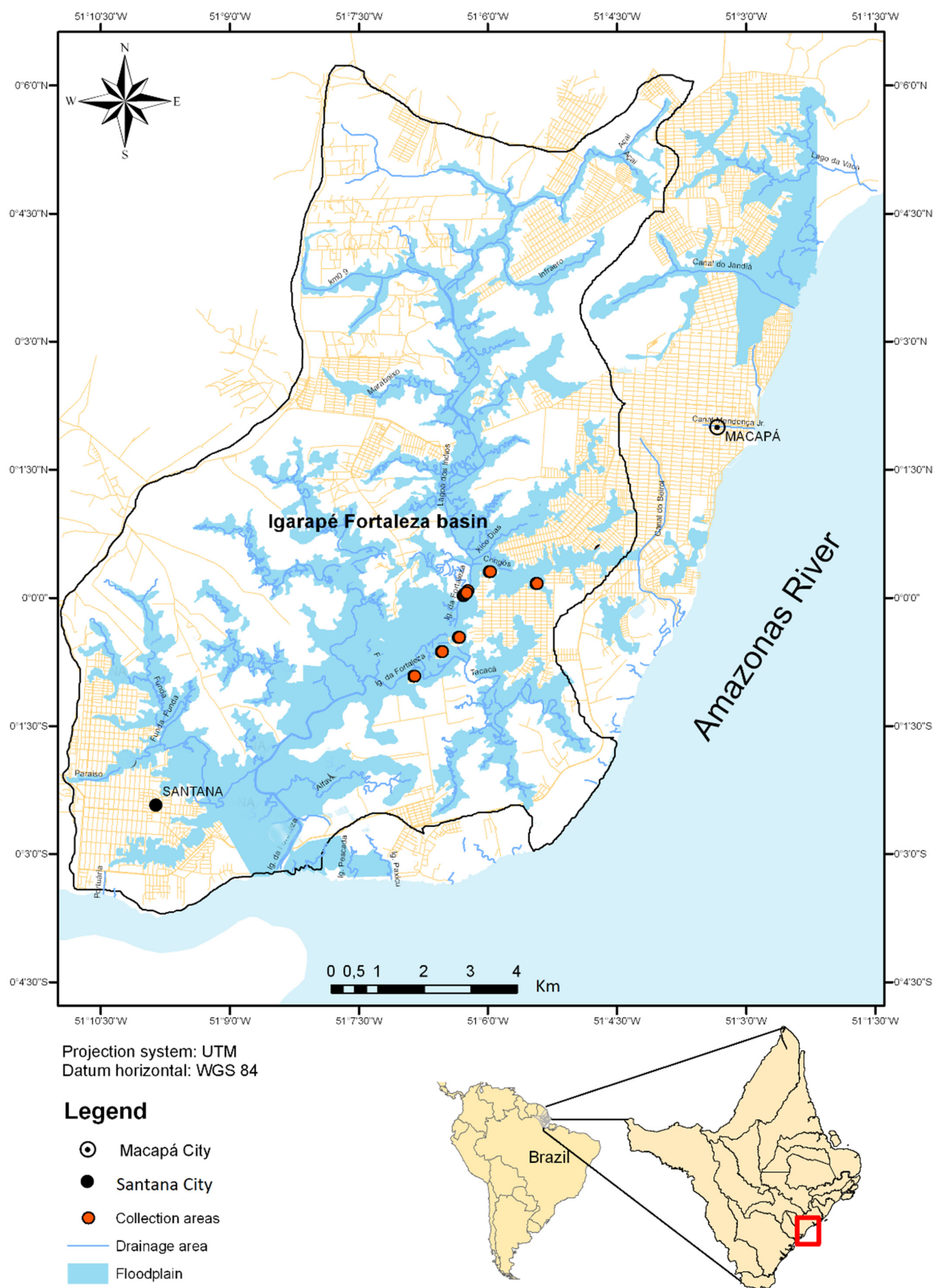


Figure 1. Collection sites of the two *Acestrorhynchidae* species from Brazilian Amazon.

Parasite collection and analysis procedures

The fish caught were weighed (g) and their total length was measured (cm). Each specimen was then evaluated macroscopically, observing the body surface, mouth, eyes, opercula and gills. The gills were removed to collect ectoparasites. The gastrointestinal tract and viscera was removed and examined in order to collect endoparasites. The parasite specimens were collected, fixed, quantified and stained for identification (EIRAS et al., 2006).

The ecological terms used followed previous recommendations (ROHDE et al., 1995; BUSH et al., 1997). The Shannon diversity index (H), evenness (E), Berger-Parker dominance index (d) and species richness (MAGURRAN, 2004) were calculated for each component parasite community, using the Diversity software (Pisces Conservation Ltd, UK). The dispersion index (DI) and discrepancy index (D) were calculated using the Quantitative Parasitology 3.0 software, in order to detect the distribution pattern of the parasite infracommunities (RÓZSA, et al., 2000), for species with prevalence >10%. The significance of DI, for each infracommunity, was tested using the d statistic (LUDWIG & REYNOLDS, 1988). For similarity measurements on parasites abundance between *A. falcirostris* and *A. falcatus*, the Jaccard index (qualitative index) and Bray-Curtis index (quantitative index) were used. These parameters take into account differences in abundance among the shared parasite species (LUDWIG & REYNOLDS, 1988; MAGURRAN, 2004). Both similarity indices were calculated using the Past software (HAMMER et al., 2001).

The body weight (Wt; g) and total length (L; cm) data were used to calculate the relative condition factor (Kn) of the host fish, as well as for determining the length-weight relationship ($W = aL^b$) after logarithmic transformation of length and weight and subsequent adjustment of two straight lines, thereby obtaining $\ln y = \ln A + B \ln x$ (LE CREN, 1951).

Data analysis

The Kolmogorov-Smirnov test was used to determine whether the parasite abundance data followed a normal distribution pattern. The differences in prevalence for each parasite infracommunity between *A. falcatus* and *A. falcirostris* were evaluated using the chi-square test (χ^2) with Yates correction; and the differences in parasite abundance using the Mann-Whitney test (U). The comparisons of parasite prevalence and abundance were only made for parasites with prevalence >10%. For each host species, the differences in species richness, Shannon index (H), evenness (E) and Berger-Parker dominance index (d) were determined using the Mann-Whitney test (U). Spearman's coefficient (r_s) was used to investigate the correlation between length and the species richness, H , E , d and abundance of the parasites, and the correlation between Kn and the abundance of the parasites (ZAR, 2010).

Principal component analysis (PCA) was carried out to compare the influence of body and diversity indices on the parasite communities of *A. falcirostris* and *A. falcatus*. This was done using the Past-Paleontological Statistics software, version 3.0 (HAMMER et al., 2001).

Results

All the specimens of *A. falcirostris* and *A. falcatus* were parasitized with one or more species and a total of 10 metazoan species occurred (Table 1). In *A. falcatus*, the dominant species was *Piscinoodinium pillulare*, while in *A. falcirostris*, it was *Ichthyophthirius multifiliis* (Table 2). The abundance of *I. multifiliis*, *P. pillulare* and *Clinostomum marginatum* (gills) had differences between *A. falcirostris* and *A. falcatus*. Prevalence of *P. pillulare* and *Diaphorocleidus* sp. was highest in *A. falcatus*. *Braga patagonica* was

Table 1. Parasites infracommunities for two species of Acestrorhynchidae from Brazilian Amazon.

Host species		<i>Acestrorhynchus falcatus</i> (n=61)			<i>Acestrorhynchus falcirostris</i> (n = 33)		
Parasite species	SI	P (%)	MA ± SD	P (%)	MA ± SD	χ^2	U
<i>Ichthyophthirius multifiliis</i> Fouquet, 1876	Gills	82.0	479.3 ± 520.3	78.8	6,125.7 ± 10,162.4	0.14	650.0*
<i>Piscinoodinium pillulare</i> Lom, 1981	Gills	78.7	3,205.5 ± 4951.9	3.0	163.7 ± 883.4	49.12**	258.0**
<i>Diaphorocleidus</i> sp.	Gills	95.1	20.8 ± 17.8	72.7	30.6 ± 33.4	9.63**	990.0
Dactylogyridae gen. sp.	Gills	59.0	2.9 ± 5.2	36.4	2.3 ± 4.7	4.40	793.0
<i>Clinostomum marginatum</i> Rudolphi, 1819 (metacercariae)	Gills	83.6	5.8 ± 5.3	84.8	57.1 ± 192.7	0.02	761.5*
<i>Clinostomum marginatum</i> (metacercariae)	Intestine	13.1	0.9 ± 4.3	-	-	-	-
Digenea gen. sp. (metacercariae)	Intestine	-	-	3.0	0.03 ± 0.17	-	-
<i>Contracaecum</i> sp. (larvae)	Intestine and liver	93.4	9.4 ± 29.3	81.8	9.1 ± 14.6	3.25	924.5
<i>Philometra</i> sp. (larvae)	Intestine	4.9	0.1 ± 0.5	3.0	0.03 ± 0.17	-	-
<i>Procamallanus</i> (<i>Spirocamallanus</i>) <i>inopinatus</i> Travassos, Artigas and Pereira, 1928 (larvae and adults)	Intestine	18.0	0.5 ± 1.8	39.4	1.2 ± 2.5	17.6**	770.5
<i>Neochinorhynchus pterodoridis</i> Thatcher, 1981 (adults)	Intestine	6.6	0.4 ± 2.8	6.1	0.9 ± 3.7	-	-
<i>Braga patagonica</i> Schödtte and Meinert, 1884	Gills	1.6	0.03 ± 0.26	-	-	-	-
<i>Ergasilus turucuyus</i> Malta and Varela, 1996	Gills	-	-	3.0	0.03 ± 0.17	-	-

The prevalence between hosts was compared using Chi-square test, and the abundance using Mann-Whitney test. P: Prevalence; MA: Mean abundance; SI: Site of infection; χ^2 : Chi-square test; U: Mann-Whitney test; *p < 0.05 and **p < 0.001.

only observed in *A. falcatus*, while *Ergasilus turucuyus* occurred only in *A. falcistrostris* (Table 1).

The parasite species of *A. falcatus* and *A. falcistrostris* presented an aggregated dispersion pattern, which is common among parasites of freshwater fish. Some species had higher values for the discrepancy index (Table 2), thus indicating greater parasite aggregation. The Shannon diversity index and the dominance index were similar for *A. falcistrostris* and *A. falcatus*, but the mean species richness of parasites and the evenness were greater for *A. falcatus* (Table 3).

In both hosts, individuals parasitized by 4-7 species predominated (Figure 2). *Acestrorhynchus falcatus* presented greater species richness per host and greater evenness, and the two hosts had similar structures for their parasite communities (Table 3).

In *A. falcatus*, length did not show any correlation with Shannon diversity index ($r_s = 0.13$, $p = 0.333$), species richness of parasites ($r_s = 0.08$, $p = 0.54$), evenness index ($r_s = 0.11$, $p = 0.39$) or Berger-Parker dominance ($r_s = -0.12$, $p = 0.35$). However, in *A. falcistrostris*, length was positively correlated with Shannon index ($r_s = 0.35$, $p = 0.04$), species richness ($r_s = 0.68$, $p = 0.0001$) and evenness index ($r_s = 0.36$, $p = 0.04$), but not with the Berger-Parker dominance index ($r_s = -0.33$, $p = 0.05$).

In *A. falcistrostris*, Shannon index ($r_s = -0.53$, $p = 0.001$) presented a significant negative correlation with Kn, as did the richness of

parasite species ($r_s = -0.48$, $p = 0.004$) with the Kn of the hosts. In *A. falcatus*, there was no significant correlation between Shannon index ($r_s = -0.04$, $p = 0.76$) and Kn, or between the richness of parasite species ($r_s = -0.18$, $p = 0.16$) and the Kn of the hosts.

In *A. falcatus*, length did not show any correlation with the abundance of *I. multifiliis* ($r_s = 0.22$, $p = 0.08$), *P. pillulare* ($r_s = 0.01$, $p = 0.91$), *Diaphorocleidus* sp. ($r_s = 0.15$, $p = 0.25$), undetermined Dactylogyridae ($r_s = 0.07$, $p = 0.61$), *C. marginatum* ($r_s = 0.08$, $p = 0.52$), *Procamallanus* (*Spirocamallanus*) *inopinatus* ($r_s = -0.05$, $p = 0.69$) or larvae of *Contracaecum* sp. ($r_s = -0.02$, $p = 0.85$). The Kn also did not show any correlation with the abundance of *I. multifiliis* ($r_s = 0.14$, $p = 0.27$), *P. pillulare* ($r_s = 0.03$, $p = 0.76$), *Diaphorocleidus* sp. ($r_s = 0.09$, $p = 0.48$), undetermined Dactylogyridae ($r_s = -0.16$, $p = 0.21$), *C. marginatum* ($r_s = -0.03$, $p = 0.81$), *P. (S.) inopinatus* ($r_s = -0.20$, $p = 0.12$) or *Contracaecum* ($r_s = 0.007$, $p = 0.95$).

In *A. falcistrostris*, length presented a positive correlation with the abundance of *Diaphorocleidus* sp. ($r_s = 0.50$, $p = 0.003$), undetermined Dactylogyridae ($r_s = 0.47$, $p = 0.005$), *C. marginatum* ($r_s = 0.49$, $p = 0.004$), *Contracaecum* sp. ($r_s = 0.40$, $p = 0.02$) and *P. (S.) inopinatus* ($r_s = 0.58$, $p = 0.0004$). Length did not show any correlation with the abundance of *I. multifiliis* ($r_s = 0.14$, $p = 0.44$). The Kn presented a positive correlation with the abundance of *I. multifiliis* ($r_s = 0.49$, $p = 0.003$). The Kn showed

Table 2. Dispersion index (DI), d -statistic, discrepancy index (D) and frequency of dominance (FD) for the parasite infracommunities of two species of Acestrorhynchidae from Brazilian Amazon.

Parasites	<i>Acestrorhynchus falcatus</i> (n = 61)				<i>Acestrorhynchus falcistrostris</i> (n = 33)			
	DI	d	D	FD (%)	DI	d	D	FD (%)
<i>Ichthyophthirius multifiliis</i>	6.766	17.59	0.439	0.1286	5.498	10.83	0.465	0.9584
<i>Piscinoodinium pillulare</i>	9.041	22.03	0.478	0.8603	-	-	-	-
<i>Diaphorocleidus</i> sp.	5.168	14.00	0.389	0.0056	2.865	5.61	0.441	0.0048
Dactylogyridae gen. sp.	3.040	8.19	0.596	0.0008	3.221	6.43	0.723	0.0004
<i>Clinostomum marginatum</i> (gills)	2.959	7.94	0.424	0.0016	2.877	5.64	0.386	0.0089
<i>Clinostomum complanatum</i> (intestine)	2.603	6.77	0.877	0.0003	-	-	-	-
<i>Contracaecum</i> sp. (intestine)	3.497	9.58	0.598	0.0012	3.252	6.49	0.549	0.0008
<i>Contracaecum</i> sp. (liver)	3.368	9.20	0.846	0.0004	2.509	4.74	0.840	0.0004
<i>Contracaecum</i> sp. (mesentery)	4.224	11.61	0.723	0.0010	3.014	5.95	0.780	0.0004
<i>Procamallanus</i> (<i>Spirocamallanus</i>) <i>inopinatus</i>	1.929	4.31	0.856	0.0001	1.554	2.04	0.706	0.0001

Table 3. Characteristics of the component communities of parasites and body parameters for two species of Acestrorhynchidae from Brazilian Amazon.

Mean indices	<i>Acestrorhynchus falcatus</i>	<i>Acestrorhynchus falcistrostris</i>	U	p
Body length (cm)	19.5 ± 1.5 (16.0-23.0)	22.1 ± 4.8 (12.5-28.5)	561.5	0.0004
Body weight (g)	76.5 ± 16.1 (26.0-115.0)	87.8 ± 44.4 (15.0-156.0)	719.0	0.023
Relative condition factor (Kn)	1.00 ± 0.04 (0.76-1.07)	1.00 ± 0.06 (0.80-1.15)	990.0	0.896
Species richness	5.7 ± 1.5 (3-9)	4.5 ± 2.2 (1-8)	6.75	0.0086
Shannon index (H')	1.55 ± 1.33 (1.01-1.625)	1.35 ± 1.40 (1.00-1.342)	977.0	0.8152
Evenness (E')	0.22 ± 0.14 (0.004-0.648)	0.14 ± 0.16 (0.000-0.550)	1397.0	0.0002
Berger-Parker (d')	0.77 ± 0.18 (0.303-0.999)	0.88 ± 0.16 (0.452-1.000)	884.5	0.3338
Total number of parasites	227,278	198,137	-	-
Number of ectoparasites	6	6	-	-
Number of endoparasites	5	5	-	-
Endoparasites (adult)	2	2	-	-
Endoparasites (larval)	5	5	-	-

U = Mann-Whitney test, p = Probability.

a negative correlation with the abundance of *Diaphorocleidus* sp. ($r_s = -0.46$, $p = 0.007$), Dactylogyridae gen. sp. ($r_s = -0.38$, $p = 0.02$), *Contracaecum* sp. ($r_s = -0.42$, $p = 0.01$) and *C. marginatum* ($r_s = -0.47$, $p = 0.006$). The Kn also did not show any correlation with the abundance of *P. (S.) inopinatus* ($r_s = -0.21$, $p = 0.25$).

The multivariate analysis of PCA based on the body and diversity parameters of *A. falcirostris* and *A. falcatus* showed that together the two main factors accounted for approximately 66% of the variance. The results showed that species richness of parasites, Shannon index and evenness were the main factors relating to *A. falcatus*, while the Kn and Berger-Parker dominance index were the factors most strongly relating to *A. falcirostris* (Figure 3).

The Jaccard index ($S_j = 0.75$) and Bray-Curtis dissimilarity ($BC = 0.17$) demonstrated a high similarity between the parasite communities of *A. falcirostris* and *A. falcatus*. The multivariate analysis based on component communities of parasites for *A. falcirostris* and *A. falcatus* led to the observation that there was only a little separation between these species of hosts, caused by *P. pillulare* and *I. multifiliis* (Figure 4).

For *A. falcatus*, the equation for the weight/length relationship was $Wt = 0.207L^{1.9853}$ ($r^2 = 0.438$), while for *A. falcirostris*, it was $Wt = 0.0151L^{2.7561}$ ($r^2 = 0.892$); both of these were allometrically negative, thus indicating that there was a greater increase in body weight than in size. The relative condition factor (Kn) of two hosts

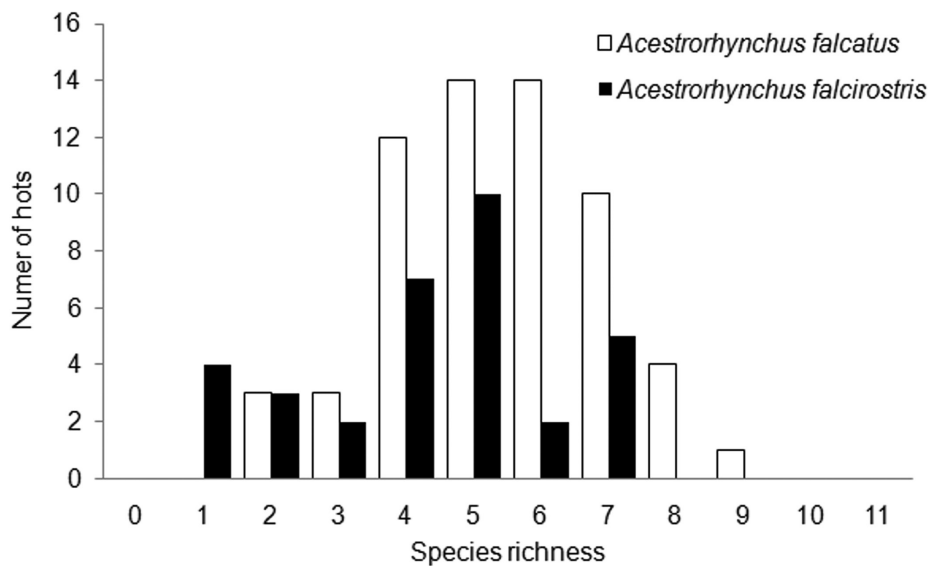


Figure 2. Species richness of parasites in two Acestrorhynchidae species from Brazilian Amazon.

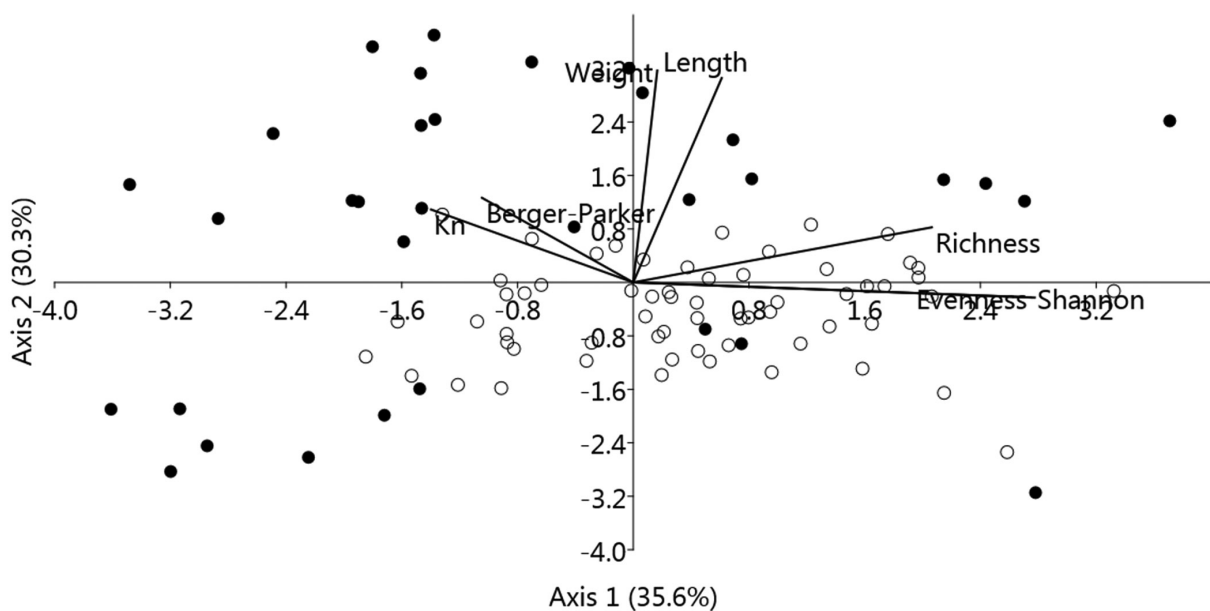


Figure 3. Scatterplot scores of the principal component analysis (PCA) on factors influencing the parasite communities of *Acestrorhynchus falcatus* (○) and *Acestrorhynchus falcirostris* (●) from Brazilian Amazon. Kn: Relative condition factor, Shannon: Shannon diversity, Richness: species richness of parasites.

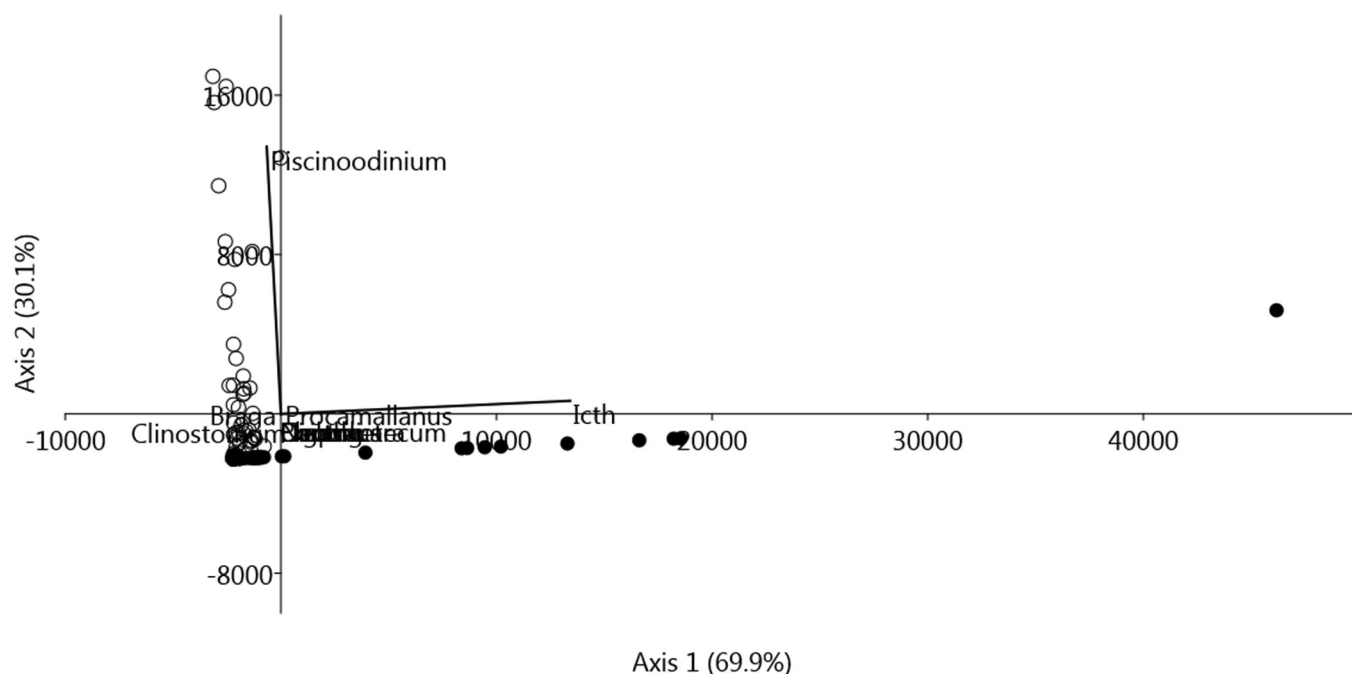


Figure 4. Scatterplot scores of the principal component analysis (PCA) on parasite communities of *Acestrorhynchus falcatus* (○) and *Acestrorhynchus falcistrostris* (●) from Brazilian Amazon. Ichth: *Ichthyophthirius multifiliis*, Piscinoodinium: *Piscinoodinium pillulare*, Diaph: *Diaphorocleidus* sp., Dacty: Dactylogyridae gen. sp., Clinostomum: *Clinostomum marginatum*, Contraecum: *Contraecum* sp.; Procamallanus: *Procamallanus* (*Spirocamallanus*) *inopinatus*, Braga: *Braga patagonica*, Ergasilus: *Ergasilus turucuyus*, Neoch: *Neochinorhynchus pterodoriidis*.

was not different ($H = 1.126$, $p = 0.599$) of the standard value ($K_n = 1.00$), indicating therefore good body conditions despite the parasitism levels found.

Discussion

The parasite community structure of fish is mostly influenced by host physiology, ecology and phylogeny. Thus, hosts that live in the same environment and are phylogenetically related, with similar ecology, can be expected to have a certain degree of homogeneity in their parasite assemblages (MUÑOZ et al., 2006; ALARCOS & TIMI, 2012; PANTOJA et al., 2016). This is particularly so for endohelminths. However, ectoparasites and endoparasites species tend to respond differently both to environmental and to host-related variability (GONZÁLEZ & OLIVA, 2006; ALARCOS & TIMI, 2012; PANTOJA et al., 2016), given that transmission of ectoparasites is unaffected by feeding habits and ontogenetic changes in the hosts, or the availability of different prey species in the environment (GONZÁLEZ & OLIVA, 2006; PANTOJA et al., 2016). This first investigation on the parasite communities in *A. falcatus* and *A. falcistrostris*, which are species that live in the same environment and are phylogenetically related, showed that the parasite community was characterized by the presence of species with high prevalence, low abundance and predominance of ectoparasites (protozoans, monogeneans and metacercariae of digeneans), but with similar endoparasites communities at the larval stage. The presence of adult endoparasites may be mostly related to the hosts' diet,

while the presence of larval endoparasites may be due both to the hosts' diet and to predator-prey interactions, since this is the path through which many endoparasites species complete their heteroxenic life cycle (MUÑOZ et al., 2006; GUIDELLI et al., 2006; BITTENCOURT et al., 2014; PANTOJA et al., 2016). Therefore, these results indicate that for ectoparasites diversity, taxonomic proximity and phylogenetic proximity of the hosts are factors that are more important than host species-wide characteristics, and suggest that there is a similar relationship for endoparasites diversity.

Even if fish species that are phylogenetically related and live in the same environment have similar diversity of endoparasites, they may show differentiated levels of infection (MUÑOZ et al., 2006; GUIDELLI et al., 2006; TAVARES & LUQUE, 2008) in relation to ectoparasites species (PANTOJA et al., 2016). In *A. falcatus*, the dominant parasite species was *P. pillulare*, followed by *I. multifiliis*, but in *A. falcistrostris*, the dominant species was *I. multifiliis*. Thus, both of the host species presented high prevalence and abundance of these protozoan ectoparasites. The size of the parasites is information of relevance for understanding how much parasite biomass can be sustained by hosts, and how this can determine the parasite community structure. Thus, the protozoa *I. multifiliis* and *P. pillulare* exhibited much higher parasite burden than did the other parasite species, probably due to the small size, low specificity and high reproduction and infection rates of these ectoparasites (NEVES et al., 2013).

In the gills of *A. falcistrostris*, the abundance of *I. multifiliis* and metacercariae of *C. marginatum* was higher than in *A. falcatus*. The latter host showed higher prevalence and abundance of *P. pillulare* and

higher prevalence of *Diaphorocleidus* sp. These differences between the hosts may have been caused by heterogeneity of the host-parasite relationship. This would have been influenced by biological factors, especially those relating to the reproduction period, since fish suffer higher stress levels during this period (SCHRECK, 2010; ROHLENOVÁ & SIMKOVÁ, 2010) and are more susceptible to parasitic infections, especially due to opportunistic parasites like the protozoa found here. For *A. falcistrostris*, the abundance of monogenoideans, *Contracaecum* sp., *P. (S.) inopinatus* and *C. marginatum* increased with the length of the hosts, indicating therefore that larger individuals accumulated greater quantities of these parasites. Moreover, hosts with lower body conditions showed higher abundance of monogenoideans, *Contracaecum* sp. and *C. marginatum*. In contrast, the length of the population of *A. falcatus* did not show any correlation of abundance with any of the parasites, or between the condition factor and the diversity and richness of parasite species, because of the homogeneity of size of specimens of this host that were examined. Nonetheless, the presence of larger parasite infracommunities in larger fish has been a common characteristic in host-parasite systems (POULIN & FITZGERALD, 1987; CARVALHO et al., 2003; MUNÓZ et al., 2006; GONZÁLEZ & OLIVA, 2006; GUIDELLI et al., 2006; PANTOJA et al., 2016).

Diversity values depend on the balance within species richness between dominant species (ALARCOS & TIMI, 2012). Larvae of *Contracaecum* sp. were the endoparasites with most importance as contributors towards abundance and species richness in *A. falcatus* and *A. falcistrostris*, while *C. marginatum*, *P. (S.) inopinatus*, *Philometra* sp. and *Neoechinorhynchus pterodoriidis* had little importance. *Acestrorhynchus falcistrostris* showed lower parasite richness per host and a lower evenness, besides the Shannon diversity index and evenness showed a weak correlation with the size of the hosts. Principal component analysis proved that the Kn and Berger-Parker dominance index were the main factors relating to *A. falcistrostris*. In contrast, there was no correlation between the size of *A. falcatus* and the diversity indexes, since the population sampled comprised a larger number of individuals and the body size was relatively homogenous and bigger than in the population of *A. falcistrostris*, although *A. falcatus* is a fish of smaller body size (SOARES et al., 2011). Moreover, principal component analysis showed that in *A. falcatus*, Shannon diversity, species richness of and evenness were the main factors relating to this host. Despite these differences in body size between *A. falcistrostris* and *A. falcatus*, there were no differences in body condition, prevalence, abundance, richness and diversity of endoparasites in these two populations if piscivorous hosts of similar age. Moreover, the Kn indicated good body condition and general health for *A. falcistrostris* and *A. falcatus*. In host-parasite networks, the host body provides a microhabitat for ectoparasites and endoparasites species (GUIDELLI et al., 2006; BELLAY et al., 2013). It has been suggested that high parasite species diversity may be an indicator of individual fish growth and body condition. In addition, correlations between the richness and abundance of parasites and host body size have commonly been reported in studies on component communities (POULIN & FITZGERALD, 1987; CARVALHO et al., 2003; GUIDELLI et al., 2006; GONZÁLEZ & OLIVA, 2006; ALARCOS & TIMI, 2012; NEVES et al., 2013; PANTOJA et al., 2016). Nevertheless, it has

been argued that wild fish populations with higher density may provide more trophic host-parasite interactions, besides sustaining a larger population of endoparasites (TAKEMOTO et al., 2005; GUIDELLI et al., 2006).

Parasites are typically found in communities in which different species coexist in fish of the same host species or in different host species (MUÑOZ et al., 2006; TAVARES & LUQUE, 2008; BELLAY et al., 2013; ALCÂNTARA & TAVARES-DIAS, 2015). Protozoan and monogenoidean ectoparasites are parasites typical of lentic environments, but their reproduction is strongly favored by eutrophized environments like the hydrographic basin of this study (TAKIYAMA et al., 2012). Some crustaceans are host and site-specific, especially in relation to fish with particular habitats and lifestyles, while other parasites frequently have no preference (TAVARES-DIAS et al., 2015). However, isopods of *B. patagonica* were found only on the gills of *A. falcatus*, and those of *E. turucuyus* only in *A. falcistrostris*. Consequently, these isopod species were the parasites of lowest prevalence and abundance in these hosts, because they found their hosts using their ability to swim or their close relationship with them (TAVARES-DIAS et al., 2015). In *A. falcatus*, the dominant species was *P. pillulare*, while in *A. falcistrostris*, it was *I. multifiliis*. The ectoparasites and endoparasites found in these two hosts presented an aggregated dispersion pattern, but *I. multifiliis*, *P. pillulare*, *Contracaecum*, *C. marginatum* and *P. (S.) inopinatus* presented the greatest aggregation (> discrepancy), in both hosts. This overdispersion is a common pattern of ectoparasites and endoparasites infracommunities in different fish populations (POULIN & FITZGERALD, 1987; NEVES et al., 2013; TAVARES-DIAS et al., 2015; PANTOJA et al., 2016), and is caused by a diversity of factors (see ROHDE et al., 1995; POULIN, 2013; TAVARES-DIAS et al., 2015).

The parasite communities in *A. falcistrostris* and *A. falcatus* were formed by parasite species common to other species of freshwater fish (CARVALHO et al., 2003; GUIDELLI et al., 2006; NEVES et al., 2013; BITTENCOURT et al., 2014; TAVARES-DIAS et al., 2015; PANTOJA et al., 2016). *Neoechinorhynchus pterodoriidis* collected from *A. falcatus* and *A. falcistrostris* is an acanthocephalan species that was described originally from *Pterodoras granulosus* Humboldt and Valenciennes 1821, a siluriform Doradidae (THATCHER, 1981). For the two host populations studied here, the structure of the component community was constituted especially by species of generalist parasites, with a direct or indirect life cycle. Therefore, our findings suggest that the interaction pattern of *A. falcatus* and *A. falcistrostris* with the endoparasites found (acanthocephalans, digeneans and nematodes) is closely associated with their diet. In addition, they suggest that the observed parasites diversity is unrelated to host-parasite coevolution, therefore indicating that there is no need to focus future research towards the historical processes shaping these relationships. This is the first record of *I. multifiliis*, *P. pillulare*, *C. marginatum*, *Diaphorocleidus* sp., *Contracaecum*, *Philometra* sp., *P. (S.) inopinatus* and *N. pterodoriidis* in *A. falcistrostris* and *A. falcatus*.

In conclusion, the present study demonstrated that there was low diversity of parasites and similar parasites communities in *A. falcatus* and *A. falcistrostris*. The parasites infracommunities of both were rich in species, with presence of protozoans, crustaceans, monogenoideans, digeneans, nematodes and acanthocephalans.

In general, larval helminths that are transmitted trophically, with low or nonexistent parasitic specificity among fish species, they infected these fish as paratenic hosts or as intermediate hosts. They may live in these hosts for long periods, thus persisting in the food chain. Thereby, these characteristics make them potentially available for any fish host, independent of its trophic level. It is clear that parasite species with the highest abundance are not the largest species, and that few species of helminths are numerically important. This can be connected to resources and the niche needed for each species. Endoparasites abundance and diversity were mostly influenced by the similar amounts and types of prey that they were eating (crustaceans and mollusks). Finally, the same geographic occurrence area (sympatry) of these two related hosts also played an important role in the parasite communities found. Therefore, these two host fish competes with each other, because they do not exhibit differences in feeding.

Acknowledgements

Dr. Marcos Tavares-Dias was supported by research fellowships from the Conselho Nacional de Pesquisa e Desenvolvimento Tecnológico (CNPq, Brazil).

References

- Alarcos AJ, Timi JT. Parasite communities in three sympatric flounder species (Pleuronectiformes: Paralichthyidae): similar ecological filters driving toward repeatable assemblages. *Parasitol Res* 2012; 110(6): 2155-2166. <http://dx.doi.org/10.1007/s00436-011-2741-5>. PMID:22167375.
- Alcântara NM, Tavares-Dias M. Structure of the parasites communities in two Erythrinidae fish from Amazon River system (Brazil). *Rev Bras Parasitol Vet* 2015; 24(2): 183-190. <http://dx.doi.org/10.1590/S1984-29612015039>. PMID:26083690.
- Bellay S, Oliveira EF, Almeida-Neto M, Lima DP Jr, Takemoto RM, Luque JL. Developmental stage of parasites influences the structure of fish-parasite networks. *PLoS One* 2013; 8(10): e75710. <http://dx.doi.org/10.1371/journal.pone.0075710>. PMID:24124506.
- Bittencourt LS, Pinheiro DA, Cárdenas MQ, Fernandes BMM, Tavares-Dias M. Parasites of native Cichlidae populations and invasive *Oreochromis niloticus* (Linnaeus, 1758) in tributary of Amazonas River (Brazil). *Rev Bras Parasitol Vet* 2014; 23(1): 44-54. <http://dx.doi.org/10.1590/S1984-29612014006>. PMID:24728360.
- Bush AO, Lafferty KD, Lotz JM, Shostak AW. Parasitology meets ecology on its own terms: Margolis et al. revisited. *J Parasitol* 1997; 83(4): 575-583. <http://dx.doi.org/10.2307/3284227>. PMID:9267395.
- Carvalho S, Guidelli GM, Takemoto RM, Pavanelli GC. Ecological aspects of endoparasite fauna of *Acestrorhynchus lacustris* (Lütken, 1875) (Characiformes, Acestrorhynchidae) on the upper Paraná River floodplain, Brazil. *Acta Sci Biol Sci* 2003; 25(2): 479-483. <http://dx.doi.org/10.4025/actascibiols.v25i2.2043>.
- Eiras JC, Takemoto RM, Pavanelli GC. *Métodos de estudo e técnicas laboratoriais em parasitologia de peixes*. Maringá: Eduem; 2006.
- Froese R, Pauly D, editors. *FishBase version 1/2016* [online]. FishBase; 2016 [cited 2015 Oct 10]. Available from: www.fishbase.org
- Gama CS, Halboth DA. Ictiofauna das ressacas das bacias do Igarapé da Fortaleza e do Rio Curiaú, In: Takiyama LR, Silva AQ. *Diagnóstico das ressacas do estado do Amapá: bacias do Igarapé da Fortaleza e Rio Curiaú*. Macapá: CPAQ/IEPA/DGEO/SEMA; 2004. p. 23-52.
- González MT, Oliva ME. Similarity and structure of the ectoparasite communities of rockfish species from the southern Chilean coast in a temporal scale. *Parasitology* 2006; 133(3): 335-343. <http://dx.doi.org/10.1017/S0031182006000370>. PMID:16753080.
- Guidelli G, Tavechio WLG, Takemoto RM, Pavanelli GC. Fauna parasitária de *Leporinus lacustris* e *Leporinus friderici* (Characiformes, Anostomidae) da planície de inundação do alto Rio Paraná, Brasil. *Acta Sci Biol Sci* 2006; 28(3): 281-290.
- Hammer O, Harper DAT, Ryan PD. PAST: paleontological statistics software package for education and data analysis. *Palaeontol Electronica* 2001; 4(1): 1-9.
- Le Cren ED. The length-weight relationship and seasonal cycle in gonad weight and condition in the perch (*Perca fluviatilis*). *J Anim Ecol* 1951; 20(2): 201-219. <http://dx.doi.org/10.2307/1540>.
- Ludwig JA, Reynolds JF. *Statistical ecology: a primer on methods and computing*. New York: Wiley-Interscience; 1988.
- Magurran AE. *Measuring biological diversity*. Oxford: Blackwell Science; 2004.
- Muñoz G, Grutter AS, Cribb TH. Endoparasite communities of five fish species (Labridae: Cheiliniinae) from Lizard Island: how important is the ecology and phylogeny of the hosts? *Parasitology* 2006; 132(3): 363-374. <http://dx.doi.org/10.1017/S0031182005009133>. PMID:16332286.
- Neves LR, Pereira FB, Tavares-Dias M, Luque JL. Seasonal influence on the parasite fauna of a wild population of *Astronotus ocellatus* (Perciformes: Cichlidae) from the Brazilian Amazon. *J Parasitol* 2013; 99(4): 718-721. <http://dx.doi.org/10.1645/12-84.1>. PMID:23421456.
- Pantoja WME, Silva LVF, Tavares-Dias M. Are similar the parasite communities structure of *Trachelyopterus coriaceus* and *Trachelyopterus galeatus* (Siluriformes: Auchenipteridae) in the Amazon basin? *Rev Bras Parasitol Vet* 2016; 25(1): 46-53. <http://dx.doi.org/10.1590/S1984-29612016016>. PMID:27007248.
- Poulin R, Fitzgerald GJ. The potential of parasitism in the structuring of a salt marsh stickleback community. *Can J Zool* 1987; 65(11): 2793-2798. <http://dx.doi.org/10.1139/z87-421>.
- Poulin R. Explaining variability in parasite aggregation levels among host samples. *Parasitology* 2013; 140(4): 541-546. <http://dx.doi.org/10.1017/S0031182012002053>. PMID:23343821.
- Pretti VQ, Calcagnotto D, Toledo-Piza M, Almeida-Toledo LF. Phylogeny of the Neotropical genus *Acestrorhynchus* (Ostariophysi: Characiformes) based on nuclear and mitochondrial gene sequences and morphology: A total evidence approach. *Mol Phylogenet Evol* 2009; 52(2): 312-320. <http://dx.doi.org/10.1016/j.jmpev.2008.12.025>. PMID:19168140.
- Rohde K, Hayward C, Heap M. Aspects of the ecology of metazoan ectoparasites of marine fishes. *Int J Parasitol* 1995; 25(8): 945-970. [http://dx.doi.org/10.1016/0020-7519\(95\)00015-T](http://dx.doi.org/10.1016/0020-7519(95)00015-T). PMID:8550295.
- Rohlenová K, Simková A. Are the immunocompetence and the presence of metazoan parasites in cyprinid fish affected by reproductive efforts of Cyprinid fish? *J Biomed Biotechnol* 2010; 2010: 418382. <http://dx.doi.org/10.1155/2010/418382>. PMID:20145709.

- Rózsa L, Reiczigel J, Majoros G. Quantifying parasites in samples of hosts. *J Parasitol* 2000; 86(2): 228-232. [http://dx.doi.org/10.1645/0022-3395\(2000\)086\[0228:QPISOH\]2.0.CO;2](http://dx.doi.org/10.1645/0022-3395(2000)086[0228:QPISOH]2.0.CO;2). PMID:10780537.
- Schreck CB. Stress and fish reproduction: the roles of allostasis and hormesis. *Gen Comp Endocrinol* 2010; 165(3): 549-556. <http://dx.doi.org/10.1016/j.ygcen.2009.07.004>. PMID:19596332.
- Silva AT, Goitein R. Diet and feeding activity of *Acestrorhynchus lacustris* (Lütken, 1875) (Characiformes, Acestrorhynchidae) in the water reservoir at Ribeirão Claro, SP. *Braz J Biol* 2009; 69(3): 757-762. <http://dx.doi.org/10.1590/S1519-69842009000400002>. PMID:19802434.
- Soares MGM, Costa EL, Siqueira-Souza FK, Anjos HDB, Yamamoto KC, Freitas CEC. *Peixes de lagos do médio Rio Solimões*. 2nd ed. Manaus: Reggo Edições; 2011.
- Takemoto RM, Pavanelli GC, Lizama MA, Luque JL, Poulin R. Host population density as the major determinant of endoparasite species richness in floodplain fishes of the upper Paraná River, Brazil. *J Helminthol* 2005; 79(1): 75-84. <http://dx.doi.org/10.1079/JOH2004264>. PMID:15831117.
- Takiyama LR, Silva URL, Jimenez EA, Pereira RA, Zacardi DM, Fernandes EF, et al. *Zoneamento ecológico econômico urbano das áreas de ressacas de Macapá e Santana, Estado do Amapá*. Macapá: IEPA; 2012.
- Tavares LER, Luque JL. Similarity between metazoan parasite communities of two sympatric brackish fish species from Brazil. *J Parasitol* 2008; 94(4): 985-989. <http://dx.doi.org/10.1645/GE-1460.1>. PMID:18576803.
- Tavares-Dias M, Dias-Júnior MB, Florentino AC, Silva LM, Cunha AC. Distribution pattern of crustacean ectoparasites of freshwater fish from Brazil. *Rev Bras Parasitol Vet* 2015; 24(2): 136-147. <http://dx.doi.org/10.1590/S1984-29612015036>. PMID:26154954.
- Thatcher VE. *Neochinorhynchus pterodoris* n. sp. (Acanthocephala: Neochinorhynchidae) do bacu liso (*Pterodorus granulosus*) da Amazônia Brasileira. *Acta Amazon* 1981; 11(3): 445-448.
- Toledo-Piza M. Phylogenetic relationships among *Acestrorhynchus* species (Ostariophysi: Characiformes: Acestrorhynchidae). *Zool J Linn Soc* 2007; 151(4): 691-757. <http://dx.doi.org/10.1111/j.1096-3642.2007.00355.x>.
- Zar JH. *Biostatistical analysis*. 5th ed. New Jersey: Prentice Hall; 2010.