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Probiotics and prebiotics in prevention and treatment of diseases in infants and children

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Abstract

Objective: To evaluate the impact of probiotics and prebiotics on the health of children.

Sources: MEDLINE and LILACS were searched for relevant English and French-language articles.

Summary of the findings: Human milk is rich in prebiotic oligosaccharides and may contain some probiotics. No data suggest that addition of probiotics to infant formula may be harmful, but evidence of its efficacy is insufficient for its recommendation. Since data suggest that addition of specific prebiotic oligosaccharides may reduce infections and atopy in healthy infants, their addition to infant formula seems reasonable. Long-term health benefits of pro- and prebiotics on the developing immune system remain to be proven. Selected probiotics reduce the duration of infectious diarrhea by 1 day, but evidence in prevention is lacking, except in antibiotic-associated diarrhea. Some specific probiotics prevent necrotizing enterocolitis, and other microorganisms may be beneficial in *Helicobacter pylori* gastritis and in infantile colic. Evidence is insufficient to recommend probiotics in prevention and treatment of atopic dermatitis. The use of probiotics in constipation, irritable bowel syndrome, inflammatory bowel disease, and extra-intestinal infections requires more studies.

Conclusions: Duration of administration, microbial dosage, and species used need further validation for both pro- and prebiotics. Unjustified health claims are a major threat for the pro- and prebiotic concept.

J Pediatr (Rio J). 2011;87(4):292-300: Gastrointestinal flora, intestinal microbiota, oligosaccharide, prebiotic, probiotic.

Introduction

Exclusive breastfeeding during at least the first 4 months of life is the preferred infant feeding method. Since this is not possible to achieve in all infants, artificial formula feeding is an alternative, that is, a second infant feeding choice. Exclusive breastfeeding, and not the composition of mother's milk, should be considered as the gold standard or the reference. Therefore, the goal of any alternative feeding method should be to mimic the effects of mother's milk on the

baby as closely as possible. There are many immunological components in mother's milk, but prebiotic oligosaccharides and the recently discovered probiotics are among the most important. Although the literature comparing the effects of breast milk to those of artificial feeding is limited, all studies show a health benefit to breastfeeding. Breast- and formulafed infants differ in physical growth and cognitive, emotional and social development. Health-care cost of formula-fed

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babies is much higher than that of breastfed babies. 1 Pediatric diseases for which the Agency for Healthcare Research and Quality reported risk ratios that favor breastfeeding are: necrotizing enterocolitis (NEC), otitis media, gastroenteritis, lower respiratory tract infection, atopic dermatitis, sudden infant death syndrome, asthma, leukemia, type 1 diabetes mellitus, and childhood obesity. 1 The first few years of life are a vulnerable period, during which the child's immature immune system is still developing and maturing. A child with frequent episodes of infection will not fully benefit from this window of opportunity and will not develop its full potential.

Mother's milk and artificial feeding

The composition of mother's milk is a very dynamic process and changes according to the region where the mother lives, the duration of breastfeeding, the moment of the day, and even during one feeding. It will never be possible to mimic this dynamic process. The macroand micronutrient composition of cow's milk differs substantially from the composition of mother's milk. The amounts and quality of proteins, carbohydrates, and lipids differ. Besides that, one of the most striking differences is the significant amount of prebiotic oligosaccharides in mother's milk (the third most important component, after carbohydrates and lipids), and the virtual absence of these oligosaccharides in animal milk.^{2,3} The amount and quality of oligosaccharides in mother's milk is a dynamic process, as happens with all the other constituents. More than 130 different oligosaccharides have been identified in breast milk.4

Gastrointestinal flora

The relevance of the composition and of the function of the gastrointestinal (GI) tract flora has been neglected for long. The GI flora, or GI microbiota, of an adult consists of more than 1,000 species. 5 Microbiota refers to a population of microscopic organisms that inhabit a bodily organ or portion of a person's body.6 Human microbiome refers to the unique entire population of microorganisms, and their complete genetic elements, which inhabit one's body.6 Adults have one trillion bacteria in the gut, that is, 10 to 100 times more bacteria than their own human cells. However, at birth, the GI tract is sterile. The presence of bacteria in the gut is mandatory for the development of different functions of the GI tract. If animals live in a sterile environment, adequate peristalsis does not develop. In other words, in the absence of an intestinal flora, the motor function of the gut is impaired. Bacteria are needed for the development of the gut-associated lymphoid tissue. It is often overlooked that the gut contains 60-70% of all the immune cells that a human being possesses.⁷

Because of the differences in composition between human milk and standard infant formula, there is broad consensus now that GI flora composition differs substantially in breastand formula-fed infants.⁸ although there is also literature concluding that breast- and bottle-fed babies have similar counts of bifidobacteria.9-12

Although bifidobacteria are the most prevalent bacteria found in the GI flora of both feeding groups, the amount is significantly higher in breastfed than in formula-fed infants.8 As early as in 1906, Tissier noted that significant stool colonization with bifidobacteria was protective against the likelihood of the development of diarrhea. The amount of Escherichia coli and bacteroids is significantly higher in formula-fed than in breastfed infants. Formula-fed infants have a more adult-type flora. After weaning (introduction of solids), the flora becomes more complex in both breastand formula-fed infants.

Bifidobacteria are the most important constituent of the dominant active flora.8 Lactobacilli are part of the sub-dominant flora and are under control by the dominant flora. Dietary and environmental changes constitute the transient flora, which is exogenous and does not colonize the GI tract. Lactobacilli and bifidobacteria inhibit the growth of exogenous and/or harmful bacteria, stimulate immune functions, aid in the digestion and/or absorption of food ingredients and minerals, and contribute to the synthesis of vitamins.

Infant's gestational age at birth, mode of delivery (vaginal birth vs. cesarean section), and diet seem to have significant effects on the intestinal microbiota.6,13 When infants are born by cesarean section, they do not swallow the mother's vaginal and intestinal flora. Mode of delivery, sterile foods, decreased consumption of naturally fermented food, increased hygiene measures, urban life, increased use of antibiotics, and many other factors decrease the exposure of the GI mucosa to microbes, which results in an altered intestinal microbiota.13

How can the dietary intake change the intestinal flora?

The composition of intestinal microflora does not change significantly after infancy. The composition of fecal flora in older children and adults is less variable and not as dependent on diet. However, during infancy, diet is a major player in the development of the intestinal microbiota. The abundant presence of prebiotic oligosaccharides in breast milk and their virtual absence in cow's milk are major determinants to explain the differences in intestinal microbiota in infants. The quality and amount of peptides¹⁴ and lactose¹⁵ are bifidogenic factors as well. During the first days or weeks of life, lactase is not yet fully developed.16 Therefore, undigested lactose reaches the colon, where it is fermented and has a bifidogenic effect.

The prebiotic concept means that non-digestible food ingredients are added to the dietary intake to beneficially affect the host by selectively stimulating the growth and/or activity of one or a limited number of bacteria in the colon that can improve the health of the host. Prebiotics evade digestion in the small intestine and must be selectively fermented in the colon. Prebiotics are usually in the form of oligosaccharides, which may occur naturally but can also be added as dietary supplements to foods, beverages, and infant formula. Although dietary nucleotides do not fit the exact definition of a prebiotic, they are prebiotic-like agents, and have immunomodulating and direct intestinal biological properties.¹⁷

Among many possible prebiotic oligosaccharides, galactooligosaccharides (GOS) and fructo-oligosaccharides (FOS) are the best known. GOS are short-chain oligosaccharides, resulting in fermentation in the cecum and right colon, are side products of lactose hydrolysis, and show lower incidence of side effects such as gas production and bloating. FOS are long-chain oligosaccharides, resulting in fermentation over the entire colon, and are derived from natural carbohydrates present in many plants like artichoke, leek, chicory, wheat, and banana. A mixture of FOS and GOS promotes the growth of healthy bacteria, and brings the GI flora composition of formula-fed infants close to that of breastfed infants. 18-20 A specific FOS/GOS mixture has also been shown to increase fecal IgA secretion.²⁰

Another possibility is to add probiotics, living microorganisms, to the food, or to administer them as food supplements or even medications. Probiotics are nonpathogenic live microorganisms that resist normal digestion to reach the colon alive, and which, when consumed in adequate amounts, have a positive effect on the health of the host. Probiotic microorganisms are typically members of the genera Lactobacillus, Bifidobacterium, and Streptococcus. However, yeast such as Saccharomyces boulardii is also a probiotic microorganism. A postbiotic is a metabolic by-product generated by a probiotic microorganism that influences the host's biological functions.²¹

Prebiotics change the intestinal flora of the host; probiotics, in turn, are specific strains of micro-organisms added as supplements, and belong to the transient flora. The healthy balance of the gut flora is constantly challenged by many elements, such as environmental factors (for example, age and stress), diseases (for example, infectious gastroenteritis), medications (for example, antibiotics and antacids), and many other factors. As with antibiotics, the use and efficacy of pro- and prebiotics need evidence-based medicine support.

Prevention with pre- and probiotics

The longer an infant is breastfed and the longer breastfeeding is exclusive, the better the protection from infectious diseases such as gastroenteritis will be. Promotion

of exclusive breastfeeding has to be maximally endorsed. During recent years, the composition of the second choice infant feeding, cow's milk-based formula, was adapted to better mimic the immune development of breastfed infants. To recreate the benefits of breastfeeding, probiotics and/or prebiotic oligosaccharides have been added to infant formula. The study from Weizman et al. illustrates the specificity of the strains used for this purpose: Lactobacillus reuteri resulted in a greater health benefit than Bifidobacterium BB12.²² Compared to the B. BB12 group, the L. reuteri group had a reduction in number of health care contacts, absent days from day care, febrile episodes, diarrheal episodes, and antibiotic prescriptions, and a reduction in the duration of diarrhea as well.²² Three large randomized controlled trials (RCTs) provide evidence of a modest effect, statistically significant but of questionable clinical importance, of some probiotic strains (Lactobacillus casei GG, L. reuteri, and Bifidobacterium lactis) on the prevention of community-acquired diarrhea.²³ Results of RCTs later published have indicated that there is modest benefit to administer probiotics to prevent acute GI tract infections in healthy infants and children.⁶ With a number needed to treat between 7 and 15 for gastroenteritis, and of 30 for respiratory tract infections, L. casei GG reduces nosocomial infections.²⁴⁻²⁶ Most of the studies were conducted in childcare centers. The strains of probiotics used include L. casei GG, Streptococcus thermophilus, B. lactis, and L. reuteri mixed with milk or infant formula, or given as an oral supplement.6 However, to date, the available data do not support routine use of probiotics to prevent nosocomial diarrhea in childcare centers. 6 In comparison to placebo, daily dietary probiotic supplementation with either a single strain (Lactobacillus acidophilus NCFM) or two strains (L. acidophilus NCFM in combination with Bifidobacterium animalis subspecies lactis Bi-07) for 6 months reduces fever, rhinorrhea, cough, antibiotic prescription incidence, and the number of missed school days attributable to illness in children 3 to 5 years of age.²⁷ Saran et al. showed that feeding fermented milk to Indian infants over a period of 6 months resulted in a significantly better weight gain and a 50% reduction of infectious diarrhea.²⁸ Bifidobacterium lactis HN019 and GOS fortified milk resulted in a better iron status, although both groups were fed isocaloric diets with the same iron content as a control formula.²⁹ In a doubleblind, prospective, randomized trial in 3,758 children aged 1-5 years living in an urban slum community in Kolkata (India), the health benefit of the daily intake of a probiotic drink with Lactobacillus casei strain Shirota or a nutrient drink during 12 weeks, with a follow-up of another 12 weeks without intake of any study product, was tested. The results showed a 14% decrease in episodes of diarrhea.³⁰

In the prevention of antibiotic-associated diarrhea (AAD), meta-analyses of published results of RCTs provide evidence for efficacy of a number of probiotic strains, such as S. boulardii.31-35 Approximately one in seven cases of AAD was prevented by the use of a probiotic.31 According to a recent review, administration of lactobacilli reduces AAD in adults, but not in children.³² A Cochrane review from 2007 concluded that there was not enough evidence to recommend probiotics to prevent AAD, because there was no statistical benefit, according to the intention-totreat analysis.36

A limited number of data suggest that probiotics reduce the incidence of atopic dermatitis. This effect is long-lasting, resulting in a reduced incidence of atopic dermatitis even at the age of 7, with an intervention that started during pregnancy and stopped at 6 months of age.³⁷ The evidence in prevention of atopic disease is limited to selected strains. 38,39 As concluded in a review by Prescott & Björkstén and in the 2007 Cochrane review, despite the encouraging results of some studies, there is insufficient evidence to recommend the routine supplementation of probiotics to either pregnant women or infants to prevent atopic diseases in childhood. 40,41 Van der Aa et al. reviewed seven RCTs on prevention of atopic dermatitis (and 12 on treatment), and found that the results of these trials are conflicting. 42 The authors conclude that there is not enough evidence to support the use of pro-, pre- or synbiotics for prevention or treatment of atopic dermatitis in children.⁴² Explanations for varied study results include host factors such as genetic susceptibility, environmental factors such as geographic region and diet, and study variables such as probiotic strains and doses used. 40,43

Different probiotics strains reduce the risk for NEC.44 Although the first papers on probiotics strains reducing the incidence of NEC date only from about 12 year ago,⁴⁵ literature on this topic has recently exploded. The old and newest data report that administration of lactobacilli and bifidobacteria reduces the incidence of NEC, especially in hospital units with a high incidence of this condition.^{45,46} However, as the number of studies increase, negative data (with L. GG) are published as well.⁴⁷ The number needed to treat to prevent one case of NEC is over 20.48,49 Improved intestinal motility may be one of the mechanisms of action of NEC prevention.⁴⁵ In newborns and infants, L. reuteri has recently been shown to improve gastric emptying and reduce regurgitation.50,51

It is estimated that approximately 40-70% of children and adult patients suffering from inflammatory bowel disease routinely use alternative medicines, including probiotics, as adjunctive or replacement therapy for prescribed medications.⁵² Although the experimental basis to expect clinical efficacy of probiotics in inflammatory bowel disease is rather convincing, clinical evidence for the benefit of probiotics is currently not convincing in Crohn's disease, either in adults or in children.⁵³ In ulcerative colitis, several probiotic products, especially VSL#3 (a high-concentration mixture of seven strains), have been found effective

as adjuvant therapy, both in inducing and maintaining remission.53 Recent literature suggested a reduction in relapse of ulcerative colitis with VSL#3 in children.54

There is some evidence that some lactobacilli prevent recurrent urinary tract infection in women. However, data in children are lacking. The same is true for recurrent vulvovaginitis. There are also some reports on probiotics in otitis media and asthma.

Most of the information regarding prebioticoligosaccharides results from studies with specific FOS/GOS mixtures. These oligosaccharides lead to a GI flora development close to the flora in breastfed infants. Stool consistency and frequency are similar in breast- and formula-fed infants if prebiotic oligosaccharides were added to the formula. Neutral and acidic oligosaccharides in preterm infants reduce serious infections in this high-risk population.⁵⁵ Moro et al. showed that a prebiotic mixture results in a decrease in atopic dermatitis, and could relate this finding to the number of bifidobacteria⁵⁶. In a similar study, a reduction of infections not only during the 6-month intervention period, but persisting up to the age of 2 years, was demonstrated. 57,58 Another group confirmed these data in a similar study with a follow-up to the age of 12 months.⁵⁹ Higher levels of secretory IgA in the feces are measured in relation to the presence of FOS/GOS in formula.60 However, immune parameters are not different at week 8 and 26 in infants fed with breast milk or formula, with or without GOS/FOS prebiotics.61

In a community-based RCT, children 1-3 years of age were randomly allocated to receive either control milk (n = 312) or the same milk fortified with 2.4 g/day of a prebiotic oligosaccharides and 1.9x107 colony forming unit/ day of B. lactis HN019 (n = 312).62 Biweekly household surveillance was conducted to gather information on compliance to trial plan and morbidity. Overall, there was no effect of the prebiotic and probiotic on diarrhea (6% reduction, 95% confidence interval [95%CI] -1 to 12%, p = 0.08). The incidence of dysentery episodes was reduced by 21% (95%CI 0-38%, p = 0.05), while the incidence of pneumonia and severe acute lower respiratory infection was reduced by 24 (95%CI 0-42%, p = 0.05) and 35% (95%CI 0-58%, p = 0.05), respectively. Compared to children in the control group, children in the intervention group had 16 (95%CI 5-26%, p = 0.004) and 5% (95%CI 0-10%, p = 0.05) reduction in days with severe illness and high fever, respectively.⁶² The authors concluded that milk can be a good medium for delivery of pre- and probiotics.62

Treatment with pre- and probiotics

Most of the evidence is provided by studies evaluating the efficacy of probiotics in the treatment of acute and chronic infectious gastroenteritis. While some studies with probiotics in acute gastroenteritis are negative, a significant number

of them show a shortening of the duration of diarrhea by approximately 24 hours.²³ A 24-hour shortening in the length of hospitalization was also reported. 63 The probiotic veast S. boulardii was shown to be more effective than fermented food. 64 Lactobacillus rhamnosus GG is associated with reduced diarrheal duration and severity. 65,66 However, recent studies with this lactobacillus reported no doseefficacy or were negative.67-69 Several meta-analyses70-72 and a Cochrane review⁷³ concluded that, in the treatment of acute infectious diarrhea in children, probiotics reduce the number of diarrheal stools and the duration of the diarrhea by approximately 1 day. Probiotic efficacy was also demonstrated in infections with Giardia lamblia and amebiasis.74,75 Seven studies on S. boulardii were included in one meta-analysis. ⁷⁶ The Iberic-Latin American guideline for acute gastroenteritis management in infants and preschoolers recommends racecadotril, zinc, smectite, L. GG or S. boulardii.77 The European Society of Paediatric Gastroenterology, Hepatology and Nutrition concluded that the use of specific probiotics in the treatment of acute gastroenteritis may be indicated in some situations.78 The benefit is strain-dependent.6

There have been no published RCTs of children that have investigated the effect of probiotics for treatment of AAD. However, recently published data suggest some efficacy in treatment as well.⁷⁹

A number of studies evaluated the efficacy of probiotics as add-on treatment in the eradication of Helicobacter pylori.80,81 While some studies are negative, the majority of the data shows a reduced incidence of adverse effects of the eradication therapy. Moreover, a number of studies showed that probiotics, even when administered as a dairy product, result in an additional eradication rate of about 10%.80 As a consequence, the use of such probiotic strains in eradication-resistant cases should be further evaluated.80,81

Probiotics have been clinically tested in many other indications such as colic, constipation, and atopic dermatitis. A number of studies provided limited evidence for a beneficial effect of different probiotics in the management of allergic diseases (atopic eczema and allergic rhinitis). 37,39 A 2008 Cochrane review concluded that probiotics have not yet been proven to be effective in the treatment of atopic dermatitis.82 A RCT concluded that synbiotics (Bifidobacterium breve M-16V and a GOS/FOS mixture) did not reduce atopic dermatitis.83 In theory, probiotics may be beneficial in the treatment of inflammatory bowel disease. A recent Cochrane review indicated that there is no proven benefit for maintaining remission by administering probiotics to adults or children with Crohn's disease.84 Nor are probiotics recommended in ulcerative colitis at this time, although there is more evidence of benefit for this disease. There is no evidence to recommend the routine use of probiotics in children with constipation or irritable bowel

syndrome. One open and one double-blind study, from the same group, reported an efficacy of over 90% of L. reuteri in colicky breastfed infants.85,86 In the open trial, success rate of the probiotic was 95% compared to a 93% failure rate of simethicone treatment.86 In the double-blind trial, crying time decreased by 90% in the probiotic group, while the decrease was 70% in the control group, which did not receive any intervention.85 Responders, defined as patients that presented a decrease of crying time by more than 50%, were 96% in the intervention group and 71% in the control group.86 On the basis of limited information, probiotics may be of benefit in treatment of colic in exclusively breastfed infants, but more studies are needed before this attitude can be recommended.6

Choice of probiotic strains, as well as timing of the intervention, are important variables.³⁷ Fecal colonization has been shown to be dose-dependent.87 Only very few data have been published regarding dose-efficacy of probiotic strains. Recently, a study with L. GG did not report a dose-effect relationship.69 A dose-related efficacy has been demonstrated for L. reuteri.88 Indirect evidence for a dose-efficacy relationship for S. boulardii is shown by the negative results in the trial by Lewis et al., in which a low dose (250 mg/day) was used to prevent AAD.90 If probiotics are used as medication, a medication-like approach should be stronger developed. In this situation, dose-response studies are required, 90 especially because there is literature suggesting that small doses can be more effective than higher ones. 91 Equally, clinical studies with the product in its commercial presentation and dosage are mandatory, since certain in vitro effects are seen only at small bacterial doses, 92 and high doses may produce opposite effects to those obtained at small doses.93 A few studies suggest that administration of probiotics (Lactobacillus rhamnosus 19070-2, Lactobacillus reuteri DSM 12246, and S. boulardii) may have beneficial effects in certain conditions, but data are too limited to make recommendations.94-96 There is even less evidence regarding the optimal duration of treatment; to the knowledge of the authors, there are no studies comparing the outcome for several durations of probiotic administration.

Gastrointestinal flora and immune function

The intestinal flora is a major determinant of the development of the intestinal mucosal defense system. It is thought that the occurrence of many diseases, both intestinal and non-intestinal, can be related to dysregulation or interference in the early development of the intestinal mucosal defense system.⁶ These diseases can be atopic or autoimmune. Although genetic predisposition is the major determining factor, GI flora plays an important role. Breakdown products from the diet, such as nucleotics and oligosaccharides, and intestinal bacteria give orientation to the development of mature T lymphocytes, which are needed for the development of the acquired and innate immune system.97 T-lymphocyte recognition of specific oligosaccharides bound to intestinal pathogens plays an important role in preventing GI illness. 6 Therefore, the early intestinal microbial development of an infant is considered to be a pivotal factor for later health. Probiotic bacteria, postbiotic bacterial by-products, and dietary prebiotics may exert positive effects on the development of the mucosal immune system.6 As a consequence, contact with "nonbeneficial" microorganisms and antimicrobial agents during the neonatal period may result in immune dysregulation in susceptible individuals, and may be related to some diseases.⁶ There is evidence that human milk contains mononuclear cells that traffic intestinally derived bacterial components from the mother to her infant.^{6,98} Mother's milk contains bacterial components derived from the mother's intestinal flora. This process is termed "bacterial imprinting,"6,99

Cost, safety and adverse effects

Cost of pre- and probiotics should be considered as well. Since probiotics shorten the duration of infectious diarrhea by about 24 hours, the patient should decide if the cost of a probiotic compensates for 24 hours less of diarrhea. However, if the few data showing a 24-hour decrease in the duration of hospitalization are confirmed, probiotics offer a socio-economic benefit to the society in case of hospitalization. 100

Probiotics are generally regarded as safe, and side effects in ambulatory care are seldom. Large-scale epidemiological studies in countries where probiotic use is endemic demonstrate low rates of systemic infection in adults, between 0.05 and 0.40%.101 Saccharomyces fungemia secondary to use of the probiotic has been described in critically ill patients who were receiving nutrition enterally or with a central venous catheter. 102 Before use of a probiotic is considered in hospitalized patients, careful assessment of the risk-benefit ratio must be made. 102 To ensure patient safety, probiotics should be properly handled during administration. 102

Regarding probiotics, strain specificity is important. Until a few years ago, probiotics were discussed primarily in the context of alternative medicine, but they are now entering mainstream medical practice. 103 As a consequence, probiotics are more frequently used in critically ill patients. The use of probiotics in cancer patients is an area of ongoing intensive research. In adult critically ill patients, probiotic administration and intestinal decontamination seem to have a similar preventive effect on infection and mortality. 104 A multispecies probiotic preparation did not reduce the risk of infectious complications and was associated with an increased risk of mortality in patients

with a predicted high risk for severe acute pancreatitis. 105 This study illustrates that probiotic administration is not free of adverse events per se. Bifidobacterium animalis, a traditional probiotic species that is tested in experimental colitis in a GF IL-10-/- mice model, is capable to induce marked duodenal and mild colonic inflammation, and also TH1/TH17 immune responses. 106 This suggests a potential pathogenic role for this commensal bacterial species in a susceptible host. 103 Although reduction of allergic sensitization has been shown with several probiotic strains, an increased incidence of sensitization has been shown as well. 107 There are reports of trials that had to be stopped because of the high incidence of GI side effects and even of heat-killed microorganisms. 108 There are indications that certain in vitro effects are seen only at low bacterial doses, 92 and that high doses may produce opposite effects to those obtained at low doses.93

Conclusion

Western medicine has only recently discovered that intestinal microbiota is a major determinant of the wellbeing of the host. Exclusive human milk is the preferred feeding for infants until 4 to 6 months of age. Human milk is rich in oligosaccharides, which are part of its prebiotic components. Human milk may also contain some naturally occurring probiotic bacteria. There are no data to suggest that addition of probiotics to infant formula may be harmful to healthy term infants. On the other hand, evidence of clinical efficacy for addition of probiotics is insufficient to actively recommend the routine use of these formulas. Specific prebiotics are capable of reducing common infections and atopy in otherwise healthy children. Addition of specific oligosaccharides to infant formula seems reasonable. Longterm health benefits of pro- or prebiotics providing evidence of beneficial effects on the developing immune system beyond early infancy remain to be proven.

There is some evidence that manipulation of the intestinal microbiota with food and food supplements containing preand probiotics contributes to a possible health benefit if the initial flora was abnormal. There is evidence in otherwise healthy infants and children to support the use of selected probiotics early in the course of infectious diarrhea, because of a reduction of its duration by 1 day. However, there is no evidence to support the routine use of probiotics to prevent infectious diarrhea. There is some evidence for using selected probiotics in the prevention of AAD, but not in its treatment. Evidence is insufficient to recommend probiotic administration in the prevention and treatment of atopic dermatitis. There is some evidence that specific probiotics prevent NEC. There may be some benefit for administration of selected probiotics as add-on treatment of Helicobacter pylori gastritis and in infantile colic, but further studies are necessary. The benefit of probiotics for treating

conditions such as constipation, irritable bowel syndrome, inflammatory bowel disease (although data are slightly better for ulcerative colitis than for Crohn's disease), and extra-intestinal infections requires more RCTs.

Although probiotics can be helpful in specific disorders, they have been broadly prescribed for disorders without clear evidence to support their use. Probiotic effects are target specific. Optimal duration of administration, preferred microbial dosage, and species used need further validation for both pro and prebiotics. The major threat for the concept of allocating a major role to the manipulation of the intestinal microbiota on health is the commercialization of products claiming health benefits that have insufficiently been validated. The effect of probiotic microorganisms vary with factors such as age, health, gender, diet, residence, and environment. The consequence of this variation is that results from studies in children/aged subjects, in sick people, or people from the Third World cannot be transferred without further examination to adults, healthy people or people from industrialized countries, respectively.6

References

- 1. Bartick M, Reinhold A. The burden of suboptimal breastfeeding in the United States: a pediatric cost analysis. Pediatrics. 2010;125: e1048-56.
- Vandenplas Y. Oligosaccharides in infant formula. Br J Nutr. 2002;87 Suppl 2:S293-6.
- 3. Picciano MF. Nutrient composition of human milk. Pediatr Clin North Am. 2001;48:53-67.
- 4. Coppa GV, Zampini L, Galeazzi T, Gabrielli O. Prebiotics in human milk: a review. Dig Liver Dis. 2006;38 Suppl 2:S291-4.
- Zoetendal EG, Vaughan EE, de Vos WM. A microbial world within us. Mol Microbiol. 2006;59:1639-50.
- 6. Thomas DW, Greer FR; American Academy of Pediatrics Committee on Nutrition; American Academy of Pediatrics Section on Gastroenterology, Hepatology, and Nutrition. Probiotics and prebiotics in pediatrics. Pediatrics. 2010;126:1217-31.
- Round JL, Mazmanian SK. The gut microbiota shapes intestinal immune responses during health and disease. Nat Rev Immunol. 2009;9:313-23.
- 8. Harmsen HJ, Wildeboer-Veloo AC, Raangs GC, Wagendorp AA, Klijn N, Bindels JG, et al. Analysis of intestinal flora development in breast-fed and formula-fed infants by using molecular identification and detection methods. J Pediatr Gastroenterol Nutr. 2000;30:61-7.
- 9. Kleessen B, Bunke H, Tovar K, Noack J, Sawatzki G. Influence of two infant formulas and human milk on the development of the faecal flora in newborn infants. Acta Paediatr. 1995;84:1347-56.
- 10. Penders J, Vink C, Driessen C, London N, Thijs C, Stobberingh EE. Quantification of Bifidobacterium spp., Escherichia coli and Clostridium difficile in faecal samples of breast-fed and formula-fed infants by real-time PCR. FEMS Microbiol Lett. 2005;243:141-7.
- 11. Sakata S, Tonooka T, Ishizeki S, Takada M, Sakamoto M, Fukuyama M. et al. Culture-independent analysis of fecal microbiota in infants. with special reference to Bifidobacterium species. FEMS Microbiol Lett. 2005;243:417-23.
- 12. Satokari RM, Vaughan EE, Favier CF, Doré J, Edwards C, de Vos WM. Diversity of Bifidobacterium and Lactobacillus spp. in breastfed and formula-fed infants as assessed by 16S rDNA sequence differences. Microb Ecol Health Dis. 2002;14:97-105.

- 13. Rautava S, Ruuskanen O, Ouwehand A, Salminen S, Isolauri E. The hygiene hypothesis of atopic disease--an extended version. J Pediatr Gastroenterol Nutr. 2004;38:378-88.
- 14. Shadid R, Haarman M, Knol J, Theis W, Beermann C, Rjosk-Dendorfer D, et al. Effects of galactooligosaccharide and longchain fructooligosaccharide supplementation during pregnancy on maternal and neonatal microbiota and immunity - a randomized, double-blind, placebo-controlled study. Am J Clin Nutr. 2007;86:1426-37.
- 15. Heyman MB; Committee on Nutrition. Lactose intolerance in infants, children, and adolescents. Pediatrics. 2006;118:1279-86.
- 16. McClure RJ, Newell SJ. Randomized controlled study of digestive enzyme activity following trophic feeding. Acta Paediatr. 2002;91:292-6.
- 17. Quan R, Barness LA. Do infants need nucleotide supplemented formula for optimal nutrition? J Pediatr Gastroenterol Nutr. 1990;11:429-34.
- 18. Moro G, Minoli I, Mosca M, Fanaro S, Jelinek J, Stahl B, et al. Dosagerelated bifidogenic effects of galacto- and fructooligosaccharides in formula-fed term infants. J Pediatr Gastroenterol Nutr. 2002:34:291-5.
- 19. Haarman M, Knol J. Quantitative real-time PCR assays to identify and quantify fecal Bifidobacterium species in infants receiving a prebiotic infant formula. Appl Environ Microbiol. 2005:71:2318-24.
- 20. Scholtens PA, Alliet P, Raes M, Alles MS, Kroes H, Boehm G, et al. Fecal secretory immunoglobulin A is increased in healthy infants who receive a formula with short-chain galacto-oligosaccharides and long-chain fructo-oligosaccharides. J Nutr. 2008;138:1141-7.
- 21. Falony G, Vlachou A, Verbrugghe K, De Vuyst L. Cross-feeding between Bifidobacterium longum BB536 and acetate-converting, butyrate-producing colon bacteria during growth on oligofructose. Appl Environ Microbiol. 2006;72:7835-41.
- 22. Weizman Z, Asli G, Alsheikh A. Effect of a probiotic infant formula on infections in child care centers: comparison of two probiotic agents. Pediatrics. 2005;115:5-9.
- 23. Szajewska H, Setty M, Mrukowicz J, Guandalini S. Probiotics in gastrointestinal diseases in children: hard and not-so-hard evidence of efficacy. J Pediatr Gastroenterol Nutr. 2006;42:454-75.
- 24. Szajewska H, Mrukowicz JZ. Use of probiotics in children with acute diarrhea. Paediatr Drugs. 2005;7:111-22.
- 25. Hojsak I, Abdović S, Szajewska H, Milosević M, Krznarić Z, Kolacek S. Lactobacillus GG in the prevention of nosocomial gastrointestinal and respiratory tract infections. Pediatrics. 2010;125:e1171-7.
- 26. Szajewska H, Kotowska M, Mrukowicz JZ, Armanska M, Mikolajczyk W. Efficacy of Lactobacillus GG in prevention of nosocomial diarrhea in infants. J Pediatr. 2001;138:361-5.
- 27. Leyer GJ, Li S, Mubasher ME, Reifer C, Ouwehand AC. Probiotic effects on cold and influenza-like symptom incidence and duration in children. Pediatrics. 2009;124:e172-9.
- 28. Saran S, Gopalan S, Krishna TP. Use of fermented foods to combat stunting and failure to thrive. Nutrition. 2002;18:393-6.
- 29. Sazawal S, Dhingra U, Sarkar A, Dhingra P, Deb S, Marwah D, et al. Efficacy of milk fortified with a probiotic Bifidobacterium lactis (DR-10TM) and prebiotic galacto-oligosaccharides in prevention of morbidity and on nutritional status. Asia Pac J Clin Nutr. 2004;13: S28.
- 30. Sur D, Manna B, Niyogi SK, Ramamurthy T, Palit A, Nomoto K, et al. Role of probiotic in preventing acute diarrhoea in children: a community-based, randomized, double-blind placebo-controlled field trial in an urban slum. Epidemiol Infect. 2010;30:1-8.
- 31. Szajewska H, Ruszczynski M, Radzikowski A. Probiotics in the prevention of antibiotic-associated diarrhea in children: a meta-analysis of randomized controlled trials. J Pediatr. 2006;149:367-72.
- 32. Corrêa NB, Péret Filho LA, Penna FJ, Lima FM, Nicoli JR. A randomized formula controlled trial of Bifidobacterium lactis and Streptococcus thermophilus for prevention of antibiotic-associated diarrhea in infants. J Clin Gastroenterol. 2005;39:385-9.
- 33. Hawrelak JA, Whitten DL, Myers SP. Is Lactobacillus rhamnosus GG effective in preventing the onset of antibiotic-associated diarrhoea: a systematic review. Digestion. 2005;72:51-6.

- D'Souza AL, Rajkumar C, Cooke J, Bulpitt CJ. Probiotics in prevention of antibiotic associated diarrhoea: meta-analysis. BMJ. 2002;324:1361.
- 35. Kale-Pradhan PB, Jassal HK, Wilhelm SM. Role of Lactobacillus in the prevention of antibiotic-associated diarrhea: a meta-analysis. Pharmacotherapy. 2010;30:119-26.
- Johnston BC, Supina AL, Ospina M, Vohra S. Probiotics for the prevention of pediatric antibiotic-associated diarrhea. Cochrane Database Syst Rev. 2007:CD004827.
- Kalliomäki M. Salminen S, Poussa T, Isolauri E. Probiotics during the first 7 years of life: a cumulative risk reduction of eczema in a randomized, placebo-controlled trial. J Allergy Clin Immunol. 2007;119:1019-21.
- 38. Kalliomäki M, Antoine JM, Herz U, Rijkers GT, Wells JM, Mercenier A. Guidance for substantiating the evidence for beneficial effects of probiotics: prevention and management of allergic diseases by probiotics. J Nutr. 2010;140:713S-21S.
- Lee J, Seto D, Bielory L. Meta-analysis of clinical trials of probiotics for prevention and treatment of pediatric atopic dermatitis. J Allergy Clin Immunol. 2008;121:116-21.
- 40. Prescott SL, Björkstén B. Probiotics for the prevention or treatment of allergic diseases. J Allergy Clin Immunol. 2007;120:255-62.
- Osborn DA, Sinn JK. Probiotics in infants for prevention of allergic disease and food hypersensitivity. Cochrane Database Syst Rev. 2007:CD006475.
- 42. van der Aa LB, Heymans HS, van Aalderen WM, Sprikkelman AB. Probiotics and prebiotics in atopic dermatitis: review of the theoretical background and clinical evidence. Pediatr Allergy Immunol. 2010;21:e355-67.
- 43. Penders J, Stobberingh EE, van den Brandt PA, Thijs C. The role of the intestinal microbiota in the development of atopic disorders. Allergy. 2007;62:1223-36.
- Alfaleh K, Anabrees J, Bassler D. Probiotics reduce the risk of necrotizing enterocolitis in preterm infants: a meta-analysis. Neonatology. 2010;97:93-9.
- 45. Hoyos AB. Reduced incidence of necrotizing enterocolitis associated with enteral administration of Lactobacillus acidophilus and Bifidobacterium infantis to neonates in an intensive care unit. Int J Infect Dis. 1999;3:197-202.
- 46. Braga TD, da Silva GA, de Lira PI, de Carvalho Lima M. Efficacy of Bifidobacterium breve and Lactobacillus casei oral supplementation on necrotizing enterocolitis in very-low-birth-weight preterm infants: a double-blind, randomized, controlled trial. Am J Clin Nutr. 2010;93:81-6.
- 47. Luoto R, Matomäki J, Isolauri E, Lehtonen L. Incidence of necrotizing enterocolitis in very-low-birth-weight infants related to the use of Lactobacillus GG. Acta Paediatr. 2010;99:1135-8.
- 48. Lin HC, Su BH, Chen AC, Lin TW, Tsai CH, Yeh TF, et al. Oral probiotics reduce the incidence and severity of necrotizing enterocolitis in very low birth weight infants. Pediatrics. 2005;115:1-4.
- 49. Lin HC, Hsu CH, Chen HL, Chung MY, Hsu JF, Lien RI, et al. Oral probiotics prevent necrotizing enterocolitis in very low birth weight preterm infants: a multicenter, randomized, controlled trial. Pediatrics. 2008;122:693-700.
- Indrio F, Riezzo G, Raimondi F, Bisceglia M, Cavallo L, Francavilla R. Effects of probiotic and prebiotic on gastrointestinal motility in newborns. J Physiol Pharmacol. 2009;60 Suppl 6:27-31.
- Indrio F, Riezzo G, Raimondi F, Bisceglia M, Filannino A, Cavallo L, et al. Lactobacillus reuteri accelerates gastric emptying and improves regurgitation in infants. Eur J Clin Invest. 2011;41:417-22.
- Sartor RB. Therapeutic manipulation of the enteric microflora in inflammatory bowel diseases: antibiotics, probiotics, and prebiotics. Gastroenterology. 2004;126:1620-33.
- Guandalini S. Update on the role of probiotics in the therapy of pediatric inflammatory bowel disease. Expert Rev Clin Immunol. 2010;6:47-54.
- 54. Miele E, Pascarella F, Giannetti E, Quaglietta L, Baldassano RN, Staiano A. Effect of a probiotic preparation (VSL#3) on induction and maintenance of remission in children with ulcerative colitis. Am J Gastroenterol. 2009;104:437-43.

- 55. Westerbeek EA, van den Berg JP, Lafeber HN, Fetter WP, Boehm G, Twisk JW, et al. Neutral and acidic oligosaccharides in preterm infants: a randomized, double-blind, placebo-controlled trial. Am J Clin Nutr. 2010;91:679-86.
- 56. Moro G, Arslanoglu S, Stahl B, Jelinek J, Wahn U, Boehm G. A mixture of prebiotic oligosaccharides reduces the incidence of atopic dermatitis during the first six months of age. Arch Dis Child. 2006;91:814-9.
- 57. Arslanoglu S, Moro GE, Boehm G. Early supplementation of prebiotic oligosaccharides protects formula-fed infants against infections during the first 6 months of life. J Nutr. 2007;137:2420-4.
- Arslanoglu S, Moro GE, Schmitt J, Tandoi L, Rizzardi S, Boehm G. Early dietary intervention with a mixture of prebiotic oligosaccharides reduces the incidence of allergic manifestations and infections during the first two years of life. J Nutr. 2008:138:1091-5.
- Bruzzese E, Volpicelli M, Squeglia V, Bruzzese D, Salvini F, Bisceglia M, et al. A formula containing galacto- and fructo-oligosaccharides prevents intestinal and extra-intestinal infections: an observational study. Clin Nutr. 2009;28:156-61.
- 60. Scholtens PA, Alliet P, Raes M, Alles MS, Kroes H, Boehm G, et al. Fecal secretory immunoglobulin A is increased in healthy infants who receive a formula with short-chain galacto-oligosaccharides and long-chain fructo-oligosaccharides. J Nutr. 2008;138:1141-7.
- Raes M, Scholtens PA, Alliet P, Hensen K, Jongen H, Boehm G, et al. Exploration of basal immune parameters in healthy infants receiving an infant milk formula supplemented with prebiotics. Pediatr Allergy Immunol. 2010;21:e377-85.
- 62. Sazawal S, Dhingra U, Hiremath G, Sarkar A, Dhingra P, Dutta A, et al. Prebiotic and probiotic fortified milk in prevention of morbidities among children: community-based, randomized, double-blind, controlled trial. PLoS One. 2010;5:e12164.
- Szajewska H, Setty M, Mrukowicz J, Guandalini S. Probiotics in gastrointestinal diseases in children: hard and not-so-hard evidence of efficacy. J Pediatr Gastroenterol Nutr. 2006;42:454-75.
- 64. Kurugöl Z, Koturoğlu G. Effects of Saccharomyces boulardii in children with acute diarrhoea. Acta Paediatr. 2005;94:44-7.
- 65. Eren M, Dinleyici EC, Vandenplas Y. Clinical efficacy comparison of Saccharomyces boulardii and yogurt fluid in acute non-bloody diarrhea in children: a randomized, controlled, open label study. Am J Trop Med Hyg. 2010;82:488-91.
- 66. Guarino A, Lo Vecchio A, Canani RB. Probiotics as prevention and treatment for diarrhea. Curr Opin Gastroenterol. 2009;25:18-23.
- Szajewska H, Skórka A, Ruszczyński M, Gieruszczak-Białek D. Meta-analysis: Lactobacillus GG for treating acute diarrhoea in children. Aliment Pharmacol Ther. 2007;25:871-81.
- 68. Misra S, Sabui TK, Pal NK. A randomized controlled trial to evaluate the efficacy of lactobacillus GG in infantile diarrhea. J Pediatr. 2009;155:129-32.
- 69. Ritchie BK, Brewster DR, Tran CD, Davidson GP, McNeil Y, Butler RN. Efficacy of Lactobacillus GG in aboriginal children with acute diarrhoeal disease: a randomised clinical trial. J Pediatr Gastroenterol Nutr. 2010;50:619-24.
- Basu S, Paul DK, Ganguly S, Chatterjee M, Chandra PK. Efficacy
 of high-dose Lactobacillus rhamnosus GG in controlling acute
 watery diarrhea in Indian children: a randomized controlled trial.
 J Clin Gastroenterol. 2009;43:208-13.
- Szajewska H, Mrukowicz JZ. Probiotics in the treatment and prevention of acute infectious diarrhea in infants and children: a systematic review of published randomized, double-blind, placebocontrolled trials. J Pediatr Gastroenterol Nutr. 2001;33 Suppl 2: S17-25.
- 72. Van Niel CW, Feudtner C, Garrison MM, Christakis DA. Lactobacillus therapy for acute infectious diarrhea in children: a meta-analysis. Pediatrics. 2002;109:678-84.
- Huang JS, Bousvaros A, Lee JW, Diaz A, Davidson EJ. Efficacy of probiotic use in acute diarrhea in children: a meta-analysis. Dig Dis Sci. 2002;47:2625-34.
- Allen SJ, Okoko B, Martinez E, Gregorio G, Dans LF. Probiotics for treating infectious diarrhoea. Cochrane Database Syst Rev. 2004:CD003048.

- 75. Dinleyici EC, Eren M, Yargic ZA, Dogan N, Vandenplas Y. Clinical efficacy of Saccharomyces boulardii and metronidazole compared to metronidazole alone in children with acute bloody diarrhea caused by amebiasis: a prospective, randomized, open label study. Am J Trop Med Hyg. 2009;80:953-5.
- Besirbellioglu BA, Ulcay A, Can M, Erdem H, Tanyuksel M, Avci IY, et al. Saccharomyces boulardii and infection due to Giardia lamblia. Scand J Infect Dis. 2006;38:479-81.
- 77. Szajewska H, Skórka A. Saccharomyces boulardii for treating acute gastroenteritis in children: updated meta-analysis of randomized controlled trials. Aliment Pharmacol Ther. 2009;30:960-1.
- 78. Gutiérrez Castrellón P, Polanco Allué I, Salazar Lindo E. An evidence based Iberic-Latin American guideline for acute gastroenteritis management in infants and prescholars. An Pediatr (Barc). 2010;72:220.e1-220.e20.
- 79. Guarino A, Albano F, Ashkenazi S, Gendrel D, Hoekstra JH, Shamir R, et al; ESPGHAN/ESPID Evidence-Based Guidelines for the Management of Acute Gastroenteritis in Children in Europe Expert Working Group. European Society for Paediatric Gastroenterology, Hepatology, and Nutrition/European Society for Paediatric Infectious Diseases evidence-based guidelines for the management of acute gastroenteritis in children in Europe: executive summary. J Pediatr Gastroenterol Nutr. 2008;46:619-21.
- Rohde CL, Bartolini V, Jones N. The use of probiotics in the prevention and treatment of antibiotic-associated diarrhea with special interest in Clostridium difficile-associated diarrhea. Nutr Clin Pract. 2009;24:33-40.
- Sachdeva A, Nagpal J. Effect of fermented milk-based probiotic preparations on Helicobacter pylori eradication: a systematic review and meta-analysis of randomized-controlled trials. Eur J Gastroenterol Hepatol. 2009;21:45-53.
- 82. Sýkora J, Valecková K, Amlerová J, Siala K, Dedek P, Watkins S, et al. Effects of a specially designed fermented milk product containing probiotic Lactobacillus casei DN-114 001 and the eradication of H. pylori in children: a prospective randomized double-blind study. J Clin Gastroenterol. 2005;39:692-8.
- Boyle RJ, Bath-Hextall FJ, Leonardi-Bee J, Murrell DF, Tang ML. Probiotics for treating eczema. Cochrane Database Syst Rev. 2008:CD006135.
- 84. van der Aa LB, Heymans HS, van Aalderen WM, Sillevis Smitt JH, Knol J, Ben Amor K, et al; Synbad Study Group. Effect of a new synbiotic mixture on atopic dermatitis in infants: a randomized-controlled trial. Clin Exp Allergy. 2010;40:795-804.
- Rolfe VE, Fortun PJ, Hawkey CJ, Bath-Hextall F. Probiotics for maintenance of remission in Crohn's disease. Cochrane Database Syst Rev. 2006:CD004826.
- 86. Savino F, Pelle E, Palumeri E, Oggero R, Miniero R. Lactobacillus reuteri (American Type Culture Collection Strain 55730) versus simethicone in the treatment of infantile colic: a prospective randomized study. Pediatrics. 2007;119:e124-30.
- 87. Savino F, Cordisco L, Tarasco V, Palumeri E, Calabrese R, Oggero R, et al. Lactobacillus reuteri DSM 17938 in infantile colic: a randomized, double-blind, placebo-controlled trial. Pediatrics. 2010;126:e526-33.
- Elmer GW, McFarland LV, Surawicz CM, Danko L, Greenberg RN. Behaviour of Saccharomyces boulardii in recurrent Clostridium difficile disease patients. Aliment Pharmacol Ther. 1999;13:1663-8.
- Shornikova AV, Casas IA, Mykkänen H, Salo E, Vesikari
 Bacteriotherapy with Lactobacillus reuteri in rotavirus gastroenteritis. Pediatr Infect Dis J. 1997;16:1103-7.
- Lewis SJ, Potts LF, Barry RE. The lack of therapeutic effect of Saccharomyces boulardii in the prevention of antibiotic-related diarrhoea in elderly patients. J Infect. 1998;36:171-4.
- 91. Borchers AT, Selmi C, Meyers FJ, Keen CL, Gershwin ME. Probiotics and immunity. J Gastroenterol. 2009;44:26-46.
- 92. Gill HS, Rutherfurd KJ, Cross ML, Gopal PK. Enhancement of immunity in the elderly by dietary supplementation with the probiotic Bifidobacterium lactis HN019. Am J Clin Nutr. 2001;74:833-9.

- Smits HH, Engering A, van der Kleij D, de Jong EC, Schipper K, van Capel TM, et al. Selective probiotic bacteria induce IL-10-producing regulatory T cells in vitro by modulating dendritic cell function through dendritic cell-specific intercellular adhesion molecule 3-grabbing nonintegrin. J Allergy Clin Immunol. 2005;115:1260-7.
- Zhang L, Li N, Caicedo R, Neu J. Alive and dead Lactobacillus rhamnosus GG decrease tumor necrosis factor-alphainduced interleukin-8 production in Caco-2 cells. J Nutr. 2005;135:1752-6.
- Villarruel G, Rubio DM, Lopez F, Cintioni J, Gurevech R, Romero G, et al. Saccharomyces boulardii in acute childhood diarrhoea: a randomized, placebo-controlled study. Acta Paediatr. 2007:96:538-41.
- Rosenfeldt V, Michaelsen KF, Jakobsen M, Larsen CN, Møller PL, Tvede M, et al. Effect of probiotic Lactobacillus strains on acute diarrhea in a cohort of nonhospitalized children attending daycare centers. Pediatr Infect Dis J. 2002;21:417-9.
- Rosenfeldt V, Michaelsen KF, Jakobsen M, Larsen CN, Møller PL, Pedersen P, et al. Effect of probiotic Lactobacillus strains in young children hospitalized with acute diarrhea. Pediatr Infect Dis J. 2002;21:411-6.
- Yuan Q, Walker WA. Innate immunity of the gut: mucosal defense in health and disease. J Pediatr Gastroenterol Nutr. 2004;38:463-73.
- Martín R, Heilig GH, Zoetendal EG, Smidt H, Rodríguez JM. Diversity of the Lactobacillus group in breast milk and vagina of healthy women and potential role in the colonization of the infant gut. J Appl Microbiol. 2007;103:2638-44.
- 100. Perez PF, Doré J, Leclerc M, Levenez F, Benyacoub J, Serrant P, et al. Bacterial imprinting of the neonatal immune system: lessons from maternal cells? Pediatrics. 2007;119:e724-32.
- Kurugöl Z, Koturoğlu G. Effects of Saccharomyces boulardii in children with acute diarrhoea. Acta Paediatr. 2005;94:44-7.
- Fedorak RN, Madsen KL. Probiotics and prebiotics in gastrointestinal disorders. Curr Opin Gastroenterol. 2004;20:146-55.
- Venugopalan V, Shriner KA, Wong-Beringer A. Regulatory oversight and safety of probiotic use. Emerg Infect Dis. 2010;16:1661-5.
- 104. Vandenplas Y, Salvatore S, Vieira M, Devreker T, Hauser B. Probiotics in infectious diarrhoea in children: are they indicated? Eur J Pediatr. 2007;166:1211-8.
- 105. Oudhuis GJ, Bergmans DC, Dormans T, Zwaveling JH, Kessels A, Prins MH, et al. Probiotics versus antibiotic decontamination of the digestive tract: infection and mortality. Intensive Care Med. 2011;37:110-7.
- 106. Besselink MG, van Santvoort HC, Buskens E, Boermeester MA, van Goor H, Timmerman HM, et al; Dutch Acute Pancreatitis Study Group. Probiotic prophylaxis in predicted severe acute pancreatitis: a randomised, double-blind, placebo-controlled trial. Lancet. 2008;371:651-9.
- 107. Moran JP, Walter J, Tannock GW, Tonkonogy SL, Sartor RB. Bifidobacterium animalis causes extensive duodenitis and mild colonic inflammation in monoassociated interleukin-10-deficient mice. Inflamm Bowel Dis. 2009;15:1022-31.
- 108. Taylor AL, Dunstan JA, Prescott SL. Probiotic supplementation for the first 6 months of life fail to reduce the risk of atopic dermatitis and increases the risk of allergen sensitization in high-risk children: a randomized controlled trial. J Allergy Clin Immunol. 2007;119:184-91.

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