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Levels of metabolizable energy and digestible lysine for broiler chicks 1-10 days of age

Níveis de energia metabolizável e Lysina digestível para pintos de corte de 1 a 10 dias de idade

Ricardo Vianna Nunes^{1*}; Thais Lorana Savoldi²; Cláudio Yuji Tsutsumi³; Cleison de Souza⁴; Jomara Broch⁴; Sharon Karla Luders Meza⁵; Liliane Borsatti⁶; José Luiz Schneiders⁵

Abstract

This study evaluated the effect of different levels of metabolizable energy (ME) and digestible lysine (DL) on the rate of protein deposition, rate deposition body fat and intestinal morphology of broiler chicks 1 to 10 days of age. The study design consisted of 1,152 broilers, weighing 52 ± 5 g, distributed in a completely randomized, 4x4 factorial design, consisting of four levels of ME (2700; 2825; 2950 and 3075 kcal/kg) and four levels of DL (1,080; 1,187; 1,295 and 1,403%), with 16 treatments and three replicates, with 24 poultry in each experimental unit. The protein deposition rate (PDR) and fat deposition rate (FDR) were not affected ($P > 0.05$) by the ME, however, PDR was influenced linearly ($P < 0.05$) by increasing lysine. There was an interaction ($P < 0.05$) between the levels of ME and DL, whose increase resulted in a higher villus height of the duodenum, jejunum and a greater relation villus: crypt ratio of the ileum. Therefore, levels of DL and ME had positive effects on the intestinal development of broiler chicks in the pre-starter phase.

Key words: Amino acid. Poultry farming. Energy density. Morphology.

Resumo

O experimento foi realizado com o objetivo de avaliar o efeito dos níveis de energia metabolizável (EM) e de lisina digestível (LD), sobre a taxa de deposição de proteína, taxa de deposição gordura corporal e morfometria intestinal de pintos de corte de 1 a 10 dias de idade. Foram utilizados 1.152 pintos de corte, com peso médio de 52 ± 5 g, distribuídos em delineamento inteiramente casualizado, em esquema fatorial 4x4, constituídos quatro níveis de EM (2700; 2825; 2950; e 3075 kcal.kg⁻¹) e quatro níveis de LD (1,080; 1,187; 1,295 e 1,403%), com 16 tratamentos, três repetições e com 24 aves ME cada unidade experimental. A taxa de deposição de proteína (TDP) e de gordura (PDR) não foi influenciada ($P > 0,05$) pelos níveis de EM, no entanto, a PDR foi influenciada de forma linear ($P < 0,05$) pelos níveis crescentes de lisina. Houve interação ($P < 0,05$) entre os níveis de EM e DL, cujo aumento nesses níveis

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proporcionaram maior altura das vilosidades do duodeno, jejuno e na relação vilo:cripta do íleo. Os níveis de lisina digestível e energia metabolizável apresentam efeito positivo sobre o desenvolvimento intestinal dos pintos de corte na fase pré-inicial.

Palavras-chave: Aminoácido. Avicultura. Densidade energética. Morfometria.

Introduction

Advances in genetics research have enabled an improvement in feed conversion rates and carcass yield of broilers, which are determining factors from a nutritional point of view, considering that the phenotypic expression of genetic potential is influenced by the environment and nutrition. Thus, knowledge of the nutritional requirements for growth and development is important to obtain the maximum performance from the poultry.

Newly hatched poultry in the pre-initial phase undergo changes in their nutrition, as their nutritional requirements were previously obtained from the yolk sac, which is rich in proteins and lipids. Once hatched, they obtain their nutrition from supplied feed based on protein and complex carbohydrates (SAKOMURA et al., 2004).

According to Gonzales and Sartori (2002), the growth of muscle tissue is closely related to nutritional factors including the level of dietary amino acids, energy, vitamins and minerals, since the excess or deficiency of these nutrients can influence protein synthesis or degradation.

The correct ratio of amino acids that comprises the starter diets is currently formulated based on the ideal protein concept, which is to provide the proper amount of essential amino acids, without deficiency or excess. The amino acid lysine is used as a reference in the early stages, and is preferably intended for muscle deposition although some body parts respond better to increases in amino acid intake. High performance broilers require greater concentrations of lysine in the diet due to higher daily protein synthesis (TAKEARA et al., 2010).

Metabolizable energy (ME) is the term used to express the energy available and is commonly used in calculations of the nutritional value of poultry feed (NUNES et al., 2008). This is considered a strategic nutritional factor in production systems that use fed ad libitum, because food consumption is mainly regulated by the caloric density of rations, which may have implications on the productive and economic efficiency of poultry activity. The requirements for crude protein, amino acids and other nutrients are usually expressed in terms of levels of ME of feed (SILVA et al., 2003).

This study was conducted in order to establish the best level of ME and digestible lysine (DL) for broiler chickens in the pre-initial phase of 1 to 10 d of age, for the rate of fat deposition, rate of body protein deposition and morphometric features of the small intestine.

Materials and Methods

The experiment was conducted at the Experimental Station of poultry farming at Universidade Estadual do Oeste do Paraná-UNIOESTE, and was approved by the ethics committee and the institution's biosafety protocol, Protocol 04511. The poultry were housed in boxes (experimental unit - UE), which had a tubular feeder, nipple-type water dispenser, hood heating (250 watt infrared lamps) and concrete floor, which was lined with pine shavings.

One thousand one hundred and fifty-two male broiler chicks, one day old, from the commercial line Cobb 500 and with an average weight of 52 ± 5 g were used. The treatments were distributed in a completely randomized design in a 4x4 factorial

design, consisting of four levels of Metabolizable energy (ME) (2700; 2825; 2950 and 3075 kcal/kg) and four levels of digestible lysine (DL) (1,080; 1,187; 1,295 and 1,403%) totaling 16 treatments with three repetitions of 24 poultry each. After the distribution of poultry, an additional group of five broilers were fasted for six hours and then killed by cervical dislocation and frozen at -200 °C for later processing.

The experimental diets were formulated based on corn and soybean meal (Table 1), according to the composition of the food and nutritional requirements proposed by Rostagno et al. (2011), maintaining the relation ideal protein for methionine + cysteine (71%), tryptophan (16%), threonine (65%), arginine (105%), isoleucine (65%) and valine (75%).

At 10 days old, two animals per EU, with a range of $\pm 10\%$ by weight processing, were fasted for six hours and weighed and killed by cervical dislocation for evaluation of body composition.

Carcasses of the additional group and the poultry slaughtered at 10 days were individually crushed and homogenized, and a sample of each bird was sent to an animal nutrition laboratory for pre-drying in a forced air oven at 550 °C for 72 hours, with subsequent pre-degreasing. The degreased carcasses were again crushed, and then submitted for analysis of dry matter, ether extract and crude protein, according to the methodology described by Silva and Queiroz (2004).

For determination of protein deposition rates (PDR) and fat deposition rates (FDR) in the carcass (g/day) the methodology adapted by Fraga et al. (2008) was used. A PDR was measured by comparing the chicks slaughtered at the end of the trial period with the additional group of five chicks slaughtered in the housing.

On the tenth day, two more poultry with the UE average weight ($\pm 10\%$) were slaughtered for the histomorphometric evaluation of the small intestine. The segments of the small intestine (duodenum, jejunum and ileum) were opened on the mesenteric border, and fragments of approximately 2.0 cm in length were carefully collected, washed in distilled water, extended the tunica serosa and fixed in buffered 10% formalin solution for 24 hours. After this period, they were subjected to several washes with 70% ethanol and kept in this solution to make the slides.

Subsequently, the samples were dehydrated in increasing alcohol solutions, cleared in xylene, embedded in paraffin wax and cut with a rotary microtome to 7 μm . Three blades were prepared per animal and each blade inserted semiserial four cuts. The sections were stained with hematoxylin-eosin.

The morphometric analysis of histologic sections was performed in the Motic Advanced 2.0 image analyzer, Animal Reproduction Sector CAU-MEU. Straight lengths were selected and measured in accordance with the adopted unit (μm), 30 villi and 30 crypts and oriented. The villus height measurements were taken from the upper base of the crypt to the apex of the villus, and crypts were measured between the lower base of the villi to the upper base of the crypt.

Statistical analyses were performed with assistance of the Statistical Analysis System version 9 (SAS, 2002). Statistical analysis of the parameters was performed by PROC GLM ($P \leq 0.05$). The parameters in which significant differences were found ($P \leq 0.05$) were estimated by polynomial regression using the PROC REG. For interactions of ME x DL, significant ($P \leq 0.05$) response surface regressions were estimated using the PROC RSREG procedure as adjustment data.

Table 1. Composition of experimental diets for broiler chicks in the pre-initial phase (1-10 days of age).

ME	2700				2825				2950				3075			
Lys Dig %	1.080	1.187	1.295	1.403	1.080	1.187	1.295	1.403	1.080	1.187	1.295	1.403	1.080	1.187	1.295	1.403
Ingredients %																
Corn	52.87	53.59	54.30	54.91	57.16	57.87	58.59	58.87	59.23	59.94	60.47	58.99	60.61	59.13	57.65	56.17
Soybean meal	24.37	27.33	30.30	33.26	23.91	26.87	29.83	32.92	24.52	27.48	30.46	33.65	24.37	27.56	30.76	33.95
Glutamic acid	7.976	5.500	2.600	0.000	7.744	5.064	2.374	0.000	8.897	6.217	3.537	0.967	8.823	6.261	3.689	1.119
Inert	6.600	5.296	4.400	3.300	3.030	1.930	0.830	0.000	2.107	1.004	0.000	0.000	0.000	0.000	0.000	0.000
Wheat bran	3.000	3.000	3.000	3.000	3.000	3.000	3.000	2.713	0.100	0.100	0.100	0.100	0.100	0.100	0.100	0.100
Dicalcium phospho	2.032	2.00	1.968	1.937	2.018	1.986	1.954	1.927	2.048	2.017	1.985	1.951	2.032	1.998	1.964	1.931
Limestone	0.953	0.954	0.956	0.957	0.961	0.963	0.964	0.963	0.946	0.948	0.949	0.952	0.957	0.959	0.962	0.965
L-Lysine HCl	0.427	0.466	0.505	0.545	0.431	0.469	0.509	0.547	0.423	0.461	0.501	0.539	0.424	0.461	0.499	0.537
Premix APP 1	0.400	0.400	0.400	0.400	0.400	0.400	0.400	0.400	0.400	0.400	0.400	0.400	0.400	0.400	0.400	0.400
Salt	0.400	0.400	0.400	0.400	0.400	0.400	0.400	0.400	0.400	0.400	0.400	0.400	0.400	0.400	0.400	0.400
DL-Methionine	0.317	0.358	0.401	0.443	0.307	0.349	0.392	0.434	0.305	0.347	0.390	0.436	0.302	0.348	0.396	0.442
L-Threonine	0.169	0.191	0.213	0.235	0.165	0.187	0.209	0.231	0.160	0.182	0.204	0.229	0.159	0.183	0.207	0.232
L-Valine	0.140	0.161	0.182	0.204	0.134	0.155	0.175	0.199	0.130	0.151	0.172	0.198	0.128	0.152	0.176	0.202
L-Arginine	0.134	0.148	0.165	0.181	0.133	0.148	0.164	0.181	0.100	0.150	0.167	0.183	0.135	0.150	0.168	0.184
Soy oil	0.100	0.100	0.100	0.100	0.100	0.100	0.100	0.100	0.100	0.100	0.164	0.890	1.066	1.794	2.520	3.246
Choline chloride	0.060	0.060	0.060	0.060	0.060	0.060	0.060	0.060	0.060	0.060	0.060	0.060	0.060	0.060	0.060	0.060
Isoleucine	0.020	0.026	0.032	0.038	0.017	0.023	0.029	0.035	0.015	0.021	0.027	0.034	0.014	0.021	0.028	0.036
BHT	0.020	0.020	0.020	0.020	0.020	0.020	0.020	0.020	0.020	0.020	0.020	0.020	0.020	0.020	0.020	0.020
Calculated																
ME kcal/kg	2700	2700	2700	2700	2825	2825	2825	2825	2950	2950	2950	2950	3075	3075	3075	3075
Crude protein g/kg	215	215	215	215	215	215	215	215	221.1	221.1	221.1	221.1	221.1	221.1	221.1	221.1
Lysine Dig g/kg	10.80	11.87	12.95	14.03	10.80	11.87	12.95	14.03	10.80	11.87	12.95	14.03	10.80	11.87	12.95	14.03
Met+Cyst Dig g/kg	7.67	8.43	9.20	9.96	7.67	8.43	9.20	9.96	7.67	8.43	9.20	9.96	7.67	8.43	9.20	9.96
Met Dig g/kg	5.41	6.00	6.60	7.19	5.36	5.95	6.55	7.15	5.36	5.95	6.56	7.17	5.35	5.96	6.58	7.20
Threonine Dig g/kg	7.02	7.72	8.42	9.12	7.02	7.72	8.42	9.12	7.02	7.72	8.42	9.12	7.02	7.72	8.42	9.12
Arginine Dig g/kg	11.34	12.46	13.60	14.73	11.34	12.46	13.60	14.73	11.34	12.46	13.60	14.73	11.34	12.46	13.60	14.73
Isoleucine Dig g/kg	7.02	7.72	8.42	9.12	7.02	7.72	8.42	9.12	7.02	7.72	8.42	9.12	7.02	7.72	8.42	9.12
Valine Dig g/kg	8.10	8.90	9.70	10.52	8.10	8.90	9.70	10.52	8.10	8.90	9.70	10.52	8.10	8.90	9.70	10.52
Potassium	0.620	0.576	0.732	0.788	0.623	0.679	0.735	0.790	0.615	0.672	0.728	0.782	0.617	0.670	0.725	0.779
Sodium	0.177	0.177	0.177	0.177	0.177	0.177	0.177	0.177	0.177	0.177	0.177	0.177	0.176	0.176	0.176	0.176
Calcium	0.932	0.932	0.932	0.932	0.932	0.932	0.932	0.932	0.932	0.932	0.932	0.932	0.932	0.932	0.932	0.932
Phosphorus Disp.	0.468	0.468	0.468	0.468	0.468	0.468	0.468	0.468	0.468	0.468	0.468	0.468	0.468	0.467	0.467	0.467

¹2,500 Vit A IU/kg; Vit D₃ 625 KIU/kg; Vit E 6,250 IU/kg; Vit K₃ 500 mg/kg; Vit B₁ 625 mg/kg; Vit B₂ 1,625 mg/kg; Vit B₆ 875 mg/kg; Vit B₁₂ 4,500 µg/kg; Ac. Pant 3,750 mg/kg; Niacin 10,500 mg/kg; Ac Fol 300 mg/kg; Biotin 20,000 µg/kg; Choline 83,531.250 mg/kg; Mn 18,738.600 PPM; Zn 27,499.998 PPM; Zn.Org 10,000PPM; Fe 11,250.001 PPM; Cu 1996.371; I 187.50 PPM; Se 100.00 PPM; Org 25,000 PPM; Narazina 12,500 mg/kg; Nicarbazin 12,500 mg/kg; BHT 37,500 mg/kg; Enramycin 2,500 mg/kg.

Results and Discussion

There was no interaction ($P>0.05$) between the levels of metabolizable energy (ME) and

digestible lysine (DL) on the protein deposition rates (PDR) and fat deposition rates (FDR) in the carcasses of broiler chicks the pre-starter period (Table 2).

Table 2. Protein deposition rate and fat deposition rate in 10-day-old broilers.

	PDR (g/day)	FDR (g/day)
ME		
2700	3.114	1.891
2825	2.93	1.721
2950	3.09	2.014
3075	3.076	2.051
Regression	ns	ns
LYS		
1.080%	3.199	1.73
1.187%	2.985	1.818
1.295%	3.051	2.039
1.403%	2.993	2.114
Regression	ns	L*
CV(%)	24.08	26.69
Average	3.057	1.946
Polynomial regression equation		
$FDR=0.33945+1.27416*LYS; R^2= 0.94$		

CV = coefficient of variation, ns = not significant; L* = linear effect at 0.05 probability.

The levels of ME and DL did not influence ($P>0.05$) the PDR. These results were different from those found by Leandro et al. (2003), who evaluated different nutritional plans (energy and protein) and observed a linear increase in the PDR. The increase in protein deposition was related to the gradual improvement in the energy / protein ratio of feed, where the lowest levels were insufficient to ensure proper deposition rate.

DL levels had a linear influence ($P<0.05$) on FDR ($FDR = 0.33945 + 1.27416*LYS; R^2 = 0.94$), but no differences were observed ($P>0.05$) in the levels of ME. These data differ from those found by Cella et al. (2009), who did not find a significant effect of different levels of DL in

broiler chicks ranging from 1 to 21 days of age. However, Toledo et al. (2007) also did not find a significant effect of different levels of DL in the diet on the PDR and FDR of broiler chicks in the pre-initial phase.

In a study conducted by Nunes et al. (2015), the authors observed that the PDR and FDR were influenced by an increase in the levels of ME and DL in the feed, with the increase of ME being the main factor responsible for increasing the FDR. According to Silva et al. (2001), the accumulation of fat in the carcass is the result of energy intake beyond the needs for maintenance and muscle growth of poultry, as well as the increase of the ME: crude protein ratio.

According to Vasconcellos et al. (2012), by reducing the concentration of protein and / or dietary amino acids, there is an increase in carcass fat content; therefore, it was understood that diets with high levels of protein and amino acids provide a lower body fat ratio, although the same was not observed in this experiment.

Different levels of ME and DL interacted to affect the duodenum villus height ($P < 0.05$; Table 3), with the best results obtained with reduced levels of ME and increased levels of DL ($DVH = -8315.6429 + 4.4169*ME + 4678.6016*LYS - 0.0003*ME^2 - 2.2669*ME*LYS + 864.125*LYS^2$; $R^2 = 0.30$) (Figure 1).

Table 3. Means of intestinal morphometry of broiler chickens from 1 to 10 days of age treated with different levels of metabolizable energy and digestible lysine.

	Duodenum			Jejunum			Ileum		
	Villus (µm)	Crypt (µm)	Villus:Crypt	Villus (µm)	Crypt (µm)	Villus:crypt (µm)	Villus (µm)	Crypt (µm)	Villus: crypt
ME									
2700	1188.06	223.30	5.35	754.56	199.14	3.83	596.84	170.35	3.60
2825	1203.90	261.78	4.66	769.64	205.58	3.79	623.31	196.39	3.19
2950	1204.83	247.42	4.97	794.71	214.08	3.75	631.87	197.15	3.22
3075	1203.74	223.16	5.40	850.28	231.42	3.71	677.60	201.71	3.37
Regression ME	ns	ns	ns	ns	ns	ns	L*	Q*	ns
Lysina									
1.08	1162.06	230.29	5.10	718.82	210.29	3.47	588.09	185.11	3.21
1.187	1184.49	232.85	5.10	753.74	212.36	3.60	607.80	187.36	3.30
1.295	1195.67	238.62	5.18	794.80	213.36	3.75	652.68	194.15	3.42
1.403	1258.32	253.89	5.01	901.83	214.21	4.25	681.05	198.97	3.46
Regression DL	ns	ns	ns	ns	ns	L**	L**	ns	ns
ME x DL	RS	RS	RS	RS	RS	ns	ns	ns	ns
CV	6.02	10.13	9.90	6.69	11.54	12.04	10.48	12.48	3.34
Average	1200.13	238.91	5.09	792.29	212.55	3.76	191.39	191.39	14.45
Polynomial regression equations									
HIV= 52.946+0.2007*ME; $R^2= 0.89$									
DIC= -2886.3+2.0606*ME-0.0003*ME ² ; $R^2= 0.93$									
RVCJ= 0.90467+2.3069*LYS; $R^2= 0.88$									
DVH= 259.18453+300.68148*LYS; $R^2=0.98$									
Response surface equations									
DVH= -8315.6429+4.4169*ME+4678.6016*LYS-0.0003*ME ² -2.2669*ME*LYS+864.125*LYS ² ; $R^2= 0.30$									
DCD= -8422.5127+6.0187*ME-63.3877*LYS-0.001*ME ² -0.1882*ME*LYS+273.0148*LYS ² ; $R^2= 0.31$									
RVCD= 5.6713-0.4519*ME+1171.798*LYS+0.0002*ME ² -0.3789*ME*LYS-26.5946*LYS ² ; $R^2= 0.23$									
VHJ= 263.0311-0.9277*ME+2673.6663*LYS+0.0006*ME ² -2.0659*ME*LYS+1546.4099*LYS ² ; $R^2= 0.71$									
CDJ= 5.6713-0.4519*ME+1171.798*LYS+0.0002*ME ² -0.3789*ME*LYS-26.5946*LYS ² ; $R^2= 0.23$									

CV = coefficient of variation; RS = response surface; L = linear effect; Q = quadratic effect; s * and ** = not significant, meaningful $P < 0.01$ and $P < 0.05$, respectively; HIV = height of ileal villus; DIC = depth of ileal crypt; RVCJ = villus: crypt of jejunum; DVH= villus height of duodenum; DCD = depth of duodenal crypt; RVCD = villus: crypt of duodenum; VHJ= height of jejunum villus; CDJ = depth of jejunum crypt.

Similarly, an interaction was observed ($P < 0.05$) between the levels of ME and DL regarding the jejunum villus height ($VHJ = 263.0311 - 0.9277*ME + 2673.6663*LYS + 0.0006*ME^2 - 2.0659*ME*LYS + 1546.4099*LYS^2$; $R^2 = 0.71$), with an increase in the size of villi observed with a reduction in the level of DL and an increased level of ME (Figure 2). The same behavior was observed for the ileal villus height ($P < 0.05$).

Figure 1. Duodenal villus height depending on the levels of metabolizable energy and digestible lysine.
 $DVH = -8315.6429 + 4.4169*ME + 4678.6016*LYS - 0.0003*ME^2 - 2.2669*ME*LYS + 864.125*LYS^2$, ($R^2 = 0.30$)

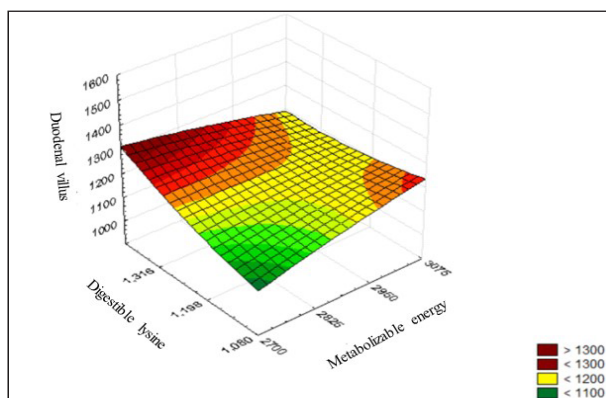
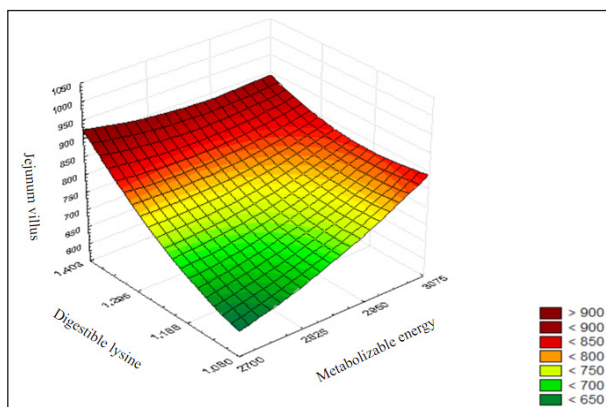


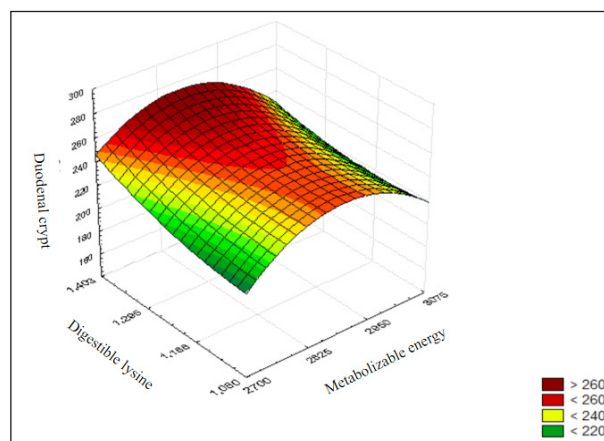
Figure 2. Villus height of the jejunum based on the levels of metabolizable energy and digestible lysine.
 $VHJ = 263.0311 - 0.9277*ME + 2673.6663*LYS + 0.0006*ME^2 - 2.0659*ME*LYS + 1546.4099*LYS^2$, ($R^2 = 0.71$)



In the duodenum, an interaction was observed ($P < 0.05$) between the levels of ME and DL regarding

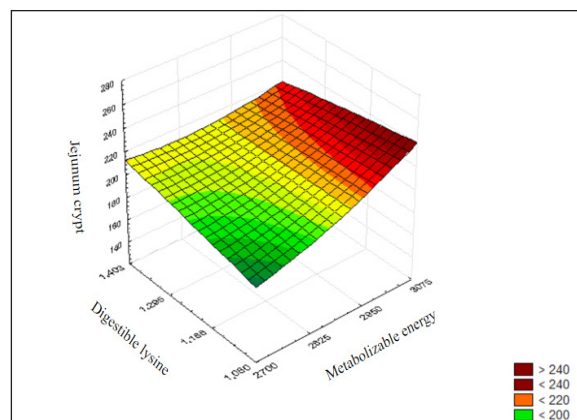
crypt depth ($DCD = -8,422.5127 + 6.0187*ME - 63.3877*LYS - 0.001*ME^2 - 0.1882*ME*LYS + 273.0148*LYS^2$; $R^2 = 0.31$) (Figure 3).

Figure 3. Duodenal crypt depth in terms of levels of metabolizable energy and digestible lysine.
 $DCD = -8422.5127 + 6.0187*ME - 63.3877*LYS - 0.001*ME^2 - 0.1882*ME*LYS + 273.0148*LYS^2$, ($R^2 = 0.31$)



The jejunum crypt depth also showed an interaction ($P < 0.05$) according to the levels of ME and DL ($CDJ = 5.6713 - 0.4519*ME + 1171.798*LYS + 0.0002*ME^2 - 0.3789*ME*LYS - 26.5946*LYS^2$; $R^2 = 0.23$), where the depths were observed with an increase in ME and a reduction in DL (Figure 4).

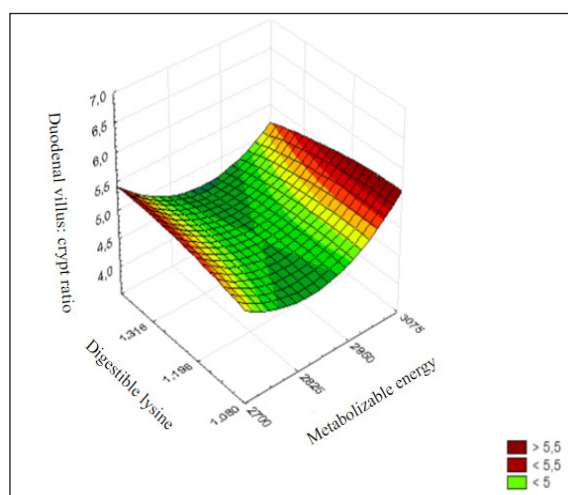
Figure 4. Crypt depth of the jejunum as a function of the levels of metabolizable energy and digestible lysine.
 $CDJ = 5.6713 - 0.4519*ME + 1171.798*LYS + 0.0002*ME^2 - 0.3789*ME*LYS - 26.5946*LYS^2$, ($R^2 = 0.23$)



Regarding the duodenal villus height to crypt depth ratio, an interaction ($P < 0.05$) was obtained between the levels of ME and DL ($RVDC = 5.6713 - 0.4519 \cdot ME + 1171.798 \cdot LYS + 0.0002 \cdot ME^2 - 0.3789 \cdot ME \cdot LYS - 26.5946 \cdot LYS^2$; $R^2 = 0.23$), which showed a better relationship with increasing levels of ME and DL (Figure 5).

Figure 5. The duodenal villus: crypt ratio based on metabolizable energy and digestible lysine.

$RVDC = 5.6713 - 0.4519 \cdot ME + 1171.798 \cdot LYS + 0.0002 \cdot ME^2 - 0.3789 \cdot ME \cdot LYS - 26.5946 \cdot LYS^2$, ($R^2 = 0.23$)



In the jejunum, the villus: crypt ratio showed a linear increase ($P < 0.05$) with increasing levels of DL ($RVCJ = 0.90467 + 2.3069 \cdot LYS$, $R^2 = 0.88$). There were no differences ($P > 0.05$) for the villus: crypt ratio in the ileum.

After poultry hatch, the morphologic and functional development of organs occurs, with a consequent increase in the area of absorption and secretion of enzymes, suggesting that the morphometric increase in digestive organs and their enzymatic activity are coordinated by the efficient supply of nutrients.

According Maiorka et al. (2003), the number of villi per area was higher in the duodenum 48 hours, and in the duodenum and jejunum 72 hours post hatch, in poultry that suffering food and water

restriction in poultry, compared to poultry that received feed and water ad libitum. The authors concluded that this increase was related to the reduction in gut size of these poultry.

The intestinal mucosa is responsible for the absorption of food, which is held in the villi through the enterocytes. The number of villi is associated with the enterocytes of the villi, and their size increase reflects a greater absorption of food (MACARI; MAIORKA, 2000). Freitas et al. (2008) reported that most villus height is related to performance results, and that poultry with larger villi have higher weight gains and better feed conversion. Thus, the larger the size of the villi, the greater the ability of digestion and absorption of nutrients, due to the larger contact area and the level of enzyme effectiveness between the mucosa and intestinal lumen.

Pelicano et al. (2003) studied the intestinal morphometry of poultry on diets with different probiotics, and found that the highest value of crypt depth was related to an increased cell proliferative activity, the purpose of which is to ensure an adequate rate of epithelial renewal, offsetting losses in the heights of the villi. These results partially corroborate with an experiment conducted by Duarte et al. (2012) who evaluated different energy levels (3200 and 3600 kcal/kg) and feeding programs, and noted that the highest level of energy led to a higher villus height and improved performance of the poultry.

Conclusions

Diets with 3075 kcal/kg and 1.403 DL% provided better intestinal morphometric characteristics of broiler chickens 1 to 10 days of age.

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