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Stomatal density, leaf area and plant size variation of *Rhizophora mangle* (Malpighiales: Rhizophoraceae) along a salinity gradient in the Mexican Caribbean

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Abstract: In community ecology, the knowledge of abiotic factors, that determine intraspecific variability in ecophysiological and functional traits, is important for addressing major questions, such as plant community assembly and ecosystem functioning. Mangroves have several mechanisms of resistance to salinity and most species exhibit some xeromorphic features in order to conserve water. Leaf area and stomatal density play an important role in maintaining water balance, and gas exchange is regulated by their aperture and density, two traits that vary intraspecifically in response to environmental conditions, such as water stress and salinity. In this study, we evaluated the effects of salinity on stomatal density, leaf area and plant size in *R. mangle* and we tested for associations among the three variables, across three sites along a natural salinity gradient in the Xel-Há Park, Quintana Roo, Mexico. We hypothesized that high salinity sites would produce smaller plants, with smaller leaves, and fewer stomata. Three sampling sites with different environmental conditions were chosen and salinities were monitored monthly. A total of 542 plants were tagged and tree heights and diameters were measured for each individual within each of the three sampling sites. Three leaves from 20 trees from each site were measured to determine leaf area. Stomatal densities were determined in each leaf using nail polish casts, examining ten 1 mm squares per leaf under an optical microscope. A principal component analysis was used to assess association between tree height, leaf area, and stomatal density for each plot. The salinity gradient was reflected in plant size, producing smaller plants at the higher salinity site. The largest leaves were found at the low salinity site ($51.2 \pm 24.99 \text{ cm}^2$). Leaf length was not correlated to plant size (LL vs. tree height: $r = 0.02$, $P = 0.8205$; LL vs. trunk diameter: $r = 0.03$, $P = 0.7336$), so we concluded that leaf length is an environmentally plastic trait of red mangroves that may vary as a function of environmental conditions, such as hydric stress caused by elevated salinity. The larger leaves from the low salinity site had lower densities of stomata (65.0 stomata. mm^2 SD= 12.3), and increasing salinities did not decrease stomatal density (intermediate salinity site: 73.4 stomata. mm^2 SD= 13.5; high salinity site: 74.8 stomata. mm^2 SD= 17.3). Our results confirm that stomatal density is inversely related to leaf area ($r = -0.29$, $P < 0.001$), especially leaf width ($r = -0.31$, $P < 0.001$), and that salinity may increase stomatal density by causing reduction of leaf size. Rev. Biol. Trop. 65 (2): 701-712. Epub 2017 June 01.

Key words: leaf plasticity, leaf area, mangroves, tree height, stomatal densities, water stress.

Mangroves are an ecological group of tropical plants which inhabit coastal environments (Tomlinson, 1986). They are normally found in flooded areas or close to water, where they can experience variable mixtures of fresh

and seawater, and often experience salt concentrations higher than seawater in the substrate, which forces mangroves to absorb water against an osmotic pressure gradient (Connor, 1969; Downton, 1982; Ball & Pidsley, 1995;

Aziz & Khan, 2001; Saenger, 2002). All mangroves show some traits of salt resistance that require metabolic energy, which may include salt exclusion, salt extrusion, salt storage, succulence, compartmentalization and osmo-compensation (Saenger, 2002). Many mangroves have also developed some xeromorphic features in order to conserve water, including a thick-walled multi-layered epidermis, thick waxy lamellar cuticle, and hairs or scales on the abaxial surface which cover salt glands and stomata (Ball, 1988; Das & Ghose, 1993; Saenger, 2002).

Four dominant species of mangroves can be found in Mexican coasts: *Rhizophora mangle* L., *Laguncularia racemosa* (L.) C.F. Gaertn., *Avicennia germinans* L. and *Conocarpus erectus* L. (López-Portillo & Ezcurra, 2002; CONABIO, 2009). The red mangrove (*R. mangle*) is an important species in tropical North and South America and West Africa, and is one of the most dominant species within its range (Mehlig, 2006), nevertheless, it is threatened by habitat conversion and pollution, which has resulted in declines throughout the wider Caribbean region (Ellison, Farnsworth, & Moore 2015). The extended distribution and dominance of this species may be partially attributed to its extreme tolerance to a range of environmental conditions, resisting waterlogged soils and high salinities up to 90 ppt (Hill, 2001). Salt uptake of *R. mangle* is mainly regulated by the root system, which excludes most salts during water uptake, while the other mangrove species (e. g. *Avicennia germinans* and *Laguncularia racemosa*) take up seawater through their roots but excrete excess salt through pores or salt glands on the leaf surface (Duke, 1983). Growth of *R. mangle* appears to peak at moderate salinities (10 PSU). At higher salinities (32 PSU) leaf area and photosynthetic potential decrease (Krauss & Allen 2003), indicating that the leaves of *R. mangle* play a major role in osmoregulation. Larger leaves have thicker boundary layers of still air lowering convective heat loss, enabling a high variation between ambient and leaf temperature. This may be a disadvantage as respiration increases

more than photosynthesis with temperature and the effect is more important for leaves under stronger radiation and where water is in short supply (Westoby, Falster, Moles, Vesk, & Wright, 2002). Leaves of the red mangrove are oval shaped or elliptic, 6-10 cm long, hairless, and tend to be relatively uniform (Tomlinson, 1986). However, leaf area may be variable (Araujo, Jaramillo, & Snedaker, 1997) and reduced under suboptimal conditions and so can be an indicator of chronic environmental stress (Canoy, 1975; Snedaker & Brown, 1981). Known stressors include high salinity, latitudinal location, exposure to pollutants (Lugo & Snedaker, 1974; Lugo, Cintrón, & Goenaga, 1981) and height induced water stress (Zhang, Zheng, & Tyree, 2012), so therefore, leaf area can be considered an indicator of chronic environmental stress (Araujo et al., 1997).

Gas exchange is mediated on one hand by regulating stomata aperture, and on the other, by controlling the number of stomata (Hetherington & Woodward 2003). Several environmental conditions can affect the morphogenesis of stomata, such as water availability (Salisbury, 1928; Xu & Zhou, 2008), temperature (Ciha & Brun, 1975; Huner, Palta, Li, & Carter, 1981), light exposure (Gay & Hurd, 1975), and environmental CO₂ concentrations (e.g. Salisbury, 1928; Bristow & Looi, 1968; Woodward & Bazzaz, 1988; Beerling & Kelly, 1997). Water stress has been associated with a reduction of stomatal density in leaves (Salisbury, 1928; Pääkkönen, Vahala, Pohjolal, Holopainen, & Kärenlampi, 1998; Sam, Jeréz, Dell'Amico, & Ruiz-Sanchez 2000; Romero-Aranda, Soria, & Cuartero, 2001; Xu & Zhou, 2008; Barbien et al., 2012), however, it may also lead to higher stomatal densities if there is an overall reduction of leaf area (Ciha & Brun, 1975; Romero-Aranda et al., 2001; Pearce, Millard, Bray, & Rood, 2005; Xu & Zhou, 2008). Mangrove stomata are similar in terms of frequency and dimension to those of other plants from other environments, but many species have stomatal crypts (e.g. *Avicennia*, *Aegiceras*, *Bruguiera*, *Ceriops*, *Heritiera*, *Lumnitzera*, *Nypa*), resulting in the formation

of substomatal chambers, in order to deal with the water stress of a saline environment. (Das & Ghose, 1993; Saenger, 2002).

The aim of this paper was to evaluate the effects of salinity on stomatal density, leaf area, plant height and diameter of *Rhizophora mangle*, and to determine if there was an association between these variables in three different sites that provide a natural salinity gradient. We hypothesized that lower salinity would lead to higher stomatal density, increased leaf area and greater tree height, because plants would be less water-stressed. In contrast, high salinity environments would produce smaller plants, with smaller leaves and fewer stomata. Support for this would contribute to our understanding of the environmental factors that promote intraspecific variability in ecophysiological and functional traits that are critical for plant function regarding water-gas exchange and biomass accumulation. Intraspecific trait variation may account for 25 % of the total trait variation of a community (Siefert et al., 2015). Knowledge of intraspecific trait variation in leaf area, stomatal density and tree height might help us improve our forecasts of carbon ecosystem budgets and our evaluations of the global and regional water cycle.

MATERIALS AND METHODS

Study site: The study was carried out in Xel-Há cove (20°19'15" N - 87°21'41" W & 20°18'50" N - 87°21'15" W) in the state of Quintana Roo, in the East coast of Yucatan Peninsula, 16 km Northeast from the city of Tulum. The area has an intermediate tropical and sub-humid climate, with rains somewhat distributed throughout the year, with a tendency of higher rainfall during summer, and rain during winter (Ax'(w_i)iw, Orellana-Lanza, Espadas, Conde, & Gay, 2009). The cove is connected to the Caribbean Sea by a channel that is about 90 m wide and 170 m long, has a central canal and two tributaries that drain towards the sea: North Arm and South Arm. The area is characterized by a pronounced and persistent salinity gradient over a short distance, the result of a mix of

underground freshwater and seawater (Organismo de Cuenca Península de Yucatán, 2008).

In January 2014 the entire area was explored, salinities were determined at several points using an YSI 650 MDS multiparameter probe (YSI Inc. Yellow Springs, OH, USA) and accessibility of *R. mangle* mangrove populations were assessed. Three sampling sites with different environmental conditions were chosen for their accessibility and salinity: *River*, *Lagoon* and *Beach*. The sites are separated from each other by at least 300 m. Plot limits were based on the natural limits and features of each site. Salinities were monitored monthly from May 2014 to February 2015 at 5 different points within each of the sampling plots.

The *River* site was located in the Southern arm section (20°18'52"N-87°21'40"W), where a 10 m x 30 m sampling plot was established parallel to the river. Salinity was measured as 15.5 PSU in the water surrounding the plants. The substrate was black to red mud mixed with high amounts of litterfall produced by the mangroves in decomposition. The *Lagoon* site was located in the Northern part of the cove (20°19'11" N - 87°21'30.39" W), where a 3 m x 6 m sampling plot was established. The salinity initially registered was 26.5 PSU in the water surrounding the mangrove plants. The soil was black mud with high amounts of litterfall and some scattered boulders. The *Beach* site (20°18'52" N - 87°21'23" W) is a small bay within the channel that connects the cove to the ocean in which salinities of 31.5 PSU were recorded in the 10 x 3 m plot. At this site, mangroves develop directly on the sandy beach.

Stomatal densities and plant morphometry: We tagged all plants within each plot, and for each plant, we measured basal stem circumference with a standard measuring tape, taking measurements just above the first pair of aerial roots. Diameters were then calculated from stem circumference. The total height of the plant was also measured along the main axis of the trees, using a standard measuring tape and a telescopic pole.

Three healthy mature leaves (i.e., fully expanded, non-senescent) of 20 different individuals in each plot were collected from the distal ends of branches exposed to full sunlight. Leaf length (LL) and width (LW) were measured for each leaf, and nail polish casts were prepared for stomatal observations (Brewer, 1992). The complete abaxial surface of each leaf was covered with transparent nail polish and left to dry for about 5 min. Once dry, the cast was removed using transparent adhesive tape, resulting in an impression of the leaf surface, which was then observed under a Leica Zoom 2000 stereoscope 45 \times using a millimeter grid. All stomata within 1 mm squares were counted, taking measurements from 10 squares on the leaf surface, avoiding the leaf edge and areas close to the central rib.

Statistical analysis: Average salinities and standard error were obtained for each month at each site. ANOVA was used to determine whether salinity differed significantly between sites over the duration of the study. The analysis was performed with InfoStat S (Universidad Nacional de Córdoba, Argentina). Stomatal counts were averaged per leaf for further analysis. Averages and standard deviation were calculated for stomatal density, leaf length and leaf width for each site for comparison. Leaf area ($LA = \Pi \cdot (LL/2) \cdot (LW/2)$) and leaf slenderness ($LS = LL/LW$) were calculated per individual / site / date. All variables were tested for normality using Shapiro-Wilks normality tests. An analysis of variance (ANOVA) was used to detect significant differences between leaf width, leaf length, leaf area, slenderness and stomatal density between sampling sites, followed by multiple comparison Tukey HSD tests. Tree heights and stem diameters did not show normal distribution within sites, and differences between sites were therefore tested using nonparametric Kruskal-Wallis ANOVA. Association between variables was tested through Pearson correlations.

A principal component analysis (JMP ver. 10.0.0, SAS Institute Inc., Cary, NC, USA) was used to determine the relationship between all

parameters among sites. All leaf traits (LL, LW, LA, LS and stomatal densities) were averaged per plant before carrying out the analysis. All variables were correlated with the resulting PCA axis to determine their contribution to the resulting components.

RESULTS

Salinities differed across sites ($F_{2, 134} = 214.77$; $P < 0.0001$) but were stable throughout the year within sites (Fig. 1). *River* had the lowest salinities, with an average salinity of 8.4 (SD = 1.20), followed by *Lagoon* with an average salinity of 13.7 (SD = 3.71), and *Beach* had the highest salinities (Average = 18.8 SD = 1.42). A total of 542 plants were tagged at the three sampling plots. The highest density of plants was recorded at the *Beach* site (11.5 ind/m²) followed by the *Lagoon* site (density = 9.05 ind/m²) while the *River* site only contained 33 plants (0.11 ind/m²). The salinity gradient was reflected in plant size and differed significantly between sites (Table 1, Fig. 2). The *Beach* site (highest salinity) had plants shorter than 3.5 m, while the *Lagoon* site had individuals mostly shorter than 3 m with some exceptions (individuals could reach 7 m). Finally, at the *River* site most individuals (90 %) were consistently taller than 5 m.

Significant differences were found for all the measured variables between sites (Table 1). Leaf width ranged between 1.9 cm (*Beach* site) and 6.8 cm (*River* site) (Table 1). Leaf length varied between 6.1 cm and 15.9 cm (Table 1), and significant differences between sites were found regarding leaf slenderness ($F_{2, 179} = 46.3$; $P < 0.0001$) and leaf area ($F_{2, 179} = 28.5$; $P < 0.0001$; Table 1 and Fig. 3), with the thinnest leaves collected at the *River* site (Fig. 3) and the largest leaves (highest leaf area) at the *River* site, while the smallest leaves corresponded to the *Lagoon* site (Fig. 3).

Within each of the sites, stomatal densities (stomata.mm⁻²) showed a wide range 54.5-102.5 stomata.mm⁻² on *Beach* leaves, 52.9-97.0 stomata.mm⁻² on *Lagoon* leaves, and 45.5-87.4 stomata.mm⁻² on *River* leaves (Table 1,

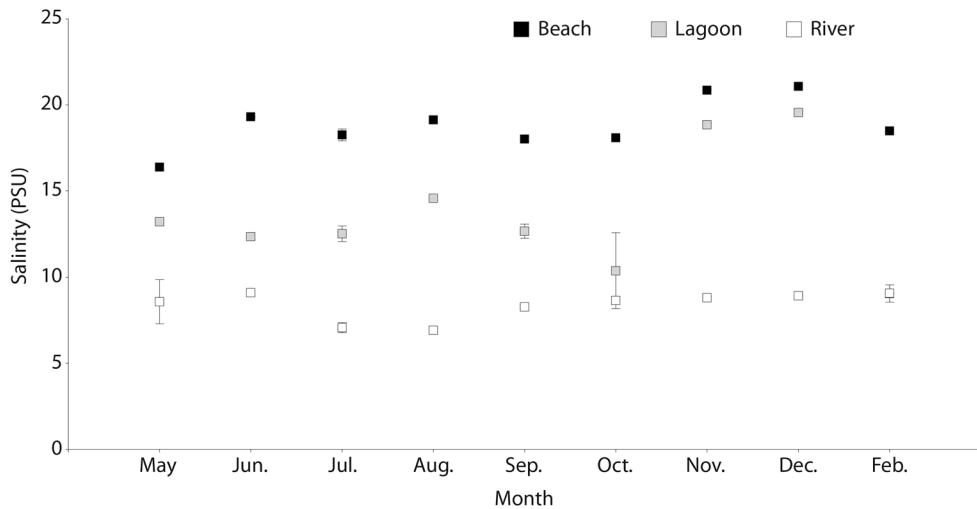


Fig. 1. Average salinities (PSU) and standard error, registered at the three sampling sites *River*; *Lagoon*; and *Beach* at the Xel-Há cove (XH), Quintana Roo, Mexico.

TABLE 1
Range and average (\pm standard deviation) salinities, leaf width, leaf length, leaf area, leaf slenderness, tree height, stem diameter and stomatal density (stomata/mm²), of *Rhizophora mangle* collected at three different sites (*River*, *Lagoon* and *Beach*) of the Xel-Há cove, Quintana Roo, Mexico, and statistical differences between the measured variables between sites

		Sites			Statistical differences between sites
		River	Lagoon	Beach	
Salinity	Mean \pm SD	8.4 \pm 1.20	13.7 \pm 3.71	18.8 \pm 1.42	F2, 134 = 214.77
	Range	4.6 - 12.8	1.6 - 19.7	16.2 - 21.2	P < 0.001
Tree height (m)	Mean \pm SD	7.5 \pm 2.36	1.4 \pm 1.77	1.3 \pm 0.61	H = 133.18
	Range	1.5 - 12.3	0.1 - 7.1	0.1 - 3.2	P < 0.001
Stem diameter (cm)	Mean \pm SD	8.7 \pm 5.00	1.1 \pm 1.48	1.6 \pm 0.51	H = 169.15
	Range	1.9 - 28.0	0.3 - 9.2	0.3 - 3.2	P < 0.001
Leaf width (cm)	Mean \pm SD	5.0 \pm 0.78	3.8 \pm 0.73	3.9 \pm 0.86	F2, 179 = 40.88
	Range	3.7 - 6.8	2.6 - 5.7	1.9 - 5.7	P < 0.001
Leaf length (cm)	Mean \pm SD	12.6 \pm 1.92	11.2 \pm 1.70	12.2 \pm 1.57	F2, 179 = 14.05
	Range	6.1 - 15.9	7.9 - 15.0	8.2 - 14.9	P < 0.001
Leaf area (cm ²)	Mean \pm SD	51.2 \pm 24.99	34.4 \pm 10.90	38.2 \pm 12.69	F _{2,179} = 28.49
	Range	25.0 - 84.4	16.1 - 59.1	12.9 - 64.9	P < 0.001
Leaf slenderness	Mean \pm SD	2.6 \pm 0.22	2.9 \pm 0.37	3.2 \pm 0.48	F _{2,179} = 46.28
	Range	2.1 - 3.1	2.2 - 4.0	2.5 - 4.8	P < 0.001
Stomatal density (stomata mm ⁻²)	Mean \pm SD	65.0 \pm 12.32	73.4 \pm 13.49	74.8 \pm 17.32	F2, 179 = 13.59
	Range	45.5 - 87.4	52.9 - 97.0	54.5 - 102.5	P < 0.001

Fig. 3). Differences in stomatal density could be explained by differences in the leaves from the *River* (P < 0.05), which had significantly lower stomatal densities (stomata.mm⁻²), while there were no significant differences between the *Lagoon* and *Beach* sites (P > 0.05; Fig. 3).

Stomatal density showed significant (P < 0.001) negative correlations with LW, LL, LA, stem diameter and tree height (Table 2). Leaf width was positively correlated with LL, LA, diameter and tree height (Table 2). However, LL was not correlated with neither

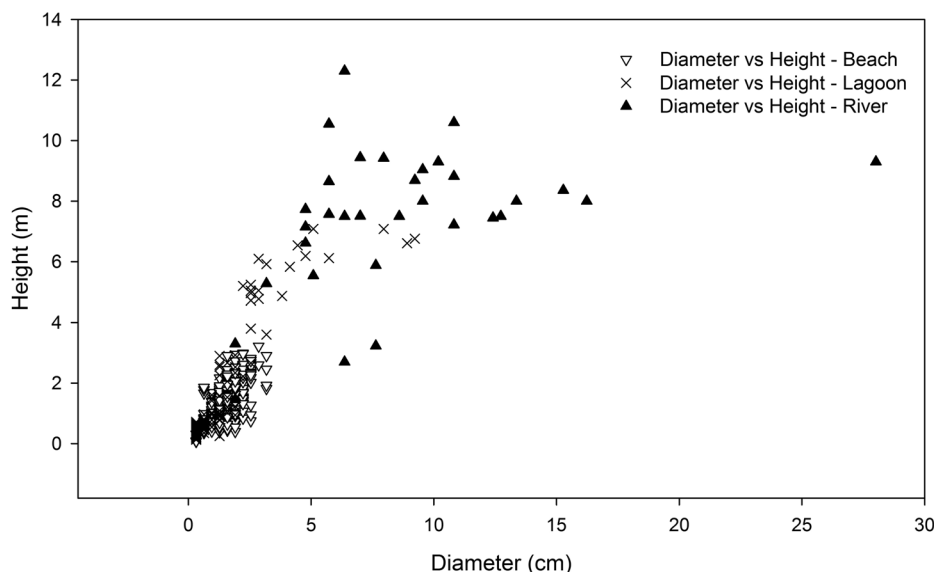


Fig. 2. Trunk diameter (cm) vs. tree height (m) of *Rhizophora mangle* plants at the three sampling plots *River* (triangle), *Lagoon* (crosses) and *Beach* (inverted triangle), in the Xel-Há Park, Quintana Roo, Mexico.

height nor diameter (Table 2). LA and LS were collinear, both being derived from LL and LW. We feel that LA was the more informative variable and so included LA and not LS in the PCA (Table 2). Tree diameter and height were also found to be strongly associated, so height was chosen to represent plant size, since it generally showed stronger association with the leaf trait variables (Table 2).

The principal component analysis (Fig. 4) differentiated among sites, especially segregating the trees from the *River* and *Beach* sites. The first principal component accounted for 53.7 % of the total variation. It was inversely correlated with stomatal density (-0.29), and directly correlated with tree height (0.30) and diameter (0.29), as well as, leaf area (0.51), leaf width (0.52), and length (0.44). The second principal component accounted for 27.40 % of the total variability of the data and showed strong inverse correlation with tree height (-0.55) and diameter (-0.56). The strongest positive association of the second component was with leaf length (0.44) (Fig. 4). The principal component scores were considered as an

index summarizing all measured variables and were used to correlate each parameter in order to determine their contribution to each of the PCA axes (Table 3). All variables were significantly correlated with component 1. The highest correlations were with leaf length, trunk diameter, and inversely with leaf area. Only three variables were significantly correlated with component 2: leaf length, trunk diameter and height (Table 3).

DISCUSSION

Plant size varied among sampled sites and reflected the salinity gradient, with the largest plants at the lowest salinity site and smaller plants at the higher salinity sites. This result is consistent with previous findings that plant growth in mangroves is affected by salinity. For example, in *R. mucronata* sapling growth declines above 50‰ seawater and in *Bruguiera parviflora* growth declines, with further increases in salinity (Aziz & Khan, 2001). In *B. parviflora*, plant growth decreases with 200 and 400 mM NaCl treatments

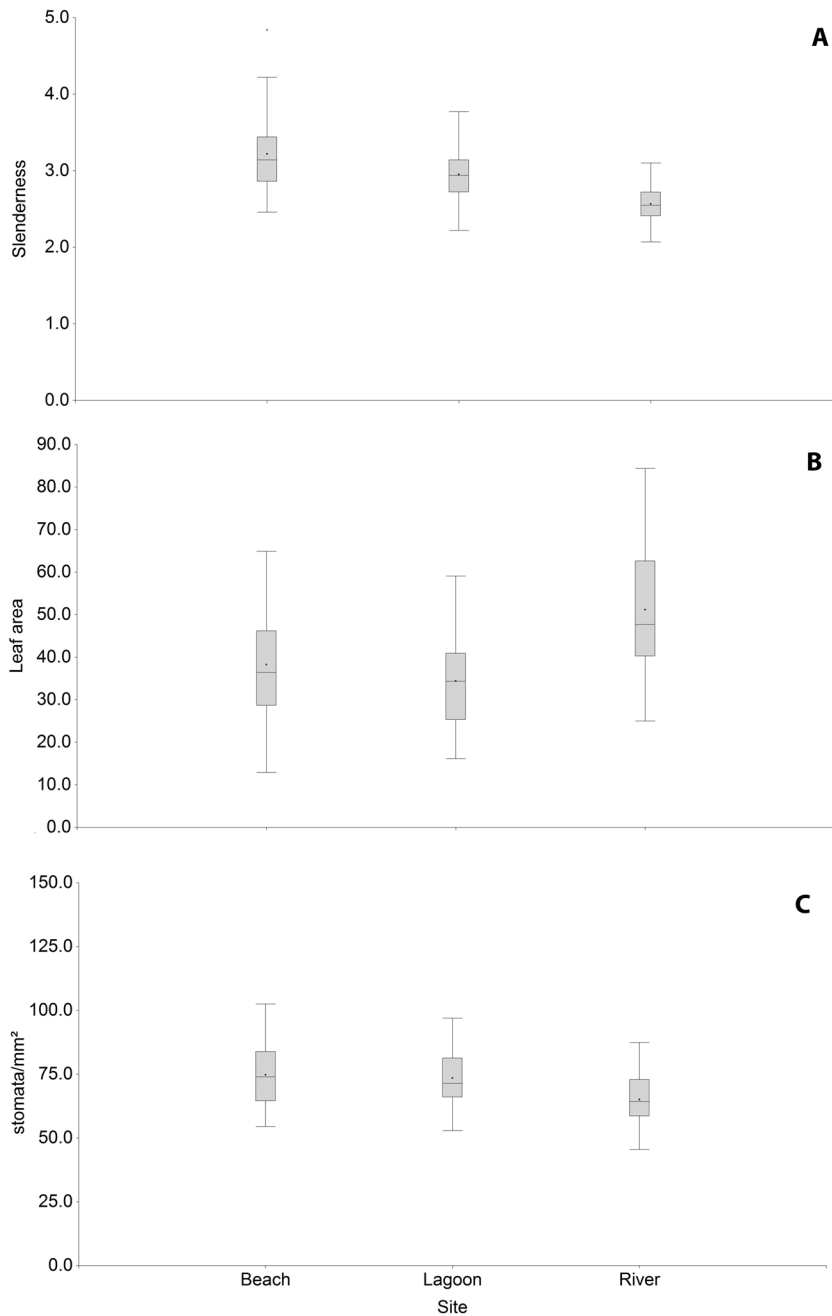


Fig. 3. Average, median and standard deviation of (A) leaf slenderness ($S = \text{Length/width}$), (B) leaf area, and (C) stomatal density/mm² of *Rhizophora mangle*, sampled at three different sites (*River*, *Lagoon*, *Beach*) of the Xel-Há Park, Quintana Roo, Mexico, and Tukey contrasts (*A*, *B*, *C*, smallest to largest value, means with a common letter are not significantly different ($P > 0.05$)).

TABLE 2
Pearson coefficients (r) and P -values of correlations between stomatal density (stomata mm⁻²), leaf width, leaf length, leaf slenderness and leaf area, tree height and diameter of *Rhizophora mangle* trees, sampled in three different environments of the Xel-Há Park, Quintana Roo, Mexico

Variable (1)	Variable (2)	Pearson	P-value
Stomatal density	Leaf width	-0.31	<0.0001
	Leaf length	-0.25	0.0009
	Area	-0.29	0.0001
	Slenderness	0.27	0.0003
	Diameter	-0.33	<0.0001
	Height	-0.32	<0.0001
Leaf width	Leaf length	0.79	<0.0001
	Area	0.96	<0.0001
	Slenderness	-0.74	<0.0001
	Diameter	0.33	<0.0001
	Height	0.37	<0.0001
Leaf length	Area	0.91	<0.0001
	Slenderness	-0.21	0.0056
	Diameter	0.03	0.7336
	Height	0.02	0.8205
Area	Slenderness	-0.55	<0.0001
	Diameter	0.22	0.0031
	Height	0.25	0.0008
Slenderness	Diameter	-0.46	<0.0001
	Height	-0.54	<0.0001
Diameter	Height	0.71	<0.0001

in long-term hydroponic culture (Parida, Das, & Mittra, 2004).

Leaves from the low salinity *River* site were the longest, however, there was no association between the length of the leaves and the size of the plants (height and stem circumference), which suggests that this variable has environmental plasticity independent of plant size. Leaf width on the other hand, showed a positive relationship with plant height and stem diameter. Mangrove leaves collected at the Xel-Há cove were comparable in length with typical mangrove leaves, which have lengths of 10-15 cm under optimum conditions (Canoy, 1975; Snedaker & Brown, 1981). Only leaves from the *River* site were wider than 5 cm, while leaves from the *Lagoon* and *Beach* sites were narrower than the reported standard of 5-7 cm (Canoy, 1975; Snedaker & Brown, 1981). Leaf slenderness also reflected the salinity gradient and had the lowest values at the *River* site and the highest value at the *Beach* site. Leaf area was lowest at the *Lagoon* site, given that the shortest and narrowest leaves were collected here, indicating some kind of environmental stress affecting leaf expansion. Mangrove leaf size can be affected by salinity, as shown by Parida et al. (2004) long-term hydroponic culture of *Bruguiera parviflora*, in which leaf size reduction with NaCl treatment was attributed

TABLE 3
Correlation coefficients (R) and their significance value (P) between the biological data set (stomatal density, leaf width, leaf length, stem diameter, tree height, slenderness, and leaf area) and the first and second principal components that synthesize them. The eigenvalue (λ) and percent of the data variability accounted by principal each component are shown in the second upper row

Variable	Principal component 1 ($\lambda=3.22$; 53.7 %)		Principal component 2 ($\lambda=1.64$; 27.4 %)	
	r	P	r	P
Stomatal density	-0.57	<.0001	0.36	0.005
Leaf width (cm)	-0.52	<.0001	0.19	0.14
Leaf length (cm)	0.95	<.0001	0.57	<.0001
Trunk diameter (cm)	0.78	<.0001	-0.72	<.0001
Height (m)	0.52	<.0001	-0.71	<.0001
Slenderness	0.54	<.0001	0.29	0.03
Leaf area (cm ²)	-0.70	<.0001	0.35	0.006

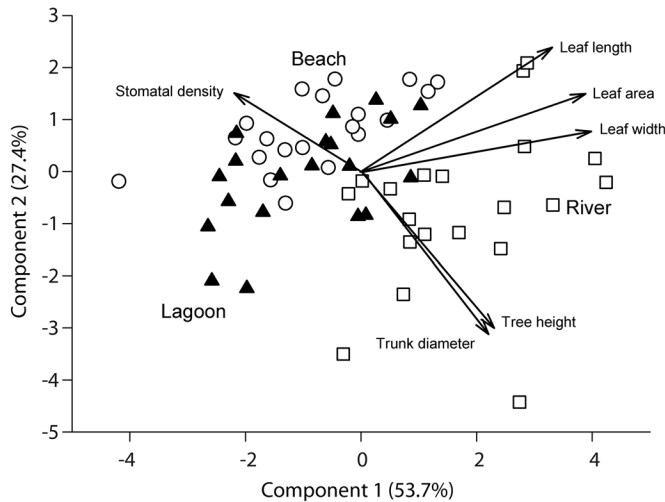


Fig. 4. Biplot indicating the distribution of the scores from the Principal Component Analysis and the variables used for *Rhizophora mangle* tree and leaf characterization in three sampling sites (*River*, squares; *Lagoon*, triangles; *Beach*, circles) within the Xel-Há cove, Quintana Roo, Mexico. Each data point represents a tree.

to reduction of intercellular space. *R. stylosa* saplings grown at salinities greater than 25 ‰ seawater, have sharp and significant decreases in leaf, stem and root dry weights (Clough, 1984). In *Avicennia germinans* high salinities increase leaf mortality and reduce leaf expansion significantly (Suárez & Medina, 2005). A comparative study in *A. germinans*, *L. racemosa* and *R. mangle* showed that elevated environmental salinities lead to increased osmolality of the leaf sap and leaf area reduction, which was especially pronounced in *R. mangle* (Medina & Francisco, 1997).

Initially we hypothesized that leaves from environments with higher salinities would have lower stomatal density in order to reduce water loss. This is based on observations made on the mangrove *Kandelia candel* (Rhizophoraceae) (Hwang & Chen, 1995) in which leaf size and stomatal density were reduced at high salinity (> 400 mM NaCl, approx. 23 PSU) in comparison with intermediate and low salinity (Hwang & Chen, 1995). Surprisingly, we observed the highest stomatal densities at the highest salinity sites, *Lagoon* and *Beach*. Leaf size responded to salinity as expected: smaller leaves in higher salinities. Water deficit can affect leaf size by

decreasing cell size and causing an increase in stomatal density (Quarrie & Jones, 1977; Spence, Wu, Sharpe, & Clark, 1986; Martínez, Silva, Ledent, & Pinto, 2007; Xu & Zhou, 2008). A study on *L. racemosa* and *A. germinans* did not show reductions of stomatal density with increases in salinity (Lovelock & Feller, 2003), which was confirmed by Schwarzbachl and Ricklefs (2001). Dahdouh-Guebas et al. (2004) also failed to find significant differences in stomatal density with elevated salinities in *A. germinans*. Stomatal density may increase with leaf size reduction to overcome a simple geometric practicality of fitting enough functional stomatal units per unit of leaf surface area to meet the desired CO₂ flux and to service photosynthetic capacity (Franks & Farquhar, 2007). Hetherington and Woodward (2003) for example demonstrated that there was a strong relationship between stomatal size and density. In addition, stomatal length seems to decrease with increasing droughts (Aasamaa, Sober, & Rahi, 2001). Smaller stomata have faster dynamic characteristics, which has implications for improved long-term water use efficiency and lower risk of disruption of the leaf hydraulic system (Drake, Froend, & Franks,

2013), allowing the stomata to open and close faster and thus respond rapidly to environmental change (Hetherington & Woodward, 2003). Salisbury (1928) found that stomatal density was inversely related to leaf size, due to a higher cell insertion (number of cells per unit area) in smaller leaves. However, at a cellular level the stomatal index remains constant. Stomata are produced early in leaf expansion so their density is at its maximum when young, and then decreases as the leaf expands to a relatively stable density, independently of the level of cell insertion or area (Gay & Hurd, 1975). Our results confirm that stomatal density is inversely related to leaf area, especially leaf width, and that salinity may increase stomatal density by causing reduction of leaf size.

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RESUMEN

Variación en la densidad estomática, área foliar y tamaño de *Rhizophora mangle* (Malpighiales: Rhizophoraceae) a lo largo de un gradiente de salinidad en el Caribe Mexicano. Los manglares tienen varios mecanismos de resistencia salina y la mayoría de las especies presentan algunas características xeromórficas con el fin de conservar el agua. El tamaño de la hoja y la densidad de los estomas desempeñan un papel importante en el

mantenimiento del equilibrio hídrico. El intercambio de gases puede medirse mediante la regulación de la apertura de los estomas, así como el número de estomas sobre la epidermis, dos características que pueden variar intraespecíficamente en función las condiciones ambientales, tales como el estrés hídrico. *Rhizophora mangle* es una de las especies de mayor importancia en América del Norte y Sur, y de África occidental. El objetivo de este trabajo fue evaluar los efectos de la salinidad sobre la densidad de los estomas, el tamaño de la hoja y el tamaño de las plantas de *Rhizophora mangle* y determinar si existe una relación entre las tres variables, comparándose tres ambientes diferentes a lo largo de un gradiente natural de salinidad en Xel-Há, Quintana Roo, México. La hipótesis fue que los ambientes de alta salinidad producirían plantas más pequeñas, con hojas más pequeñas y menos estomas. Se seleccionaron tres sitios de estudio con condiciones ambientales diferentes y se midió la salinidad cada mes. Un total de 542 plantas fueron etiquetadas en los tres sitios, y se midió su altura y diámetro del tronco. Se recolectaron tres hojas de 20 árboles en cada uno de los sitios, y se obtuvo el área de cada hoja. La densidad estomática se midió mediante la técnica de microrelieve con barniz de uñas, observando diez 1 mm cuadrados bajo un microscopio óptico. Se utilizó un análisis de componentes principales para determinar la asociación entre altura de árbol, área de hoja y densidad estomática. El gradiente de salinidad se vio reflejado en el tamaño de las plantas, produciendo plantas más pequeñas en el sitio de alta salinidad. El largo de las hojas no se correlacionó con el tamaño de las plantas, por lo cual se concluyó que esta variable tiene plasticidad ambiental particular. Las hojas más grandes fueron encontradas en el sitio de baja salinidad y tuvieron densidades estomáticas menores. No se pudo observar que la densidad de estomas disminuyera con las altas salinidades. Estos resultados confirman que la densidad estomática es inversamente relacionada con el tamaño de la hoja, especialmente el ancho, y que la densidad estomática incrementa con la salinidad debido a la reducción del tamaño de hoja.

Palabras clave: altura, área foliar, densidad estomática, estrés hídrico, manglar, plasticidad de hoja.

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