



Journal of Applied Research and  
Technology

ISSN: 1665-6423

[jart@aleph.cinstrum.unam.mx](mailto:jart@aleph.cinstrum.unam.mx)

Centro de Ciencias Aplicadas y  
Desarrollo Tecnológico  
México

Belhamdi, Badreddine; Merzougui, Zoulikha; Trari, Mohamed; Addoun, Abdelhamid  
A kinetic, equilibrium and thermodynamic study of L-phenylalanine adsorption using  
activated carbon based on agricultural waste (date stones)

Journal of Applied Research and Technology, vol. 14, núm. 5, 2016, pp. 354-366

Centro de Ciencias Aplicadas y Desarrollo Tecnológico  
Distrito Federal, México

Available in: <http://www.redalyc.org/articulo.oa?id=47448461009>

- How to cite
- Complete issue
- More information about this article
- Journal's homepage in [redalyc.org](http://redalyc.org)

[redalyc.org](http://redalyc.org)

Scientific Information System

Network of Scientific Journals from Latin America, the Caribbean, Spain and Portugal

Non-profit academic project, developed under the open access initiative



Review

# A kinetic, equilibrium and thermodynamic study of L-phenylalanine adsorption using activated carbon based on agricultural waste (date stones)

Badreddine Belhamdi<sup>a,\*</sup>, Zoulikha Merzougui<sup>a</sup>, Mohamed Trari<sup>b</sup>, Abdelhamid Addoun<sup>a</sup>

<sup>a</sup> Laboratory of Physical and Chemical Study of Materials and Applications in the Environment, Faculty of Chemistry (USTHB), BP 32-16111 EL-Alia, Algeria

<sup>b</sup> Laboratory of Storage and Valorization of Renewable Energies, Faculty of Chemistry (USTHB), BP 32-16111 EL-Alia, Algeria

## Abstract

The main purpose of this work is to produce low cost activated carbons from date stones wastes for the adsorption of L-phenylalanine. The activated carbons were prepared by chemical activation with KOH (ACK) and ZnCl<sub>2</sub> (ACZ) and characterized by scanning electron microscopy, N<sub>2</sub> adsorption–desorption isotherms and FT-IR spectroscopy. Both The activated carbons ACK and ACZ have high specific surface areas and large pore volumes, favorable for the adsorption. Batch experiments were conducted to determine the adsorption capacities. A Strong dependence of the adsorption capacity on pH was observed, the capacity decreases with increasing pH up to optimal value of 5.7. The adsorption follows a pseudo-second order kinetic model. Additionally, the equilibrium adsorption data were well fitted to the Langmuir isotherm, and the maximum adsorption capacities of L-phenylalanine onto ACK and ACZ were 188.3 and 133.3 mg g<sup>-1</sup> at pH 5.7, respectively. The thermodynamic study revealed that the adsorption of L-phenylalanine onto activated carbons was exothermic in nature. The proposed adsorption mechanisms take into account the hydrophobic and electrostatic interactions which played the critical roles in the L-phenylalanine adsorption.

© 2016 Universidad Nacional Autónoma de México, Centro de Ciencias Aplicadas y Desarrollo Tecnológico. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

**Keywords:** Activated carbon; L-phenylalanine; Size distribution; Sorbent surface; Adsorption isotherm; Thermodynamic

## 1. Introduction

Adsorption of amino acids onto solid surfaces has received much attention because of its scientific importance and applications in the separation and purification processes (Han & Yun, 2007; Hong & Bruening, 2006; Kostova & Bart, 2007; O'Connor et al., 2006; Sánchez-Hernández, Bernal, del Nozal, & Toribio, 2016). Amino acids are biomolecules of great relevance that are widely used in many industries such as food, cosmetic, medicine, biochemistry and others (Bourke & Kohn, 2003; Hartmann, 2005; Infante et al., 2004; Oshima, Saisho, Ohe, Baba, & Ohto, 2009; Palit & Moulik, 2001). They are non-toxic and are used as building blocks for the production of pharmaceutical and agrochemical compounds. In addition,

they are interesting molecules as adsorbates because of their molecular size and zwitterionic nature (O'Connor et al., 2006). Similar to many amino acids, L-phenylalanine is essential for animals and the human body. It is extensively used as ingredient in food or feed additive, in infusion fluids, nutraceutical and pharmaceutical (Pimentel, Alves, Costa, Fernandes, et al., 2014; Pimentel, Alves, Costa, Torres, et al., 2014; Zhou, Liao, Wang, Du, & Chen, 2010). Generally, the amino acids have been studied by adsorption on well-ordered surfaces of solids. On the other hand, most of the current methods employed for the removal of L-phenylalanine from protein hydrolysates are based on the adsorption on activated carbon, polymeric resins, zeolites and ion exchangers (Lopes, Delvivo, & Silvestre, 2005; Outinen et al., 1996; Shimamura et al., 2002). These studies give information for practical researches on the purification and separation of amino acids. Over the last years, several studies have been reported for the adsorption of amino acids on porous solids (Casado et al., 2012; El Shafei, 2002; El Shafei & Moussa, 2001; Ghosh, Badruddoza, Uddin, & Hidajat, 2011;

\* Corresponding author.

E-mail address: [Badrobg@gmail.com](mailto:Badrobg@gmail.com) (B. Belhamdi).

Peer Review under the responsibility of Universidad Nacional Autónoma de México.

Goscianska, Olejnik, & Pietrzak, 2013b, 2013c; Jiao, Fu, Shuai, & Chen, 2012; Long et al., 2009; Mei, Min, & Lü, 2009; Palit & Moulik, 2001; Silvério, Dos Reis, Tronto, & Valim, 2008; Titus, Kalkar, & Gaikar, 2003; Wu, Zhao, Nie, & Jiang, 2009). However, their adsorption capacity is still low because of the small pore volume or wide pores of these adsorbents, which are consequently inappropriate to the molecular size of amino acids. Moreover, the major drawback of the adsorption process is the high cost for the production and regeneration of adsorbents. Such inconvenient resulted in growing research on inexpensive adsorbents (Alves, Franca, & Oliveira, 2013a, 2013b; Clark, Alves, Franca, & Oliveira, 2012; Goscianska, Nowicki, & Pietrzak, 2014; He, Lin, Long, Liang, & Chen, 2015; Sebben & Pendleton, 2015).

In this study, porous activated carbon-based materials obtained from date stones (seeds) are potential adsorbents for L-phenylalanine amino acid. This is mainly due to their physical and chemical characteristics such as highly developed porous structure, good thermal stability, low cost and more accessibility. Date stones are among the most common agricultural by products available in palms growing in the Mediterranean countries like Algeria, which is one of the largest producers in the world. Algeria produces more than 400 different varieties of dates with an annual production of about 400,000 tons (Chandrasekaran & Bahkali, 2013). Date stones constitute roughly 10% of the date weight and this lignocellulosic-based agricultural waste is a good precursor for preparing activated carbon because of its excellent natural structure and low ash content (Bouchenafa-Saib, Grange, Verhasselt, Addoun, & Dubois, 2005; Merzougui & Addoun, 2008). As it is well known, two methods are commonly used for the preparation of activated carbon: physical and chemical activations. Compared with the physical process, the chemical activation presents some advantages like low activation temperature, short activation time, high surface area, well developed microporosity of activated carbon, simple operation and low energy consumption (Deng, Yang, Tao, & Dai, 2009; Pereira et al., 2014). Therefore, the date stones can be activated with chemical agents such as KOH, ZnCl<sub>2</sub>, H<sub>3</sub>PO<sub>4</sub>, K<sub>2</sub>CO<sub>3</sub> and NaOH, to obtain activated carbons with well-developed textural characteristics. To the best of our knowledge, the use of activated carbons for the L-phenylalanine recovery from aqueous solutions by adsorbents based on date stones are not available in the open literature. Thus, the principal objective of this work was to prepare porous activated carbons with high surface areas from date stones by chemical activation with KOH and ZnCl<sub>2</sub>. The activated carbons proved to be good candidates for the adsorption of L-phenylalanine in an aqueous medium.

## 2. Materials and methods

### 2.1. Materials

The date stones used in this study were from Algerian origin. The following reagents were used: L-phenylalanine standard (>98%, Fluka, France), potassium hydroxide (>98%, Sigma Aldrich, USA), zinc chloride (>98%, Sigma Aldrich, USA), KH<sub>2</sub>PO<sub>4</sub> (>99%, Fluka, France), K<sub>2</sub>HPO<sub>4</sub> (>99%, Fluka,

France), NaHCO<sub>3</sub> (>99%, Sigma Aldrich, USA), Na<sub>2</sub>CO<sub>3</sub> (>99%, Sigma Aldrich, USA), HCl (37%, Sigma Aldrich, USA), KCl (>98%, Fluka, France), NaCl (>99%, Sigma Aldrich, USA), NaOH (>99%, Sigma Aldrich, USA). Ultrapure water was obtained from milli-Q system (Millipore, France).

### 2.2. Preparation of the activated carbons

The activated carbons were prepared from date stones. At first, the stones were thoroughly washed with distilled water and dried in an air oven at 120 °C; such protocol was effective to facilitate crushing and grinding. A fraction particle size of between 0.5 and 1 mm was used for the preparation of activated carbons by impregnation with ZnCl<sub>2</sub> and KOH. The precursor was impregnated with a chemical activating agent in a solid form. The impregnated precursor was carbonized in a horizontal tubular furnace under nitrogen flow with a heating rate of 5 °C min<sup>-1</sup>, to allow free evolution of volatiles, up to the hold temperature for 1 h. The resulting activated carbon was immersed in HCl solution (0.1 mol L<sup>-1</sup>) under reflux ebullition (3 h) in order to extract the compound formed and reagent excess. Then, the solution was filtered and the black solid was washed with hot distilled water until the test with AgNO<sub>3</sub> became negative. The adsorbent was dried at 120 °C, and kept in tightly closed bottles until use. The activated carbons were named ACZ (1 g ZnCl<sub>2</sub>: 1 g date stones, activated at 600 °C), ACK (9 mmol KOH: 1 g date stones, activated at 800 °C).

### 2.3. Characterization

The specific surface area and pore structure of the activated carbons were characterized by nitrogen adsorption-desorption isotherms at -196 °C using the ASAP 2010 Micromeritics equipment. All the activated carbons were outgassed at 150 °C overnight. The specific surface area was calculated by the Brunauer–Emmett–Teller (BET) equation (Brunauer, Emmett, & Teller, 1938). The external surface area, micropore area and micropore volume were calculated by the *t*-plot method. The total pore volume was evaluated from the liquid volume of N<sub>2</sub> at a high relative pressure near unity 0.99 (Guo & Lua, 2000). The mesopore volume was calculated by subtracting the micropore volume from the total volume. The pore size distribution (PSD) was determined using the density functional theory (DFT) model. The morphology of activated carbons was visualized by scanning electron microscopy (SEM) using a Philips XL 30 equipped with an energy dispersive spectrometer (EDS). The Fourier transform infrared (FT-IR) spectroscopy was used to determine the functional groups of the activated carbons; the spectra were recorded over the range (400–4000 cm<sup>-1</sup>) on a Perkin-Elmer spectrum two spectrometer using KBr pellets.

### 2.4. Determination of zero point charge pH<sub>PZC</sub>

The determination of the point of zero charge (pH<sub>PZC</sub>) was conducted to investigate how the surface charge of ACK and ACZ adsorbents depends on pH. pH<sub>PZC</sub> of the activated carbons was determined using the procedure described elsewhere

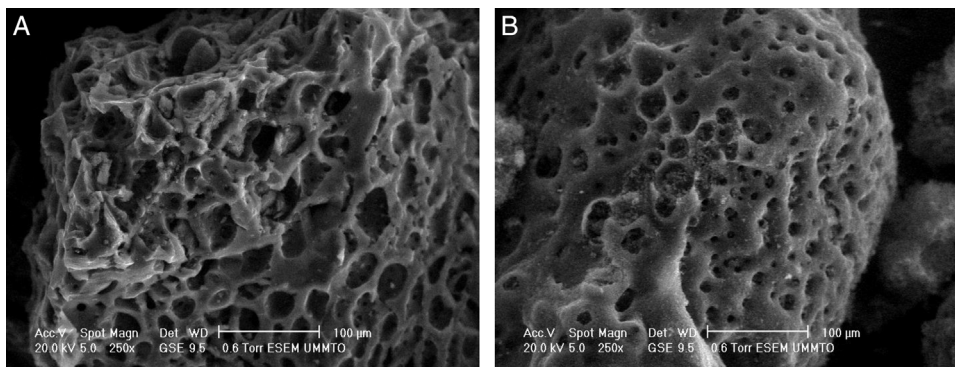


Fig. 1. SEM images of ACZ (A) and ACK (B) before adsorption.

(Prahas, Kartika, Indraswati, & Ismadji, 2008): 0.01 M of NaCl was prepared and the initial pH was adjusted between 2 and 12 using HCl or NaOH solution (0.1 M). 50 mL of NaCl solution was placed in Erlenmeyer flasks with 0.1 g of adsorbent. The flasks were kept under agitation (150 rpm, 48 h), and the final pH of the solution was measured. The intersection point of the curves  $\text{pH}_{\text{final}}$  vs.  $\text{pH}_{\text{initial}}$  and the bisector was taken as  $\text{pH}_{\text{PZC}}$ .

## 2.5. Adsorption experiments

The batch adsorption experiments were performed in 100 mL Erlenmeyer flasks containing a mass of adsorbent mixed with a known volume of L-phenylalanine solution ( $200 \text{ mg L}^{-1}$ ) under agitation (150 rpm). The effect of the contact time was studied to determine the time required for equilibrium at natural pH at  $20^\circ\text{C}$ . For the temperature effect, 50 mg of activated carbon was added to 50 mL of L-phenylalanine solutions with concentrations ranging from 50 to  $1000 \text{ mg L}^{-1}$ , prepared by dissolving appropriate amounts of L-phenylalanine in ultrapure water ( $18.2 \text{ M}\Omega \text{ cm}$ ), the flasks were maintained under constant agitation at various temperatures (20, 25, 35 and  $40^\circ\text{C}$ ) for 300 min. The effect of pH on the adsorption was performed by mixing 50 mg of activated carbon into 50 mL of L-phenylalanine solutions in the pH range (2.0–9.4), under constant agitation at  $20^\circ\text{C}$  for 300 min. The pH value of the L-phenylalanine solutions was changed by using different buffer solutions (pH 5.7–7.2 potassium phosphate buffer; pH 2.0 HCl–KCl buffer; pH 9.4 bicarbonate buffer). The concentrations of L-phenylalanine remaining in the supernatant solutions were filtered using a hydrophilic syringe filter with a pore size of  $0.45 \mu\text{m}$ . The adsorbed amount was determined using a UV–vis spectrophotometer at 257 nm. (The adsorption capacity) The equilibrium adsorption capacity per unit mass of activated carbons  $q_e$  ( $\text{mg g}^{-1}$ ) and the removal percentage of the L-phenylalanine  $\eta$  (%) were calculated from the following equation:

$$q_e = \frac{(C_0 - C_e)V}{W} \quad (1)$$

$$\eta = \frac{(C_0 - C_e)}{C_0} \times 100 \quad (2)$$

where  $C_0$  and  $C_e$  are the initial and equilibrium L-phenylalanine amino acid concentrations in the liquid phase ( $\text{mg L}^{-1}$ ),  $W$  (g) the weight of adsorbent and  $V$  (L) the volume of solution.

## 2.6. Desorption study

Desorption of L-phenylalanine was investigated in order to explore the regeneration and recycling ability of the two activated carbons. For this, 50 mg of ACK and ACZ were mixed with 50 mL of L-phenylalanine solution at a saturated concentration, and stirred at 150 rpm at optimum adsorption temperature ( $20^\circ\text{C}$ ) for 300 min. The amount of adsorbed amino acid was determined by the same equation used in the adsorption experiments (see Section 2.5). Thereafter, ACK and ACZ were washed with ultrapure water until the residual concentration of L-phenylalanine becomes negligible. The loaded activated carbons were then allowed to be in contact with 50 mL of two eluent solutions (M NaOH and M HCl,  $10^{-2} \text{ M}$ ) for 300 min. The desorbed carbons were again subjected to the next batch in order to check desorption and reusability of ACK and ACZ. The amount of desorbed amino acid was calculated from the concentration of desorbed L-phenylalanine in liquid phase using UV–vis spectrophotometer at 257 nm. The percentage of desorbed L-phenylalanine from the activated carbons was calculated according to the following equation:

$$\text{Desorption (\%)} = \left( \frac{\text{Mass desorbed}}{\text{Mass adsorbed}} \times 100 \right) \quad (3)$$

## 3. Results and discussion

### 3.1. Characterization of activated carbons

#### 3.1.1. Scanning electron microscopy (SEM)

The SEM micrographs show the effects of  $\text{ZnCl}_2$  and KOH on the surface pore structures of the activated carbons (Fig. 1). The external morphology shows more or less homogeneous cavities on the surfaces of ACK and ACZ. These cavities resulted from the evaporation of chemical agents during the activation process, leaving space previously occupied by KOH and  $\text{ZnCl}_2$ . Figure 1 suggests that the large pores on the surface are connected to a whole network of smaller pores inside the activated

Table 1  
Textural characteristics of ACK and ACZ.

Adsorbent	Surface area ( $\text{m}^2 \text{g}^{-1}$ )			Pore volume ( $\text{cm}^3 \text{g}^{-1}$ )			DFT pore size (nm)
	$S_{\text{BET}}$	$S_{\text{ext}}$	$S_{\text{mic}}$	$V_T$	$V_{\text{mes}}$	$V_{\text{mic}}$	
ACK	1209	276	933	0.550	0.180	0.370	1.48
ACZ	1235	525	710	0.630	0.341	0.289	1.59

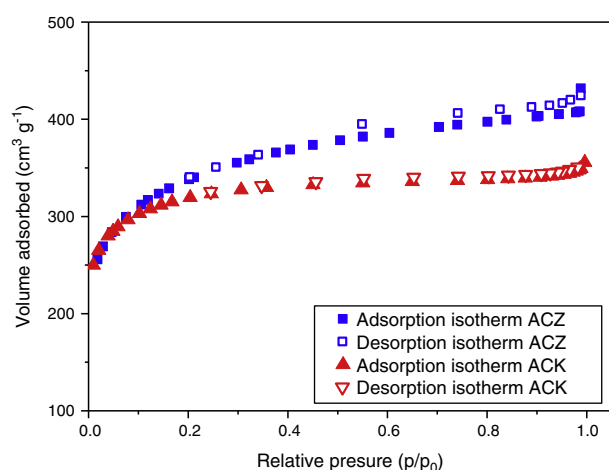


Fig. 2. Nitrogen adsorption–desorption isotherms of ACK and ACZ samples at  $-196^\circ\text{C}$ .

carbon. Therefore, the ACK and ACZ have great potential as good adsorbents for L-phenylalanine.

### 3.1.2. Nitrogen sorption

The  $\text{N}_2$  adsorption–desorption isotherms of activated carbons ACK and ACZ are given in Figure 2. The isotherms are type I, according to the IUPAC classification, assigned to microporous materials, and they present at a very low relative pressure ( $P/P_0 < 0.2$ ) a significant increase of  $\text{N}_2$  adsorption corresponding to the micropores filling (Fig. 2). However, the amount of adsorbed nitrogen is reduced at higher pressures, suggesting the development of both micro and meso-porosity in ACK and ACZ. The presence of hysteresis loops indicates that some mesoporosity starts to be developed by capillary condensation. The textural parameters of activated carbons determined from nitrogen adsorption-desorption are gathered in Table 1. It can be concluded that the activation of date stones by  $\text{ZnCl}_2$  and KOH leads to active coals of a well-developed surface area and a high pore volume (Table 1). Among the activated carbons, ACZ exhibits the highest surface area ( $1235 \text{ m}^2 \text{g}^{-1}$ ) and pore volume ( $0.63 \text{ cm}^3 \text{g}^{-1}$ ) while ACK has a smaller surface area ( $1209 \text{ m}^2 \text{g}^{-1}$ ) and pore volume ( $0.55 \text{ cm}^3 \text{g}^{-1}$ ). As shown in Table 1, the activated carbon prepared by KOH is essentially microporous, with 77% of its surface area. Similar trends have been found for the influence of the chemical activating agent on the development of the surface area and pore volume of activated carbons obtained through  $\text{ZnCl}_2$  and KOH activation of other lignocellulosic materials (Angin, 2014; Bagheri & Abedi, 2009; Foo & Hameed, 2011; Sreńscek-Nazzal, Kamińska, Michalkiewicz, & Koren, 2013; Yorgun, Vural, &

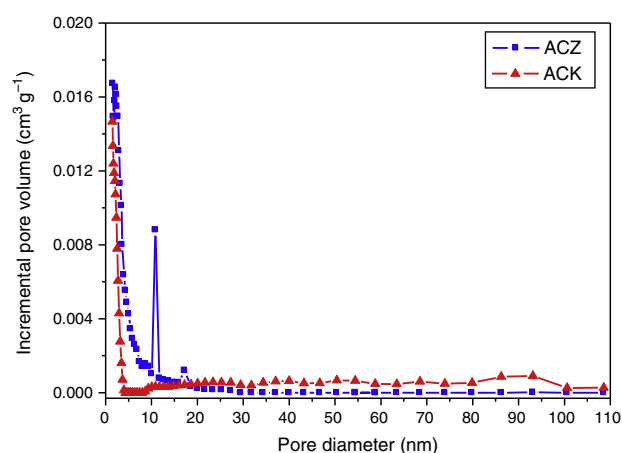


Fig. 3. DFT pore size distribution for ACK and ACZ samples.

Demiral, 2009; Zhu, Wang, Peng, Yang, & Yan, 2014). The pore size distribution is an important property in the adsorption mechanism because the adsorption of molecules of different sizes and shapes is directly related to the pore size of adsorbents. According to the classification adopted by IUPAC, adsorbent pores are classified as micropores ( $<2 \text{ nm}$ ), mesopores ( $2\text{--}50 \text{ nm}$ ) and macropores ( $>50 \text{ nm}$ ). Figure 3 shows the pore size distribution for the activated carbons, which clearly indicates that the pore diameter is in the micropore range. It is important to mention that a L-phenylalanine molecule is relatively small with a size of  $0.7 \text{ nm} \times 0.5 \text{ nm} \times 0.5 \text{ nm}$  (Alves et al., 2013b; Long et al., 2009). Therefore, such micropores with a size of ( $<2 \text{ nm}$ ) are accessible for L-phenylalanine molecules.

### 3.1.3. Infrared spectroscopy (FT-IR)

The FT-IR is an important technique to qualitatively determine the characteristic functional groups of the adsorbents. The spectra of porous activated carbons (Fig. 4) show multiple functions which can also be observed in other carbons activated by KOH and  $\text{ZnCl}_2$  (Huang, Ma, & Zhao, 2015; Lua & Yang, 2005; Saka, 2012). The FT-IR analysis indicates that ACK and ACZ exhibit a similar shape and the same functional groups. The broad band in the range ( $3000\text{--}3500 \text{ cm}^{-1}$ ) is ascribed to the O–H stretching mode of hydroxyl groups with hydrogen bending of adsorbed water. Bands ( $2900\text{--}2950 \text{ cm}^{-1}$ ) are assigned to asymmetric and symmetric stretching vibrations of aliphatic bond  $-\text{CH}$ ,  $-\text{CH}_2$  and  $-\text{CH}_3$  while the bands around  $1580 \text{ cm}^{-1}$  may be due to the presence of aromatic  $\text{C}=\text{C}$  ring stretching vibration. The band at  $1400 \text{ cm}^{-1}$  is associated with  $-\text{COO}-$  asymmetric vibration of carboxylic groups while that at  $1384 \text{ cm}^{-1}$  is due to stretching vibration of  $-\text{CH}_3$  group. Finally, the vibration band centered at  $1115 \text{ cm}^{-1}$  is attributed to C–O



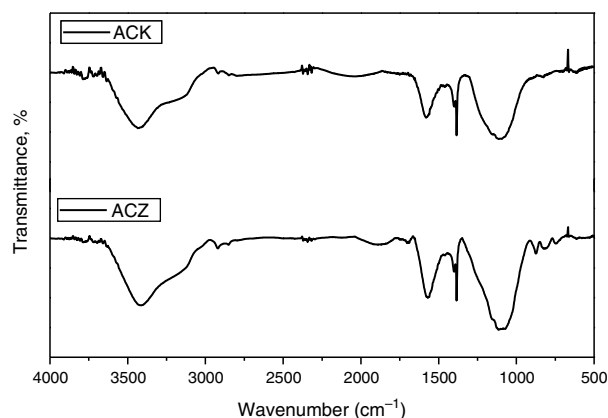


Fig. 4. The FTIR spectra of ACK and ACZ samples before adsorption in the range 4000–500  $\text{cm}^{-1}$ .

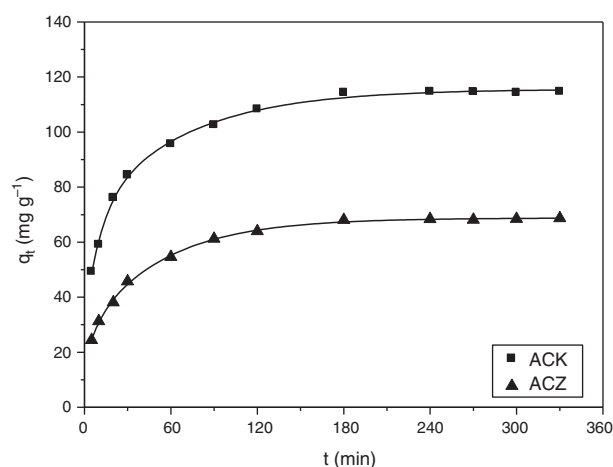


Fig. 5. Effect of contact time on the L-phenylalanine adsorption onto ACK and ACZ (L-phenylalanine concentration =  $200 \text{ mg L}^{-1}$ , adsorbent dose =  $1 \text{ g L}^{-1}$ , agitation speed = 150 rpm, contact time = 300 min, temperature =  $20^\circ\text{C}$ , natural pH).

stretching vibrations, as in alcohols, phenols, acids, ethers or esters. The presence of the functional groups such as carboxyl and hydroxyl are potential adsorption sites for L-phenylalanine amino acid.

### 3.2. Adsorption studies

#### 3.2.1. Effect of contact time

The contact time is a fundamental parameter in any transfer phenomena such as adsorption. The equilibrium adsorption capacity of L-phenylalanine on activated carbon was investigated to determine the time required to reach the equilibrium between adsorbents (50 mg) and L-phenylalanine solution, ( $200 \text{ mg L}^{-1}$ ) (Fig. 5); it can be observed that the adsorption capacities of activated carbons gradually increase with the contact time and does not stop until an equilibrium state is reached (180 min). No obvious variation in the amount of adsorbed L-phenylalanine was observed; the adsorbed mass at equilibrium reflects the maximum adsorption capacity of the activated carbons under the operating conditions; the equilibrium adsorption capacity of L-phenylalanine on ACK ( $114 \text{ mg g}^{-1}$ ) is higher than ACZ

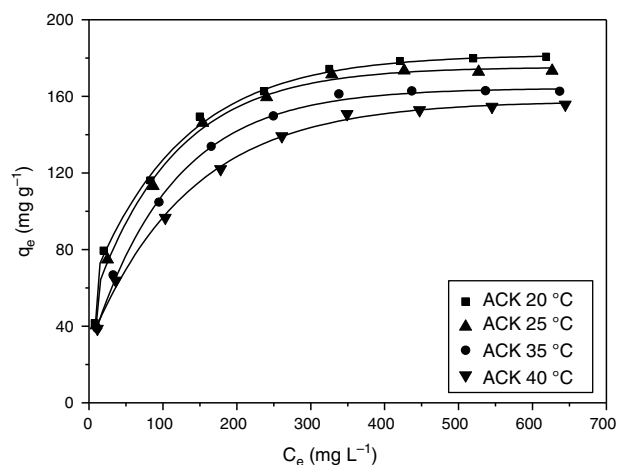
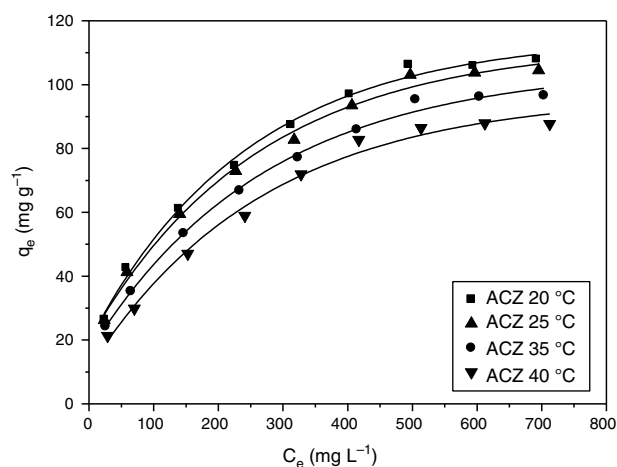


Fig. 6. Effect of temperature on L-phenylalanine adsorption onto ACK and ACZ (adsorbent dose =  $1 \text{ g L}^{-1}$ , agitation speed = 150 rpm, contact time = 300 min, natural pH).

( $68 \text{ mg g}^{-1}$ ) (Fig. 5). Comparing the results obtained in this work (porous activated carbons) with mesoporous materials, such as the SBA-3 mesoporous silica tested elsewhere (Goscianska, Olejnik, & Pietrzak, 2013a), it can be concluded that the mesoporous materials need a much longer period to reach the equilibrium; in this case the pore size distribution of ACK and ACZ, centered at 1.48 and 1.59 nm, respectively, are beneficial for the adsorption because L-phenylalanine molecules have easy access to the pores.

#### 3.2.2. Temperature effect

The temperature has a direct influence on the adsorption of amino acids. Figure 6 shows the temperature effect on the adsorption of L-phenylalanine by activated carbons. The same behavior is observed for ACK and ACZ and the isotherms plotted at various temperatures show that the equilibrium adsorption capacity decreases with increasing temperature from 20 to  $40^\circ\text{C}$ , indicating that the adsorption of L-phenylalanine is of exothermic nature. These results show that the decrease of adsorption at a high temperature can be ascribed to the greater tendency of L-phenylalanine molecules to form hydrophobic bonds in an aqueous medium, thus hindering their hydrophobic interactions with the adsorbent surface (El Shafei & Moussa, 2001).

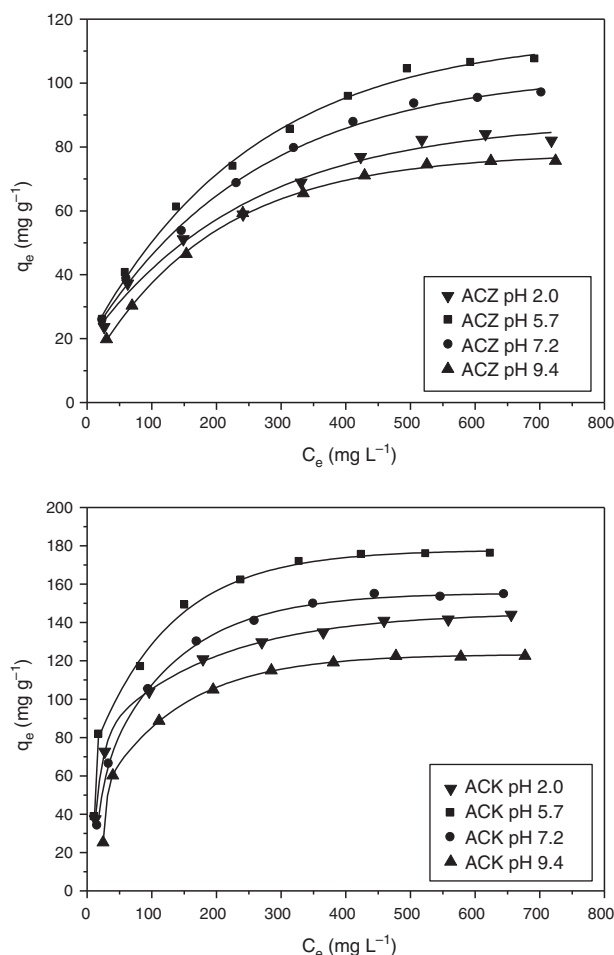


Fig. 7. Effect of pH on L-phenylalanine adsorption onto ACK and ACZ (adsorbent dose = 1 g L<sup>-1</sup>, agitation speed = 150 rpm, contact time = 300 min, temperature = 20 °C).

On the other hand, the decreased adsorption at equilibrium is due to decreased surface activity at higher temperatures. The best uptake of L-phenylalanine was obtained at 20 °C which is selected as an optimal adsorption temperature and will be used for further experiments.

### 3.2.3. Effect of pH

pH is known to be a crucial parameter that affects the adsorption behavior at water–solid interfaces. Its effect on the amino acid adsorption on ACK and ACZ at different buffer solutions ranging from 2.0 to 9.4 at 20 °C is illustrated in Figure 7. All equilibrium isotherms are of the L-type (Langmuir isotherms) and the amount of adsorbed L-phenylalanine increases with raising the initial concentration. At a low concentration, the amino acid is randomly deposited on the adsorbent, and the fast uptake can be attributed to a large number of empty sites on the surface (Goscianska et al., 2014). In contrast, at high concentrations, the nonpolar groups of amino acids are close to each other until they touch inside their van der Waals radii, leading to a dense packing of molecules on the active sites of the adsorbent. The removal of L-phenylalanine from an aqueous solution is strongly dependent on pH (Fig. 7) and this can be explained by pH<sub>pzc</sub>. pH<sub>pzc</sub> (Fig. 8) is found to be 6.18 (ACZ) and 6.80 (ACK). The surface

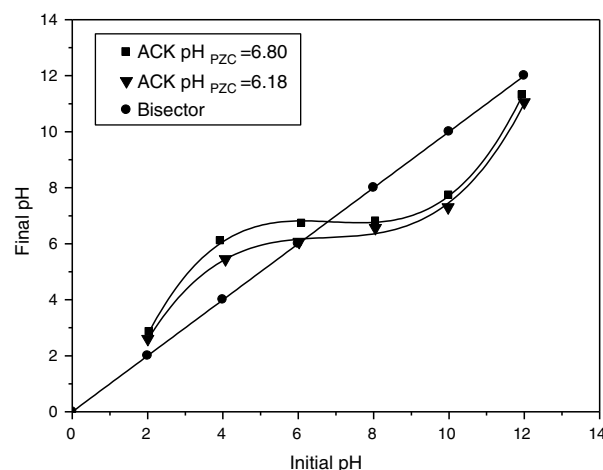


Fig. 8. Determination of the pH of zero point charge (PZC) of ACZ and ACK.

of the activated carbon is positively charged below pH<sub>pzc</sub> and negatively charged above pH<sub>pzc</sub>. The maximum adsorption capacity of both activated carbons was obtained at pH 5.7, which is close to the isoelectric point (PI = 5.48) of L-phenylalanine. The latter is known as a Zwitterion containing both amine and carboxylic groups, near to the isoelectric point and presents both negative and positive charges (Jiao et al., 2012). Therefore, the Coulomb repulsive interaction between the L-phenylalanine molecules is almost negligible. Furthermore, the strong hydrophobic interactions amino acid/adsorbents, and intra-molecular interaction between amino acid molecules are responsible of close packing of L-phenylalanine in the micropores of adsorbents, leading to the highest adsorption capacity at this pH. At low pH (<2), the surface charge of the activated carbon is negative and the cationic form of the amino acid is not favorable for the adsorption because of the electrostatic repulsion; this explains the decrease in the adsorption efficiency. Figure 7 also shows that the adsorbed amount of L-phenylalanine decreases with increasing pH from 5.7 to 9.4 and this can be explained by the strong electrostatic repulsion adsorbent/amino acid molecules, negatively charged. This effect is similar to that reported previously (Alves et al., 2013b; Goscianska et al., 2013c); as consequence, the adsorption of L-phenylalanine is inhibited above and below pH 5.7. Based on the experimental results, pH 5.7 was selected as an optimum value.

### 3.2.4. Adsorption kinetic study

Several kinetic models were proposed to understand the behavior of adsorbents and to study the mechanisms controlling the adsorption. In this study, the experimental data of L-phenylalanine adsorption are examined using a pseudo-first and pseudo-second order kinetic model. The pseudo first-order kinetic model is expressed in its linear form by the following equation (He et al., 2015):

$$\ln(q_e - q_t) = \ln q_e - k_1 t \quad (4)$$

Table 2  
Kinetic parameters for L-phenylalanine adsorption on ACK and ACZ.

Model	Parameters	Adsorbent	
		ACK	ACZ
Pseudo-first order	$q_{e,exp}$ (mg g <sup>-1</sup> )	114.36	68.03
	$\eta$ (%)	56.89	33.85
	$q_{e,cal}$ (mg g <sup>-1</sup> )	62.49	45.01
	$k_1$ (min <sup>-1</sup> )	0.0194	0.0205
	$R^2$	0.982	0.995
Pseudo-second order	$\Delta q$ (%)	30.63	23.00
	$q_{e,cal}$ (mg g <sup>-1</sup> )	119.05	72.46
	$k_2$ (g mg <sup>-1</sup> min <sup>-1</sup> )	0.0007	0.0009
	$R^2$	0.999	0.999
	$\Delta q$ (%)	2.65	4.00

While the pseudo second-order kinetic model is based on the assumption of a chemisorption of the adsorbate on the adsorbent (He et al., 2015):

$$\frac{t}{q_t} = \frac{1}{k_2 q_e^2} + \frac{t}{q_e} \quad (5)$$

where  $q_t$  and  $q_e$  (mg g<sup>-1</sup>) are the amount of amino acid adsorbed at time  $t$  (min) and at equilibrium, respectively,  $k_1$  (min<sup>-1</sup>) and  $k_2$  (g mg<sup>-1</sup> min<sup>-1</sup>) are the rate constant of the pseudo first order and pseudo second order adsorption models.

The best fit was validated on the base of the correlation coefficient ( $R^2$ ), the difference between the experimental and theoretical adsorption capacities and the normalized standard deviation  $\Delta q$  (%) (Sen Gupta & Bhattacharyya, 2011):

$$\Delta q(\%) = \sqrt{\frac{\sum [(q_{e,exp} - q_{e,cal})/q_{e,exp}]^2}{n - 1}} 100 \quad (6)$$

where  $n$  is the number of data points,  $q_{e,exp}$  and  $q_{e,cal}$  are the experimental and calculated equilibrium adsorption capacity values (mg g<sup>-1</sup>), respectively.

The aim of this kinetic study was to find the appropriate model that better describes the experimental data and that determines the kinetic parameters of the mass transfer of L-phenylalanine. The data were examined by using Eqs. (4)–(6). The calculated kinetic parameters for L-phenylalanine adsorption on activated carbons are gathered in Table 2. The adsorption capacities calculated from the pseudo second-order model are very close to the experimental ones as evidenced from the low values  $\Delta q$  (%) and high correlation coefficient ( $R^2 > 0.99$ ). Although the  $R^2$  values for the pseudo first-order model are all above 0.98 (Table 2), the eminent variances (the relative error of L-phenylalanine onto ACK and ACZ are 45.35% and 33.84%, respectively) between the experimental and calculated adsorption capacities reflect the poor fitting of the pseudo first-order model. Hence, L-phenylalanine adsorption kinetics on both ACZ and ACK is well described by the pseudo second-order kinetics.

### 3.2.5. Adsorption isotherms

The adsorption isotherm is generally applied to analyze the experimental data at equilibrium. The isotherms were further investigated by performing batch adsorption experiments over

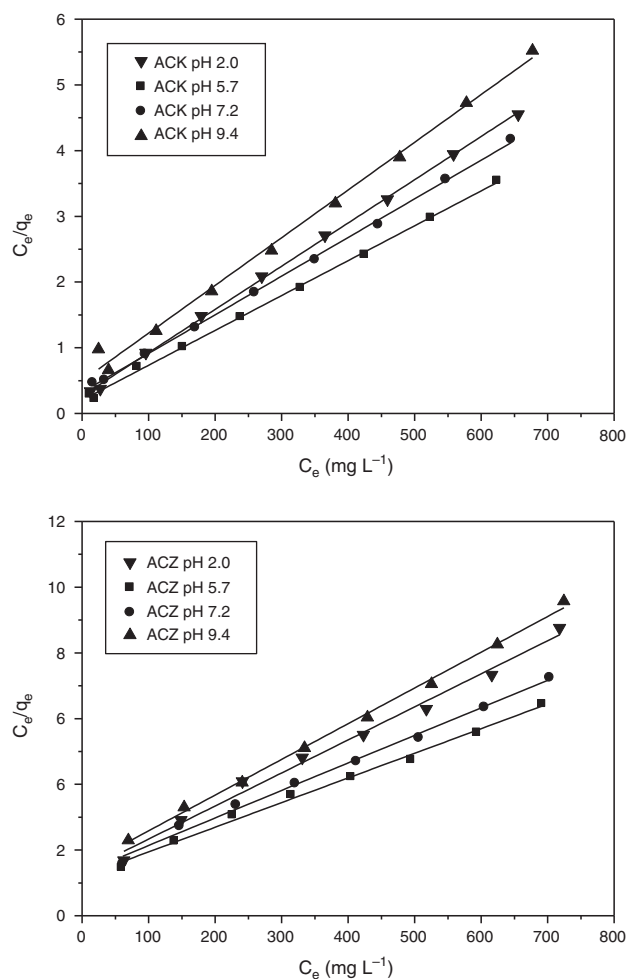


Fig. 9. Langmuir isotherm for adsorption of L-phenylalanine on ACK and ACZ at different values of pH (adsorbent dose = 1 g L<sup>-1</sup>, agitation speed = 150 rpm, contact time = 300 min, temperature = 20 °C).

the pH range (2–9.4) at the optimal temperature of 20 °C. The curves were fitted by the most used models namely the Langmuir and Freundlich ones. The Langmuir model is based on the assumption that the maximum adsorption corresponds to a saturated monolayer of adsorbate molecules on homogeneous sites of adsorbent with a constant energy, and no interaction between adsorbed species. The linear form is expressed as follows (Yang, Yu, & Chen, 2015):

$$\frac{C_e}{q_e} = \frac{1}{q_{max} K_L} + \frac{C_e}{q_{max}} \quad (7)$$

where  $q_{max}$  and  $q_e$  are the equilibrium and maximum adsorption capacities (mg g<sup>-1</sup>), respectively;  $K_L$ , the Langmuir constant related to the affinity of the binding sites (L mg<sup>-1</sup>); and  $C_e$ , the equilibrium concentration of adsorbate in an aqueous phases (mg L<sup>-1</sup>). The values of  $q_{max}$  and  $K_L$  are calculated from the slopes ( $1/q_{max}$ ) and intercept ( $1/q_{max} K_L$ ) of the linear plots of ( $C_e/q_e$ ) vs.  $C_e$  (Fig. 9).

The main characteristics of the Langmuir isotherm can be expressed by a dimensionless separation factor,  $R_L$ , which is



defined by the following formula (Liu, Zheng, Wang, Jiang, & Li, 2010):

$$R_L = \frac{1}{1 + K_L C_0} \quad (8)$$

The  $R_L$  value indicates the possibility of the adsorption process being favorable ( $0 < R_L < 1$ ), unfavorable ( $R_L > 1$ ), linear ( $R_L = 1$ ), or irreversible ( $R_L = 0$ ).

The Freundlich isotherm model is based on heterogeneous adsorbent surface (Yang et al., 2015):

$$\ln q_e = \ln K_F + \frac{1}{n} \ln C_e \quad (9)$$

The linear plot of  $\ln q_e$  vs.  $\ln C_e$  (Fig. 10) enables the determination of the Freundlich constants  $K_F$  and  $n$  from the intercept and slope, respectively.

The isotherm parameters of L-phenylalanine are calculated from the Langmuir and Freundlich models (Table 3), all the  $R^2$  values of the Langmuir model are greater than 0.99. These values are much higher than those of the Freundlich model, whatever the pH, suggesting the applicability of the Langmuir model which reveals a monolayer coverage of L-phenylalanine on homogeneous sites for both adsorbents. The  $R_L$  values are between 0 and 1, indicating that the L-phenylalanine adsorption on ACK and ACZ is favorable under the operating conditions. As the pH increases from 2.0 to 9.4,  $q_{\max}$  shows a significant decrease, reflecting that the adsorption is more favorable at pH 5.7, which is close to the isoelectric point ( $PI = 5.48$ ). ACK exhibits a maximum monolayer adsorption of L-phenylalanine ( $188.3 \text{ mg g}^{-1}$ ) compared to ACZ ( $133.3 \text{ mg g}^{-1}$ ) due to its largest microporous surface area. As the pore size of ACK is 1.48 nm, most pores including micropores are easily accessible to L-phenylalanine with a size of  $0.7 \times 0.5 \times 0.5 \text{ nm}^3$ . Consequently, the microporous surface area plays an important role for determining the adsorption capacity of the small biomolecule L-phenylalanine. The maximum adsorption capacities of ACK and ACZ for L-phenylalanine are compared to those values reported

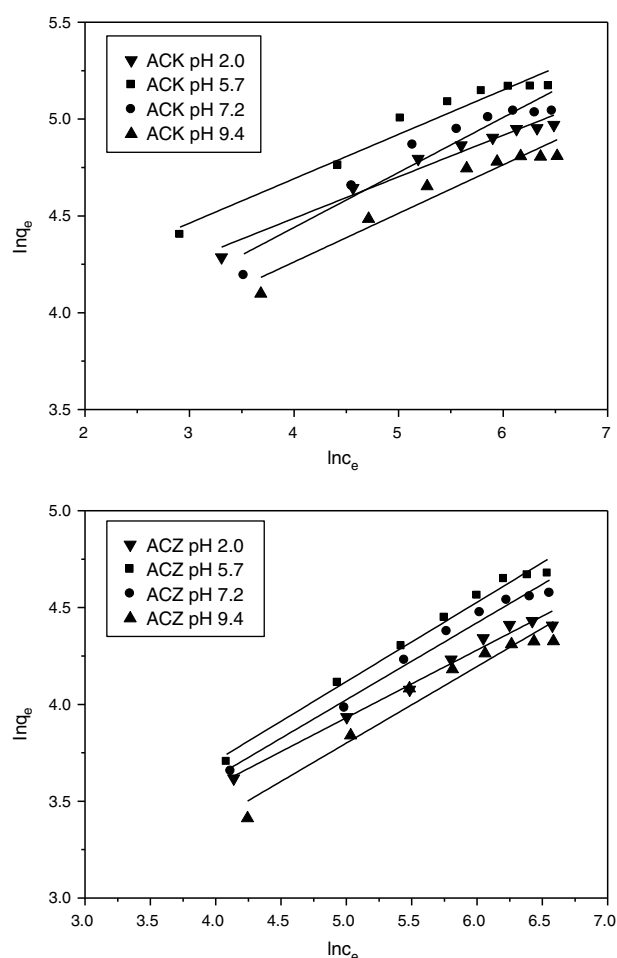


Fig. 10. Freundlich isotherm for adsorption of L-phenylalanine on ACK and ACZ at different values of pH (adsorbent dose =  $1 \text{ g L}^{-1}$ , agitation speed = 150 rpm, contact time = 300 min, temperature =  $20^\circ \text{C}$ ).

Table 3  
Isotherm parameters for L-phenylalanine adsorption on ACK and ACZ.

Model	Adsorbent	Parameters	pH			
			2.0	5.7	7.2	9.4
Langmuir	ACK	$q_{e,\text{exp}} (\text{mg g}^{-1})$	144.07	176.01	154.68	122.66
		$q_{\max} (\text{mg g}^{-1})$	151.97	188.32	170.06	137.74
		$k_L (\text{L mg}^{-1})$	0.0246	0.0262	0.0181	0.0146
		$R_L$	0.0483	0.0455	0.0646	0.0789
		$R^2$	0.999	0.998	0.998	0.993
	ACZ	$q_{e,\text{exp}} (\text{mg g}^{-1})$	84.01	107.45	96.94	75.65
		$q_{\max} (\text{mg g}^{-1})$	99.40	133.33	119.33	91.99
		$k_L (\text{L mg}^{-1})$	0.0076	0.0063	0.0065	0.0073
		$R_L$	0.1412	0.1655	0.1613	0.1462
		$R^2$	0.988	0.993	0.993	0.997
Freundlich	ACK	$k_F$	37.738	43.613	27.222	26.016
		$n$	4.667	4.364	3.520	3.986
		$R^2$	0.963	0.952	0.911	0.921
	ACZ	$k_F$	8.874	7.927	7.667	6.191
		$n$	2.860	2.443	2.516	2.529
		$R^2$	0.978	0.985	0.980	0.944

Table 4

Comparison of the maximum adsorption capacities ( $Q_m$ ) of various adsorbents for L-phenylalanine.

Adsorbent	$Q_m$ (mg g <sup>-1</sup> )	Reference
Polymeric adsorbent	115.6	(Grzegorzczuk and Carta, 1996)
Polymeric resins	65.9–100.8	(Díez et al., 1998)
Carbonated calcium phosphates	44.0	(Bihi et al., 2002)
NAZSM-5 zeolite	41.3	(Titus et al., 2003)
Commercial activated carbon	100.0	(Garnier et al., 2007)
Organic-inorganic hybrid membranes	1.2	(Wu et al., 2009)
Spherical carbon aerogels	66.1	(Long et al., 2009)
Macroporous resins	12.8–84.0	(Mei et al., 2009)
Activated defective coffee beans	69.5	(Clark et al., 2012)
Calcined CuZnAl-CO <sub>3</sub> layered double hydroxides	46.4	(Jiao et al., 2012)
Activated corn cobs	109.2	(Alves et al., 2013a, 2013b)
Mesoporous materials CSBA-15, CSBA-16 and CKIT-6	0.27–0.30	(Goscianska et al., 2013b)
Mesoporous silica	36.0–69.0	(Goscianska et al., 2013a)
Mesoporous carbon CMK-3	273.0	(Goscianska et al., 2014)
Multi-walled carbon nanotubes CNTs	233.0	(Goscianska et al., 2014)
Activated carbon ACK	188.3	This study
Activated carbon ACZ	133.3	This study

in the literature for other adsorbents (Table 4). In comparison with various adsorbents, our activated carbons ACK and ACZ have high adsorption capacities and can be considered as effective adsorbents for the recovery of L-phenylalanine from aqueous solutions.

### 3.2.6. Adsorption thermodynamics

A thermodynamic study was performed for the determination of the free energy change ( $\Delta G^\circ$ ), entropy ( $\Delta S^\circ$ ) and enthalpy ( $\Delta H^\circ$ ). A previous study of the temperature effect on L-phenylalanine adsorption over ACZ and ACK enabled us to determine the thermodynamic parameters at 20–40 °C and 800 mg L<sup>-1</sup>. They were calculated from the following equations (Bouguettoucha, Reffas, Chebli, Mekhalif, & Amrane, 2016; Milonjic, 2007):

$$K_C = \frac{q_e}{C_e} \quad (10)$$

$$\Delta G^\circ = -RT \ln(\rho K_C) \quad (11)$$

$$\ln(\rho K_C) = \frac{\Delta S^\circ}{R} - \frac{\Delta H^\circ}{RT} \quad (12)$$

where  $K_C$  is the equilibrium constant (L g<sup>-1</sup>),  $T$  the absolute temperature (K),  $R$  the universal gas constant (8.314 J mol<sup>-1</sup> K<sup>-1</sup>) and  $\rho$  the density of water (g L<sup>-1</sup>).  $\Delta H^\circ$  and  $\Delta S^\circ$  are calculated from the slope and intercept of the plots of  $\ln(\rho K_C)$  vs.

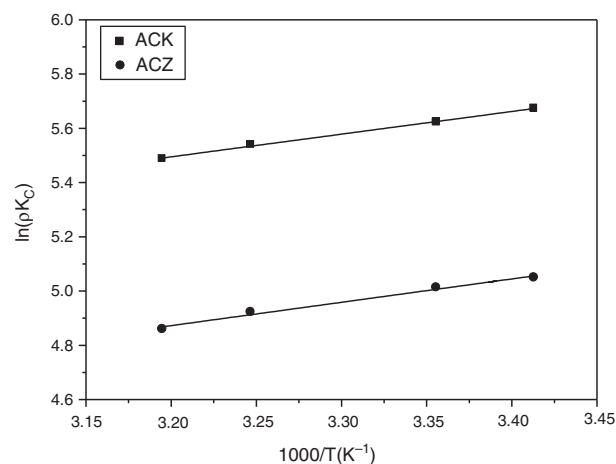


Fig. 11. Regressions of Van't Hoff for thermodynamic parameters of L-phenylalanine adsorption on ACK and ACZ.

$1/T$ , respectively (Fig. 11 and Table 5). The negative enthalpies ( $\Delta H^\circ$ ) indicate the exothermic nature of the adsorption, in agreement with the temperature effect study (see Section 3.2.2). It has been reported that  $\Delta H^\circ$  is in the range (2.1–20.9 kJ mol<sup>-1</sup>), indicating a physisorption (Liu, 2009).  $\Delta H^\circ$  (–6.947 and –7.153 kJ mol<sup>-1</sup> for ACK and ACZ, respectively) showed a physisorption of L-phenylalanine, with weak interactions while the negative free enthalpy ( $\Delta G^\circ$ ) indicates the spontaneous nature of L-phenylalanine uptake over the studied temperature range. The variation of  $\Delta G^\circ$  for physisorption is in the range (0–20.9 kJ mol<sup>-1</sup>), whereas this energy ranges from 80 to 200 kJ mol<sup>-1</sup> for a chemisorption (Liu, 2009). In our case,  $\Delta G^\circ$  (Table 5) is characteristic of a physical adsorption. The entropy ( $\Delta S^\circ$ ) is used to describe the randomness at the solid–solution interface during the recovery process. The positive values of  $\Delta S^\circ$  demonstrate an increase in randomness during the adsorption of L-phenylalanine on ACK and ACZ.

### 3.3. Proposed mechanism of adsorption

To further understand the adsorption behavior and select a desorption approach, the adsorption mechanism of L-phenylalanine amino acid was discussed. The adsorption mechanisms occurred mainly because of the hydrogen bonding formation, hydrophobic and electrostatic interactions of amino acid molecules with the activated carbons surface. The main active sites for binding of L-phenylalanine by the activated carbons are the hydroxyl and carboxyl groups on the surface of ACK and ACZ, which react with polar molecules and various functional groups. The surface of porous activated carbon can include electrically charged groups (ACZ/ACK surface –OH<sub>2</sub><sup>+</sup>: below pH<sub>pzc</sub>) and (ACZ/ACK surface –O<sup>-</sup>: above pH<sub>pzc</sub>), electrically neutral groups (ACZ/ACK surface –OH: near pH<sub>pzc</sub>). L-phenylalanine amino acid has dissociation constants (pK<sub>1</sub> = 1.83 and pK<sub>2</sub> = 9.13) and isoelectric point (PI = 5.48) (Jiao et al., 2012). The molecule is positively charged (<sup>+</sup>NH<sub>3</sub>–R–COOH) for pH < PI and negatively charged (NH<sub>2</sub>–R–COO<sup>-</sup>) for pH > PI,

Table 5

Thermodynamic parameters for L-phenylalanine adsorption on ACZ and ACK.

Adsorbent	$\Delta H^\circ$ (kJ mol <sup>-1</sup> )	$\Delta S^\circ$ (kJ mol <sup>-1</sup> K <sup>-1</sup> )	$\Delta G^\circ$ (kJ mol <sup>-1</sup> )			
			293 K	298 K	303 K	313 K
ACK	-6.947	0.019	-13.820	-13.930	-14.184	-14.280
ACZ	-7.153	0.017	-12.300	-12.418	-12.604	-12.646

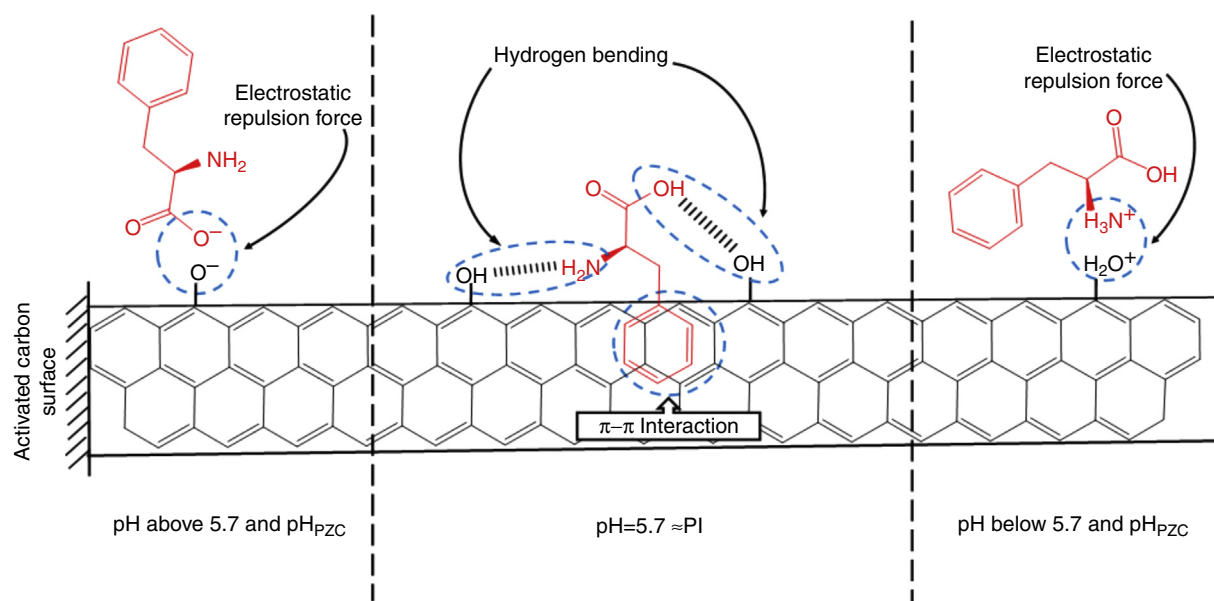


Fig. 12. Proposed adsorption mechanism of L-phenylalanine onto ACZ and ACK adsorbents.

and behaves in an aqueous medium as a dipolar zwitterion ( $^+\text{NH}_3\text{-R-COO}^-$ ) at pH-PI. In acid solution, the presence of  $\text{H}_3\text{O}^+$  ions in the surface of ACZ and ACK causes repulsion of protonated amino groups with the surface functional groups, and thus lower the adsorption efficiency. In a basic solution,  $\text{OH}^-$  ions present on the adsorbent surface compete with anionic carboxylic groups for L-phenylalanine molecules (repulsion effect) and inhibit the adsorption. The highest uptake of L-phenylalanine at pH 5.7 indicates a dominant hydrophobic interaction with  $\pi$ - $\pi$  type between the phenyl rings of amino acid molecules and graphene rings of the activated carbons surface (Doulia, Rigas, & Gimouhopoulos, 2001; Rajesh, Majumder, Mizuseki, & Kawazoe, 2009). Electrostatic attraction between anionic carboxylic groups of L-phenylalanine molecules and  $\text{OH}^-$  ions in the surface of activated carbons also accounts for the increased adsorption. Additionally, such strong bindings between L-phenylalanine and the adsorbent surface can be explained by the formation of hydrogen binding between oxygenated groups at the activated carbons surface and amino groups of L-phenylalanine. According to these results, the adsorption mechanism is proposed in Figure 12.

#### 3.4. Desorption behavior of L-phenylalanine from activated carbons

Regeneration and reuse of adsorbents for further cycles is important from the economic perspective. Desorption of L-phenylalanine from the activated carbons was evaluated using two different eluents: NaOH and HCl (0.01 M). The highest desorption was achieved in the NaOH solution with almost 95.7% for ACZ and 88.8% for CAK against 21 and 6.5% in the HCl solution. This may be due to the enhancement of the number of negatively charged sites at high pH which increases the electrostatic repulsion, which liberates L-phenylalanine from ACK and ACZ. To check the adsorption efficiency, the desorbed ACK and ACZ were dried overnight and subjected to a new adsorption/desorption cycle. During the second cycle, the adsorption capacities obtained were 29.4 (ACK) and 23.5 mg g<sup>-1</sup> (ACZ). A significant decay in the adsorption capacity of both activated carbons was observed and may be attributed to the depletion of active sites of the adsorbents being occupied by the amino acid. With the increase of the repeated cycle, the rate of desorption was also greatly decreased (Fig. 13).

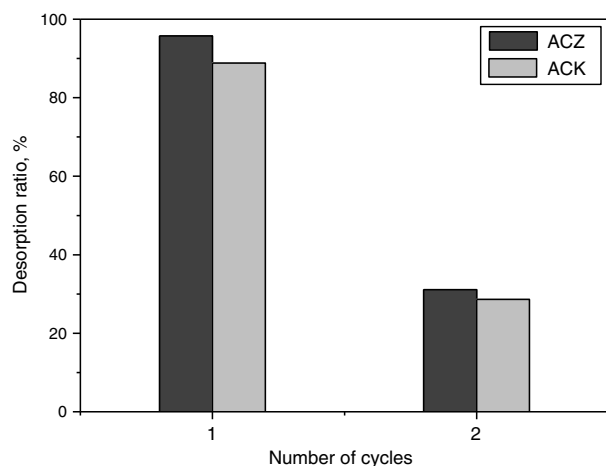


Fig. 13. Desorption ratio (%) of L-phenylalanine from ACZ and ACK adsorbents using  $0.01 \text{ mol L}^{-1}$  NaOH after two cycles.

#### 4. Conclusions

Porous activated carbons, namely ACK and ACZ, were successfully synthesized using the chemical activation method from agricultural wastes (date stones), which were later characterized using various analytical techniques such as SEM, FT-IR and  $\text{N}_2$  adsorption–desorption isotherms. The prepared activated carbons showed well-developed textural characteristics, with high BET surface areas, large pore volumes and tight pore size distribution. The pseudo second-order model was more suitable to describe the adsorption kinetics. The results showed that the temperature range ( $20\text{--}40^\circ\text{C}$ ) and pH ( $2\text{--}9.4$ ) exhibited remarkable influences on the adsorption of L-phenylalanine on activated carbons. The adsorption equilibrium data were well described by the Langmuir model, suggesting homogeneous adsorption. The maximum adsorption capacity was obtained at pH 5.7, which can be ascribed to the hydrogen bonding formation, hydrophobic and electrostatic interactions. The thermodynamic analysis indicated that L-phenylalanine adsorption was spontaneous, exothermic in nature and followed a physisorption mechanism. For desorption of the L-phenylalanine from the activated carbons, NaOH and HCl solutions with same concentration of  $0.01 \text{ M}$  were used. The desorption of the amino acid was better in the NaOH solution. Because of the high adsorption efficiency and desorption possibility, the present activated carbons from date palm seed wastes could be successfully applied as low-cost adsorbents in amino acid purification and separation.

#### Conflict of interest

The authors have no conflicts of interest to declare.

#### Acknowledgments

This work was financially supported by the Faculty of Chemistry (USTHB, Algiers). The authors gratefully acknowledge Pr. Y. Boucheffa and Pr. Z. Benabdelghani for their technical assistance.

#### References

- Alves, C. C., Franca, A. S., & Oliveira, L. S. (2013a). Evaluation of an adsorbent based on agricultural waste (corn cobs) for removal of tyrosine and phenylalanine from aqueous solutions. *BioMed Research International*, 978256.
- Alves, C. C., Franca, A. S., & Oliveira, L. S. (2013b). Removal of phenylalanine from aqueous solutions with thermo-chemically modified corn cobs as adsorbents. *LWT-Food Science and Technology*, 51(1), 1–8.
- Angin, D. (2014). Production and characterization of activated carbon from sour cherry stones by zinc chloride. *Fuel*, 115, 804–811.
- Bagheri, N., & Abedi, J. (2009). Preparation of high surface area activated carbon from corn by chemical activation using potassium hydroxide. *Chemical Engineering Research and Design*, 87(8), 1059–1064.
- Bihi, N., Bennani-Ziatni, M., Taitai, A., & Lebugle, A. (2002). Adsorption d'acides aminés sur des phosphates de calcium carbonatés analogues aux tissus calcifiés. *Annales de Chimie Science des Matériaux*, 27(2), 61–70.
- Bouchenafa-Saib, N., Grange, P., Verhasselt, P., Addoun, F., & Dubois, V. (2005). Effect of oxidant treatment of date pit active carbons used as Pd supports in catalytic hydrogenation of nitrobenzene. *Applied Catalysis A: General*, 286(2), 167–174.
- Bouguettoucha, A., Reffas, A., Chebli, D., Mekhalif, T., & Amrane, A. (2016). Novel activated carbon prepared from an agricultural waste. Stipa tenacissima, based on  $\text{ZnCl}_2$  activation—characterization and application to the removal of methylene blue. *Desalination and Water Treatment*, 1–14.
- Bourke, S. L., & Kohn, J. (2003). Polymers derived from the amino acid L-tyrosine: Polycarbonates, polyarylates and copolymers with poly (ethylene glycol). *Advanced Drug Delivery Reviews*, 55(4), 447–466.
- Brunauer, S., Emmett, P. H., & Teller, E. (1938). Adsorption of gases in multimolecular layers. *Journal of the American Chemical Society*, 60(2), 309–319.
- Casado, C., Castán, J., Gracia, I., Yus, M., Mayoral, A., Sebastián, V., et al. (2012). L- and D-proline adsorption by chiral ordered mesoporous silica. *Langmuir*, 28(16), 6638–6644.
- Chandrasekaran, M., & Bahkali, A. H. (2013). Valorization of date palm (Phoenix dactylifera) fruit processing by-products and wastes using bioprocess technology – Review. *Saudi Journal of Biological Sciences*, 20(2), 105–120.
- Clark, H. M., Alves, C. C., Franca, A. S., & Oliveira, L. S. (2012). Evaluation of the performance of an agricultural residue-based activated carbon aiming at removal of phenylalanine from aqueous solutions. *LWT-Food Science and Technology*, 49(1), 155–161.
- Deng, H., Yang, L., Tao, G., & Dai, J. (2009). Preparation and characterization of activated carbon from cotton stalk by microwave assisted chemical activation—application in methylene blue adsorption from aqueous solution. *Journal of Hazardous Materials*, 166(2), 1514–1521.
- Díez, S., Leitão, A., Ferreira, L., & Rodrigues, A. (1998). Adsorption of phenylalanine onto polymeric resins: equilibrium, kinetics and operation of a parametric pumping unit. *Separation and Purification Technology*, 13(1), 25–35.
- Douliat, D., Rigas, F., & Gimouhopoulos, C. (2001). Removal of amino acids from water by adsorption on polystyrene resins. *Journal of Chemical Technology and Biotechnology*, 76(1), 83–89.
- El Shafei, G. M. (2002). Effect of pore structure on surface characteristics of zirconium phosphate-modified silica. *Journal of Colloid and Interface Science*, 250(2), 394–399.
- El Shafei, G. M., & Moussa, N. A. (2001). Adsorption of some essential amino acids on hydroxyapatite. *Journal of Colloid and Interface Science*, 238(1), 160–166.
- Foo, K. Y., & Hameed, B. H. (2011). Preparation of activated carbon from date stones by microwave induced chemical activation: Application for methylene blue adsorption. *Chemical Engineering Journal*, 170(1), 338–341.
- Garnier, C., Görner, T., Villieras, F., De Donato, P., Polakovič, M., Bersillon, J. L., et al. (2007). Activated carbon surface heterogeneity seen by parallel probing by inverse liquid chromatography at the solid/liquid interface and by gas adsorption analysis at the solid/gas interface. *Carbon*, 45(2), 240–247.



- Ghosh, S., Badruddoza, A. Z. M., Uddin, M. S., & Hidajat, K. (2011). Adsorption of chiral aromatic amino acids onto carboxymethyl- $\beta$ -cyclodextrin bonded Fe<sub>3</sub>O<sub>4</sub>/SiO<sub>2</sub> core-shell nanoparticles. *Journal of Colloid and Interface Science*, 354(2), 483–492.
- Goscianska, J., Nowicki, W., & Pietrzak, R. (2014). Physicochemical and sorption properties of multi-walled carbon nanotubes decorated with silver nanoparticles. *Chemical Engineering Journal*, 250, 295–302.
- Goscianska, J., Olejnik, A., & Pietrzak, R. (2013a). Adsorption of L-phenylalanine onto mesoporous silica. *Materials Chemistry and Physics*, 142(2), 586–593.
- Goscianska, J., Olejnik, A., & Pietrzak, R. (2013b). Comparison of ordered mesoporous materials sorption properties towards amino acids. *Adsorption*, 19(2–4), 581–588.
- Goscianska, J., Olejnik, A., & Pietrzak, R. (2013c). In vitro release of L-phenylalanine from ordered mesoporous materials. *Microporous and Mesoporous Materials*, 177, 32–36.
- Grzegorzczak, D. S., & Carta, G. (1996). Adsorption of amino acids on porous polymeric adsorbents—I. Equilibrium. *Chemical Engineering Science*, 51(5), 807–817.
- Guo, J., & Lua, A. C. (2000). Preparation of activated carbons from oil-palm-stone chars by microwave-induced carbon dioxide activation. *Carbon*, 38(14), 1985–1993.
- Han, M. H., & Yun, Y. S. (2007). Mechanistic understanding and performance enhancement of biosorption of reactive dyestuffs by the waste biomass generated from amino acid fermentation process. *Biochemical Engineering Journal*, 36(1), 2–7.
- Hartmann, M. (2005). Ordered mesoporous materials for bioadsorption and biocatalysis. *Chemistry of Materials*, 17(18), 4577–4593.
- He, J., Lin, R., Long, H., Liang, Y., & Chen, Y. (2015). Adsorption characteristics of amino acids on to calcium oxalate. *Journal of Colloid and Interface Science*, 454, 144–151.
- Hong, S. U., & Bruening, M. L. (2006). Separation of amino acid mixtures using multilayer polyelectrolyte nanofiltration membranes. *Journal of Membrane Science*, 280(1), 1–5.
- Huang, Y., Ma, E., & Zhao, G. (2015). Thermal and structure analysis on reaction mechanisms during the preparation of activated carbon fibers by KOH activation from liquefied wood-based fibers. *Industrial Crops and Products*, 69, 447–455.
- Infante, M. R., Pérez, L., Pinazo, A., Clapés, P., Morán, M. C., Angelet, M., et al. (2004). Amino acid-based surfactants. *Comptes Rendus Chimie*, 7(6–7), 583–592.
- Jiao, F. P., Fu, Z. D., Shuai, L., & Chen, X. Q. (2012). Removal of phenylalanine from water with calcined CuZnAl-CO<sub>3</sub> layered double hydroxides. *Transactions of Nonferrous Metals Society of China*, 22(2), 476–482.
- Kostova, A., & Bart, H. J. (2007). Preparative chromatographic separation of amino acid racemic mixtures: I. Adsorption isotherms. *Separation and Purification Technology*, 54(3), 340–348.
- Liu, Q. S., Zheng, T., Wang, P., Jiang, J. P., & Li, N. (2010). Adsorption isotherm, kinetic and mechanism studies of some substituted phenols on activated carbon fibers. *Chemical Engineering Journal*, 157(2), 348–356.
- Liu, Y. (2009). Is the free energy change of adsorption correctly calculated? *Journal of Chemical & Engineering Data*, 54(7), 1981–1985.
- Long, D., Zhang, R., Qiao, W., Zhang, L., Liang, X., & Ling, L. (2009). Biomolecular adsorption behavior on spherical carbon aerogels with various mesopore sizes. *Journal of Colloid and Interface Science*, 331(1), 40–46.
- Lopes, D. C. F., Delvivo, F. M., & Silvestre, M. P. C. (2005). Use of activated carbon for removing phenylalanine from reconstituted skim milk powder hydrolysates. *LWT-Food Science and Technology*, 38(5), 447–453.
- Lua, A. C., & Yang, T. (2005). Characteristics of activated carbon prepared from pistachio-nut shell by zinc chloride activation under nitrogen and vacuum conditions. *Journal of Colloid and Interface Science*, 290(2), 505–513.
- Mei, J., Min, H., & Lü, Z. (2009). Enhanced biotransformation of L-phenylalanine to 2-phenylethanol using an in situ product adsorption technique. *Process Biochemistry*, 44(8), 886–890.
- Merzougui, Z., & Addoun, F. (2008). Effect of oxidant treatment of date pit activated carbons application to the treatment of waters. *Desalination*, 222(1), 394–403.
- Milonjić, S. K. (2007). A consideration of the correct calculation of thermodynamic parameters of adsorption. *Journal of the Serbian Chemical Society*, 72(12), 1363–1367.
- O'Connor, A. J., Hokura, A., Kisler, J. M., Shimazu, S., Stevens, G. W., & Komatsu, Y. (2006). Amino acid adsorption onto mesoporous silica molecular sieves. *Separation and Purification Technology*, 48(2), 197–201.
- Oshima, T., Saisho, R., Ohe, K., Baba, Y., & Ohto, K. (2009). Adsorption of amino acid derivatives on calixarene carboxylic acid impregnated resins. *Reactive and Functional Polymers*, 69(2), 105–110.
- Outinen M. T., Tossavainen O., Harju M., & Linko P. (1996). Method for removing phenylalanine from proteinaceous compositions, a product so obtained and use thereof. US Patents 5547687.
- Palit, D., & Moulik, S. P. (2001). Adsorption behaviors of L-histidine and DL-tryptophan on cholesterol, silica, alumina, and graphite. *Journal of Colloid and Interface Science*, 239(1), 20–26.
- Pereira, R. G., Veloso, C. M., da Silva, N. M., de Sousa, L. F., Bonomo, R. C. F., de Souza, A. O., et al. (2014). Preparation of activated carbons from cocoa shells and siriguela seeds using H<sub>3</sub>PO<sub>4</sub> and ZnCl<sub>2</sub> as activating agents for BSA and  $\alpha$ -lactalbumin adsorption. *Fuel Processing Technology*, 126, 476–486.
- Pimentel, F. B., Alves, R. C., Costa, A. S., Fernandes, T. J., Torres, D., Almeida, M. F., et al. (2014). Nutritional composition of low protein and phenylalanine-restricted dishes prepared for phenylketonuric patients. *LWT-Food Science and Technology*, 57(1), 283–289.
- Pimentel, F. B., Alves, R. C., Costa, A. S., Torres, D., Almeida, M. F., & Oliveira, M. B. P. (2014). Phenylketonuria: Protein content and amino acids profile of dishes for phenylketonuric patients. The relevance of phenylalanine. *Food Chemistry*, 149, 144–150.
- Prahas, D., Kartika, Y., Indraswati, N., & Ismadji, S. (2008). Activated carbon from jackfruit peel waste by H<sub>3</sub>PO<sub>4</sub> chemical activation: Pore structure and surface chemistry characterization. *Chemical Engineering Journal*, 140(1), 32–42.
- Rajesh, C., Majumder, C., Mizuseki, H., & Kawazoe, Y. (2009). A theoretical study on the interaction of aromatic amino acids with graphene and single walled carbon nanotube. *The Journal of Chemical Physics*, 130(12), 124911.
- Saka, C. (2012). BET, TG-DTG, FT-IR, SEM, iodine number analysis and preparation of activated carbon from acorn shell by chemical activation with ZnCl<sub>2</sub>. *Journal of Analytical and Applied Pyrolysis*, 95, 21–24.
- Sánchez-Hernández, L., Bernal, J. L., del Nozal, M. J., & Toribio, L. (2016). Chiral analysis of aromatic amino acids in food supplements using sub-critical fluid chromatography and Chirobiotic T2 column. *The Journal of Supercritical Fluids*, 107, 519–525.
- Sebben, D., & Pendleton, P. (2015). (Amino acid + silica) adsorption thermodynamics: Effects of temperature. *The Journal of Chemical Thermodynamics*, 87, 96–102.
- Sen Gupta, S., & Bhattacharyya, K. G. (2011). Kinetics of adsorption of metal ions on inorganic materials: A review. *Advances in Colloid and Interface Science*, 162(1–2), 39–58.
- Shimamura S., Tamura Y., Miyakawa H., Saito H., Kawaguchi Y., Isomura N., Akazome Y., Ochi H., & Kawamoto M. (2002). *Peptide mixture and products thereof*. US Patents 6395508.
- Silvério, F., Dos Reis, M. J., Tronto, J., & Valim, J. B. (2008). Adsorption of phenylalanine on layered double hydroxides: Effect of temperature and ionic strength. *Journal of Materials Science*, 43(2), 434–439.
- Sreńscek-Nazzal, J., Kamińska, W., Michalkiewicz, B., & Koren, Z. C. (2013). Production, characterization and methane storage potential of KOH-activated carbon from sugarcane molasses. *Industrial Crops and Products*, 47, 153–159.
- Titus, E., Kalkar, A. K., & Gaikar, V. G. (2003). Equilibrium studies of adsorption of amino acids on NaZSM-5 zeolite. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 223(1), 55–61.
- Wu, H., Zhao, Y., Nie, M., & Jiang, Z. (2009). Molecularly imprinted organic-inorganic hybrid membranes for selective separation of



- phenylalanine isomers and its analogue. *Separation and Purification Technology*, 68(1), 97–104.
- Yang, J., Yu, M., & Chen, W. (2015). Adsorption of hexavalent chromium from aqueous solution by activated carbon prepared from longan seed: Kinetics, equilibrium and thermodynamics. *Journal of Industrial and Engineering Chemistry*, 21, 414–422.
- Yorgun, S., Vural, N., & Demiral, H. (2009). Preparation of high-surface area activated carbons from Paulownia wood by  $\text{ZnCl}_2$  activation. *Microporous and Mesoporous Materials*, 122(1), 189–194.
- Zhou, H., Liao, X., Wang, T., Du, G., & Chen, J. (2010). Enhanced L-phenylalanine biosynthesis by co-expression of pheA<sub>fabr</sub> and aroF<sub>wt</sub>. *Bioresource Technology*, 101(11), 4151–4156.
- Zhu, X. L., Wang, P. Y., Peng, C., Yang, J., & Yan, X. B. (2014). Activated carbon produced from paulownia sawdust for high-performance  $\text{CO}_2$  sorbents. *Chinese Chemical Letters*, 25(6), 929–932.