



Journal of the Mexican Chemical Society

ISSN: 1870-249X

editor.jmcs@gmail.com

Sociedad Química de México

México

Zaky, M.; El-Sayedl, Mohamed Y.; El-Megharbel, Samy M.; Abo Taleb, Sameh; Refat, Moamen S.
Synthesis, Chemical Structure Elucidation and Biological Studies on the Effect of Some Vital Metal
Ions on Lisinopril

Journal of the Mexican Chemical Society, vol. 58, núm. 2, 2014, pp. 142-151

Sociedad Química de México

Distrito Federal, México

Available in: <http://www.redalyc.org/articulo.oa?id=47531764009>

- How to cite
- Complete issue
- More information about this article
- Journal's homepage in redalyc.org

redalyc.org

Scientific Information System

Network of Scientific Journals from Latin America, the Caribbean, Spain and Portugal

Non-profit academic project, developed under the open access initiative

Synthesis, Chemical Structure Elucidation and Biological Studies on the Effect of Some Vital Metal Ions on Lisinopril

M. Zaky,¹ Mohamed Y. El-Sayed^{1,2}, Samy M. El-Megharbel,^{1,3} Sameh Abo Taleb,¹ and Moamen S. Refat^{3,4}

¹ Department of Chemistry, Faculty of Science, Zagazig University, Egypt

² Faculty of Applied Medical Science, Al Jouf University-Al Qurayate

³ Department of Chemistry, Faculty of Science, Taif University, 888 Taif, Kingdom Saudi Arabia

⁴ Department of Chemistry, Faculty of Science, Port Said, Port Said University, Egypt msrefat@yahoo.com

Received October 28th, 2013; Accepted February 12th, 2014

Abstract. Complexes of lisinopril as a pharmaceutical ligand with Ca(II), Mg(II), Zn(II), Fe(III) and VO(II), were synthesized and characterized by microanalysis, conductance, infrared and thermogravimetric (TGA/DTG and DTA) measurements. The lisinopril ligand acts as a monodentate feature via one carboxylate oxygen atom. Lisinopril and their complexes have been checked against some kinds of bacteria and fungi which gave a significant effect. The kinetic thermodynamic parameters such as: activation of E^* , ΔH^* , ΔS^* and ΔG^* were estimated using Coats and Redfern as well as Horowitz-Metzger equations.

Key words: Lisinopril complexes, Infrared spectra, electronic spectra, thermal analysis and antimicrobial activity.

Resumen. En este trabajo se sintetizaron los complejos de lisinopril como ligando farmacéutico con iones Ca(II), Mg(II), Zn(II), Fe(III) and VO(II). Los complejos obtenidos fueron caracterizados por microanálisis, mediante mediciones de conductividad, espectros de absorción IR y termogravimetría (TGA/DTG y DTA). El lisinopril actúa como el ligando monodentado mediante el átomo de oxígeno del grupo carboxilo. La actividad biocida del lisinopril y sus complejos fue demostrada mediante ensayos con diferentes cepas de bacterias y micro hongos. Se estimaron parámetros cinéticos y termodinámicos tales como activación de E^* , ΔH^* , ΔS^* y ΔG^* , utilizando ecuaciones de Coats y Redfern y también de Horowitz-Metzger.

Palabras clave: Complejos de lisinopril, espectros de infrarrojo, espectros electrónicos, termoanálisis, actividad antimicrobiana.

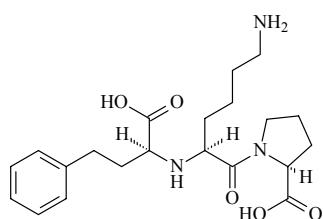
Introduction

Metal ions are required for many critical functions in humans [1-5]. Scarcity of some metal ions can lead to disease. Well-known examples include pernicious anemia resulting from iron deficiency, growth retardation arising from insufficient dietary zinc, and heart disease in infants owing to copper deficiency [1]. Metals and metal complexes have played key role in the development of modern chemotherapy [6]. For example, anti-cancer platinum drugs appear in more chemotherapy regimens than any other class of anticancer agents and have contributed substantially to the success achieved in treating cancer over the past three decades. This is allowing the drug to be released in a controlled fashion or at specific location [7]. This approach may lead to the rescue of drugs that have failed because of poor pharmacology or high toxicity. For example, complexation of non-steroidal anti-inflammatory drugs to copper overcomes some of the gastric side effects of these drugs [8]. The release of cytotoxins such as nitrogen mustards from redox-active metals such as cobalt in the hypoxic regions of solid tumors has the potential to improve drug activity and reduce toxicity [9]. The metal based drugs are also being used for the treatment of a variety of ailments viz. diabetes, rheumatoid arthritis, inflammatory and cardiovascular diseases as well as diagnostic [10-12]. A number of drugs and potential pharmaceutical agents also contain metal-binding or metal-recognition sites, which were influence on their bioactivities [13-24]. Numerous examples of these metallodrugs and metallopharmaceuticals and their actions can be found in the literature, for instance: (a) several

anti-inflammatory drugs, such as aspirin and its metabolite salicylglycine [13-16], suprofen [17], and paracetamol [18]; (b) the potent histamine-H₂-receptor antagonist cimetidine [19] can form complexes with Cu²⁺ and Fe³⁺, and the histidine blocker antiulcer drug famotidine can also form stable complex with Cu²⁺ [20, 21]; (c) the anthelmintic and fungistatic agent thia-bendazole, which is used for the treatment of several parasitic diseases, forms a Co²⁺ complex of 1:2 metal to drug ratio [22] (d) the Ru²⁺ complex of the anti-malaria agent chloroquine exhibits an activity two to five times higher than the parent drug against drug-resistant strains of *Plasmodium falciparum* [23]. However, it is known that some drugs act as potential ligands, a lot of studies are being carried out to ascertain how metal binding influences the activities of the drugs [24]. Metal-organic frameworks are not only stems from their tremendous potential applications in areas such as catalysis, molecular adsorption, magnetism, nonlinear optics, and molecular sensing, but also from their novel topologies and intriguing structural diversities [25-28]. On the other hand, many organic drugs, which possess modified pharmacological and toxicological properties administered in the form of metallic complexes [29], have the potential to act as ligands and the resulting metal-drug complexes are particularly important both in coordination chemistry and biochemistry [30-34] however, the study of metal-drug complexes is still in its early stages, thus representing a great challenge in current synthetic chemistry and coordination chemistry.

Lisinopril *N*²-[(1*S*)-1-carboxy-3-phenylpropyl]-L-lysyl-L-proline is a lysine analog of enalaprilat. It is a long-acting, nonsulphydryl angiotensin-converting enzyme inhibitor (For-

mula 1), treatment of hypertension and congestive heart failure in daily dosages of 10-80 mg [35]. Pharmacological activity of lisinopril has been proved in previous experimental and clinical studies [36-37]. Historically, lisinopril was the third angiotensin-converting enzyme inhibitor (after captopril and enalapril) and was introduced into therapy in the early 1990. A number of properties distinguish it from other angiotensin-converting enzyme inhibitors: It is hydrophilic, has a long half-life and tissue penetration, and is not metabolized by the liver. Successful clinical trials in essential hypertension, renovascular hypertension and congestive heart failure have been conducted with lisinopril. Lisinopril is not significantly metabolized in humans; the absorbed drug is primarily excreted unchanged in urine. Peak serum concentrations of lisinopril are reached in about 6 h after administration. Mean maximum serum concentrations of lisinopril are about 80-140 ng/mL after a single oral dose of 20 mg [38]. Its indications, contraindications and side effects are as those for all angiotensin-converting enzyme inhibitors. Lisinopril is a subject of monograph in the United States Pharmacopoeia, which recommends an HPLC method for its analysis [39]. A variety of analytical techniques have been developed for the determination of lisinopril in pharmaceutical preparations, such as HPLC [40], GC [41], capillary electrophoresis [42-43], spectrophotometry [44-45], spectrofluorimetric [40], micellar electrokinetic chromatography [46] and polarographic [45]. Techniques have been described for determination of lisinopril in biologic fluids including GC [47], HPLC [48], LC-MS [49], fluoroimmunoassay [50], radioimmunoassay [51], fluorozmatic assay [52] and ion selective electrodes [53].



Formula 1. Structure of lisinopril (Lis).

Lisinopril possesses are poor in electromagnetic absorbance due to weak benzene chromophore; as a consequence, poor sensitivity can be achieved by UV spectrophotometric method. Therefore, most of these assays employ pre-column derivatization reactions to enhance the determination sensitivity. However, the preparation of derivatives is a time consuming procedure, and the derivatization frequently generates products that are unstable. Moreover, peak splitting owing to slow cis-trans isomerization may exist in the RP-HPLC [54]. GC and LC with mass spectrometric detection were widely applied to determination in biological fluids. In the literature survey, the synthesis and characterization of anti-hypertensive drugs complexes with some metal ions have been described [55-57]. The scope of this article is to discuss the coordination behavior of some of vital metal ions like Ca(II), Mg(II), Zn(II), Fe(III) and VO(II) with one of common blood pressure drug as lisinopril. The antibacterial and antifungal activities of lisinopril and its complexes were also evaluated.

Results and Discussion

The reactions of lisinopril (Lis) with the metal ions Ca(II), Mg(II), Zn(II), Fe(III) and VO(II) gave a colored solid complexes in moderate to good yields (60-80%). The physical and analytical data, colors, percentage (carbon, hydrogen and nitrogen) and melting/decomposition temperatures of the compounds are presented in Table 1. The found and calculated percentages of elemental analysis CHN are in a well agreement with each other and prove the suggested molecular formula of the resulted lisinopril complexes. The complexes have high melting points (226-266) °C. The lisinopril ligand behaves as monodentate ligand and coordinates to the metal ions via one the oxygen atom of the carboxylate group. The isolated lisinopril complexes are 1:1 molar ratio of (M:Lis) where M=Ca(II), Mg(II), Zn(II), Fe(III) and VO(II).

Molar conductivities: The molar conductivity values for the Ca(II), Mg(II), Zn(II), Fe(III) and VO(II) complexes of lisinopril in DMSO solvent (1.00×10^{-3} M) were found to be in the range $5\text{-}27 \Omega^{-1}\text{cm}^2\text{mol}^{-1}$ at 25 °C, suggesting them to

Table 1. Elemental analysis and physical data of lisinopril complexes.

Complex	M. wt. (g/mol)	mp/°C	color	% C		% H		% N		% M		$\Delta m(\Omega^{-1}\text{cm}^2\text{mol}^{-1})$
				Calc.	found	Calc.	found	Calc.	found	Calc.	found	
[Ca(Lis)(Cl)(NH ₄)(H ₂ O) ₄].2H ₂ O (C ₂₁ H ₄₅ N ₄ O ₁₁ ClCa)	604.5	233	Yellow	41.69	42.23	7.44	7.25	9.26	9.49	6.62	6.60	22
[Mg(Lis)(Cl)(NH ₄)(H ₂ O) ₄].9H ₂ O (C ₂₁ H ₅₉ N ₄ O ₁₈ ClMg)	714.5	240	Yellow	35.27	35.22	8.26	8.28	7.84	7.06	3.36	3.24	27
[Zn(Lis)(SO ₄)(NH ₄)(H ₂ O) ₃].9H ₂ O (C ₂₁ H ₅₇ N ₄ O ₂₁ SZn)	798.4	234	Pale brown	31.56	31.35	7.14	7.10	7.01	7.88	8.19	8.04	7
[Fe(Lis)(NO ₃) ₂ (NH ₄)(H ₂ O)].H ₂ O (C ₂₁ H ₃₇ N ₆ O ₁₃ Fe)	637	266	Brown	39.56	40.28	5.81	5.47	13.19	13.60	8.79	8.51	5
[VO(Lis)(SO ₄)(NH ₄) ₂].4H ₂ O (C ₂₁ H ₄₅ N ₅ O ₁₃ SVO)	674	226	Dark green	37.39	36.63	6.68	6.53	10.39	10.78	9.94	9.87	8

be non-electrolytes [58] as shown in Table 1. Hence the molar conductance values indicate that no ions are present outside the coordination sphere so the Cl^- , SO_4^{2-} and NO_3^- ions may be exhibit inside the coordination sphere or absent. The obtained results were strongly matched with the qualitative analyses where Cl^- , SO_4^{2-} and NO_3^- ions are detected in case of Ca(II) , Mg(II) , Zn(II) , Fe(III) and VO(II) complexes after degradation of these complexes by using nitric acid then precipitation of Cl^- and SO_4^{2-} by addition of AgNO_3 and BaCl_2 solutions, respectively to the solutions of the mentioned complexes were detected by using infrared spectral data. The low conductivity values [59] are in agreement with the low solubility of the above complexes in water, alcohol, chloroform, acetone and most of organic solvents. On other hand, they are soluble in DMSO, DMF and concentrated acids.

Electronic absorption spectra of lisinopril complexes: Electronic spectra of the lisinopril complexes were recorded in the 200-900 nm regions in DMSO. There are three detected absorption bands at (225 and 270) and 320 nm in the electronic spectrum of the free lisinopril ligand, these bands assigned to $\pi-\pi^*$ and $n-\pi^*$ transitions, respectively. These transitions occur in case of unsaturated hydrocarbons, which contain carbon atom attached with oxygen atoms as in carboxylic and ketone groups [60]. These bands of the free lisinopril ligand are bathochromically affected (red shifted) clearly in the electronic spectra of the Fe(III) and VO(II) complexes which show absorption bands at (259 and 325 shoulder) nm for Fe(III) complex and at (257 and 326 shoulder) nm for VO(II) complex which assigned to $\pi-\pi^*$ and $n-\pi^*$ transitions, respectively. This bathochromically change in the spectra of Fe(III) and VO(II) complexes indicated the association metal-to-ligand chelations. These results are clearly in accordance with the results of FT-IR and $^1\text{H-NMR}$ spectra.

Infrared spectra of lisinopril complexes: The infrared spectral data of lisinopril and its complexes under investigation are recorded in Table 2. The spectra are similar but there are some differences which could give indication on the type of coordination. The infrared spectra of the free lisinopril show

two broad bands observed at 3554 and 3396 cm^{-1} , these bands are assigned due to the $\nu_{\text{as}}(\text{OH})$ and $\nu_{\text{s}}(\text{OH})$ stretching vibration of the -OH group. The infrared spectral bands of lisinopril complexes have a broad band's observed at 3400-3416 cm^{-1} , these bands are assigned to the $\nu(\text{OH})$ stretching vibration of the coordinated H_2O molecules. The IR spectrum of lisinopril free ligand show strong absorption band at 1675 cm^{-1} due to $\nu(\text{C=O})$ stretching vibration of the carboxylic group [61]. The Ca(II) , Mg(II) , Zn(II) , Fe(III) and VO(II) complexes show no absorption band at 1675 cm^{-1} $\nu(\text{C=O})$, that is indicative of the deprotonation of COOH group and involvement of the carboxyl group in the formation of M-O bonds [62]. The spectra of the complexes show two characteristic bands at (1404-1187), (1409-1187), (1400-1107), (1415-1155) and (1400-1105) cm^{-1} for Ca(II) , Mg(II) , Zn(II) , Fe(III) and VO(II) complexes respectively, assigned as $\nu(\text{COO}^-)$ asymmetric and symmetric stretching vibrations of the ligated carboxylate anion respectively. The criteria that can be used to distinguish between the three binding states of the carboxylate complexes have been studied previously by Deacon and Phillips [63]. These criteria are: (a) $\Delta\nu > 200 \text{ cm}^{-1}$ (where $\Delta\nu = [\nu_{\text{as}}(\text{COO}^-) - \nu_{\text{s}}(\text{COO}^-)]$) this relation was found in case of monodentate carboxylate complexes, (b) bidentate or chelating carboxylate complexes exhibit $\Delta\nu$ significantly smaller than ionic values ($\Delta\nu < 100 \text{ cm}^{-1}$), and finally, (c) bridging complexes show $\Delta\nu$ comparable to ionic values ($\Delta\nu \sim 150 \text{ cm}^{-1}$). Therefore, the difference value $\Delta\nu$ is a useful characteristic for determining the coordination mode of the carboxylate group of the ligands. The observed $\Delta\nu$ for Ca(II) , Mg(II) , Zn(II) , Fe(III) and VO(II) complexes (Table 2) fall in the range 217-295 cm^{-1} indicating a monodentate coordination mode of the carboxylate group [64, 65]. The spectrum of lisinopril free ligand show absorption bands at 3100 and 3260 cm^{-1} due to $\nu(\text{NH})$ stretching vibration of -NH and $-\text{NH}_2$ groups respectively, these peaks are found in the spectra of the metal complexes with no significant change in their intensity and position. This observation suggests that both -NH and $-\text{NH}_2$ groups haven't involved in the coordination. The $\nu(\text{V=O})$ stretching vibration in the vanadyl complex is observed as ex-

Table 2. IR frequencies (cm^{-1}) of lisinopril and its metal complexes.

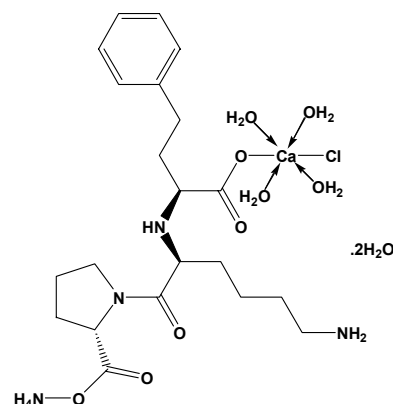
Compound	$\nu(\text{O-H});$ H_2O	$\nu(\text{NH});$ NH_2 and NH	$\nu(\text{C=O})$ COOH	$\Delta(\text{NH}_2)$	$\nu_{\text{as}}(\text{COO}^-)$	$\nu_{\text{s}}(\text{COO}^-)$	$\Delta\nu$	$\nu(\text{M-O})$	$\nu(\text{M-N})$
lisinopril	—	3265 3100	1675	1653	—	—	—	—	—
$[\text{Ca}(\text{Lis})(\text{Cl})(\text{NH}_4)(\text{H}_2\text{O})_4] \cdot 2\text{H}_2\text{O}$	3416	3260 3106	—	1632	1404	1187	217	551	490
$[\text{Mg}(\text{Lis})(\text{Cl})(\text{NH}_4)(\text{H}_2\text{O})_4] \cdot 9\text{H}_2\text{O}$	3407	3250 3099	—	1636	1409	1187	222	604	432
$[\text{Zn}(\text{Lis})(\text{SO}_4)(\text{NH}_4)(\text{H}_2\text{O})_3] \cdot 9\text{H}_2\text{O}$	3400	3247 3105	—	1608	1400	1107	293	615	428
$[\text{Fe}(\text{Lis})(\text{NO}_3)_2(\text{NH}_4)(\text{H}_2\text{O})] \cdot \text{H}_2\text{O}$	3415	3270 3107	—	1632	1415	1155	260	550	463
$[\text{VO}(\text{Lis})(\text{SO}_4)(\text{NH}_4)_2] \cdot 4\text{H}_2\text{O}$	3409	3210 3101	—	1618	1400	1105	295	615	490

pected band at 960 cm^{-1} , which is a good agreement with those known for many vanadyl complexes [66]. The coordination of nitrate group for the Fe(III) ions were also supported by check the IR spectrum of the ferric(III) complex, where the nitrate complex displayed two stretching vibration bands at 1350 cm^{-1} and 1100 cm^{-1} assigned to $\nu_{\text{as}}(\text{NO}_2)$ and $\nu_{\text{s}}(\text{NO}_2)$ respectively, the stretching motion of $\nu(\text{N}=\text{O})$ is observed at 1415 cm^{-1} as a very strong band, while the two bending motion of the type $\Delta(\text{NO}_2)$ are well resolved and observed at 790 and 697 cm^{-1} suggesting that bidentate nitrate group [67, 68]. Clearly the test against the presence of sulfate group in the VO(II) and Zn(II) complexes gave a positive result, this conclusion was supported by detected the two infrared frequencies bands at about 1100 and 600 cm^{-1} overlapping with angular deformation motions of the coordinated water molecules. New vibrating absorption bands were observed in the range of $(550\text{--}615)\text{ cm}^{-1}$ and $(428\text{--}490)\text{ cm}^{-1}$ are assigned to the stretching absorption bands of M-O and M-N bands, respectively [69]. According to the IR data, the lisinopril coordinated to the metal ions as monodentate ligand via the carboxylate oxygen atom [30].

¹H-NMR of lisinopril and its complexes: The ¹H-NMR data of free lisinopril and its Ca (II) complex, as an example are listed in Table (3). Upon comparison the free lisinopril ligand with its metal complex Ca(II), the significant change is the disappearance of the characteristic peak for hydrogen of -COOH at $\Delta = 11.0$ ppm in Ca(II) complex indicates that the coordination of lisinopril ligand to Ca(II) through the deprotonated carboxylic group [70]. On other hand there is no significant change in the characteristic peaks of hydrogen for -NH, -NH₂, -CH₂ and -CH groups of aromatic ring in the free lisinopril ligand and its Ca(II) complex. This discussion is supported that these groups are not involved in the complexation [70]. The appearance of new peaks at 3.65 and 3.76 ppm in the spectra of Ca(II) complex which can assigned to the coordinated and uncoordinated of H₂O molecules. In general, the structures of investigated complexes can be formulated as shown in Formulas (2-6).

Thermal analysis: The obtained lisinopril complexes were studied by thermogravimetric (TG), differential thermogravimetric (DTG) and (DTA) analysis from ambient temperature to 800°C under nitrogen atmosphere. The TG curves were redrawn as mg mass loss versus temperature and DTG curves were redrawn as rate of loss of mass versus temperature. The thermal decomposition results are summarized in Table 4.

[Ca(Lis)(Cl)(NH₄)(H₂O)₄].2H₂O complex: The thermal decomposition of Ca(II) complex of lisinopril with general formula [Ca(Lis)(Cl)(NH₄)(H₂O)₄].2H₂O is occurs at four steps. The first degradation step is in the range of $23\text{--}174^\circ\text{C}$ at $\text{DTG}_{\text{max}} = 61^\circ\text{C}$ and its assigned to the loss of $4(\text{H}_2\text{O})$ mol-



Formula 2. Suggested structures of Ca(II) complex of lisinopril.

ecules of hydration water with an observed weight loss 11.69% (calculated = 11.91%), the activation energy of this step is 25.3 kJmol^{-1} . The second step is occurred within a temperature range $174\text{--}342^\circ\text{C}$ at $\text{DTG}_{\text{max}} = 296^\circ\text{C}$ due to the loss of $2(\text{H}_2\text{O})$ molecules of coordinated water + $\text{NH}_4\text{Cl} + \text{C}_2\text{H}_6$ with a weight loss (Found = 20.57% , Calculated = 19.77%), the activation energy of this step is 75.9 kJmol^{-1} . The third step is occurred within a temperature range $342\text{--}483^\circ\text{C}$ at $\text{DTG}_{\text{max}} = 410^\circ\text{C}$ and $\text{DTA} = 445^\circ\text{C}$ (exo) due to the loss $\text{C}_{10}\text{H}_{18}\text{N}_2\text{O}_2$ (organic moiety) with an observed weight loss 33.07% (calculated = 32.75%), the activation energy of this step is 120 kJmol^{-1} . The fourth step is occurred within a temperature range $483\text{--}591^\circ\text{C}$ at $\text{DTG}_{\text{max}} = 538^\circ\text{C}$ and $\text{DTA} = 533^\circ\text{C}$ (exo) due to the loss of $\text{C}_3\text{H}_5\text{NO}_2$ (organic moiety) with an observed weight loss 13.99% (Calculated = 14.39%), the activation energy of this step is 216 kJmol^{-1} . The CaO + residual carbon atoms are remaining stable till 800°C as final residual.

[Mg(Lis)(Cl)(NH₄)(H₂O)₄].9H₂O complex: The thermal decomposition of Mg(II) complex of lisinopril with general formula [Mg(Lis)(Cl)(NH₄)(H₂O)₄].9H₂O is occurs at four steps. The first degradation step is in the range of $54\text{--}196^\circ\text{C}$ at $\text{DTG}_{\text{max}} = 92^\circ\text{C}$ and its assigned to the loss of $5.5(\text{H}_2\text{O})$ molecules of hydration water with an observed weight loss 14.13% (Calculated = 13.86%), the activation energy of this step is 37.1 kJmol^{-1} . The second step is occurred within a temperature range $196\text{--}333^\circ\text{C}$ at $\text{DTG}_{\text{max}} = 302^\circ\text{C}$ and $\text{DTA} = 277^\circ\text{C}$ (exo) assigned to the loss of $7.5(\text{H}_2\text{O})$ molecules of hydration water with a weight loss (Found = 17.86% , Calculated = 18.89%), the activation energy of this step is 82.2 kJmol^{-1} . The third step is occurred within a temperature range $333\text{--}460^\circ\text{C}$ at $\text{DTG}_{\text{max}} = 399^\circ\text{C}$ and $\text{DTA} = 332^\circ\text{C}$ (exo) assigned to the loss of $\text{NH}_4\text{Cl} + \text{C}_7\text{H}_{13}\text{N}$ (organic moiety) with an observed weight loss 23.39%

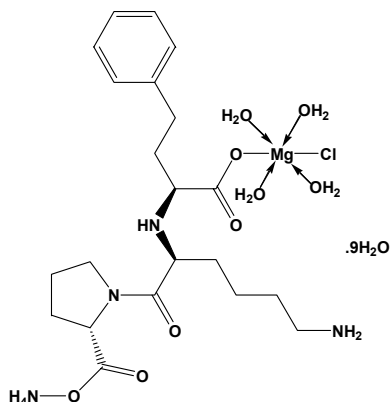
Table 3. ¹H-NMR spectral data of lisinopril and its Ca(II) complex.

Compound	δ ppm of hydrogen					
	H; -CH ₂	H; -NH ₂	H; -NH-	H; -CH aromatic	H; -OH ₂	H; -COOH
Lisinopril	1.55, 1.97, 2.20	2.0	2.6	7.12, 7.21	—	11.0
Ca(II) complex	1.44, 1.97, 2., 11	2.1	2.7	7.17, 7.27	3.65, 3.76	—

Table 4. Thermal data of lisinopril complexes.

Compound	Steps	TG, temp. range (°C)	DTG _{max} (°C)	DTA (°C)	TG mass loss (%)		Assignments
					Calcd.	Found	
[Ca(Lis)(Cl)(NH ₄)(H ₂ O) ₄].2H ₂ O (C ₂₁ H ₄₅ N ₄ O ₁₁ ClCa)	1	23-174	61	—	11.91	11.69	4H ₂ O
	2	174-342	296	—	19.77	20.57	2H ₂ O+NH ₄ Cl+C ₂ H ₆
	3	342-483	410	445 (exo)	32.75	33.07	C ₁₀ H ₁₈ N ₂ O ₂ (organic moiety)
	4	483-591	538	533 (exo)	14.39	13.99	C ₃ H ₅ NO ₂ (organic moiety)
Final residue = CaO + 6C (found = 20.68%, Calcd. = 21.17%)							
[Mg(Lis)(Cl)(NH ₄)(H ₂ O) ₄].9H ₂ O (C ₂₁ H ₅₉ N ₄ O ₁₈ ClMg)	1	54-196	92	—	13.86	14.13	5.5H ₂ O
	2	196-333	302	277 (exo)	18.89	17.86	7.5H ₂ O
	3	333-460	399	332 (exo)	23.02	23.39	NH ₄ Cl+ C ₇ H ₁₃ N (organic moiety)
	4	460-601	510	513 (exo)	30.23	30.83	C ₉ H ₁₆ N ₂ O ₄ (organic moiety)
Final residue = MgO + 5C (found = 13.79%, Calcd. = 13.99%)							
[Zn(Lis)(SO ₄)(NH ₄)(H ₂ O) ₃].9H ₂ O (C ₂₁ H ₅₇ N ₄ O ₂₁ SNZn)	1	30-127	81	82 (endo)	11.27	11.09	5H ₂ O
	2	198-303	275	—	27.68	27.26	7H ₂ O+C ₆ H ₆ +NH ₃ C ₅ H ₁₂ N (organic moiety)
	3	303-443	360	276 (exo)	10.77	10.91	H ₂ SO ₄ +C ₃ H ₆ N ₂ O ₄ (organic moiety)
	4	485-593	556	558 (exo)	29.06	28.49	CH ₄
	5	711-788	753	—	2.00	2.28	
Final residue = ZnO + 6C (found = 19.97%, Calcd. = 19.21%)							
[Fe(Lis)(NO ₃) ₂ (NH ₄)(H ₂ O)].H ₂ O (C ₂₁ H ₅₇ N ₆ O ₁₃ Fe)	1	43-138	73	—	4.23	3.86	1.5H ₂ O
	2	138-311	250	278 (exo)	36.26	37.26	½ H ₂ O+NH ₄ NO ₃ +C ₆ H ₈ NO ₃ (organic moiety)
	3	311-462	407	416 (exo)	38.77	38.59	C ₁₀ H ₂₁ N ₃ O ₄ (organic moiety)
Final residue = FeO + 5C (found = 20.29%, Calcd. = 20.72%)							
[VO(Lis)(SO ₄)(NH ₄) ₂].4H ₂ O (C ₂₁ H ₄₅ N ₅ O ₁₃ SVVO)	1	49-122	76	—	5.34	4.68	2H ₂ O
	2	122-322	237	257 (exo)	32.94	32.84	2H ₂ O+2NH ₃ +H ₂ SO ₄ +C ₃ H ₄ N (organic moiety)
	3	322-410	362	357 (endo)	14.54	14.88	C ₆ H ₁₀ O (organic moiety)
	4	410-528	476	478 (exo)	36.05	36.47	C ₁₀ H ₁₅ N ₂ O ₅ (organic moiety)
Final residue = V + 2C (found = 11.13%, Calcd. = 11.13%)							

exo=exothermic peak, endo=endothermic peak.



Formula 3. Suggested structures of Mg(II) complex of lisinopril.

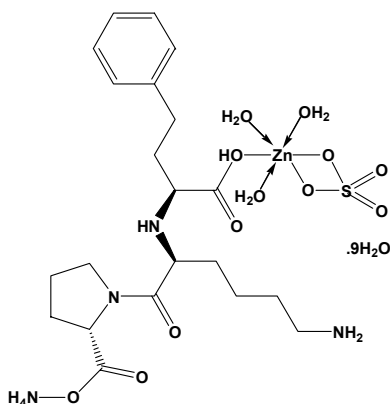
(calculated = 23.02%), the activation energy of this step is 108 kJmol⁻¹. The fourth step is occurred within a temperature range 460-601 °C at DTG_{max} = 510 °C and DTA = 513 °C (exo) due to the loss of C₉H₁₆N₂O₄ (organic moiety) with an observed weight loss 30.83% (calculated = 30.23%), the activation energy of this step is 188 kJmol⁻¹. The MgO residual carbon atoms are remaining stable till 800 °C as final residual.

[Zn(Lis)(SO₄)(NH₄)(H₂O)₃].9H₂O complex: The thermal decomposition of Zn(II) complex of lisinopril with general formula [Zn(Lis)(SO₄)(NH₄)(H₂O)₃].9H₂O is occurs at five steps. The first degradation step is in the range of 30-127 °C at DTG_{max} = 81 °C and DTA = 82 °C (exo) and its assigned to the loss of 5(H₂O) molecules of hydration water with an observed weight loss 11.09% (Calculated = 11.27%), the activation energy of this step is 55.3 kJmol⁻¹. The second step is occurred within a temperature range 198-303 °C at DTG_{max} = 275 °C assigned to the loss of another 7(H₂O) molecules of hydration water + C₆H₆ + NH₃ gas with a weight loss (Found = 27.26%, Calculated = 27.68%), the activation energy of this step is 109 kJmol⁻¹. The third step is occurred within a temperature range 303-443 °C at DTG_{max} = 360 °C and DTA = 276 °C (exo) assigned to the loss of C₅H₁₂N (organic moiety) with observed weight loss 10.91% (calculated = 10.77%), the activation energy of this step is 77.5 kJmol⁻¹. The fourth step is occurred within a temperature range

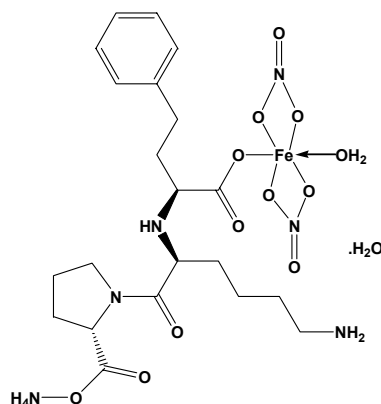
485-593 °C at DTG_{max} = 556 °C and DTA = 558 °C (exo) assigned to the loss of H₂SO₄ + C₃H₆N₂O₄ (organic moiety) with an observed weight loss 28.49% (calculated = 29.06%), the activation energy of this step is 296 kJmol⁻¹. The fifth step is occurred within temperature range 711-788 °C at DTG_{max} = 753 °C assigned to the loss of CH₄ with an observed weight loss 2.28% (calculated = 2.00%). The ZnO + residual carbon atoms are remaining stable till 800 °C as final residual.

[Fe(Lis)(NO₃)₂(NH₄)(H₂O)].H₂O complex: The thermal decomposition of Fe(III) complex of lisinopril with general formula [Fe(Lis)(NO₃)₂(NH₄)(H₂O)].H₂O is occurs at three steps. The first degradation step is in the range of 43-138 °C at DTG_{max} = 73 °C and its assigned to the loss of 1.5(H₂O) molecules of hydration water with an observed weight loss 3.86% (Calculated = 4.23%), the activation energy of this step is 49.9 kJ mol⁻¹. The second step is occurred within a temperature range 138-311 °C at DTG_{max} = 250 °C and DTA = 278 °C (exo) assigned to the loss of 1/2(H₂O) molecules of hydration water + NH₄NO₃ + C₆H₈NO₃ (organic moiety) with observed weight loss 37.26% (calculated = 36.26%), the activation energy of this step is 69.1 kJ mol⁻¹. The third step is occurred within a temperature range 311-462 °C at DTG_{max} = 407 °C and DTA = 416 °C (exo) assigned to the loss of C₁₀H₂₁N₃O₄ (organic moiety) with observed weight loss 38.59% (Calculated = 38.77%), the activation energy of this step is 111 kJ mol⁻¹. The FeO residual carbon atoms are remaining stable till 800 °C as final residual.

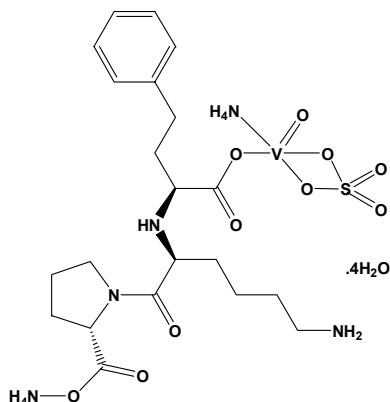
[VO(Lis)(SO₄)(NH₄)₂].4H₂O complex: The thermal decomposition of VO(II) complex of lisinopril with general formula [VO(Lis)(SO₄)(NH₄)₂].4H₂O is occurs at four steps. The first degradation step is in the range of 49-122 °C at DTG_{max} = 76 °C and it is assigned to the loss of 2(H₂O) molecules of hydration water with an observed weight loss 4.68% (calcd. = 5.34%), the activation energy of this step is 62.5 kJ mol⁻¹. The second step is occurred within a temperature range 122-322 °C at DTG_{max} = 237 °C and DTA = 257 °C (exo) assigned to the loss of another 2(H₂O) molecules of hydration + 2NH₃ gas + H₂SO₄ + C₃H₄N (organic moiety) with observed weight loss 32.84% (Calculated = 32.94%), the activation energy of this step is 39.6 kJ mol⁻¹. The third step is occurred within a temperature range



Formula 4. Suggested structures of Zn(II) complex of lisinopril.



Formula 5. Suggested structures of Fe(III) complex of lisinopril.



Formula 6. Suggested structures of VO(II) complex of lisinopril.

322-410 °C at $DTG_{max}=362$ °C and $DTA = 357$ °C (endo) assigned to the loss of $C_6H_{10}O$ (organic moiety) with observed weight loss 14.88% (Calculated = 14.54%), the activation energy of this step is 130 kJ mol^{-1} . The fourth step is occurred within a temperature range 410-528 °C at $DTG_{max} = 476$ °C and $DTA = 478$ °C (exo) assigned to the loss of $C_{10}H_{15}N_2O_5$ (organic moiety) with observed weight loss 36.47% (Calculated = 36.05%), the activation energy of this step is 219 kJ mol^{-1} . The V + residual carbon atoms are remaining stable till 800 °C as final residual.

Kinetic studies: In the present investigation, the general thermal behaviors of the lisinopril complexes in terms of stability ranges, maximum temperature peaks and values of kinetic parameters, are shown in Table (5). The kinetic and thermodynamic parameters are evaluated using the Coats-Redfern and Horowitz-Metzger equations [71-73]. The entropy of activation, ΔS^* , is calculated. The enthalpy activation, ΔH^* , and Gibbs free energy, ΔG^* , are calculated from $\Delta H^* = E^* - RT$ and $\Delta G^* = \Delta H^* - T\Delta S^*$, respectively. The thermodynamic behavior of most lisinopril complexes with Ca(II), Mg(II), Zn(II), Fe(III) and VO(II) metal ions is non-spontaneous (more ordered) reactions (ΔS is negative value), endothermic reactions ($\Delta H > 0$) and endergonic ($\Delta G > 0$), during the reactions. The thermodynamic data obtained with the two methods are in harmony with each other. The correlation coefficients of the Arrhenius plots of the thermal decomposition steps were found to lie in the range 0.9637-0.9996, showing a good fit with linear function. The thermograms and the calculated thermal parameters for the complexes show that the stability of these complexes depends on the nature of the central metal ion. The thermal stability of the metal complexes was found to increase periodically with increase in atomic number of the metal and the larger value of charge/radius ratio [74].

Microbiological investigation of lisinopril complexes: Antibacterial and antifungal activities of lisinopril complexes were carried out against of bacteria as *Escherichia coli* (Gram -ve) and *Staph albus* (Gram +ve) as well as fungi as *Aspergillus niger* and *Aspergillus flavus*. The antimicrobial activities based on the size of inhibition zone around dishes were estimated. The free lisinopril has the lowest activity against four types of

bacteria and fungi, while the Zn(II) complex was found to have the highest activity. The biological activities increase in the following order: Zn(II)/Lis > VO(II)/Lis > Fe(III)/Lis > Ca(II)/Lis > Mg(II)/Lis. The data are listed in Table 6.

Experimental part

Materials: All chemicals used in this investigation were of highest purity grade (Merck). Selected metal salts as $CaCl_2$, $MgCl_2 \cdot 6H_2O$, $ZnSO_4 \cdot H_2O$, $Fe(NO_3)_3 \cdot 9H_2O$ and $VOSO_4 \cdot H_2O$ were used. Lisinopril were received from Egyptian International Pharmaceutical Industrial Company.

Preparation of solid lisinopril complexes: $[Ca(Lis)(Cl)(NH_4)(H_2O)_4] \cdot 2H_2O$ (I), $[Mg(Lis)(Cl)(NH_4)(H_2O)_4] \cdot 9H_2O$ (II), $[Zn(Lis)(SO_4)(NH_4)(H_2O)_3] \cdot 9H_2O$ (III), $[Fe(Lis)(NO_3)_2(NH_4)(H_2O)] \cdot H_2O$ (IV) and $[VO(Lis)(SO_4)(NH_4)_2] \cdot 4H_2O$ (V) were prepared, employing a 1:1 (metal:Lis) ratio. The complexes were prepared by mixing equal volumes (20 mL) of distilled water solution of $CaCl_2$ (0.111 g, 1.0 mmol), $MgCl_2 \cdot 6H_2O$ (0.203g, 1.0 mmol), $ZnSO_4 \cdot H_2O$ (0.180 g, 1.0 mmol), $Fe(NO_3)_3 \cdot 9H_2O$ (0.404 g, 1.0 mmol) and $VOSO_4 \cdot H_2O$ (0.163 g, 1.0 mmol) with a methanol solution of lisinopril (0.405g, 1.0 mmol). The mixtures are neutralized at pH (7.0-9.0) using 5% alcoholic ammonia solution, then the mixtures are heating at about ~60 °C for about one hour with stirring and left to evaporate slowly at room temperature overnight. The obtained precipitates were filtered off, washed several times by minimum amount of hot methanol and dried at 60 °C over anhydrous $CaCl_2$. The melting points of the five lisinopril complexes are observed within the range of 226-266 °C. The Ca(II), Mg(II), Zn(II), Fe(II) and VO(II) complexes are soluble in dimethyl sulfoxide (DMSO) and N,N-dimethylformamide (DMF) with gently heating. The yield of lisinopril complexes is in the range of 60-75%.

Preparations of stock: Ten g of barium chloride dihydrate $BaCl_2 \cdot 2H_2O$ is weighted and dissolved in the least amount of distilled water. The volume was completed to 100 mL in a measuring flask to give 10% solution.

A weight of 0.1701 g of $AgNO_3$ was dissolved in 100 mL distilled water in a dark measuring flask to obtain an approximate 0.01 M solution.

The stock solution of NH_4OH was prepared by taking 15.15 mL of the concentrate NH_3 (33% v/v) in 35 mL distilled water. The volume was then completed to 100 mL by methanol to give approximately (5% v/v) solution.

A weight of 0.2 g of the solid lisinopril complex was dissolved in the least amount of 2 mol nitric acid. The solution was evaporated near dryness. This process is repeated twice and the residue was dissolved in about 50 mL of hot distilled water to obtain a clear solution. The volume was completed to 100 mL in a measuring flask.

Apparatus and experimental conditions: Carbon, hydrogen and nitrogen content were determined using a Perkin-Elmer CHN Elemental Analyzer model 2400. The metal content was found gravimetrically by converting the compounds into their corresponding carbides or oxides. The Ca(II), Mg(II), Zn(II),

Table 5. Thermodynamic parameters of the lisinopril complexes.

Complex	Stage	Method	Parameter					r
			E* (J mol ⁻¹)	A (s ⁻¹)	ΔS* (J mol ⁻¹ K ⁻¹)	ΔH* (J mol ⁻¹)	ΔG* (J mol ⁻¹)	
Ca(II)	1 st	CR	2.54E+04	1.95E+01	-2.21E+02	2.26E+04	9.65E+04	0.97071
		HM	2.52E+04	5.97E+01	-2.12E+02	2.25E+04	9.32E+04	0.96377
	2 nd	CR	6.56E+04	6.17E+03	-1.78E+02	6.09E+04	1.62E+05	0.99968
		HM	8.63E+04	6.73E+05	-1.39E+02	8.16E+04	1.61E+05	0.99612
	3 rd	CR	1.13E+05	2.35E+06	-1.30E+02	1.08E+05	1.96E+05	0.99721
		HM	1.27E+05	3.92E+07	-1.06E+02	1.21E+05	1.94E+05	0.99414
Mg(II)	4 th	CR	2.09E+05	1.83E+11	-3.76E+01	2.03E+05	2.33E+05	0.99376
		HM	2.24E+05	3.05E+12	-1.42E+01	2.18E+05	2.29E+05	0.99107
	1 st	CR	3.70E+04	4.10E+02	-1.97E+02	3.40E+04	1.06E+05	0.97315
		HM	3.72E+04	1.76E+03	-1.84E+02	3.42E+04	1.02E+05	0.97133
	2 nd	CR	7.11E+04	1.98E+04	-1.68E+02	6.63E+04	1.63E+05	0.99989
		HM	9.33E+04	2.52E+06	-1.28E+02	8.85E+04	1.62E+05	0.99814
	3 rd	CR	1.01E+05	3.63E+05	-1.45E+02	9.55E+04	1.93E+05	0.99529
		HM	1.15E+05	6.58E+06	-1.21E+02	1.09E+05	1.91E+05	0.99368
	4 th	CR	1.86E+05	1.18E+10	-6.02E+01	1.80E+05	2.27E+05	0.9958
		HM	1.90E+05	4.65E+10	-4.87E+01	1.84E+05	2.22E+05	0.99408
Zn(II)	1 st	CR	5.25E+04	4.87E+05	-1.37E+02	4.95E+04	9.82E+04	0.99396
		HM	5.81E+04	5.16E+06	-1.18E+02	5.52E+04	9.69E+04	0.98749
	2 nd	CR	9.86E+04	2.41E+07	-1.09E+02	9.41E+04	1.54E+05	0.9968
		HM	1.21E+05	4.38E+09	-6.54E+01	1.17E+05	1.53E+05	0.99506
	3 rd	CR	7.38E+04	4.15E+03	-1.82E+02	6.85E+04	1.84E+05	0.99118
		HM	8.13E+04	3.10E+04	-1.65E+02	7.60E+04	1.81E+05	0.99103
	4 th	CR	2.82E+05	8.52E+15	5.16E+01	2.75E+05	2.32E+05	0.98965
		HM	3.11E+05	5.27E+17	8.58E+01	3.04E+05	2.33E+05	0.99382
Fe(III)	1 st	CR	4.92E+04	7.89E+04	-1.52E+02	4.63E+04	9.91E+04	0.97195
		HM	5.07E+04	5.70E+05	-1.36E+02	4.78E+04	9.49E+04	0.97058
	2 nd	CR	6.08E+04	5.29E+03	-1.78E+02	5.64E+04	1.50E+05	0.99715
		HM	7.75E+04	4.61E+05	-1.41E+02	7.31E+04	1.47E+05	0.99231
	3 rd	CR	1.00E+05	4.16E+05	-1.44E+02	9.48E+04	1.93E+05	0.99587
		HM	1.22E+05	1.82E+07	-1.13E+02	1.16E+05	1.93E+05	0.9978
VO(II)	1 st	CR	6.06E+04	5.85E+06	-1.17E+02	5.77E+04	9.85E+04	0.98217
		HM	6.45E+04	7.09E+07	-9.59E+01	6.16E+04	9.51E+04	0.97997
	2 nd	CR	4.21E+04	5.00E+01	-2.17E+02	3.78E+04	1.48E+05	0.99562
		HM	3.71E+04	2.71E+01	-2.22E+02	3.29E+04	1.46E+05	0.99018
	3 rd	CR	1.27E+05	1.42E+08	-9.51E+01	1.21E+05	1.82E+05	0.99186
		HM	1.34E+05	9.74E+08	-7.91E+01	1.28E+05	1.79E+05	0.99196
	4 th	CR	2.10E+05	4.48E+12	-1.04E+01	2.04E+05	2.12E+05	0.99745
		HM	2.29E+05	1.06E+14	1.59E+01	2.22E+05	2.10E+05	0.99901

ΔS*: entropy of activation, ΔH*: enthalpy of activation, ΔG*: Gibbs free energy, E*: activation energy, A: pre-exponential factor and r = correlation coefficient of the linear plot.

(CR): Coats and Redfern and (HM): Horowitz-Metzger equations.

Fe(III) and VO(II) contents were determined gravimetrically by the direct ignition of the complexes at 800 °C for 3 hours till constant weight. The residue was then weighted in the forms of metal oxides. Molar conductivities of freshly prepared 1.0×10^{-3} mol DMSO solutions of the complexes were measured using Jenway 4010 conductivity meter. IR spectra were recorded on Bruker FTIR Spectrophotometer (4000-400 cm⁻¹) in KBr

pellets. The electronic spectra were measured in the DMSO solvent with concentration (1.0×10^{-3} M) for the free ligands and their complexes using Jenway 6405 spectrophotometer with 1 cm quartz cell, in the range 200-900 nm. ¹H-NMR spectra of the free ligands and their complexes were recorded on Varian Gemini 200 MHz Spectrophotometer using DMSO-d₆ as solvent and TMS as internal reference. Thermogravimetric analy-

Table 6. Antimicrobial data of lisinopril and its complexes.

Compound	Diameter of inhibition zone (cm)			
	<i>E. coli</i>	<i>Staph albus</i>	<i>Aspergillus niger</i>	<i>Aspergillus flavus</i>
Control	0	0	0	0
lisinopril	0	0	0	0
Ca(II)/Lis complex	0	0	0	0.4
Mg(II)/Lis complex	0	0	0	0.3
Zn(II)/Lis complex	0	0.2	0.3	0.5
Fe(III)/Lis complex	0	0	0	0.5
Vo(II)/Lis complex	0	0	0.2	0.4

sis (TGA, DTG and DTA) was carried out in the temperature range from 25 to 800 °C in a steam of nitrogen atmosphere by using Shimadzu TGA-50 H thermal analyzer. The experimental conditions were: platinum crucible, nitrogen atmosphere with a 30 mL/min flow rate and a heating rate of 10 °C/min.

Most commonly used methods are the differential method of Freeman and Carroll [71] integral method of Coat and Redfern [72] and the approximation method of Horowitz and Metzger [73]. In the present investigation, the general thermal behaviors of the lisinopril complexes in terms of stability ranges, peak temperatures and values of kinetic parameters are discussed. The kinetic parameters have been evaluated using the Coats-Redfern equation:

$$\int_0^\alpha \frac{d\alpha}{(1-\alpha)^n} = \frac{A}{\phi} \int_{T_1}^{T_2} \exp\left(-\frac{E^*}{RT}\right) dt \quad (1)$$

This equation on integration gives;

$$\ln\left[-\frac{\ln(1-\alpha)}{T^2}\right] = -\frac{E^*}{RT} + \ln\left[\frac{AR}{\phi E^*}\right] \quad (2)$$

Where ϕ is the linear heating rate, R is the gas constant, T is the DTG temperature peak, α , is the fraction of the sample decomposed at time t , A is the pre-exponential factor. A plot of left-hand side against $1/T$ was drawn. E^* is the energy of activation in J mol^{-1} and calculated from the slop and A in (s^{-1}) from the intercept value. The entropy of activation ΔS^* in $(\text{JK}^{-1}\text{mol}^{-1})$ was calculated by using the equation:

$$\Delta S^* = R \ln(Ah/k_B T_s) \quad (3)$$

Where k_B is the Boltzmann constant, h is the Plank's constant and T_s is the DTG peak temperature [75].

The Horowitz-Metzger equation is an illustrative of the approximation methods.

$$\log\{[1 - (1 - \alpha)^{1-n}]/(1 - n)\} = E^*\theta/2.303RT_s^2 \text{ for } n \neq 1 \quad (4)$$

When $n = 1$, the LHS of equation 4 would be $\log[-\log(1 - \alpha)]$. For a first-order kinetic process the Horowitz-Metzger equation may be written in the form:

$$\log[\log(w_\alpha / w_\gamma)] = E^*\theta/2.303RT_s^2 - \log 2.303$$

Where $\theta = T - T_s$, $w_\gamma = w_\alpha - w$, w_α = mass loss at the completion of the reaction. The plot of $\log[\log(w_\alpha / w_\gamma)]$ vs θ was drawn and found to be linear from the slope of which E^* was calculated. The pre-exponential factor, A , was calculated from the equation:

$$E^* / RT_s^2 = A / [\phi \exp(-E^*/RT_s)]$$

The entropy of activation, ΔS^* , was calculated from equation 3. The enthalpy activation, ΔH^* , and Gibbs free energy, ΔG^* , were calculated from; $\Delta H^* = E^* - RT$ and $\Delta G^* = \Delta H^* - T\Delta S^*$, respectively.

Microbiological investigation: According to Gupta et al. [76], the hole well method was applied. The investigated isolates of bacteria and fungi were seeded in tubes with nutrient broth (NB) and Dox's broth (DB), respectively. The seeded (NB) for bacteria and (DB) for fungi (1 mL) were homogenized in the tubes with 9 mL of melted (45 °C) nutrient agar (NA) for bacteria and (DA) for fungi. The homogenous suspensions were poured into Petri dishes. The holes (diameter 0.5 cm) were done in the cool medium. A 100 μL of the investigated compounds were taken using a micropipette. After incubation for 24 h in an incubator at 37 °C and 28 °C for bacteria and fungi, respectively, the inhibition zone diameter were measured and expressed in cm. The antimicrobial activities of the investigated compounds were tested against of bacteria as *Escherichia coli* (Gram -ve) and *Staph albus* (Gram +ve) as well as fungi as *Aspergillus flavus* and *Aspergillus niger*. At the same time the pure solvent was tested with the antimicrobial investigations of the complexes. The concentration of each solution was 1.0×10^{-3} mol/L. Commercial DMSO was employed to dissolve the tested samples.

References

1. Sekhon, B.S. J. Pharm. Educ. Res. **2011**, 2(1), 1-20.
2. Xiao, D.R.; Wang, E.B.; An, H.Y.; Su, Z.M.; Li, Y.G.; Gao, L.; Sun, C.Y.; Xu, L. Chem. Eur. J. **2005**, 11, 6673-6686.
3. Drevensek, P.; Zupancic, T.; Pihlar, B.; Jerala, R.; Kolitsch, U.; Plaper, A.; Turel, I. J. Inorg. Biochem. **2005**, 99, 432-442.
4. He, J.H.; Xiao, D.R.; Chen, H.Y.; Yan, S.W.; Sun, D.Z.; Wang, X.; Yang, J.; Yuan, R.; Wang, E.B. Inorg. Chim. Acta **2012**, 385, 170-177.

5. Laxmi, K.; Sproules, S.; Pawar, O.; Markad, G.; Haram, S.; Pura-nik, V.; Salunke-Gawali, S. *J. Mol. Struct.* **2013**, *1048*, 223-229.
6. Gielen, M.; Tiekink, E.R.T., Eds., *Metallotherapeutic Drugs and Metal-Based Diagnostic Agents, the Use of Metals in Medicine*, Wiley, Chichester, **2005**.
7. Bharti, S.K.; Singh, S.K. *Pharmacia Lett.* **2009**, *1*(2), 39-51.
8. Weder, J.E.; Dillon, C.T.; Hambley, T.W.; Kennedy, B.J.; Lay, P.A.; Biffin, J.R.; Regtop, H.L.; Daview, N.M. *Coord. Chem. Rev.* **2002**, *232*, 95-126.
9. Ware, D.C.; Brothers, P.J.; Clark, G.R. *J. Chem. Soc. Dalton Trans.* **2000**, 925-932.
10. Nakai, M.; Sekiguchi, F.; Obata, M.; Ohtsuki, C.; Adachi, Y.; Sakurai, H.; Orvig, C.; Rehder, D.; Yano, S. *J. Inorg. Biochem.* **2005**, *99*, 1275-1282.
11. Chaviara, T.; Christidis, P.C.; Papageorgiou, A.; Chrysogelou, E.; Hadjipavlou-Litina, D.J.; Bolos, C.A. *J. Inorg. Biochem.* **2005**, *99*, 2102-2109.
12. Sadler, P.J.; Guo, Z. *Pure Appl. Chem.* **1998**, *70*, 863-871.
13. Baslas, R.K.; Zamani, R.; Nomani, A.A. *Experientia* **1979**, *35*, 455-456.
14. Gonzalez, B.E.; Daeid, N.N.; Nolan, K.B.; Farkas, E. *Polyhedron* **1994**, *13*, 1495-1499.
15. Nolan, K.B.; Soudi, A.A. *Inorg. Chim. Acta* **1995**, *230*, 209-210.
16. Muller, J.G.; Burrows, C.J. *Inorg. Chim. Acta*, **1998**, *275*, 314-319.
17. Underhill, A.E.; Bougourd, S.A.; Flugge, M.L.; Gale, S.E.; Gomm, P.S. *J. Inorg. Biochem.* **1993**, *52*, 139-144.
18. Lawal, A.; Obaleye, J.A. *Biokemistri* **2007**, *19*(1), 9-15.
19. Kirkova, M.; Atanassova, M.; Russanov, E. *Gen. Pharmacol.* **1999**, *33*, 271-276.
20. Duda, A.M.; Kowalik-Jankowska, T.; Kozlowski, H.; Kupka, T. *J. Chem. Soc. Dalton Trans* **1995**, 2909-2913.
21. Kubiak, M.; Duda, A.M.; Ganadu, M.L.; Kozlowski, H. *J. Chem. Soc. Dalton Trans* **1996**, 1905-1908.
22. Umadevi, B.; Muthiah, P.T.; Shui, X.; Eggleston, D.S. *Inorg. Chim. Acta* **1995**, *234*, 149-152.
23. Sanchez-del Grado, R.A.; Navarro, M.; Perez, H.; Urbina, J.A. *J. Med. Chem.* **1996**, *39*, 1095-1099.
24. Behrens, N.B.; Diaz, G.M.; Goodgame, D.M.L. *Inorg. Chim. Acta* **1986**, *125*, 21-264.
25. Xie, Y.-M.; Wu, J.-H. *Inorg. Chim. Acta* **2014**, *412*, 15-19.
26. Cao, X.; Yu, L.; Huang, R. *J. Solid State Chem.* **2014**, *210*(1), 74-78.
27. Sutrisno, A.; Huang, Y. *Solid State Nuclear Magnetic Resonance* **2013**, *49-50*, 1-11.
28. Zheng, X.-P.; Lu, Y.; Zhang, H.; Zhang, Z.-m.; Wang, E.-b. *Inorg. Chem. Commun.* **2013**, *33*, 29-32.
29. Lpez-Gresa, M.P.; Ortiz, R.; Perell, L.; Latorre, J.; Liu-Gonzalez, M.; Garcia- Granda, M.; Pe rez-Priede, M.; Cantn, E.; *J. Inorg. Biochem.* **2002**, *92*, 65.
30. Turel, I. *Coord. Chem. Rev.* **2002**, *232*, 27-47.
31. Xiao, D.R.; Wang, E.B.; An, H.Y.; Li, Y.G.; Xu, L. *Cryst. Growth Des.* **2007**, *7*, 506-512.
32. Xiao, D.R.; He, J.H.; Sun, D.Z.; Chen, H.Y.; Yan, S.W.; Wang, X.; Yang, J.; Yuan, R.; Wang, E.B. *Eur. J. Inorg. Chem.* **2012**, 1783-1789.
33. Palumbo, M.; Gatto, B.; Zagotto, G.; Palu, G. *Trends. Microbiol.* **1993**, *1*, 232-235.
34. Sissi, C.; Andreolli, M.; Cecchetti, V.; Fravolini, A.; Gatto, B.; Palumbo, M. *Bioorg. Med. Chem.* **1998**, *6*, 1555-1561.
35. Lancaster, S.G.; Todd, P.A. *Drugs* **1988**, *35*, 646-669.
36. Rush, J.E.; Merrill, D.D. *J. Cardiovasc. Pharm.* **1987**, *9*, 99-107.
37. Gomez, H.J.; Smith, S.G.; Moncloa, F. *Am. J. Med.* **1988**, *85*, 35-37.
38. Beermann, B.; Till, A.E.; Gomez, H.J.; Hichens, M.; Bolognese, J.A.; Junggren, I.L. *Biopharm. Drug Disposit.* **1989**, *10*, 397-409.
39. The United States Pharmacopeia, 24th Revision, the National For-mulary, 19th Revision, United States Pharmacopeial Convention, Inc., Rockville, MD, **2000**.
40. El-Gindy, A.; Ashour, A.; Abdel-Fattah, L. *J. Pharm. Biomed. Anal.* **2001**, *25*, 913-922.
41. Qin, X.Z.; Nguyen, D.S.T.; Ip, D.P. *J. Liq. Chromatogr.* **1993**, *16*(17), 3713.
42. Kalal, H.S.; Rafiei, J.; Bani, F. *Int. J. Environ. Res.* **2010**, *4*, 289-296.
43. Du, J.; Ji, X.J.; Huang, H. *Chin. J. Anal. Chem.* **2009**, *37*, 681-684.
44. El-Yazbi, F.A.; Abdine, H.H.; Shaalan, R.A. *J. Pharm. Biomed. Anal.* **1999**, *19*(6), 819-827.
45. Razak, O.A.; Belal, S.F.; Bedair, M.M. *J. Pharm. Biomed. Anal.* **2003**, *31*, 701-711.
46. Bonazzi, D.; Gotti, R.; Andrisano, V. *J. Pharm. Biomed. Anal.* **1997**, *16*, 431-438.
47. Avadhamulu, A.B.; Pantulu, A.R.R. *Indian Drugs* **1993**, *30*, 646.
48. Qin, W.; Zhang, Z.; Tian, Y.; Xu, F. *Biomed. Chromatogr.* **2007**, *21*, 415-421.
49. Padua, A.A.F.; Barrientos-Astigarraga, R.E.; Rezende, V.M. *J. Chromatogr. B* **2004**, *809*, 211-216.
50. Yuan, A.S.; Gilbert, J.D. *J. Pharm. Biomed. Anal.* **1996**, *14*(7), 773-781.
51. Shepley, K.; Rocci, M.L.; Patrick, H. *J. Pharm. Biomed. Anal.* **1988**, *6*(3), 241-251.
52. Abdel-Fattah, L.; El-Kosasy, A.; Abdel-Aziz, L. *J. Am. Sci.*, **2010**, *6*(10), 1115-1121.
53. Gustafsson, S.; Erriksson, B.M.; Nilsson, I. *J. Chromatogr.* **1990**, *506*, 75-83.
54. Barbato, F.; Morrica, P.; Quaglia, F. *Il Farmaco*, **1994**, *49*, 457-460.
55. Hajnalka, J.; Pettinari, C.; Marchetti, F.; Kamu, E.; Nagy, L.; Troyanov, S.; Pellerito, L. *J. Inorg. Biochem.* **2003**, *97*, 370-376.
56. Anvarhusein, I.A.; Wazeer, M.I.M. *Spectrochim. Acta Part A* **2006**, *65*, 191-195.
57. Xiaoling, W.; Du, L.; Li, D.; Gong, Q.; Wang, L.; Lin, Y. *Spectrochim. Acta Part A* **2012**, *94*, 12-17.
58. Geary, W.J. *Coord. Chem. Rev.* **1971**, *7*, 81-122.
59. Kobelnik, M.; Cassimiro, D.L.; Santos, D.D.; Ribeiro, C.A.; Crespi, M.S. *Chinese J. Chem.* **2011**, *29*(11), 2271-2277.
60. Refat, M.S. *Spectrochim. Acta Part A* **2007**, *68*, 1393-1405.
61. Sadeek, S.A.; El-Didamony, A.M.; El-Shwiniy, W.H.; Zordok, W.A. *J. Arg. Chem. Soc.* **2009**, *97*, 51-76.
62. Adam, A.A. *J. Mat. Sci. Res.* **2012**, *1*, 167-182.
63. Deacon, G.B.; Phillips, R.J. *Coord. Chem. Rev.* **1980**, *33*, 227-250.
64. Dendrinou-Samara, C.; Tsotsou, G.; Ekateriniadou, L.V.; Kort-saris, A.H.; Raptopoulou, C.P.; Terzis, A.; Kyriakidis, D.A.; Kes-sisoglou, D.P. *J. Inorg. Biochem.* **1998**, *71*, 171-179.
65. Nakamoto, K. *Infrared and Ramman Spectra of Inorganic and Coordination Compounds*, (4th ed.), New York: Wiley. pp. 230, **1986**.
66. Bhattacharyya, S.; Mukhopadhyay, S.; Samanta, S.; Weakley, T.J.R.; Chaudhury, M. *Inorg. Chem.* **2002**, *41*, 2433-2440.
67. Nakamoto, K. *Infrared Spectra of Inorganic and Coordination Compounds*, Wiley Interscience, New York, **1978**.
68. Ross, S.D. *Inorganic Infrared and Raman Spectra*, Mc Graw Hill, London, **1972**.
69. Sharma, S.; Iqbal, S.A.; Bhattacharya, M. *Orient. J. Chem.* **2009**, *25*(4), 1101- 1104.
70. Refat, M.S. *J. Mol. Struct.* **2011**, 985, 380-390.
71. Freeman, E.S.; Carroll, B. *J. Phys. Chem.* **1958**, *62*, 394-397.
72. Coats, A.W.; Redfern, J.P. *Nature* **1964**, *201*, 68-69.
73. Horowitz, H.W.; Metzger, G. *Anal. Chem.* **1963**, *35*, 1464.
74. Flynn, J.H.F.; Wall, L.A. *J. Res. Natl. Bur. Stand.* **1966**, *70A*, 487-523.
75. Wendlandt, W.W. *Thermal Methods of Analysis*, Wiley, New York, **1974**.
76. Gupta, R.; Saxena, R.K.; Chatarvedi, P.; Viridi, J.S. *J. Appl. Bac-teriol.* **1995**, *78*, 378.