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EFFECT OF PROTEIN ON PERFORMANCE AND HAEMATOLOGY OF BROILERS FED CASSAVA PEEL-BASED DIETS

EFFECTO DE LA PROTEÍNA SOBRE EL RENDIMIENTO Y HEMATOLOGÍA DE BROILERS ALIMENTADOS A BASE DE RESIDUO DE MANDIOCA

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ADDITIONAL KEYWORDS

Unconventional feedstuff. Blood parameters.

PALABRAS CLAVE ADICIONALES

Alimentos alternativos. Parámetros sanguíneos.

SUMMARY

A experiment was conducted to investigate the influence of the protein source on the performance and haematology of broiler chickens on cassava peel-based diets using fishmeal and groundnut cake as protein sources. In a 42 days feeding trial, a total of 192 Anak, four weeks old, unsexed broiler chickens with average initial body weight of 0.685 ± 0.0027 (mean \pm SD) were divided into two groups of diets containing 0, 5, 10 and 15% dried cassava peel meal. Birds in one group had fishmeal as protein source while the other had groundnut cake. Each diet was offered to three pens of 8 birds each.

Final live weight, weekly weight gain, feed and protein intake, feed conversion and protein efficiency ratios, haematology and corpuscular indices were not significantly affected by protein source. Fishmeal inclusion resulted in higher ($p < 0.001$) serum albumin and lower ($p > 0.05$) serum cholesterol than groundnut cake inclusion. It was concluded that groundnut cake could serve as protein source for broiler chickens without any adverse effect on performance indices.

RESUMEN

En este estudio se investigó la influencia de la fuente de proteína sobre el rendimiento y hematología de broilers alimentados con dietas con harina de residuos de mandioca y harina de pescado o torta de cacahuete, como fuente de

proteína. Un total de 192 broilers Anak de un día de edad, no sexados, se distribuyeron (a la semana 5) en dos grupos de dietas que contenían 0, 5, 10 y 15% de harina de residuos de mandioca. A las aves de uno de los grupos se les suministró harina de pescado como fuente de proteína y a las del otro, torta de cacahuete. Cada dieta fue ofrecida a tres lotes de 8 aves cada uno.

El peso vivo final, la ganancia de peso semanal, los índices de conversión de pienso y de proteína, la hematología e índices corpusculares no fueron afectados por la fuente de proteína. La inclusión de harina de pescado dio lugar a mayores valores de albúmina sérica y menor cantidad de colesterol sérico que la inclusión de cacahuete. Se concluyó que la torta de cacahuete podría ser útil como fuente de proteína para broilers, sin efectos adversos sobre el rendimiento.

INTRODUCTION

That food scarcity is a plague in many developing countries of the world, including Nigeria where daily intake of animal protein per caput falls far below the normal intake as recommended by ILCA (1980) and FAO (1986), is not in doubt. To alleviate this situation, it has been realized that broiler production is the fastest and easiest route (Nworgu *et al.*, 2000; Dipeolu *et al.*, 1996;

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Larry, 1993) since they are prolific, possess a high feed conversion ratio and are accepted by all, irrespective of religion.

However, feed cost are presently very high and makes up to 60-70% (Larry, 1993) or 70-80% (Oruwari *et al.*, 1995) of the total cost of production in Nigeria compared to 50-70% in developed countries (Thackie and Flenscher, 1995). This therefore highlights the importance of feed management to broiler producers. Thus it is necessary to reduce the cost of feeds in order to produce cheaper products without affecting profits. Since energy source constitutes 45-60% of finished feeds for monogastric animals (Tewe and Egbunike, 1992) and birds eat to satisfy their energy requirements (Sibbald, 1982), cassava products, roots, peels and leaves, have been exploited in the replacement of cereals like maize. Although inclusion of cassava peel in the diet increased feed intake and decreased body weight in broilers (Tewe, 1983), cassava peel can be used to cut down the cost of production and lead to an active and sustainable development in livestock production (Olorede *et al.*, 2002). Some workers (Osei and Duodu, 1988; Agunbiade *et al.*, 2002; Salami *et al.*, 2003) recommended the inclusion of fermented cassava peel up to 15% in broiler diets with no adverse effects. Even the use of oven-dried cassava peels led to a reduction in weight gain but no effects on mortality and carcass quality.

However, for a reasonable performance of animals fed cassava-based diets, the rations must be nutritionally balanced and contain sufficient sulphur-containing amino acids (Tewe, 1992). Fishmeal has been widely used in poultry rations to provide the necessary amino acids although some variations still exist (Bello, 1984) especially due to processing method, type of fish and storage. Considering this variation and the fact that the cost of fishmeal has placed it out of reach to farmers in most developing countries, it becomes evident that some plant protein sources can be used in spite of

their deficiency in sulphur-containing amino acids (Antunes and Sgarbieri, 1980; Barbour *et al.*, 1993). These deficiencies are normally rectified by the addition of other protein sources or synthetic amino acids. The crude protein (%) and metabolisable energy (kcal/kg) contents of fish meal are 65 and 2580, whereas groundnut cake meal has values of 40 and 2500 respectively (NRC, 1994).

In the light of the above, therefore, this study became necessary with the aim of studying the influence of the protein source on the performance and haematology of broiler chickens on cassava peel-based diets using fishmeal and groundnut cake.

MATERIALS AND METHODS

192 Anak unsexed four week-old broiler chickens were used for the experiment. They were fed on a commercial broiler starter diet (Livestock Feeds Plc.) until the end of the 4th week and supplied drinking water *ad libitum*. They were also given all the necessary vaccinations and medication as and when due. Thereafter, the birds were randomly assigned to eight dietary treatments. Each diet was offered to three pens of 8 birds each. Diets 1 to 4 contained fishmeal (FM) as the protein source while groundnut cake (GNC) provided this nutrient in diets 5-8. In a 2 x 4 factorial design, each group of diets contained 0, 5, 10 or 15% of cassava peel meal (**table I**). Thus the treatments were designated OFM, 5FM, 10FM and 15FM for the diets with fishmeal and 0GNC, 5GNC, 10GNC and 15GNC for the diets with groundnut cake. In weeks 6 and 7 all the birds were supplied drinking water containing 3-nitro-4-hydroxylphenyl arsenic acid (prophylactic dose to control coccidiosis), at the rate of 8 mg/litres of water, after which use was discontinued.

As the birds were assigned to the treatments (at the onset of week 5), they were weighed. The initial body weight of birds was 0.695±0.0027 (mean±SD). Thereafter, feed intake and liveweight changes

DIETARY PROTEIN ON PERFORMANCE AND HAEMATOLOGY OF BROILER CHICKENS

Table 1. *Ingredients and chemical composition of the experimental diets.* (Ingredientes y composición nutritiva de las dietas experimentales).

Ingredients (%)	Fish meal				Groundnut cake			
	0FM	5FM	10FM	15FM	0GNC	5GNC	10GNC	15GNC
Maize	49	44	39	34	49	44	39	34
Cassava peels	0	5	10	15	0	5	10	15
Fish meal	3	3	3	3	0	0	0	0
Groundnut cake	0	0	0	0	14	14	14	14
Soya bean meal	15	15	15	15	6	6	6	6
Wheat offal	17.5	17.5	17.5	17.5	17.0	17.0	17.0	17.0
Palm kernel cake	7.5	7.5	7.5	7.5	7.5	7.5	7.5	7.5
Blood meal	3.5	3.5	3.5	3.5	2	2	2	2
Bone meal	2	2	2	2	2	2	2	2
Palm oil	1	1	1	1	1	1	1	1
Oyster shell	1	1	1	1	1	1	1	1
Vitamin-mineral premix*	0.25	0.25	0.25	0.25	0.25	0.25	0.25	0.25
Salt	0.25	0.25	0.25	0.25	0.25	0.25	0.25	0.25
Chemical composition								
Calculated values (%)								
ME (kcal/kg)	2817	2752	2688	2623	2820	2755	2691	2626
Crude protein	19.61	19.42	19.22	19.03	19.01	18.82	18.63	18.43
Crude fibre	4.361	4.703	5.044	5.386	4.528	4.869	5.211	5.552
Arginine	1.247	1.288	1.329	1.370	1.501	1.542	1.583	1.624
Histidine	0.542	0.532	0.522	0.512	0.444	0.434	0.424	0.413
Isoleucine	0.732	0.716	0.700	0.684	0.649	0.633	0.617	0.601
Leucine	1.780	1.727	1.674	1.621	1.556	1.503	1.450	1.397
Lysine	1.075	1.076	1.077	1.078	0.845	0.846	0.847	0.848
Methionine	0.323	0.321	0.319	0.317	0.292	0.290	0.288	0.286
Phenylalanine	0.981	0.960	0.938	0.917	0.898	0.877	0.855	0.834
Threonine	0.749	0.733	0.717	0.701	0.616	0.600	0.584	0.568
Tryptophan	0.224	0.225	0.226	0.227	0.190	0.191	0.192	0.193
Valine	1.077	1.056	1.034	1.013	0.932	0.910	0.889	0.867
Determined values (%)								
Dry matter	89.4	90.1	89.9	89.5	89.6	90.1	89.9	90.0
Crude protein	19.58	19.91	18.52	19.87	18.01	17.93	17.17	18.72
Crude fibre	4.41	4.80	5.17	5.52	4.35	4.71	5.09	5.49
Ether extract	6.71	5.41	5.84	6.71	6.27	6.31	6.74	6.75
Ash	7.15	6.53	5.84	8.95	6.50	7.21	6.52	9.27

*Supplied per kg diet: Vit. A, 10 000 i.u.; Vit. D3, 2000 i.u.; Vit. E, 2.5 i.u.; Vit. K, 2.0 mg; Riboflavin, 4.2 mg; Pantothenic acid, 5.0 mg; nicotinic acid, 20.0 mg; choline chloride, 300.0 mg; folic acid, 0.5 mg; Mn, 56.0 mg; I, 1.0 mg; Fe, 20.0 mg; Cu, 10.0 mg; Zn, 50.0 mg; Co, 1.25 mg.

were recorded weekly. At the end of the 10th week all birds were weighed, slaughtered, eviscerated, dressed and again weighed. The feeding trial lasted for 42 days. The

liver, heart and gizzard were cleaned and weighed. At slaughter, blood samples were collected into two test tubes per bird, one with EDTA as anticoagulant and the other

Table II. *Effects of dietary protein source on performance of broilers fed diets containing cassava peel. (Efectos de la fuente de proteína de la dieta sobre la productividad de broilers alimentados con dietas que contienen harina de residuos de mandioca).*

Nutrients	0FM	5FM	10FM	15FM	0GNC	5GNC	10GNC	15GNC	Pooled SD	Statistical significance		
										DPS	CPC	DPS x CPC
Initial live weight (kg/bird)	0.70	0.70	0.70	0.70	0.70	0.68	0.69	0.69	0.0027	0.3988	0.3452	0.5891
Final live weight (kg/bird)	1.75	1.62	1.60	1.56	1.54	1.53	1.56	1.75	0.0315	0.8707	0.2581	0.1655
Weight gain (kg/bird)	1.05	0.92	0.91	0.86	0.84	0.85	0.87	1.06	0.0311	0.1277	0.9850	0.4263
Weight gain (kg/bird/week)	0.33	0.30	0.30	0.27	0.22	0.23	0.25	0.30	0.0160	0.1277	0.9850	0.4263
Dressed weight (kg)	1.41 ^a	1.20 ^b	1.19 ^b	1.17 ^b	1.13 ^b	1.15 ^b	1.16 ^b	1.29 ^a	0.0329	0.14163	0.0472	0.0036
Total feed intake (kg)	8.07	7.35	7.49	7.53	7.43	7.03	7.76	8.67	0.1788	0.8602	0.8319	0.8026
Feed intake (kg/bird/week)	1.36	1.22	1.23	1.25	1.24	1.17	1.30	1.44	0.0308	0.8602	0.8319	0.8026
Protein intake (kg/bird/week)	0.29	0.27	0.26	0.28	0.25	0.23	0.25	0.30	0.0082	0.4856	0.6479	0.7463
Feed conversion ratio	4.12	4.07	4.10	4.63	5.64	5.09	5.20	4.80	0.2067	0.7876	0.8858	0.3113
Protein efficiency ratio	1.14	1.11	1.15	0.96	0.88	1.01	1.00	1.00	0.0334	0.5322	0.9241	0.9489

Means with different superscripts in the same row are significantly different ($p < 0.05$).**Table III.** *Effects of dietary protein source on proportional weight (% live weight) of organs and main cuts of broilers at 10 weeks of age. (Efecto de la fuente de proteína en la dieta sobre el peso relativo de órganos y despiece (% peso vivo) de broilers de 10 semanas).*

Parts	0FM	5FM	10FM	15FM	0GNC	5GNC	10GNC	15GNC	Pooled SD	Statistical significance		
										DPS	CPC	DPS x CPC
Liver weight (kg)	0.06	0.06	0.06	0.06	0.06	0.06	0.06	0.07	0.0130	0.7347	0.8569	0.7099
Heart weight (kg)	0.03	0.04	0.04	0.04	0.03	0.03	0.03	0.03	0.0087	0.1623	0.1710	0.3354
Gizzard weight (kg)	0.07	0.09	0.07	0.07	0.07	0.09	0.07	0.08	0.0180	0.8052	0.5678	0.8250
Shank	4.32	4.00	3.92	5.06	4.50	3.88	4.48	4.38	0.0194	0.9089	0.6177	0.8778
Breast	19.24	19.43	17.66	18.67	17.47	18.79	17.47	19.22	0.0640	0.4902	0.4666	0.5872
Wing	9.48	8.57	9.63	9.55	9.41	8.74	8.15	9.93	0.0301	0.5905	0.8469	0.7133
Back	9.26	8.77	9.52	9.16	10.55	8.91	8.51	9.82	0.0428	0.7767	0.2434	0.0209
Drumstick	21.62	20.35	20.13	21.17	20.89	21.12	21.02	20.87	0.0662	0.6977	0.8637	0.3424
Neck	4.16	4.62	4.08	4.94	4.68	4.00	4.24	4.70	0.0131	0.7259	0.2444	0.8705

Means with different superscripts in a row are significantly different ($p < 0.05$).

DIETARY PROTEIN ON PERFORMANCE AND HAEMATOLOGY OF BROILER CHICKENS

without EDTA. The first sample was used for haematological analyses while the second was centrifuged at 4,000g for 10 minutes for serum separation. Five dressed birds having a representative weight of the group were randomly picked from each treatment and cut into standard parts namely shank, breast, wings, back, drumstick and neck and weighed separately.

All the diets were chemically analysed according to the standard of AOAC (1990) methods for their proximate composition. Packed cell volume (PCV) was determined by the method of Wintrobe (1933), the red blood cell (RBC), white blood cell (WBC), mean cell volume (MCV), mean cell haemoglobin (MCH) and mean cell haemoglobin concentration (MCHC) were determined as reported by Jain (1986) while haemoglobin (Hb) was estimated as reported by Swensen (1951) and Coles (1986).

Total serum protein was estimated by the biuret method of Reinhold (1953), serum albumin according to Doumas and Briggs (1972) and Peters *et al.* (1982) and serum globulin was by the method of Rodkey (1965) and Coles (1986). Serum cholesterol was determined according to the methods of Roschlan *et al.* (1974).

Data were subjected to routine ANOVA from General Linear Model procedures using the software package SPSS 13.0 for windows. Means were separated using Duncan's Multiple Range Test from the same software.

RESULTS

The composition of the experimental diets is shown in **table I**. The crude protein content (on dry matter basis) of the diets ranged from 19.3% in 10GNC to 22.2% in 15FM while the metabolizable energy ranged from 2626 to 2820 Kcal/kg (on as fed basis).

Final liveweight, weekly weight gain, feed and protein intake as well as feed conversion and protein efficiency ratios did not vary significantly with treatment (**table II**). Thus while weekly feed intake varied

insignificantly from 1.17 kg in 5GNC to 1.44 kg in 15GNC, weight gain varied from 0.22 kg in 0GNC to 0.33 kg in 0FM. Feed conversion ratio and protein efficiency ratio were in the range of 4.07-5.64 and 0.88-1.15, respectively. However, the dressed weights of the birds were significantly ($p < 0.05$) influenced by the dietary treatments with 0FM and 15GNC birds being heavier than all the others. On the contrary, the weights of the liver, heart and gizzard as well as the chicken cuts, all of them expressed as percent of final liveweights (**table III**) were stable with the dietary treatments.

The source of dietary protein did not affect the haematology of the birds but with the increase of cassava in the diets Hb, RBC, WBC and PCV increased significantly ($p < 0.05$). As regards the corpuscular values (MCV, MCH and MCHC), all values obtained remained stable irrespective of dietary treatment (**table IV**). Total serum protein, globulin and albumin/globulin remained stable with the cassava peel-based diets regardless of the source of dietary protein. However, fishmeal inclusion in the diet resulted in a higher ($p < 0.05$) serum albumin but lower serum cholesterol ($p < 0.05$) than groundnut cake. These two indices were elevated ($p < 0.05$) with dietary inclusion of cassava peel at 5% after which there was a significant ($p < 0.05$) depression.

DISCUSSION

The nutrient composition of the experimental diets shown in **table I** is in agreement with the recommendations of Oluyemi and Roberts (1980) for broiler finishers. They were also similar to the diets formulated by Esonu and Bamgbose (2000) and so can be considered adequate for broiler finishers.

That final liveweight, weight gain, feed intake, protein intake, feed conversion ratio and protein efficiency ratio were not influenced by the cassava peel-based diets whether with fish meal or groundnut cake as protein source is in agreement with the

Table IV. Effect of dietary protein source and cassava peel content on the haematological and biochemical indices of broilers. (Efecto de la fuente de proteína en la dieta y del contenido de residuos de mandioca sobre los índices hematológicos de broilers).

Parameters	0FM	5FM	10FM	15FM	0GNC	5GNC	10GNC	15GNC	Statistical significance		
									Pooled SD	DPS	CPC
Hb (g/dl)	8.41 ^a	9.38 ^b	8.86 ^c	8.82 ^c	7.04 ^d	8.78 ^c	8.82 ^c	10.12 ^e	0.309	0.0168	<0.001
RBC (10 ⁹ /l)	2.21 ^a	2.39 ^b	2.63 ^c	2.35 ^b	1.77 ^a	2.29 ^c	2.41 ^b	2.58 ^f	0.094	0.0136	<0.001
WBC (x10 ³ /l)	29.39 ^a	34.09 ^b	24.72 ^c	25.13 ^c	22.78 ^d	27.90 ^c	26.48 ^c	29.91 ^a	1.265	0.1741	0.008
PCV (%)	25.20 ^a	28.40 ^b	30.40 ^c	27.00 ^d	26.20 ^e	26.60 ^e	26.60 ^e	30.00 ^c	0.659	<0.001	<0.001
MCV (fl)	114.20	118.00	115.60	115.00	119.60	116.00	110.80	116.00	0.920	0.9544	0.3867
MCH (Pg)	38.40	38.80	37.20	37.40	39.40	38.20	36.40	39.00	0.361	0.6753	0.1964
MCHC (g/dl)	33.40	32.60	31.80	32.40	32.80	32.20	32.60	33.00	0.173	0.7428	0.1908
Total protein (g/dl)	5.94	6.00	5.22	5.04	4.55	6.30	5.94	4.14	0.2747	0.1336	<0.001
Albumin (g/dl)	2.50 ^a	3.40 ^b	2.62 ^a	2.27 ^c	1.88 ^c	3.00 ^b	2.72 ^a	1.42 ^d	0.2205	0.001	<0.001
Globulin (g/dl)	3.44	2.60	2.60	2.77	2.67	3.30	3.22	2.78	0.1206	0.1243	0.3892
Albumin/globulin (%)	72.67	130.77	100.77	81.92	70.41	90.91	84.47	52.21	8.266	0.6572	0.0761
Cholesterol (mg/dl)	127.92 ^b	149.60 ^a	81.46 ^c	119.00 ^b	143.30 ^a	154.49 ^a	100.12 ^b	96.06 ^b	9.551	0.1384	0.0039

Means with different superscripts in a row are significantly different (p<0.05).

reports on the use of cassava peel up to 15% in broiler diets by Osei and Duodu (1988); Agunbiade *et al.* (2002) and Salami *et al.* (2003). Although final liveweight was not affected by the dietary treatment, dressing weight was best for 0FM and 15 GNC. Result is in agreement with the findings of Willis *et al.* (1997), Nworgu and Egbunike (1999) and Odunsi *et al.* (1999). This effect on dressing weight may imply changes in gut transit time of the diets and feed efficiency as shown by Ravindran (1995) and Agwunobi (1999) that could result in changes in the weight of the gastrointestinal tract as a function of dietary treatments. However, that the weights of heart, liver, gizzard and the chicken cuts were not influenced by the treatment conforms with the results of Berri (2000), Nworgu *et al.* (2001) and Egbunike and Williams (2005) but not with those of Pollock (1997). Body weight gains and feed conversions from the present study are also in good agreement with those in which conventional feed ingredients were fed to Anak strain of broiler of a similar weight range (Oruseibo and Alu, 2006; Oyedele *et al.*, 2003; Idowu *et al.*, 2003).

The protein source did not have any influence on the haematological indices and corpuscular indices indicating that the diets were ideal or adequate for broiler finishers. However, these values were significantly elevated (p<0.05) as the cassava peel level increased regardless of the protein source, but are within the normal range reported in literature (Jain, 1986). This suggests that the diets with cassava peels promoted a better haematological *milieu* as observed in rats by Adejumo (2004).

The adequacy of the diets is also shown by the stability of the serum protein, globulin and albumin/globulin ratio values of the birds on the different diets regardless of protein source and cassava peel levels. The protein source however significantly influenced the serum albumin and cholesterol levels with these increasing and

DIETARY PROTEIN ON PERFORMANCE AND HAEMATOLOGY OF BROILER CHICKENS

decreasing, respectively, with fishmeal. The values in these parameters of this study are at variance with those of Nworgu (2004) with albumin being lower and albumin/globulin ratio and cholesterol being higher. The cholesterol values are however generally within the normal range according to Sturkie *et al.* (2000). The increase in the albumin level at 5% cassava peel levels regardless of protein source may be construed as implying that the improvement

of the haematology referred to earlier was best at this level.

CONCLUSION

The results on final liveweight, weight gain, feed conversion ratio and protein efficiency ratio obtained in this study indicate that broilers could be raised on cassava peel-based diets using groundnut cake as the protein source.

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EGBUNIKE, AGIANG, OWOSIBO AND FATUFE

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