



Ciência & Saúde Coletiva

ISSN: 1413-8123

cecilia@claves.fiocruz.br

Associação Brasileira de Pós-Graduação em  
Saúde Coletiva  
Brasil

Benbrahim-Tallaa, Lamia; Waalkes, Michael  
Inorganic arsenic and human prostate cancer  
Ciência & Saúde Coletiva, vol. 14, núm. 1, janeiro-fevereiro, 2009, pp. 307-318  
Associação Brasileira de Pós-Graduação em Saúde Coletiva  
Rio de Janeiro, Brasil

Disponível em: <http://www.redalyc.org/articulo.oa?id=63014133>

- Como citar este artigo
- Número completo
- Mais artigos
- Home da revista no Redalyc

redalyc.org

Sistema de Informação Científica  
Rede de Revistas Científicas da América Latina, Caribe, Espanha e Portugal  
Projeto acadêmico sem fins lucrativos desenvolvido no âmbito da iniciativa Acesso Aberto

## Inorganic arsenic and human prostate cancer \*

### Arsênico inorgânico e câncer de próstata humano

Lamia Benbrahim-Tallaa <sup>1</sup>  
Michael Waalkes <sup>1</sup>

**Abstract** *We critically evaluated the etiologic role of inorganic arsenic in human prostate cancer. We assessed data from relevant epidemiologic studies concerning environmental inorganic arsenic exposure. Whole animal studies were evaluated as were in vitro model systems of inorganic arsenic carcinogenesis in the prostate. Multiple studies in humans reveal an association between environmental inorganic arsenic exposure and prostate cancer mortality or incidence. Many of these human studies provide clear evidence of a dose-response relationship. Relevant whole animal models showing a relationship between inorganic arsenic and prostate cancer are not available. However, cellular model systems indicate arsenic can induce malignant transformation of human prostate epithelial cells in vitro. Arsenic also appears to impact prostate cancer cell progression by precipitating events leading to androgen independence in vitro. Available evidence in human populations and human cells in vitro indicates that the prostate is a target for inorganic arsenic carcinogenesis. A role for this common environmental contaminant in human prostate cancer initiation and/or progression would be very important.*

**Key words** *Androgen-independent, Arsenic, Carcinogenesis, Human malignant transformation, Prostate, Ras*

**Resumo** *Realizamos uma avaliação crítica do papel etiológico do arsênico inorgânico no câncer de próstata humano. Avaliamos dados de estudos epidemiológicos relevantes referentes à exposição ao arsênico inorgânico ambiental. Foram avaliados estudos com animais completos, bem como sistemas de modelo in vitro de carcinogênese decorrente de arsênico inorgânico na próstata. Estudos múltiplos em seres humanos revelaram uma associação entre exposição ao arsênico inorgânico ambiental e mortalidade por ou incidência de câncer de próstata. Muitos desses estudos em seres humanos oferecem indícios claros de uma relação dose-resposta. Não se encontram disponíveis modelos animais completos relevantes que mostrem uma relação entre arsênico inorgânico e câncer de próstata. Contudo, os sistemas de modelos celulares indicam que o arsênico é capaz de levar a transformações malignas de células epiteliais da próstata humana in vitro. Aparentemente, o arsênico também tem um impacto na progressão do câncer de próstata ao precipitar eventos que levam à independência de andrógeno in vitro. Os indícios disponíveis em populações humanas e células humanas in vitro indicam que a próstata é alvo da carcinogênese de arsênico inorgânico. Um papel para esse contaminante ambiental comum na iniciação e/ou progressão do câncer de próstata humano seria de suma importância.*

**Palavras-chave** *Andrógeno-independente, Arsênico, Carcinogênese, Transformação maligna humana, Próstata, Ras*

\* This article was originally published by *Environ Health Perspect* 116:158–164 (2008). doi:10.1289/ehp.10423 available via <http://dx.doi.org> [Online 8 November 2007] and is part of the scientific collaboration between *Cien Saude Colet* and *EHP*. The authors declare they have no competing financial interests.

<sup>1</sup>Inorganic Carcinogenesis Section, Laboratory of Comparative Carcinogenesis, National Cancer Institute at the National Institute of Environmental Health Sciences, National Institutes of Health, Department of Health and Human Services. MD F0-09, 111 Alexander Dr., Research Triangle Park, NC 27709 USA. waalkes@niehs.nih.gov

Inorganic arsenic, a metalloid, is ubiquitously distributed in nature. In natural deposits, this metalloid forms a complex with pyrite, for which it has a strong affinity<sup>1</sup>. However, under certain conditions (pH, temperature, etc.), inorganic arsenic readily dissociates from its soil-bound forms and enters the aquifer<sup>2</sup>. For this reason, the major source of human exposure to arsenic is naturally contaminated drinking water from underground wells. Probably more than 100 million people are exposed to inorganic arsenic at levels above 10 µg/L, the drinking-water standard in many countries<sup>3</sup>. Arsenic is also released into the atmosphere from both natural and anthropogenic sources. Globally, natural emissions of arsenical compounds have been estimated at about 8,000 tons each year, whereas anthropogenic emissions are about 3 times higher<sup>4,5</sup>. Food, particularly vegetables and rice, may be an additional source of exposure to inorganic arsenic<sup>4,5</sup>. Occupational exposure to arsenic occurs in specific industries such as mining, smelting operations, wood preservation, and electronics<sup>3</sup>.

Arsenical exposure produces various adverse effects such as dermal lesions, hypertension, ischemic heart disease, liver disease, peripheral vascular disorders, arteriosclerosis, diabetes, neuropathy, and cancer<sup>4,5</sup>. The carcinogenic potential of inorganic arsenic exposure through drinking water in humans is a cause for considerable concern<sup>3-6</sup>. Indeed, inorganic arsenic is a potent, multisite human carcinogen most frequently associated with tumors of the skin, urinary bladder, and lung<sup>3-6</sup>. There are also human data associating inorganic arsenic exposure with cancers of the liver, prostate, and kidney. The mechanisms by which inorganic arsenic is carcinogenic are not completely defined<sup>7-10</sup>. A challenge to elucidating these mechanisms has been the difficulty encountered in the development of experimental whole animal models of arsenic carcinogenesis. In essence, it has proven difficult until recently to induce cancer in animals using inorganic arsenic as a single agent<sup>10</sup>. In place of whole animal models, cell lines such as human prostate epithelial cells<sup>11</sup>, keratinocytes<sup>12</sup>, and urothelial cells<sup>13</sup>, which may represent *in vivo* targets of arsenic, provide a relevant and reasonable *in vitro* approach to study the molecular events in inorganic arsenic carcinogenesis.

### Arsenic toxicokinetics and metabolism

The metabolism of arsenic compounds in mammals has been reviewed<sup>14-16</sup>. Inorganic arsenic is well absorbed from the gastrointestinal tract and

distributed throughout the body<sup>5</sup>. It freely crosses the rodent and human placenta<sup>4</sup>. In many tissues inorganic arsenic is biotransformed by methylation<sup>5</sup>. Some cells methylate inorganic arsenic very poorly or not at all, for example, keratinocytes<sup>17</sup> or prostate epithelial cells<sup>18</sup>. Biomethylation of arsenic is no longer considered a detoxification process, as trivalent methylated arsenical intermediates are highly toxic<sup>19-21</sup> and possibly carcinogenic<sup>22</sup>. Reduction of arsenate (As5+) to arsenite (As3+) is necessary before methylation can occur. Arsenate is rapidly reduced to arsenite by glutathione *S*-transferase omega and/or arsenate reductase. Arsenite is then methylated to form methylarsonate (MMA5+) and dimethylarsinic acid (DMA5+) by arsenic methyltransferase using *S*-adenosylmethionine (SAM) as the methyl donor. The intermediate metabolites methylarsonous acid (MMA3+) and dimethylarsinous acid (DMA3+) are generated during this process<sup>14,20</sup>. The precise role of trivalent methylated arsenical species in inorganic arsenic carcinogenesis is not fully understood, although MMA3+ can induce malignant transformation of human urothelial cells *in vitro*<sup>22</sup>.

Both arsenite and arsenate are actively transported into cells<sup>23,24</sup> by mechanisms that may involve organic ion transporters<sup>25</sup>. Recent evidence indicates that multidrug resistance protein 1 (MRP1), an ATP-binding cassette transport protein, is involved in efflux of arsenite in an ATP- and glutathione-dependent manner. It appears arsenic is effluxed as a triglutathione complex<sup>26</sup> produced by glutathione *S*-transferase pi<sup>27</sup>, which may stress cellular redox systems from continuous demands on glutathione.

### Prostate cancer

The prostate gland is characterized by the age-dependent development of abnormal proliferative diseases ranging from

benign prostate hyperplasia to overt malignancies. Prostate cancer is the most frequently diagnosed non-skin cancer among men and the second leading cause of male cancer deaths in the United States<sup>28</sup>. There were approximately 6.7 million cancer deaths worldwide in 2002, and of these, prostate cancer was the fifth most common overall and the second most common among men<sup>29</sup>. Migrant studies provide strong evidence for the role of the environment in prostate cancer. Throughout the last several centuries, major migratory movements of humans have taken place in many parts of the world and continue even today. With increasing length of

stay, cancer mortality rates among immigrants move toward those in the adopted country. This has been clearly shown for prostate cancer<sup>30</sup>. There are also intra- and interracial differences in prostate cancer incidence and mortality rates worldwide, and the environment and migration patterns seem to influence these disparities<sup>31-33</sup>. These studies provide insight into the relative contributions of heredity and environment in prostate cancer.

### Inorganic arsenic as a human carcinogen

The NTP and the IARC have concluded that arsenic is a human carcinogen<sup>3,6</sup>. Arsenic contamination of drinking water is a common occurrence and a worldwide public health issue. Some countries have truly daunting issues with arsenic contamination of drinking-water supplies, and endemic chronic arsenicalism is observed in many places in India, Bangladesh, Taiwan, and China<sup>3</sup>. Although chronic arsenic exposure produces a variety of adverse effects, its carcinogenic potential in humans is perhaps of greatest concern. Although the exact modes of action remain to be defined, it is reasonable to assume that site-specific and multifactorial mechanisms apply to inorganic arsenic.

The carcinogenic potential of arsenic was recognized over 100 years ago by Hutchinson<sup>34</sup>, a British physician who observed skin cancers occurring in patients treated with medicinal arsenicals. Further evidence for arsenic as a human carcinogen after industrial exposure comes from studies of arsenic ore smelters and pesticide workers<sup>35</sup>. In numerous countries it has been shown that people who consume arsenic-contaminated drinking water can develop various cancers<sup>3-6</sup>. Thus, arsenic is a human carcinogen after environmental, occupational, or medicinal exposures. Strong epidemiological associations exist between inorganic arsenic ingestion and cancers of skin, urinary bladder, and lung. Epidemiologic evidence has also linked arsenic in the drinking water to prostate, kidney, and liver cancers<sup>3-5</sup>. In fact, in its 2004 evaluation summary, the most recent IARC monograph on arsenic clearly states "Excess mortality from prostate cancer was found in South-West Taiwan"<sup>3</sup>.

Data on concentrations of arsenic in human target tissues, especially for internal organs, are largely lacking. This factor

becomes problematic when attempting to produce biokinetic models or when defining what are reasonable exposures for *in vitro* studies. At least some human tissues, particularly the skin, clearly

will accumulate arsenic, and skin levels in the range of 5,700 µg/kg (~ 76 µM) have been reported from arsenic intoxicated people in Bangladesh<sup>3</sup>. This is in contrast to circulating levels of up to 60 µg/L (~ 0.8 µM) in blood and 274 µg/L (~ 3.6 µM) in urine during chronic arsenic intoxication<sup>4</sup>. Thus, it is unclear if circulating or excreted levels of arsenic actually reflect target tissue or target cell burden. Perhaps most important, there is essentially no information on arsenic levels in the human prostatic tissue. Clearly, further work in this area is required.

### Arsenic carcinogenesis in animals

Until recently, inorganic arsenic in rodents was generally not carcinogenic except in model systems involving co-administration with known carcinogenic agents<sup>36,37</sup>. However, a series of studies from our laboratory [for review, see Waalkes<sup>10</sup>] has recently demonstrated that inorganic arsenite administered during the second half of gestation to pregnant mice of several strains will induce or impact the development of cancer in the offspring as adults in various tissues, including tissues that are potential human targets such as liver and lung. In studies using prenatal arsenic exposure combined with exposure to additional agents after birth, tumors of the urinary bladder can also be induced. Together these studies provide consistent evidence that *in utero* arsenic is carcinogenic in mice and targets several tissues that are concordant with human target sites.

However, prostate cancers do not develop in these mouse studies<sup>10</sup>. In this regard, the genetically unaltered mouse is not the rodent of choice for *in vivo* models of human prostate cancer<sup>38</sup>. The reasons for this include the observation that mice are resistant to the induction of prostatic tumors by chemical carcinogens as well as differences in anatomy and pathophysiology<sup>38</sup>. Transgenic mouse lines are available in which prostate carcinomas preferentially occur<sup>38,39</sup>, but arsenic has not been tested in these models. Rats generally are considered a better rodent model of prostate cancer because prostate lesions can be chemically induced and in the early stages are androgen dependent<sup>38</sup>. However, arsenic biokinetics in rats is very dissimilar to that in humans or mice, and rats are considered a poor model for human arsenic toxicology<sup>14</sup>. Furthermore, although pentavalent methylated arsenicals are complete carcinogens and tumor promoters in rats<sup>40</sup>, they do not target the prostate. Thus, at present, whole rodent prostate models of inorganic arsenic carcinogenesis are not available.

### Arsenic exposure and human prostate cancer

The first evidence that inorganic arsenic was associated with prostate cancer in humans came from Taiwan in the late 1980s<sup>41</sup> (Table 1). This was a follow-up study that focused on dose-response relationships between arsenic and cancer in a population exposed to high levels of arsenic in the drinking water from local artesian wells. The population studied was from the area of endemic "blackfoot" disease in southwest Taiwan, a disease involving the peripheral vascular dysfunction likely due, at least in part, to arsenic exposure<sup>42</sup>. Although the original study had not looked at cancer of the prostate<sup>42</sup>, the subsequent study found a remarkable association between arsenic exposure and prostate cancer mortality in this population<sup>41</sup>. In this regard, the age-standardized mortality from prostate cancer in the group exposed to the highest levels of arsenic in the drinking water ( $\geq 0.60$  ppm) was nearly 6-fold greater than that of the general population in Taiwan. In addition, when drinking-water arsenic levels were stratified ( $< 0.30$  ppm,  $0.30-0.59$  ppm and  $\geq 0.60$  ppm), a significant dose-response relationship occurred between arsenic level and age-adjusted prostate cancer mortality. The exposed population lived in a relatively small area and had similar lifestyles, diets, living conditions, and sociodemographic characteristics compared with those of nearby unaffected villages, prompting the authors to conclude that the striking differences in cancer mortality between these groups could be explained "solely by the difference in arsenic concentrations in drinking water"<sup>41</sup>.

Prostate cancer is not always fatal, particularly in its early stages, and as the cause of death was determined in this study by death certificate<sup>41</sup>, it is likely that the rate of deaths would be much lower than the incidence of prostate cancers in this population. There were also large increases in mortality from liver, lung, skin, bladder, and kidney cancers in this population due to arsenic exposure that generally exceeded the rate of prostate cancer deaths<sup>41</sup>. Therefore, other cancers may have overshadowed relatively rare cancers of the prostate. Furthermore, prostate cancer is usually a disease of older men, and because arsenic is a very effective, multisite carcinogen, perhaps some of the most sensitive subjects may have died of other arsenic-induced cancers before the development of advanced and deadly prostate cancer. Indeed, prostate cancer is considered to have a relatively low case-fatality rate<sup>3</sup>, making mortality as an end point potentially insensitive of actual disease status, at least in the early stages.

A follow-up study to those of Chen *et al.*<sup>41,42</sup> concerning arsenic and cancer mortality used some of the same population at risk but added data from additional villages in the area of endemic blackfoot disease and specifically studied dose-response relationships<sup>43</sup>. In this study, the age-adjusted mortality for prostate cancer in the population exposed to the highest arsenic levels in the drinking water ( $> 0.60$  ppm) was nearly 10-fold higher (9.18 deaths/100,000) than that at the lowest level ( $< 0.30$  ppm; 0.95 deaths/100,000) of exposure. A clear dose-response relationship also occurred between arsenic exposure and prostate cancer mortality

**Table 1.** Epidemiologic studies of arsenic exposure and prostate cancer in humans.

Study	Population location	Source of arsenic	Result	Dose-response relationship
Chen <i>et al.</i> <sup>41</sup>	Southwest Taiwan <sup>a</sup>	Drinking water	Increased mortality	Clear evidence
Wu <i>et al.</i> <sup>43</sup>	Southwest Taiwan <sup>b</sup>	Drinking water	Increased mortality	Increased mortality
Chen and Wang <sup>44</sup>	Taiwan	Drinking water	Increased mortality	Increased mortality
Tsai <i>et al.</i> <sup>45</sup>	Southwest Taiwan	Drinking water	Increased mortality	Not investigated
Lewis <i>et al.</i> <sup>46</sup>	Utah, USA	Drinking water	Increased mortality	Some evidence <sup>c</sup>
Hindwood <i>et al.</i> <sup>47</sup>	Victoria, Australia	Local water/soil	Increased incidence	No evidence <sup>d</sup>

<sup>a</sup> Study focused on the area of endemic blackfoot disease; <sup>b</sup> The Wu *et al.*<sup>43</sup> study used the Chen *et al.*<sup>41</sup> population, with expansion into additional villages in the blackfoot-endemic area; <sup>c</sup> Based on the authors' interpretation after stratification of data based on drinking-water levels; <sup>d</sup> The rate of prostate cancer incidence was significantly elevated at the highest level of exposure when arsenic exposure was stratified based on arsenic in water and/or soil. When arsenic exposure was stratified on water levels only (low, medium, high, and very high), prostate cancer incidence appeared elevated in the high and very high categories [see Figure 3 in Hindwood *et al.*<sup>47</sup>]. This did not, however, show a significant linear dose-response relationship.

when drinking-water levels of arsenic were stratified ( $< 0.30$ ,  $0.30$ – $0.59$ , and  $> 0.60$  ppm) in this study. These interpretations must be tempered by the small number of cancer deaths due to prostate cancer in this study, but, nonetheless, the findings are consistent with the prior work<sup>41</sup>. Exposure levels were determined by median village levels of arsenic in drinking water wells, and, as such, may be subject to the “ecological fallacy” that the association observed at the village level may not hold at the individual level<sup>43</sup>. Even after considering this and other confounding factors, the authors felt that arsenic content should still be strongly suspected as the main cause of excess cancer deaths in this population<sup>43</sup>.

In subsequent work from Taiwan, the study population was expanded from the area of endemic blackfoot disease used in the first two studies<sup>41,43</sup> to a much more comprehensive study of all 314 precincts and townships in Taiwan as a whole. In all, 83,656 wells were tested for arsenic<sup>44</sup>. Based on multiple regression analysis with adjustments for urbanization and age, mortality rates from cancer of the prostate again increased in correlation with increasing average drinking water level of arsenic.

In an independent study of the area of endemic blackfoot disease in southwest Taiwan, Tsai *et al.*<sup>45</sup> computed age-adjusted standardized mortality ratios (SMRs) using death certificates with national reference rates. The SMR for prostate cancer in the arsenic-exposed population was 1.96, with a 95% confidence interval (CI) of 1.4–2.6, indicating a significant increase in the number of observed cases compared with the number of expected based on the national reference rates. The number of observed cases in this arsenic-exposed population was 48, and dose-response effects were not investigated.

The role of drinking-water arsenic in prostate cancer mortality has also been studied in a U.S. population<sup>46</sup>.

Mortality was assessed in a retrospective cohort of Millard County, Utah, residents along with drinking-water arsenic exposure levels that accounted for residence time in the study area. The cohort consisted of 2,073 members with at least 20 years of exposure history and was assembled through membership records of the Church of Jesus Christ of Latter-day Saints. Arsenic exposure was stratified into low ( $< 1,000$  ppb-years), medium ( $1,000$ – $4,999$  ppb-years) and high ( $> 5,000$  ppb-years) levels<sup>46</sup>. Without considering specific arsenic exposure levels, the overall SMR for prostate cancer mortality was significantly elevated in the cohort (1.45; 95% CI, 1.07–1.91, based on 50 deaths) compared with that of Utah white males. The au-

thors indicate that SMR analysis hinted at a dose-response relationship when based on low (SMR = 1.07), medium [1.70 (significantly elevated)] and high (1.65) arsenic exposure<sup>46</sup>.

In a study from Australia, geographic areas with soil arsenic  $> 100$  mg/kg and/or drinking water concentrations  $> 0.01$  mg/L were selected and related to cancer incidence<sup>47</sup>. Standardized incidence ratios (SIRs) were generated for 22 areas of elevated arsenic exposure in Victoria and compared with all Victorian cancer rates as a baseline. For all areas with any elevated arsenic (soil or water or both), the SIR was significantly increased for prostate cancer (1.14; 95% CI, 1.05–1.23). Exposure was also stratified as only high soil or only high water arsenic (low) or both high soil and high water arsenic (high). When arsenic exposure was stratified by exposure type (i.e., high water only, high soil only, high water/high soil), the SIR for prostate cancer remained significantly elevated (1.20; 95% CI, 1.06–1.36), in the high water/high soil category. Dose-response analysis was performed on data stratified based on water content of arsenic as low ( $< 0.01$  mg/L), medium ( $0.01$ – $0.1$  mg/L), high ( $0.1$ – $0.2$  mg/L), and very high ( $> 0.2$  mg/L) levels. No linear dose response was detected for prostate cancer incidence using this water stratification, but based on graphical presentation, the SIRs for the high and very high categories appeared elevated (95% CIs did not include 1.0). The study included 619 cases of prostate cancer. The authors make the point that of those targets expected *a priori* from other studies, only prostate cancer was significantly elevated.

In a population of male copper foundry workers industrially exposed to arsenic as well as other metals, a correlative survey of plasma neoplastic biomarkers was conducted<sup>48</sup>. A strong positive correlation occurred between urinary arsenic concentration and serum prostate-specific antigen (PSA). PSA is a well-established biomarker for prostate cancer that is considered a mainstay of early prostate cancer detection. The exposure to other metals complicates interpretation of this study, but the correlation between arsenic in the urine and circulating PSA was robust. In this regard, tumors arising from human prostate epithelial cells transformed by inorganic arsenic *in vitro* also show a remarkable overexpression of PSA<sup>11</sup>.

The results of various positive studies of prostate cancer and arsenic exposure were considered as a whole by the IARC<sup>3</sup>. The specific conclusion was that “data from southwest Taiwan indicate a consistent pattern of increased mortality from prostate cancer in areas with high contamination by ar-



senic, and there is evidence of a dose-related effect"<sup>3</sup>. Although the prostate was not specifically mentioned as a human target site in the final evaluation of the monograph, the implications of the text are clear and, at least in part, are supported by the data from the United States and Australia, which make it less likely that the Taiwanese are uniquely sensitive. Whatever the conclusion, the available evidence indicates an obvious need for additional studies of arsenic as a human prostatic carcinogen.

As a potential complicating factor in dose-response analysis, evidence indicates that arsenic can adversely affect testicular function in animals, even at levels near the range for some human exposure situations. This includes loss of testicular weight, diminished sperm count, and decreased 17 $\beta$ -hydroxysteroid dehydrogenase (17 $\beta$ -HSD) activity in mice chronically given 4 ppm arsenic in the drinking water<sup>49</sup>. In this regard, 17 $\beta$ -HSD is an enzyme important in production of testosterone from its immediate precursors, such as androstenedione. Similarly, in rats chronic oral arsenic exposure decreases testicular weight, sperm count, testicular 17 $\beta$ -HSD activity, and plasma and testicular testosterone concentrations<sup>50</sup>. Prostate cancer, particularly in its early stages, is dependent typically on circulating androgens and will regress with orchiectomy and/or antiandrogen therapy, two strategies commonly used in prostate cancer treatment<sup>51</sup>. Thus, if higher doses of arsenic similarly suppressed testosterone production in humans, this could complicate the dose-response analysis by potentially diminishing carcinogenic response at higher doses. There is no direct evidence of this in humans, however.

### ***In vitro* model of arsenic-induced prostatic carcinogenesis**

***In vitro*** models can be invaluable for studies on carcinogenic mechanisms and can be applied to the various stages of oncogenesis including initiation and progression. In fact, those employing human cells can provide carefully controlled exposure circumstances that are impossible in environmentally exposed human populations. Cell model systems have been used to identify molecular markers of transformation during prostate cancer development. In this regard, the majority of commonly used human prostate cell lines are derived from biopsies of metastatic prostate cancer<sup>52,53</sup> and, as such, would be more appropriate for defining molecular events occurring during tumor progression to advanced prostate cancer. Prostate cancer

has the added aspect of acquired androgen independence, generally occurring as a progression to a deadly form of the disease. Hence, tumor derived cell lines have been used to extensively study androgen independence<sup>54</sup>. For the study of carcinogenic initiation, one would want a nontransformed ("normal") line that is nontumorigenic upon inoculation into mice. Human arsenic exposure is typically to an acutely tolerable dose over long periods of time. To use doses (concentrations) similar to human exposure, cells should be exposed to relatively low levels of arsenic for protracted periods. Hence, an immortalized cell line is essential.

The human prostate epithelial cell line RWPE-1 was originally derived from normal human prostate epithelium<sup>53,55</sup>. RWPE-1 cells are immortalized and nontumorigenic upon inoculation into immunocompromised mice, an important observation, as the ability to form tumors is a key element in the definition of cellular malignant transformation. By continuous exposure of this line to low levels of inorganic arsenic over a period of several months, a malignant transformant was developed<sup>11</sup>. Essentially, RWPE-1 cells were cultured in the presence of 5  $\mu$ M arsenic continuously for up to 30 weeks, while parallel control cultures served as passagematched controls. Cell samples were frozen periodically to allow for assessment of timecourse changes after confirmation of transformation. This chronic arsenic-exposed prostate epithelial (CAsE-PE) cell line, showed a 2.2-fold increase in matrix metalloproteinase-9 (MMP-9) secretion compared with control<sup>11</sup>. Increased MMP-9 is associated with Ras-induced or cadmium-induced malignant transformation of RWPE-1 cells<sup>56,57</sup>, occurs in human prostate tumors and in primary cultures of prostatic cancer cells, and is associated with aggressive prostatic malignancies<sup>58</sup>. When CAsE-PE cells were inoculated into the renal capsule of nude mice, all of the mice inoculated developed tumors within 10 weeks while control cells remained nontumorigenic<sup>11</sup>. The aggressive carcinoma that developed from CAsE-PE inoculation showed several characteristics in common with human prostatic cancers, including overproduction of human PSA<sup>11</sup>, clearly indicating their origin. The rapidly formed tumors resulting from CAsE-PE cell inoculation often invaded local tissue<sup>11</sup>. Because animal models for arsenic carcinogenesis are currently absent for the prostate, this ***in vitro*** system has been used to help define the molecular events in arsenic-induced prostatic carcinogenesis. Indeed, these cells and their heterotransplantation tumors show a remarkable series of characteristics in common with human prostate carcinoma (Table 2). The finding that human prostate

epithelial cells are directly susceptible to arsenic-induced malignant transformation strongly fortifies the evidence for a potential role of arsenic in human prostate cancer.

### Molecular events in arsenic-induced malignant transformation

Several studies were conducted to examine the molecular events in arsenic-induced malignant transformation in human prostate cells, including studies on DNA methylation<sup>18</sup>. Inorganic arsenic biomethylation uses SAM as the methyl donor, and SAM depletion can induce DNA hypomethylation<sup>59</sup>. Indeed, in CAsE-PE cells arsenic-induced malignant transformation also induces genomic DNA hypomethylation<sup>18</sup>. A decrease of DNA methyltransferase activity is an early event occurring before malignant transformation and may account for the subsequent genomic DNA hypomethylation<sup>18</sup>. Arsenic-induced DNA hypomethylation occurs in malignantly transformed rodent liver cells<sup>60</sup> and in the liver of mice after chronic exposure to inorganic arsenic<sup>61</sup>. Furthermore, hepatocellular carcinoma induced by transplacental exposure to inorganic arsenic in mice is associated with aberrant gene expression changes likely due, at least in part, to errors in DNA methylation including hypomethylation of steroid signaling transcription

factors<sup>62</sup>. The finding of arsenic-induced DNA hypomethylation in human prostate cells indicates this may be a plausible contributing factor for tumor development in arsenic-exposed human populations<sup>18</sup>. Carcinogenesis can result from aberrations of genomic DNA methylation that include hypomethylation of the promoter of cancer-related genes. Global hypomethylation of genomic DNA is often observed in tumors and contributes to overexpression of protooncogenes, growth factors, and genes that are involved in cancer cell proliferation, invasion, and metastasis<sup>63</sup>. DNA hypomethylation is viewed as a nongenotoxic mechanism facilitating aberrant gene expression<sup>64,65</sup>. Aberrant gene expression is a common occurrence in arsenic-exposed cells.

Studies show that both CAsE-PE and parental cells have a very poor capacity to methylate arsenic, making competition for SAM an unlikely basis for arsenic-induced DNA hypomethylation<sup>18</sup>. There is, however, emerging evidence that during cellular adaptation to chronic arsenic exposure, SAM recycling may be reduced in order to overproduce glutathione for arsenic efflux through transsulfuration of homocysteine<sup>68</sup>.

A marked overexpression of unmutated *K-ras* was also observed in CAsE-PE cells<sup>18</sup>. Although hypomethylation of the *ras* gene can lead to activation, the *K-ras* promoter region, including the major transcriptional initiation site, was essential-

**Table 2.** Characteristics in common between human prostate carcinoma cells and arsenic-transformed human prostate epithelial cells.

Characteristic	Human prostate carcinoma	Arsenic-transformed cells	References
Hyperproliferative	+	+	Achanzar <i>et al.</i> <sup>56</sup> Benbrahim-Tallaa <i>et al.</i> <sup>66</sup>
MMP-9 overexpression	+	+	Achanzar <i>et al.</i> <sup>56</sup> Benbrahim-Tallaa <i>et al.</i> <sup>18</sup>
Tumor formation	+	+	Achanzar <i>et al.</i> <sup>56</sup>
PSA overexpression	+	+	Achanzar <i>et al.</i> <sup>56</sup> Benbrahim-Tallaa <i>et al.</i> <sup>66</sup> Benbrahim-Tallaa <i>et al.</i> <sup>67</sup>
Unmutated ras overexpression	+	+	Benbrahim-Tallaa <i>et al.</i> <sup>18</sup> Benbrahim-Tallaa <i>et al.</i> <sup>67</sup>
Acquired androgen independence through AR by-pass	+	+	Benbrahim-Tallaa <i>et al.</i> <sup>66</sup> Benbrahim-Tallaa <i>et al.</i> <sup>67</sup>
HER-2/neu overexpression	+	+	Benbrahim-Tallaa <i>et al.</i> <sup>67</sup>
Invasive	+	+ <sup>a</sup>	Achanzar <i>et al.</i> <sup>56</sup>

<sup>a</sup> In tumors formed by heterotransplant of CAsE-PE cells.



ly unmethylated in both control and CAsE-PE cells<sup>18</sup>. Thus, although genomic DNA hypomethylation was observed in arsenic-transformed cells, this does not appear to be the direct cause of overexpression of K-*ras*<sup>18</sup>. Whatever the basis, K-*ras* overexpression appears to have been a key molecular change associated with arsenic-induced transformation of CAsE-PE cells. K-*ras* overexpression was observed as early as 12 weeks after arsenic exposure and reached its highest level after approximately 30 weeks of continuous arsenic exposure, the time point for malignant transformation. This is consistent with previous data suggesting that K-*ras* amplification could be an early event in the pathogenesis of prostatic carcinogenesis<sup>69</sup> and may be a critical factor that drives prostate cancer development<sup>70</sup>. Thus, the *in vitro* prostate model of arsenic carcinogenesis<sup>18</sup> duplicates this key aspect of the corollary disease in humans (Table 2).

The normal development, growth, and survival of the prostate epithelium are regulated both by systemic and local androgen and by local production of growth factors by the prostatic stroma<sup>71</sup>. However, regulatory interactions between androgens and growth factors often become distorted in prostate cancer<sup>71</sup>. Ras is a critical signaling molecule that controls several signaling pathways in prostate cancer<sup>70,72</sup>. Yet, *ras* mutations are infrequent in prostate cancer<sup>73</sup>. This is consistent with the hypothesis that wild-type *ras* is chronically activated by autocrine and paracrine factor stimulation in prostate cancer<sup>70,72</sup>. Virtually all the growth factor receptors upregulated in prostate cancer activate *ras* for their signal transduction activity<sup>72</sup>. In essence, *ras* signaling represents a convergence point for numerous diverse extracellular signals in prostate cancer<sup>72</sup>. One of the best-characterized effector pathways triggered by Ras activation is the MAPK (serine-threonine protein kinases) pathway. The activation of K-*ras* by arsenic in CAsE-PE cells is by some mechanism other than promoter region hypomethylation, perhaps involving genes upstream of *ras*<sup>18</sup>. In this regard, a series of proteins participating in protein-protein interactions are responsible for the control of *ras* activation and include Raf (c-Raf-1, A-Raf, and B-Raf), MEK (MAPK/ERK kinases 1 and 2), and ERK1/2<sup>74</sup>. The ERK1/2 signaling pathway plays an important role in cellular growth and differentiation<sup>74</sup>. Thus, molecular events upstream of *ras* have been compared in CAsE-PE and control cells. Clearly, proteins upstream of K-*ras*, including A-Raf and B-Raf showed greatly increased expression in CAsE-PE cells compared with control<sup>67</sup>. There was also an increased expression of phosphorylated MEK1/2

and ELK in CAsE-PE cells compared with control<sup>67</sup>. Thus, there is a correlation between elevated levels of active phosphor-MAPK and arsenic-induced prostate cell transformation.

Prostate cancer is a leading cause of male cancer death because in its advanced stages it acquires androgen independence and becomes resistant to androgen ablation therapy. Surgery can cure locally confined prostate cancer, but there are currently no effective treatments for androgen-independent, metastatic prostate cancer. When prostate cancer progresses in this manner, it is variously called "androgen independent" or "hormone refractory," because it is resistant to hormone ablation therapy. However, evidence indicates advanced prostate cancers often are not fully independent of androgen, but rather have become hypersensitive even to very low levels of androgen<sup>70</sup>. A majority of prostate tumors obtained from patients failing androgen ablation therapy overexpress the androgen receptor (AR), sensitizing the cells to low levels of androgen<sup>75</sup>. This overexpression is often associated with gene amplification<sup>75</sup>. Frequently, the AR is mutated in advanced prostate cancers, which results in a receptor that can be activated by nonandrogens<sup>76</sup>. Because the Ras/MAPK signaling pathway can also reduce the androgen requirement of prostate cells<sup>77</sup>, one would predict that stimulation of this signaling pathway might allow androgen-regulated gene expression even at very low levels of androgen. Evidence suggests that the Raf/MEK/ERK pathway plays a critical role in the modulation of AR activity in response to *ras*<sup>72</sup>. In addition, MAP kinase activity correlates with progression to an increasingly advanced and hormone-independent stage<sup>78</sup>.

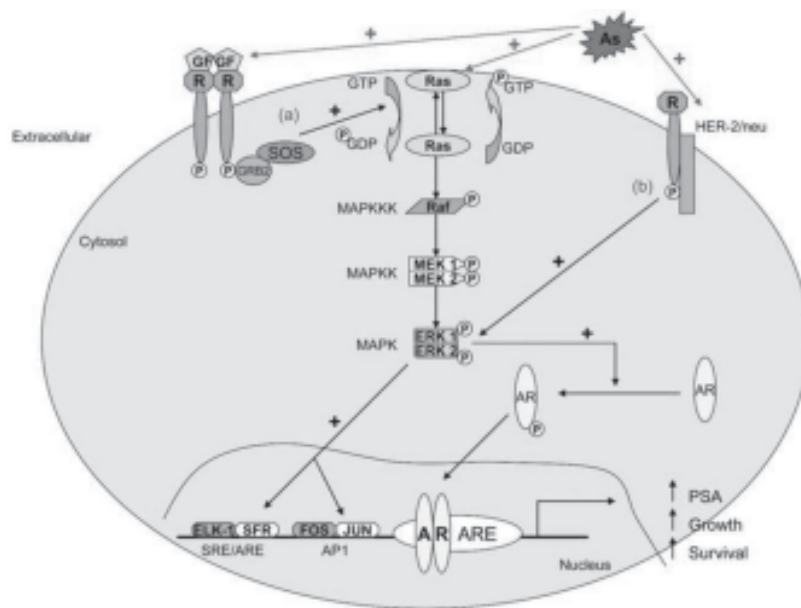
CAsE-PE cells, in which chronic arsenic exposure induced malignant transformation, hyperproliferation, and overexpression of K-*ras*<sup>18</sup>, have also been used to help define the role of arsenic in prostate cancer progression. The evidence shows CAsE-PE cells clearly acquired androgen independence during transformation that is not associated with AR overexpression<sup>66</sup>. The AR in CAsE-PE cells actually is less responsive to androgen, indicating an AR mutation that causes hypersensitivity to androgens is unlikely<sup>66,67</sup>. In addition, alterations in androgen metabolism, estrogen production, and estrogen receptor levels and sensitivity also had limited roles in this conversion<sup>66,67</sup>. However, the overexpression of *HER-2/neu* is a prominent feature<sup>67</sup> and one in which CAsE-PE cells have in common with androgen-independent human prostate carcinoma (Table 2). Thus, it appears arsenic-induced malignant transformation precipitates upregula-

tion of *ras* which in turn, allows by-pass of AR to induce androgen independence in human prostate epithelial cells (Figure 1). The fact that a common environmental contaminant such as arsenic can induce prostate tumor cells to progress to a much more lethal state could be very important in human populations exposed to this metalloid.

Overall the CAsE-PE cells and their heterotransplantation tumors show a remarkable series of traits in common with advanced human prostate carcinoma (Table 2).

## Conclusions

It has been known for over a century that inorganic arsenic is a human carcinogen. Arsenic exposure affects millions of people worldwide. Various studies in human populations exposed to arsenic via the environment provide evidence of a causal link to prostate cancer. In many cases this association is dose related, adding further evidence for an etiological role for the metalloid in this important human cancer. Rodent models of inorganic arsenic carcinogenesis generally have been slow to develop and have not specifically shown the prostate as a target of inorganic arsenic carcinogenesis. The rat, which is a



**Figure 1.** Mechanisms of arsenic-induced acquired androgen independence. Abbreviations: AR, androgen receptor; ARE, androgen responsive element; As, arsenic; GF, growth factors; MAPK, mitogen-activated protein kinase; MAPKK, MAPK kinase; MAPKKK, MAPK kinase kinase; PSA, prostate-specific antigen. It is known that exposure to arsenic initiates GF receptor signaling and Ras-dependent activation of MEK1/2 and ERK1/2. (A) As prostate cancer progresses to androgen independence, the growth factors production increases. Growth factor signal transduction pathways have been shown to stimulate AR activation. All these growth factors use the Ras/MAPK pathway for a portion of their signal transduction. Binding of GF results in dimerization, autophosphorylation of the receptor, and tyrosine phosphorylation of other proteins. The GF receptor activates ras which in turn activates Raf, which phosphorylates and activates MEK, which in turn, phosphorylates and activates ERK. Activated MAPK can regulate targets in the cytosol and also translocate to the nucleus causing phosphorylation of transcription factors such as c-Fos to create AP-1 and ELK-1, which contribute to proliferation. (B) HER-2/neu promotes phosphorylation of AR at multiple sites even in the presence of very low androgen levels. HER-2/neu indirectly activates MAPK. MAPK might phosphorylate the AR, creating an androgen-independent receptor.

species of choice for animal models of human prostate cancer, is unfortunately a poor choice for modeling human arsenic toxicity. Various studies using human prostate epithelial cells in culture have shown that low-level inorganic arsenic exposure can induce malignant transformation specifically in these cells. The finding that human prostate epithelial cells are directly sensitive to malignant transformation induced by inorganic arsenic strongly supports a potential role for arsenic in human prostate cancer. Heterotransplantation of these cells into nude mice produced aggressive carcinomas that overexpress PSA in a fashion similar to human prostate carcinoma. In addition, inorganic arsenic stimulates acquired androgen independence during this malignant transformation, a condition associated with advanced human prostate cancer and poor prognosis. The cancer risk at low doses of arsenic is a subject of considerable debate and may not be solved solely by epidemiologic means, particularly for tar-

get sites such as the prostate, for which there are currently no whole-animal models. Therefore, it is essential to learn more about arsenic's mode of action at the target cell level. Arsenic seems to have the potential for many mechanisms of action in the development of cancer, including prostate cancer. Finally, additional research is clearly needed at all levels on the role of arsenic in prostate cancer development and progression.

## Acknowledgments

We thank J Liu and E Tokar for their critical review of this manuscript. This research was supported by the Intramural Research Program of the NIH, National Cancer Institute, Center for Cancer Research. The authors declare they have no competing financial interests.

## References

1. Nordstrom DK. Worldwide occurrences of arsenic in ground water. *Science* 2002; 21:2143–2145.
2. Smedley PL, Kinniburgh DG. A review of the source, behavior and distribution of arsenic in natural waters. *Appl Geochem* 2002; 17:517–568.
3. International Agency for Research on Cancer. Some drinking-water disinfectants and contaminants, including arsenic. *IARC Monogr Eval Carcinog Risks Hum* 2004; 84:269–477.
4. National Research Council. *Arsenic in Drinking Water*: Washington, D.C.: National Academy Press; 1999.
5. National Research Council. *Arsenic in Drinking Water*: Washington, D.C.: National Academy Press; 2001.
6. National Toxicology Program. Arsenic compounds, inorganic. In: 11th Report on Carcinogens. Research Triangle Park, NC: National Toxicology Program; 2004.
7. Kitchin KT. Recent advances in arsenic carcinogenesis: modes of action, animal model systems, and methylated arsenic metabolites. *Toxicol Appl Pharmacol* 2001; 172:249–261.
8. Rossman TG. Mechanism of arsenic carcinogenesis: an integrated approach. *Mutat Res* 2003; 533:37–65.
9. Simeonova PP, Luster MI. Mechanisms of arsenic carcinogenicity: genetic or epigenetic mechanisms? *J Environ Pathol Toxicol Oncol* 2000; 19:281–286.
10. Waalkes MP, Liu J, Diwan BA. Transplacental arsenic carcinogenesis in mice. *Toxicol Appl Pharmacol* 2007; 222(3):271–280.
11. Achanzar WE, Brambila EM, Diwan BA, Webber MM, Waalkes MP. Inorganic arsenite induced malignant transformation of human prostate epithelial cells. *J Natl Cancer Inst* 2002; 94:1888–1891.
12. Pi J, Qu W, Reece JF, Kumagai Y, Waalkes MP. Transcription factor Nrf2 activation by inorganic arsenic in cultured keratinocytes: involvement of hydrogen peroxide. *Exp Cell Res* 2003; 290:234–245.
13. Sens DA, Park S, Gurel V, Sens MA, Garrett SH, Somji S. Inorganic cadmium- and arsenite-induced malignant transformation of human bladder urothelial cells. *Toxicol Sci* 2004; 79:56–63.
14. Aposhian VH, Aposhian MM. Arsenic toxicology: five questions. *Chem Res Toxicol* 2006; 19:1–15.
15. Styblo M, Drobna Z, Jaspers I, Lin S, Thomas DJ. The role of biomethylation in toxicity and carcinogenicity of arsenic: a research update. *Environ Health Perspect* 2002; 110:767–771.
16. Thomas DJ, Styblo M, Lin S. The cellular metabolism and systemic toxicity of arsenic. *Toxicol Appl Pharmacol* 2001; 176:127–144.
17. Patterson TJ, Ngo M, Aronov PA, Reznikova TV, Green PJ, Rice RH. Biological activity of inorganic arsenic and antimony reflects oxidation state in cultured human keratinocytes. *Chem Res Toxicol* 2003; 16:1624–1631.
18. Benbrahim-Tallaa L, Waterland RA, Styblo M, Achanzar WE, Webber MM, Waalkes MP. Molecular events associated with arsenic-induced malignant transformation of human prostatic epithelial cells: aberrant genomic DNA methylation and K-ras oncogene activation. *Toxicol Appl Pharmacol* 2005; 206:288–298.
19. Mass MJ, Tennant A, Roop BC, Cullen WR, Styblo M, Thomas DJ, Kligerman AD. Methylated trivalent arsenic species are genotoxic. *Chem Res Toxicol* 2001; 14:355–361.

20. Styblo M, Del Razo LM, Vega L, Germolec DR, LeCluyse EL, Hamilton GA, Reed W, Wang C, Cullen WR, Thomas DJ. Comparative toxicity of trivalent and pentavalent inorganic and methylated arsenicals in rat and human cells. *Arch Toxicol* 2000; 74:289-299.
21. Wei M, Wanibuchi H, Morimura K, Iwai S, Yoshida K, Endo G, Nakae D, Fukushima S. Carcinogenicity of dimethylarsinic acid in male F344 rats and genetic alterations in induced urinary bladder tumors. *Carcinogenesis* 2002; 23:1387-1397.
22. Bredfeldt TG, Jagadish B, Eblin KE, Mash EA, Gandolfi AJ. Monomethylarsonous acid induces transformation of human bladder cells. *Toxicol Appl Pharmacol* 2006; 216:69-79.
23. Huang RN, Lee TC. Cellular uptake of trivalent arsenite and pentavalent arsenate in KB cells cultured in phosphate-free medium. *Toxicol Appl Pharmacol* 1996; 136:243-249.
24. Liu Z, Shen J, Carbrey JM, Mukhopadhyay R, Agre P, Rosen BP. Arsenite transport by mammalian aquaglyceroporins AQP7 and AQP9. *Proc Natl Acad Sci USA* 2002; 99:6053-6058.
25. Bridges CC, Zalups RK. Molecular and ionic mimicry and the transport of toxic metals. *Toxicol Appl Pharmacol* 2005; 204:274-308.
26. Leslie EM, Haimeur A, Waalkes MP. Arsenic transport by the human multidrug resistance protein 1 (MRP1/ABCC1). Evidence that a tri-glutathione conjugate is required. *J Biol Chem* 2004; 279:32700-32708.
27. Liu J, Chen H, Miller DS, Saavedra JE, Keefer LK, Johnson DR, Klaassen CD, Waalkes MP. Overexpression of glutathione S-transferase II and multidrug resistance transport proteins is associated with acquired tolerance to inorganic arsenic. *Mol Pharmacol* 2001; 60:302-309.
28. Crawford ED. Epidemiology of prostate cancer. *Urology* 2003; 62:3-12.
29. Parkin DM, Bray F, Ferlay J, Pisani P. Global cancer statistics, 2002. *CA Cancer J Clin*. 2005; 55:74-108.
30. McKay L, Macintyre S, Ellaway A. Migration and Health: A Review of the International Literature. Occasional paper N. 12. Glasgow: MRC Social and Public Health Sciences Unit; 2003. p. 89-129.
31. Moradi T, Delfino RJ, Bergstrom SR, Yu ES, Adami HO, Yuen J. Cancer risk among Scandinavian immigrants in the US and Scandinavian residents compared with US whites, 1973-89. *Eur J Cancer Prev* 1998; 7:117-125.
32. Stemmermann GN, Nomura AM, Chyou PH, Kato I, Kuroishi T. Cancer incidence in Hawaiian Japanese: migrants from Okinawa compared with those from other prefectures. *Jpn J Cancer Res* 1991; 82:1366-1370.
33. Thomas DB, Karagas MR. Migrant studies. In: Schottenfeld D, Fraumeni JF, editors. *Cancer Epidemiology and Prevention*. 2<sup>nd</sup> ed. Oxford, UK: Oxford University Press; 1996. p. 236-254.
34. Hutchinson J. Arsenic cancer. *Br Med J* 1887; 2:1280-1281.
35. Brown KG, Ross GL. American Council on Science and Health. Arsenic, drinking water, and health: a position paper of the American Council on Science and Health. *Regul Toxicol Pharmacol* 2002; 36:162-174.
36. Germolec DR, Judson Spalding J, Yu HS, Chen JS, Simeonova PP, Humble PC, Bruccoleri A, Boorman GA, Foley JF, Yoshida T, Luster MI. Arsenic enhancement of skin neoplasia by chronic stimulation of growth factors. *Am J Pathol* 1998; 153:1175-11785.
37. Rossman TG, Uddin AN, Burns FJ. Evidence that arsenite acts as a co-carcinogen in skin cancer. *Toxicol Appl Pharmacol* 2004; 198:394-404.
38. Shirai T, Takahashi S, Cui L, Futakuchi M, Kato K, Tamano S, Imaida K. Experimental prostate carcinogenesis—rodent models. *Mutat Res* 2000; 462:219-226.
39. Green JE, Greenberg NM, Ashendel CL, Barrett JC, Boone C, Getzenberg RH, Henkin J, Matusik R, Janus TJ, Scher HI. Workgroup 3: transgenic and reconstitution models of prostate cancer. *Prostate* 1998; 36:59-63.
40. Wanibuchi H, Salim EL, Kinoshita A, Shen J, Wei M, Morimura K, Yoshida K, Kuroda K, Endo G, Fukushima S. Understanding arsenic carcinogenicity by the use of animal models. *Toxicol Appl Pharmacol* 2004; 198:366-376.
41. Chen CJ, Kuo TL, Wu MM. Arsenic and cancers. *Lancet* 1988; 1:414-415.
42. Chen CJ, Chuang YC, Lin TM, Wang CJ. Malignant neoplasms among residents of a blackfoot disease-endemic area in Taiwan: high-arsenic artesian well water and cancers. *Cancer Res* 1985; 45:5895-5899.
43. Wu MM, Kuo TL, Hwang YH, Chen CJ. Dose-response relation between arsenic concentration in well water and mortality from cancers and vascular diseases. *Am J Epidemiol* 1989; 130:1123-1132.
44. Chen CJ, Wang CJ. Ecological correlation between arsenic levels in well water and age adjusted mortality from malignant neoplasms. *Cancer Res* 1990; 50:5470-5474.
45. Tsai SM, Wang TN, Ko YC. Mortality for certain diseases in areas with high levels of arsenic in drinking water. *Arch Environ Health* 1999; 54:186-193.
46. Lewis DR, Southwick JW, Ouellet-Hellstrom R, Rensch J, Calderon RL. Drinking water arsenic in Utah: a cohort mortality study. *Environ Health Perspect* 1999; 107:359-365.
47. Hinwood AL, Jolley DJ, Sim MR. Cancer incidence and Arsenic-induced prostate cancer high environmental arsenic concentrations in rural populations: results of an ecological study. *Int J Environ Health* 1999; 9:131-141.
48. Szymanska-Chabowska A, Antonowicz-Juchniewicz J, Andrezejak R. Plasma concentration of selected neoplastic markers in persons occupationally exposed to arsenic and heavy metals. *Medycyna Pracy* 2004; 55:313-320.
49. Pant N, Murthy RC, Srivastava SP. Male reproductive toxicity of sodium arsenite in mice. *Hum Exp Toxicol* 2004; 23:399-403.
50. Jana K, Jana S, Samanta PK. Effects of chronic exposure to sodium arsenite on hypothalamo-pituitary-testicular activities in adult rats: possible an estrogenic mode of action. *Reprod Biol Endocrinol* 2006; 16:4-9.
51. Kyprianou N, Isaacs JT. Activation of programmed cell death in the rat ventral prostate. *Endocrinology* 1988; 122:552-562.
52. Webber MM, Bello D, Quader S. Immortalized and tumorigenic adult human prostatic epithelial cell lines: characteristics and applications. Part I. Cell markers and immortalized nontumorigenic cell lines. *Prostate* 1996; 29:386-394.

53. Webber MM, Bello D, Quader S. Immortalized and tumorigenic adult human prostatic epithelial cell lines: characteristics and applications. Part 2. Tumorigenic cell lines. *Prostate* 1997; 30:58–64.
54. Gustavsson H, Welén K, Damber JE. Transition of an androgen-dependent human prostate cancer cell line into an androgen-independent subline is associated with increased angiogenesis. *Prostate* 2005; 62:364–373.
55. Bello D, Webber MM, Kleinman HK, Wartinger DD, Rhim JS. 1997. Androgen responsive adult human prostatic epithelial cell lines immortalized by human papillomavirus 18. *Carcinogenesis* 18:1215–1223.
56. Achanzar WE, Diwan BA, Liu J, Quader S, Webber MM, Waalkes MP. Cadmium induced malignant transformation of human prostate epithelial cells. *Cancer Res* 2001; 61:455–458.
57. Webber MM, Waghray A, Bello D, Rhim JS. Proteases and invasion in human prostate epithelial cell lines: implications in prostate cancer prevention and intervention. *Radiat Oncol Invest* 1996; 3:358–362.
58. Hamdy FC, Fadlon EJ, Cottam D, Lawry J, Thurrell W, Silcocks PB, Anderson JB, Williams JL, Rees RC. Matrix metalloproteinase 9 expression in primary human prostatic adenocarcinoma and benign prostatic hyperplasia. *Br J Cancer* 1994; 69:177–182.
59. Loenen WA. S-Adenosylmethionine: jack of all trades and master of everything? *Biochem Soc Trans* 2006; 34:330–333.
60. Zhao CQ, Young MR, Diwan BA, Coogan TP, Waalkes MP. Association of arsenic induced malignant transformation with DNA hypomethylation and aberrant gene expression. *Proc Natl Acad Sci USA* 1997; 94:10907–10912.
61. Chen H, Li S, Liu J, Diwan BA, Barrett JC, Waalkes MP. Chronic inorganic arsenic exposure induces hepatic global and individual gene hypomethylation: implications for arsenic hepatocarcinogenesis. *Carcinogenesis* 2004; 25:1779–1786.
62. Waalkes MP, Liu J, Chen H, Xie Y, Achanzar WE, Zhou YS, Cheng ML, Diwan BA. Estrogen signaling in livers of male mice with hepatocellular carcinoma induced by exposure to arsenic in utero. *J Natl Cancer Inst* 2004; 96:466–474.
63. Stirzaker C, Song JZ, Davidson B, Clark SJ. Transcriptional gene silencing promotes DNA hypermethylation through a sequential change in chromatin modifications in cancer cells. *Cancer Res* 2004; 64:3871–3877.
64. Counts JL, Goodman JL. Hypomethylation of DNA: a nongenotoxic mechanism involved in tumor promotion. *Toxicol Lett* 1995; 82–83:663–672.
65. Vorce RL, Goodman JL. Altered methylation of ras oncogenes in benzidine-induced B6C3F1 mouse liver tumors. *Toxicol Appl Pharmacol* 1989; 100:310–398.
66. Benbrahim-Tallaa L, Webber MM, Waalkes MP. Acquisition of androgen independence by human prostate epithelial cells during arsenic-induced malignant transformation. *Environ Health Perspect* 2005; 113:1134–1139.
67. Benbrahim-Tallaa L, Webber MM, Waalkes MP. Mechanisms in acquired androgen independence during arsenic-induced malignant transformation of human prostate epithelial cells. *Environ Health Perspect* 2007; 115:243–247.
68. Coppin JF, Webber MM, Waalkes MP. Possible interactions between cellular methyl metabolism and adaptive efflux during chronic arsenic exposure in human cells [Abstract]. *Toxicol Sci* 2007; 96(suppl):303.
69. Lau AT, Li M, Xie R, He QY, Chiu JF. Opposed arsenite-induced signaling pathways promote cell proliferation or apoptosis in cultured lung cells. *Carcinogenesis* 2004; 25:21–28.
70. Weber MJ, Gioeli D. Ras signaling in prostate cancer progression. *J Cell Biochem* 2004; 91:13–25.
71. Feldman BJ, Feldman D. The development of androgen-independent prostate cancer. *Nat Rev Cancer* 2001; 1:34–45.
72. Gioeli D. Signal transduction in prostate cancer progression. *Clin Sci (Lond)* 2005; 108:293–308.
73. Carter BS, Epstein JI, Isaacs WB. Ras gene mutations in human prostate cancer. *Cancer Res* 1990; 50:6830–6832.
74. McCubrey JA, Steelman LS, Abrams SL, Lee JT, Chang LF, Bertrand FE, Navolanic PM, Terrian DM, Franklin RA, D'Assoro AB, Salisbury JL, Mazzarino MC, Stivala F, Libra M. Roles of the RAF/MEK/ERK and PI3K/PTEN/AKT pathways in malignant transformation and drug resistance. *Adv Enzyme Regul* 2006; 46:249–279.
75. Linja MJ, Savinainen KJ, Saramaki OR, Tammela TL, Vessella RL, Visakorpi T. Amplification and overexpression of androgen receptor gene in hormone-refractory prostate cancer. *Cancer Res* 2001; 61:3550–3555.
76. Culig Z, Klocker H, Bartsch G, Hobisch A. Androgen receptor mutations in carcinoma of the prostate: significance for endocrine therapy. *Am J Pharmacogenomics* 2001; 1:241–249.
77. Bakin RE, Gioeli D, Sikes RA, Bissonette EA, Weber MJ. Constitutive activation of the Ras/mitogen-activated protein kinase signaling pathway promotes androgen hypersensitivity in LNCaP prostate cancer cells. *Cancer Res* 2003; 63:1981–1989.
78. Gioeli D, Mandell JW, Petroni GR, Frierson HF Jr, Weber MJ. Activation of mitogen activated protein kinase associated with prostate cancer progression. *Cancer Res* 1999; 59:279–284.

Received 1 May 2007

Accepted 8 November 2007